

المجلة الأردنية للعلوم الحياتية
Jordan Journal of Biological Sciences (JJBS)
<http://jjbs.hu.edu.jo>

Jordan Journal of Biological Sciences (JJBS) (ISSN: 1995–6673): An International Peer- Reviewed Research Journal financed by the Scientific Research Support Fund, Ministry of Higher Education and Scientific Research, Jordan and published quarterly by the Deanship of Research and Graduate Studies, The Hashemite University, Jordan.

Editor-in-Chief

Professor Abu-Elteen, Khaled H.
The Hashemite University

Editorial Board (Arranged alphabetically):

- | | |
|---|--|
| - Professor Abdalla, Shtaywy S.
University of Jordan | - Professor Elkarmi, Ali Z.
The Hashemite University |
| - Professor Abdel-Hafez, Sami K.
Yarmouk University | - Professor Oran, Sawsan A.
University of Jordan |
| - Professor Al-Hadidi, Hakam F.
Jordan University of Science and Technology | - Professor Sallal, Abdul-Karim J.
Jordan University of Science and Technology |
| - Professor Bashir, Nabil A.
Jordan University of Science and Technology | - Professor Tarawneh, Khaled A.
Mutah University |

International Advisory Board:

- | | |
|---|---|
| Prof. Abdul-Haque, Allah Hafiz
National Institute for Biotechnology and
Genetic Engineering, Pakistan | Prof. Bamburg, James
Colorado State University, U.S.A, |
| Prof. El Makawy, Aida, I
National Research Center, Giza, Egypt | Prof. Garrick, Michael D
State University of New York at Buffalo, U.S.A. |
| Prof. Ghannoum, Mahmoud A
University Hospital of Cleveland and Case Western
Reserve University, U.S.A. | Prof. Gurib-Fakim, Ameenah F
Center for Phytotherapy and Research,
Ebene, Mauritius. |
| Prof. Hassanali, Ahmed
Kenya University, Nairobi, Kenya | Prof. Kaviraj, Anilava
India University of Kalyani, India |
| Prof. Matar, Ghassan M
American University of Beirut, Lebanon | Prof. Martens, Jochen
Institute Fur Zoologie, Germany |
| Prof. Nasher, Abdul Karim
Sanna' University, Yemen | Prof. Stanway, Glyn
University of Essex, England |
| Prof. Waitzbauer, Wolfgang
University of Vienna, Austria | Prof. Wan Yusoff, Wan Mohtar
University Kebangsaan Malaysia, Malaysia |

Submission Address

Professor Abu-Elteen, Khaled H.
Deanship of Scientific Research and Graduate Studies
The Hashemite University
P.O. Box 330127, Zarqa, 13115, Jordan
Phone: +962-5-3903333 ext. 4399
E-Mail: jjbs@hu.edu.jo

Editorial Board Support Team

Language Editor Dr. Qusai Al-Debyan	Publishing Layout Mohannad Oqdeh
--	---



Hashemite Kingdom of Jordan



The Hashemite University

Jordan Journal of Biological Sciences

*An International Peer-Reviewed Scientific Journal
Financed by the Scientific Research Support Fund*

Instructions to Authors

Scopes

Study areas include cell biology, genomics, microbiology, immunology, molecular biology, biochemistry, embryology, immunogenetics, cell and tissue culture, molecular ecology, genetic engineering and biological engineering, bioremediation and biodegradation, bioinformatics, biotechnology regulations, gene therapy, organismal biology, microbial and environmental biotechnology, marine sciences. The JJBS welcomes the submission of manuscript that meets the general criteria of significance and academic excellence. All articles published in JJBS are peer-reviewed. Papers will be published approximately one to two months after acceptance.

Type of Papers

The journal publishes high-quality original scientific papers, short communications, correspondence and case studies. Review articles are usually by invitation only. However, Review articles of current interest and high standard will be considered.

Submission of Manuscript

Manuscript, or the essence of their content, must be previously unpublished and should not be under simultaneous consideration by another journal. The authors should also declare if any similar work has been submitted to or published by another journal. They should also declare that it has not been submitted/ published elsewhere in the same form, in English or in any other language, without the written consent of the Publisher. The authors should also declare that the paper is the original work of the author(s) and not copied (in whole or in part) from any other work. All papers will be automatically checked for duplicate publication and plagiarism. If detected, appropriate action will be taken in accordance with International Ethical Guideline. By virtue of the submitted manuscript, the corresponding author acknowledges that all the co-authors have seen and approved the final version of the manuscript. The corresponding author should provide all co-authors with information regarding the manuscript, and obtain their approval before submitting any revisions. Electronic submission of manuscripts is strongly recommended, provided that the text, tables and figures are included in a single Microsoft Word file. Submit manuscript as e-mail attachment to the Editorial Office at: JJBS@hu.edu.jo. After submission, a manuscript number will be communicated to the corresponding author within 48 hours.

Peer-review Process

It is requested to submit, with the manuscript, the names, addresses and e-mail addresses of at least 4 potential reviewers. It is the sole right of the editor to decide whether or not the suggested reviewers to be used. The reviewers' comments will be sent to authors within 6-8 weeks after submission. Manuscripts and figures for review will not be returned to authors whether the editorial decision is to accept, revise, or reject. All Case Reports and short Communication must include at least one table and/or one figure.

Preparation of Manuscript

The manuscript should be written in English with simple lay out. The text should be prepared in single column format. Bold face, italics, subscripts, superscripts etc. can be used. Pages should be numbered consecutively, beginning with the title page and continuing through the last page of typewritten material.

The text, excluding the abstract, if required, can be divided into numbered sections with brief headings. Starting from introduction with section 1. Subsections should be numbered (for example 2.1 (then 2.1.1, 2.1.2, 2.2, etc.), up to three levels.

Manuscripts in general should be organized in the following manner:

- Title
- Abstract
- Keywords
- Introduction
- Materials & Methods
- Results & Discussion
- Conclusion
- Acknowledgements
- References

Brief guidelines

Title Page

The title page should contain a brief title, correct first name, middle initial and family name of each author and name and address of the department(s) and institution(s) from where the research was carried out for each author. The title should be without any abbreviations and it should enlighten the contents of the paper. All affiliations should be provided with a lower-case superscript number just after the author's name and in front of the appropriate address.

The name of the corresponding author should be indicated along with telephone and fax numbers (with country and area code) along with full postal address and e-mail address.

ABSTRACT

The abstract should be concise and informative. It should not exceed **350 words** in length for full manuscript and Review article and 150 words in case of Case Report and/ or Short Communication.. It should briefly describe the purpose of the work, techniques and methods used, major findings with important data and conclusions. No references should be cited in this part. Generally non-standard abbreviations should not be used, if necessary they should be clearly defined in the abstract, at first use.

Keywords

Immediately after the abstract, **about 4-8 keywords** should be given. Use of abbreviations should be avoided, only standard abbreviations, well known in the established area may be used, if appropriate. These keywords will be used for indexing.

Abbreviations

Non-standard abbreviations should be listed and full form of each abbreviation should be given in parentheses at first use in the text.

INTRODUCTION

Provide a factual background, clearly defined problem, proposed solution, a brief literature survey and the scope and justification of the work done.

MATERIALS AND METHODS

Give adequate information to allow the experiment to be reproduced. Already published methods should be mentioned with references. Significant modifications of published methods and new methods should be described in detail. Capitalize trade names and include the manufacturer's name and address. Subheading should be used.

RESULTS

Results should be clearly described in a concise manner. Results for different parameters should be described under subheadings or in separate paragraph. Results should be explained, but largely without referring to the literature. Table or figure numbers should be mentioned in parentheses for better understanding.

DISCUSSION

The discussion should not repeat the results, but provide detailed interpretation of data. This should interpret the significance of the findings of the work. Citations should be given in support of the findings. The results and discussion part can also be described as separate, if appropriate. The Results and Discussion sections can include subheadings, and when appropriate, both sections can be combined

CONCLUSIONS

This should briefly state the major findings of the study.

Acknowledgment

A brief acknowledgment section may be given after the conclusion section just before the references. The acknowledgment of people who provided assistance in manuscript preparation, funding for research, etc. should be listed in this section.

Tables and Figures

Tables and figures should be presented as per their appearance in the text. It is suggested that the discussion about the tables and figures should appear in the text before the appearance of the respective tables and figures. No tables or figures should be given without discussion or reference inside the text.

Tables should be explanatory enough to be understandable without any text reference. Double spacing should be maintained throughout the table, including table headings and footnotes. Table headings should be placed above the table. Footnotes should be placed below the table with superscript lowercase letters. Each table should be on a separate page, numbered consecutively in Arabic numerals.

Each figure should have a caption. The caption should be concise and typed separately, not on the figure area. Figures should be self-explanatory. Information presented in the figure should not be repeated in the table. All symbols and abbreviations used in the illustrations should be defined clearly. Figure legends should be given below the figures.

References

References should be listed alphabetically at the end of the manuscript. Every reference referred in the text must be also present in the reference list and vice versa. In the text, a reference identified by means of an author's name should be followed by the year of publication in parentheses (e.g.(Brown,2009)). For two authors, both authors' names followed by the year of publication (e.g.(Nelson and Brown, 2007)). When there are more than two authors, only the first author's name followed by "*e tal.*" and the year of publication (e.g. (Abu-Elteen *e tal.*, 2010)). When two or more works of an author has been published during the same year, the reference should be identified by the letters "a", "b", "c", etc., placed after the year of publication. This should be followed both in the text and reference list. e.g., Hilly, (2002a, 2002b); Hilly, and Nelson, (2004). Articles in preparation or submitted for publication, unpublished observations, personal communications, etc. should not be included in the reference list but should only be mentioned in the article text (e.g., Shtyawy,A., University of Jordan, personal communication). Journal titles should be abbreviated according to the system adopted in Biological Abstract and Index Medicus, if not included in Biological Abstract or Index Medicus journal title should be given in full. The author is responsible for the accuracy and completeness of the references

and for their correct textual citation. Failure to do so may result in the paper being withdrawn from the evaluation process. Example of correct reference form is given as follows:-

Reference to a journal publication:

Bloch BK. 2002. Econazole nitrate in the treatment of *Candida* vaginitis. *S Afr Med J*, **58**:314-323.

Ogunseitan OA and Ndoye IL. 2006. Protein method for investigating mercuric reductase gene expression in aquatic environments. *Appl Environ Microbiol*, **64**: 695-702.

Hilly MO, Adams MN and Nelson SC. 2009. Potential fly-ash utilization in agriculture. *Progress in Natural Sci*, **19**: 1173-1186.

Reference to a book:

Brown WY and White SR. 1985. **The Elements of Style**, third ed. MacMillan, New York.

Reference to a chapter in an edited book:

Mettam GR and Adams LB. 2010. How to prepare an electronic version of your article. In: Jones BS and Smith RZ (Eds.), **Introduction to the Electronic Age**. Kluwer Academic Publishers, Netherlands, pp. 281–304.

Conferences and Meetings:

Embabi NS. 1990. Environmental aspects of distribution of mangrove in the United Arab Emirates. Proceedings of the First ASWAS Conference. University of the United Arab Emirates. Al-Ain, United Arab Emirates.

Theses and Dissertations:

El-Labadi SN. 2002. Intestinal digenetic trematodes of some marine fishes from the Gulf of Aqaba. MSc dissertation, Hashemite University, Zarqa, Jordan.

Nomenclature and Units

Internationally accepted rules and the international system of units (SI) should be used. If other units are mentioned, please give their equivalent in SI.

For biological nomenclature, the conventions of the *International Code of Botanical Nomenclature*, the *International Code of Nomenclature of Bacteria*, and the *International Code of Zoological Nomenclature* should be followed.

Scientific names of all biological creatures (crops, plants, insects, birds, mammals, etc.) should be mentioned in parentheses at first use of their English term.

Chemical nomenclature, as laid down in the *International Union of Pure and Applied Chemistry* and the official recommendations of the *IUPAC-IUB Commission on Biochemical Nomenclature* should be followed. All biocides and other organic compounds must be identified by their Geneva names when first used in the text. Active ingredients of all formulations should be likewise identified.

Math formulae

All equations referred to in the text should be numbered serially at the right-hand side in parentheses. Meaning of all symbols should be given immediately after the equation at first use. Instead of root signs fractional powers should be used.

Subscripts and superscripts should be presented clearly. Variables should be presented in italics. Greek letters and non-Roman symbols should be described in the margin at their first use.

To avoid any misunderstanding zero (0) and the letter O, and one (1) and the letter I should be clearly differentiated.

For simple fractions use of the solidus (/) instead of a horizontal line is recommended. Levels of statistical significance such as: * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ do not require any further explanation.

Copyright

Submission of a manuscript clearly indicates that: the study has not been published before or is not under consideration for publication elsewhere (except as an abstract or as part of a published lecture or academic thesis); its publication is permitted by all authors and after accepted for publication it will not be submitted for publication anywhere else, in English or in any other language, without the written approval of the copyright-holder. The journal may consider manuscripts that are translations of articles originally published in another language. In this case, the consent of the journal in which the article was originally published must be obtained and the fact that the article has already been published must be made clear on submission and stated in the abstract. It is compulsory for the authors to ensure that no material submitted as part of a manuscript infringes existing copyrights, or the rights of a third party.

Ethical Consent

All manuscripts reporting the results of experimental investigation involving human subjects should include a statement confirming that each subject or subject's guardian obtains an informed consent, after the approval of the experimental protocol by a local human ethics committee or IRB. When reporting experiments on animals, authors should indicate whether the institutional and national guide for the care and use of laboratory animals was followed.

Plagiarism

The JJBS hold no responsibility for plagiarism. If a published paper is found later to be extensively plagiarized and is found to be a duplicate or redundant publication, a note of retraction will be published, and copies of the correspondence will be sent to the authors' head of institute.

Galley Proofs

The Editorial Office will send proofs of the manuscript to the corresponding author as an e-mail attachment for final proof reading and it will be the responsibility of the corresponding author to return the galley proof materials appropriately corrected within the stipulated time. Authors will be asked to check any typographical or minor clerical errors in the manuscript at this stage. No other major alteration in the manuscript is allowed. After publication authors can freely access the full text of the article as well as can download and print the PDF file.

Reprints

Twenty (20) reprints are provided to corresponding author free of charge within two weeks after the printed journal date. For orders of more reprints, a reprint order form and prices will be sent with article proofs, which should be returned directly to the Editor for processing.

Disclaimer

Articles, communication, or editorials published by JJBS represent the sole opinions of the authors. The publisher shoulders no responsibility or liability what so ever for the use or misuse of the information published by JJBS.

Indexing

JJBS is indexed and abstracted by the Chemical Abstract Service, CAB International Abstracts, Zoological Abstract, EBSCO Database, Directory of Open Access Journals, Index Copernicus Master List, Open J Gate, NDL Japanese Periodicals Index, Genomics Journal Seek Database, SCImago and SCIRUS.

Vj g'Hashemite University
Deanship of Scientific Research and Graduate Studies
TRANSFER OF COPYRIGHT AGREEMENT

Journal publishers and authors share a common interest in the protection of copyright: authors principally because they want their creative works to be protected from plagiarism and other unlawful uses, publishers because they need to protect their work and investment in the production, marketing and distribution of the published version of the article. In order to do so effectively, publishers request a formal written transfer of copyright from the author(s) for each article published. Publishers and authors are also concerned that the integrity of the official record of publication of an article (once refereed and published) be maintained, and in order to protect that reference value and validation process, we ask that authors recognize that distribution (including through the Internet/WWW or other on-line means) of the authoritative version of the article as published is best administered by the Publisher.

To avoid any delay in the publication of your article, please read the terms of this agreement, sign in the space provided and return the complete form to us at the address below as quickly as possible.

Article entitled:-----

Corresponding author: -----

To be published in the journal: Jordan Journal of Biological Sciences (JJBS)

I hereby assign to the Hashemite University the copyright in the manuscript identified above and any supplemental tables, illustrations or other information submitted therewith (the "article") in all forms and media (whether now known or hereafter developed), throughout the world, in all languages, for the full term of copyright and all extensions and renewals thereof, effective when and if the article is accepted for publication. This transfer includes the right to adapt the presentation of the article for use in conjunction with computer systems and programs, including reproduction or publication in machine-readable form and incorporation in electronic retrieval systems.

Authors retain or are hereby granted (without the need to obtain further permission) rights to use the article for traditional scholarship communications, for teaching, and for distribution within their institution.

- ☐ I am the sole author of the manuscript
- ☐ I am signing on behalf of all co-authors of the manuscript
- ☐ The article is a 'work made for hire' and I am signing as an authorized representative of the employing company/institution

Please mark one or more of the above boxes (as appropriate) and then sign and date the document in black ink.

Signed: _____ Name printed: _____
Title and Company (if employer representative) : _____
Date: _____

Data Protection: By submitting this form you are consenting that the personal information provided herein may be used by the Hashemite University and its affiliated institutions worldwide to contact you concerning the publishing of your article.

Please return the completed and signed original of this form by mail or fax, or a scanned copy of the signed original by e-mail, retaining a copy for your files, to:

Hashemite University
Deanship of Scientific Research and Graduate Studies
Zarqa 13115 Jordan
Fax: +962 5 3903338
Email: jjbs@hu.edu.jo

EDITORIAL PREFACE

Jordan Journal of Biological Sciences (JJBS) has had another great year. We have seen a significant increase in articles submission from both regional and international scholars. The editorial board members of JJBS have been very busy throughout the year to maintain excellence in the quality publication of accepted papers. As a result, JJBS has been indexed by CABI's Full-Text Repository, EBSCO and is currently under evaluation to be indexed in National Library of Medicine's MEDLINE\ PubMed system and Elsevier's SciVerse Scopus. As in the previous two years, this sixth volume of JJBS will include four issues, ten to twelve articles in each issue. In the coming year, it is my vision to have JJBS publishes more outstanding papers and review articles from distinguished scholars in various areas of biological sciences. In addition, I will be working on the inclusion of JJBS in ISI, which will lead to a wider readership and good impact factor. As you read throughout this inaugural volume of JJBS, I would like to remind you that the success of our journal depends directly on the number of quality articles submitted for review. Accordingly, I would like to request your participation by submitting quality manuscripts for review and by encouraging your colleagues to do the same. One of the great benefits we can provide to our prospective authors, regardless of acceptance of their manuscript or not, is the mentoring nature of our review process. JJBS provides authors with high quality, helpful reviews that are shaped to assist authors in improving their manuscripts.

I would like to thank the JJBS International Advisory Board members for their continuous support of JJBS. Furthermore, I would like to thank the JJBS Editorial Board members for their exceptional work and continuous support to JJBS. My thanks are also extended to the Hashemite University and Jordanian Scientific Research Support Fund for their continuous financial and administrative support to JJBS.

Moreover, and as always, I would like to highlight and proudly thank the group of authoritative reviewers, both local and international, who have done an outstanding work. We are honored to have you on our review list and many thanks for your valuable mentorship and contributions you provided to authors. Indeed, we count on your excellent reviews to include only high quality articles worthy of publication in JJBS. Together, we strive to make JJBS reach a remarkable rank among other international journals. I very much appreciate your support to make JJBS one of the most authoritative journals in biological sciences.

Prof. Khaled H. Abu-Elteen
Editor-in-Chief
Hashemite University
Zarqa, Jordan
March 2013

CONTENTS

Mini Review

- 246 - 251 Bionanotechnology: The Novel Nanoparticles Based Approach for Disease Therapy.
Adel M. Mahasneh

Original Articles

- 252 - 256 Levels of Chromium and Copper in Liver and Muscle Tissues of the Round Sardinella *Sardinella aurita* (Valenciennes) from the Oran Coastline, Algeria.
Nardjess Benamar and Boutiba Zitouni
- 258 - 265 Evaluation of Antioxidant Properties of *Morus nigra* L. Fruit Extracts [II].
Najlaa K. Issa and Rihan S. Abd-Aljabar
- 266 - 271 Antimicrobial Activity of Xerophytic Plant (*Cotula cinerea* Delile) Extracts Against Some Pathogenic Bacteria and Fungi.
Djamel Bensizerara, Taha Menasria, Maimouna Melouka, Lamia Cheriet and Haroun Chenchouni
- 272 - 276 Correlation Between Numerical Profiles Generated for Soil Spore Forming Bacilli and Their Inhibitory Potential Against *Staphylococcus aureus* ATCC 6538.
Qasem M. Abu Shaqra
- 278 - 282 Nitrate Reductase Assay Using Sodium Nitrate for Rapid Drug Susceptibility Testing of *Mycobacterium tuberculosis* Directly on Sputum Samples.
Mohammed Abdul- Imam Almazini
- 284 - 290 Protective Effects of *Enantia chlorantha* Stem Bark Extracts on Acetaminophen Induced Liver Damage in Rats.
Olamide E. Adebiyi and Mathew O. Abatan
- 292 - 299 Odonata of Wadi Al Mujib Catchment with Notes on the Impact of Wadi Al Mujib Dam, Jordan (Insecta: Odonata).
Zuhair S. Amr, Loay S. Al Azzam, Ahmad Katbeh-Bader and Ehab K. Eid
- 300 - 307 Comparative Studies on Anti-hyperglycemic Effects of Ethyl Acetate and Methanol Extract of *Albizia lucida* Benth Bark in Alloxan Induced Diabetic Rats.
Arumugam S. Kumar, Subramanian Kavimani and Korlakunta N. Jayaveera
- 308 - 315 Bacteriological and Mycological Assessment for Water Quality of Duhok Reservoir, Iraq.
Yahya A. Shekha, Hero M. Ismael and Akhter A. Ahmed
- 316 - 319 Effects of *Theileria lestoquardi* Infection on Haematological and Biochemical Parameters in Experimentally Infected Desert Ewes.
Aisha A. Elsadig, Yousif H. Abdalla Elmansoury, Husna M. Elbasheir, Amna E. Babiker, Aza A. Adam, Tahani O. Abdelmageed and Sabri Hussein

- 320 - 323 Computational Prediction of Binding of Methyl Carbamate, Sarin, Deltamethrin and Endosulfan Pesticides on Human Oxyhaemoglobin.
Padma Saxena
- 324 - 327 New Records of Arthropod Ectoparasites of Bats from North-Eastern Algeria.
Mohamed Lamine Bendjeddou, Idir Bitam, Awatef Abiadh, Zihad Bouslama and Zuhair S. Amr
- 328 - 333 Evaluation of Immunomodulatory Effects of Antiepileptic Drug Phenytoin.
Mohammad A. Al- Fararjeh, Mohammad H. Jaber and Yaseen S. Abdelrahman
-

Short Communication

- 334 - 336 Prevalence of Lactose Intolerance in Primary School Children in Qena Governorate, Egypt.
Sawsan M A. Abuhamdah, Ghaleb A. Oriquat, Tahia H. Saleem and Mohammed H. Hassan

Bionanotechnology: The Novel Nanoparticles Based Approach for Disease Therapy

Adel M. Mahasneh*

Department of Biological Sciences, Faculty of Science, University of Jordan, Amman 11942, Jordan

Received: July 22, 2013

Revised: September 16, 2013

Accepted: September 27, 2013

Abstract

Bionanotechnology has probably been the most developing field in the last decade as an integration of biotechnology on the molecular level and nanotechnology. It provided new avenues of research and development to produce novel and new nanomaterials of medicinal applications which paved the way to the vibrant field of nanomedicine. This is dependent on combining biological norms with unique characteristics of nano-sized particles to carry out very specific functions. This will hopefully lead to the elucidation of our understanding of disease therapy and the biology of different life forms. The prospects of using nanoparticles in such fields would include drug and gene therapy, tumor control, detection of pathogens and proteins, tissue and cell engineering, DNA probing, pest control in agricultural fields and many other applications. Knowing the expected implications of novel nanoparticles applications in nanoparticle based medicine and research, it became important to compile literature relevant to this subject in a single publication. Thus, this mini review was aimed to be a contribution to this emerging volatile area of bionanotechnology, where efforts are being exerted to apply basic scientific ideas in clinical practice.

Keywords: Bionanotechnology, Nanoparticles, Nanomaterials, Delivery Systems, Disease Therapy.

1. Introduction

Nowadays, global concern has emerged as we are entering the post antibiotic era with reduced capabilities in some areas to combat microbes and disease (Blecher *et al.*, 2011). Hence, the development of novel therapeutic approaches to these challenges constitute a focal point of modern research (Carpenter *et al.*, 2009; Hentzer and Givskov, 2003; Al-Hussaini and Mahasneh, 2009 a, b; Al-Hussaini and Mahasneh, 2011; Taleb *et al.*, 2012; Taleb and Mahasneh, 2010a; Taleb and Mahasneh, 2010b; Taleb and Mahasneh, 2012; Knipe *et al.*, 2013). The alternative to traditional therapies would be through investigating new and novel avenues of molecular therapies to include, for example, use of probiotics (Mahasneh and Abbas, 2010; Elbaz and Willner *et al.*, 2012), quorum sensing regulation where the virulence of many pathogens is regulated by quorum sensing mechanism (Abudoleh and Mahasneh 2012) and last but not least the combination therapy. In this case, chemotherapy is given in combination with other means such as probiotics and nanoparticles to target specifically the diseased cells or tissues (Taleb and Mahasneh, 2012; Day *et al.*, 2009). Such approaches will definitely improve the outcomes of both traditional, as well as, novel bionanotechnology approaches in nanomedicine, stem cell and gene therapies (Lin *et al.*, 2013). The nanomedicine arena, if exploited

properly, would have far reaching implications on medicinal professions, definition of disease, their diagnosis and treatment. Nanomedicine is based upon the use of nanoparticles at the level of 1-100 nm (Caruthers *et al.*, 2007; Pissuwan *et al.*, 2011; Knipe *et al.*, 2013).

In the last two decades, nanoscaled technologies in health sciences have witnessed ample efforts in research, development and patenting (Caruthers *et al.*, 2007). These efforts formed the cornerstone for the transfer of nanotechnological research into clinical settings (Elbaz and Willner, 2012) and it was expanded to shape what is known nowadays as nanomedicine. The aim of this activity was and still is to maximize efficiency in controlling diseases. To achieve this, nanomedicine concepts are used in the early diagnosis which leads to suggesting the treatment program with the least possible side effects and finally to evaluate its efficacy and feasibility (Park *et al.*, 2010). Conversely, the European Science Foundation defined nanomedicine as “the science and technology of diagnosing, treating and preventing disease and traumatic injury, of relieving pain, and of preserving and improving human health, using molecular tools and molecular knowledge of the human body” (Bawa *et al.*, 2005). Nanodrugs, particularly nanoparticulate drugs, offer advantages over their bulk counterparts in one or more properties such as solubility, bioavailability, half life, stability, ability to cross biological barriers, toxicity, dose delivery and specificity

* Corresponding author. e-mail: amahasneh@ju.edu.jo.

(Blecher *et al.*, 2011; Paulo *et al.*, 2010). To elucidate nanomedicine implications in medical practice, such as cancer therapy, development of specific drug delivery system (Figure 1) and nanorobots have attracted a great deal of research where now we know exactly what such systems specifications would be (Arcese *et al.*, 2012). The fully functional nanorobot should be able to carry a payload (drug), direct mobility to specific targets in patient body, adhere or attach to tumor cells and finally release the payload. Further researchers' efforts would no doubt be turned to design very specific nanorobots and nanoparticles.

The hopes surrounding nanotechnology in different fields, including medicine, are immense (Salvador-Morales *et al.*, 2012). It does not offer improvements to existing techniques only, but provides leads to new tools and applications (del-Pozo-Rodriguez *et al.*, 2013). Among these would be drug manipulation at the nanoscale with what impact such manipulation would leave upon the bioactive characteristics such as controlled release, solubility, retention time and probably environmentally triggered controlled release (Douglas *et al.*, 2012). It is also realized that nanoparticles' surface area provides increased functionality which lends itself to further biomedical as well as environmental applications (Chunbai *et al.*, 2013) and gene therapy (Pissuwan *et al.*, 2011).

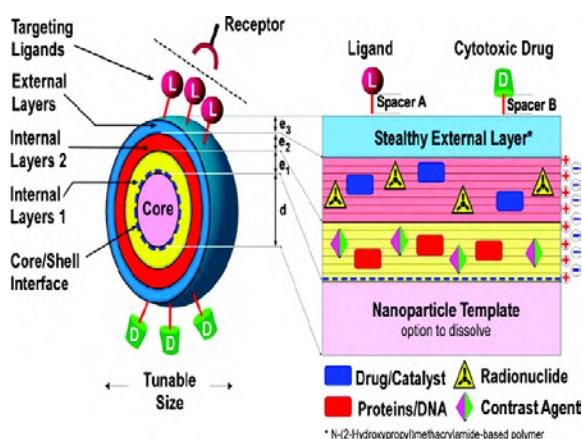


Figure 1. Core/shell drug delivery system consisting of cytotoxic stealth carrier particles based on nanoparticles. Usually the core is surrounded by several layers and chemical groups could be added to the external layer thus allowing attachment of biomolecules.

2. Applications of Biocompatible Nanoparticles

The distinct characteristics of the varying types of nanoparticles presented them for a wide variety of applications in medicine (Sih *et al.*, 2013), in biological sciences for the development of biosensors, DNA probes (Tang *et al.*, 2006), in medicine (Mukherjee *et al.*, 2007), in controlling pathogenic bacteria (Raja *et al.*, 2012) and fungi (Min *et al.*, 2009), and in ecological applications pertaining to water treatment (Lyon *et al.*, 2006) and plant diseases (Jo *et al.*, 2009; Aguilar-Mendez, 2010; Kim *et al.*, 2008).

Nanoparticles of the size 1-10 nm were capable of readily interacting with HIV virus through selective binding to glycoprotein thus inhibiting viruses from binding to host cells (Elechiguerra *et al.*, 2005). Metallic nanoparticles such as silver nanos are being used in medical implants and devices to prevent microbial infections (Geethalakshmi and Sarada, 2010). Furthermore, some nanoparticles have been tested for their probable use in burnt tissues, alternative dental materials and textile fabrics (Duran *et al.*, 2007).

Gene therapy is an emerging field where nanoparticles would no doubt play a major role in elucidating means and ways of introducing and delivering genes into target cells or tissues. In here, nucleic acids are delivered through special delivery systems thus controlling gene flow and expression. Pedro *et al.* (2008) recall the great advances in human gene therapy which were driven by interest in understanding of the molecular mechanisms of diseases and what they implicate of vector design in the quest for producing more efficient and safe drug in addition to gene delivery systems thus bridging bottlenecks through new technologies. In this context, it is worthy to mention research efforts in the search for designer bionanoparticles in the form of viral like particles (VLPs). These nanoparticles are unique in their diversity in terms of structure, shapes, architecture and production means (Figure 2). In this direction, new administration routes are being sought through oral and pulmonary means although intravenous route is the one of choice for most current nanoparticles delivery (Uchegbu and Siew, 2013). It should be noted that an array of cell types exist in the human body, which means that higher organisms, including our bodies, are just compilations of multifunctional nanosystems (Suh *et al.*, 2009). Research on nanoparticles usually involved organic, inorganic and composite nanos to be produced, characterized and then used in drug delivery, bioimaging and other applications which necessitates further biocompatibility studies (Salvador-Morales, 2012). The shortage of effective and safe *in vitro* delivery systems has been a challenging obstacle in developing new nanoparticle based therapeutics (Troiber and Wagner, 2011). To secure an acceptable level of safety, other aspects of nanoparticles have to be considered. Priority aspects include routes of administration, therapeutic dose and frequency as well as reproducibility (Mohan, 2010; Azzazy and Mansour, 2009).

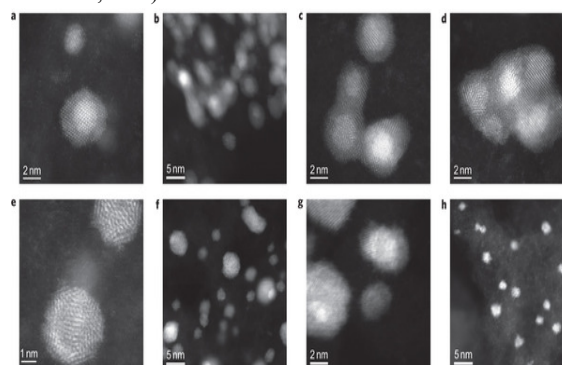


Figure 2. Images of nanoparticles showing diversity in terms of structure, shape, architecture and arrangements.

3. Nanoparticles Preparation Methods

Synthesis of nanoparticles has emerged in the last decade as an intersection between nanotechnology and biotechnology; hence the coining of the new expanding bionanotechnology (Raja *et al.*, 2012). Due to the possible wide application of nanoparticles, this emerging science put nanoparticles at the tip of modern material science research. These nanoparticles are expected to exhibit new and/or novel characteristics associated with size, morphology and distribution (Veerasamy *et al.*, 2011). Such characteristics are deemed to meet different applications of nanoparticles (Sayed and Ahmad, 2013). To improve the functionality of nanoparticles, it is mandatory to be functionalized accordingly with probably protein or biocompatible surfaces to reduce their toxicity if they are of inorganic origin such as the quantum dots (Sayed and Ahmad, 2013). Recent literature lists several possible means of nanoparticle synthesis ranging from totally chemical methods to fully biological processes (Veerasamy *et al.*, 2011) such as reduction in solutions, chemical and photochemical reactions, thermal decomposition, radiation-assisted (Bhuvanasree *et al.*, 2013), electrochemical and finally most recently green synthesis (Raja *et al.*, 2012; Sayed and Ahmad, 2013; Gopinath and Velusamy, 2013). Nanoparticles synthesis using mostly toxic chemicals with low rates of conversion and laborious purification methods with high energy physical procedures prompted investigators to search for biologically mediated methods including microorganisms such as fungi, bacteria and plant systems including plant extracts (Gopinath and Velusamy, 2013; Sayed and Ahmad, 2013). These biological processes offered great deal in developing new methods to produce nanoparticles. They offer cost-effective methods and they are easily sealed up and economize energy demands and finally they are ecofriendly (Ponarulselvam *et al.*, 2012; Song and Kim, 2009; Ahmad *et al.*, 2011). Table 1 highlights the most used methods of nanoparticle preparation.

Table 1. Brief description of the most common and emerging nanoparticles preparation methods; (+): produced; (++): experimental optimization.

Method	Advantages	Scale-up
Nanoprecipitation of polymers	Fast; reproducible	++
Polymerization of alkylcyanoacrylates	Easy to produce core-shell nanos	++
Interfacial polycondensations	Modulation of nanos thickness	++
Formation of polyelectrolyte complexes	Positive or negative charge nanos are produced	++
Nanos from neutral nanogels	Organic solvent-free; controlled release of drugs	++
Ionic gelation	Possibility of controlled drug release in response to a stimulus	++

Gelation of emulsion droplets	Hydrophilic and biocompatible	++
Emulsification-reverse salting-out	Less stress upon fragile drugs	++
Emulsification-solvent evaporation	Encapsulation of hydrophilic and lipophilic drugs	++
Colloidal mill	Controlled nano size	++
Natural organisms-Fungi	Simple; non-toxic; reliable; ecofriendly	++
Natural organisms-Bacteria	Well defined size and distinct morphology; biocompatible; ecofriendly	++
Natural products-plant extracts (green synthesis)	Cost-effective; simple; ecofriendly	++

4. Molecular Therapy and Delivery Systems

The ample amount of information pertaining to the understanding of molecular mechanisms of diseases that was gathered in the last two decades coupled with advances at the gene therapy level has prompted research for more efficient, effective and safe delivery systems (Pedro *et al.*, 2008; Bansal *et al.*, 2012; Neves *et al.*, 2012). In cancer treatment, current research include development of delivery systems that allow very specific dosing routes and unusual therapeutic targets (Brannon-Peppas and Blanchette, 2012; Mignani *et al.*, 2013).

In the last few years we observed the emergence of many new and improved intelligent nanoscale platforms which is suitable for drug and gene delivery as well as imaging. It also witnessed the search for smart functionality in delivery systems to include cell and receptor specific targeting in the context of nano-bio interaction (Lehner *et al.*, 2013) in nanomedicine (accepted). Nanotechnology-based carriers have shown very successful results in terms of low toxicity and efficient delivery to targets including cells and/or tissues of mammalian systems (Seth *et al.*, 2012; Shen *et al.*, 2012). Different nanotechnology systems would provide solutions for improved delivery with least toxicity which is the core for translating effectively basic medical sciences into enhanced clinical practice (Liu and Zhang, 2011; Singh *et al.*, 2012).

In this context, consideration should be given to the size of delivery vehicle, ease of penetration into the cellular membranes and sustainability in the cytoplasm (Li and Huang 2010). Among the best studied nanocarriers are those synthesized from liposomes, dendrimers, carbon nanotubes and nanoclusters (fullerenes), polymers and metal and metal oxides nanoparticles. Table 2 presents some delivery systems and some of their unique characteristics that make therapy more effective.

From the aforementioned table, it is rather clear that the best fit delivery system is one that exhibits characteristics of both non-viral as well as viral delivery vectors coupled with disarming such systems from undesired properties.

Table 2. Diversity of different delivery systems available and under testing and could be used for the treatment of different diseases.

Delivery system	Main characteristics	Comments
Liposomes	Biocompatible and biodegradable; most suitable for DNA, RNA delivery into mammalian cells; persistence in blood circulation; increased specificity for gene therapy	Pharmaceutical and medical applications; easy to synthesize and easy storage
Dendrimers	Highly symmetric; many functional groups at surfaces; better encapsulation of genes; high payload delivery	May act as multifunctional delivery application; modifiable surface groups thus tuning their activity
Fullerenes	Carbon nanoclusters; possibility of functionalizing it in different ways such as making it water-soluble; less toxic than cationic liposomes	It forms protective sheath over bound DNA expanding chances of incorporation into chromosomes
Carbon nanotubes	Gene and drug delivery; compatibility with aqueous environments; nontoxic in mammalian cells and tissues	Very successful in gene delivery for disease therapy
Quantum dots	Extremely small; mainly of heavy metal origin; very effective in imaging enhancement	Often they undergo leaching in biological environment leading to toxicity in mammalian cells
Gold, silver magnetic nanoparticles	Highly noted in biomedical application, bionanotechnology, imaging, gene and drug delivery for disease and gene therapy	Nontoxic nature of gold nanos; non-opsogenic and of stealth properties; used in magnetic hyperthermia therapy cancer cells
Viral systems – oncoviruses	Specific gene therapy; stability during genome integration; insert large gene segments; infect only dividing cells	During gene therapy trials, possible recombination with endogenous human retroviruses
Viral systems – Adenovirus	Gene therapy; low pathogenicity for humans; infect non-dividing cells	It induces strong immune response; no integration into the host
Viral systems – Adeno-associated-virus	No pathogenicity and toxicity; infect non-dividing cells; long-term transgene expression	No specific integration' may mutagenize cells
Viral systems – Lentivirus	Can carry large gene inserts; stable expression; infect non-dividing cells	Biosafety problems

However, more work is needed to overcome problems such as lack of specific targeting, toxicity, low transfection efficacy and expression in case of gene therapy and specificity, all of which hindered advancement in molecular and regular therapy approaches (Knipe *et al.*, 2013).

Finally, it seems that nanotechnology, especially bionanotechnology is expanding at a speedy pace exploring novel methods at both the traditional and the molecular levels of therapy.

It is also clear that progress of the new emerging field of nanomedicine is dependent on exploring bionanotechnology potential in terms of translating the laboratory based trials and observations into the real clinical settings. These necessitate further research on scale-up, inconsistencies from batch to batch, confirmation of stealth properties of nanoparticles *in vivo*, improved specificity of the different nanoparticles and finally improved targeting means through nanorobots development. The concept of smart nanosystems for new and novel biomedical applications is being exploited. Solution of all or part of these concerns would no doubt open new frontiers for the nano-based diagnostics for the detection of tumors, infections and neurological diseases among many others. At this stage of scientific discovery I would dare to say that nanoparticles-based nanomedicine would in the very near future transform the understanding of the human biological structure and function. This transformation will no doubt furnish new nanoparticle based applications to include chemotherapy, activity monitors (blood pressure, glucose monitors), biochips,

pacemakers, insulin pumps, needleless injectors, medical flow sensors, drug delivery systems and finally designer gene therapy systems. This would expand the prospective horizons of nanoparticles use as well as the need for further investigations pertaining to the final fate and probable interactions of these bionanoparticles in biological systems.

5. Conclusion

Science has greatly advanced into the area of applied nanotechnology and the internal components of living cells are of the same scale. This lead bionanotechnologists to look to cell biology for medicinal applications using biological structures, processes and information. Much of bionanotechnology is molecular biology based applications. Individual molecules, bacteria and viruses can be easily detected. Nanoparticles are already in use for drug delivery trials, biological labeling, medical imaging and various other analytical purposes for clinical use. Other new and more complex bionanodevices are being produced and investigated for novel applications.

Acknowledgements

Thanks are due to Ms. Sara Hamdan who helped in typing the manuscript.

References

- Abudoleh SM and Mahasneh AM. 2012. Quorum sensing inhibitors from epiphytic bacteria isolated from wild berries. In : Mendez-Vilas A (Eds), **Microbes in Applied Research: Current Advances and Challenges**. Formatex Research Center, Spain.
- Aguilar-Mendez A, Martin-Martinez S, Ortega-Arroyo L, Cobian-Portillo G and Sanchez-Espindola E. 2011. Synthesis and characterization of silver nanoparticles: effect on phytopathogen *Colletotrichum gloeosporioides*. *J Nanoparticles Res.*, **13(6)**:2525-2532.
- Ahmad N, Sharma S, Singh VN, Shamsi SF, Fatma A and Mehta BR. 2011. Biosynthesis of silver nanoparticles from *Desmodium triflorum*: a novel approach towards weed utilization. *Biotechnol Res Int.*, **2011**: doi:10.4061/2011/454090.
- Al-Hussaini R and Mahasneh AM. 2009a. Quorum sensing and microbial growth antagonistic activity of *Laurus nobilis*. *Jordan Medical J.*, **43(4)**:286-294.
- Al-Hussaini R and Mahasneh AM. 2009b. Microbial growth and quorum sensing activities of herbal plant extracts. *Molecules*, **14(9)**:3435-3448.
- Al-Hussaini R and Mahasneh AM. 2011. Antibacterial and antifungal activity of ethanol extract of different parts of medicinal plants in Jordan. *Jordan J Pharmaceutical Sci.*, **4(1)**:57-69.
- Arcese L, Fruchard M and Ferreira A. 2012. Endovascular magnetically guided robots: navigation modeling and optimization. *IEEE Transaction on Biomedical Eng.*, **59(4)**:977-987.
- Azzazy HM and Mansour MM. 2009. *In vitro* diagnostic prospects of nanoparticles. *Clinica Chimica Acta*, **403(1-2)**:1-8.
- Bansal SS, Kausar H, Vadhanam MV, Ravoori S and Gupta RC. 2012. Controlled systemic delivery by polymeric implants enhances tissue and plasma curcumin levels compared with oral administration. *Eur J Pharm Biopharm.*, **80(3)**:571-577.
- Bawa R, Bawa SR, Maebius SB, Flynn T and Wei C. 2005. Protecting new ideas and inventions in nanomedicine with patents. *Nanomedicine: Nanotechnology, Biology and Medicine*, **1(2)**:150-158.
- Bhuvanaree SR, Harini D, Rajaram A and Rajaram R. 2013. Rapid synthesis of gold nanoparticles with *Cissus quadrangularis* extract using microwave irradiation. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, **106**:190-196.
- Blecher K, Nasir A and Friedman A. 2011. The growing role of nanotechnology in combating infectious diseases. *Virulence*, **2(5)**:395-401.
- Brannon-Peppas L and Blanchette J. 2012. Nanoparticle and targeted systems for cancer therapy. *Advanced Drug Delivery Rev.*, **56(11)**:1649-1659.
- Carpenter MK, Frey-Vasconcells J and Rao MS. 2009. Developing safe therapies from human pluripotent stem cells. *Nature Biotechnol.*, **27(7)**:606-613.
- Caruthers SD, Wickline SA and Lanza GM. 2007. Nanotechnological applications in medicine. *Current Opinion in Biotechnol.*, **18(1)**:26-30.
- Day E S, Morton JG and West JL. 2009. Nanoparticles for thermal cancer therapy. *J Biomechanical Eng.*, **131(7)**:074001 doi:10.1115/1.3156800.
- del Pozo-Rodriguez A, Delgado D, Gascon AR and Solinis MA. 2013. Lipid nanoparticles as drug/gene delivery systems to the retina. *J Ocular Pharmacol Therapeutics*, **29(2)**:173-188.
- Douglas SM, Bachelet I and Church GM. 2012. A logic-gated nanorobot for targeted transport of molecular payloads. *Science*. **335(6070)**:831-834.
- Duran N, Marcato PD, DeSouza G, Alves OL and Esposito E. 2007. Antibacterial effect of silver nanoparticles produced by fungal process on textile fabrics and their effluent treatment. *J Biomedical Nanotechnol.*, **3(2)**:203-208.
- Elbaz J and Willner I. 2012. DNA origami: nanorobots grab cellular control. *Nature Materials*, **11**:276-277.
- Elechiguerra JL, Burt JL, Morones JR, Camacho-Bragado A, Gao X and Lara HH. 2005. Interaction of silver nanoparticles with HIV-1. *J Nanobiotechnol.*, **3**:6-11.
- Geethalakshmi R and Sarada DL. 2010. Synthesis of plant-mediated silver nanoparticles using *Trianthema decandra* extract and evaluation of their antimicrobial activities. *Inter J Eng Sci Technol.*, **2(5)**:970-975.
- Gopinath V and Velusamy P. 2013. Extracellular biosynthesis of silver nanoparticles using *Bacillus* sp. GP-23 and evaluation of their antifungal activity towards *Fusarium oxysporum*. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, **106**:170-174.
- He C, Yin L, Tang C and Yin C. 2013. Multifunctional polymeric nanoparticles for oral delivery of TNF- α siRNA to macrophages. *Biomaterials*, **34(11)**:2843-2854.
- Hentzer M and Givskov M. 2003. Pharmacological inhibition of quorum sensing for the treatment of chronic bacterial infections. *J Clinical Invest.*, **112(9)**:1300-1307.
- Jo YK, Kim BH and Jung G. 2009. Antifungal activity of silver ions and nanoparticles on phytopathogenic fungi. *Plant Dis.*, **93(10)**:1037-1043.
- Kim HS, Kang HS, Chu GJ and Byun HS. 2008. Antifungal effectiveness of nanosilver colloid against rose powdery mildew in greenhouse. *Solid State Phenomena*, **135**:15-18.
- Knipe JM, Peters JT and Peppas NA. 2013. Theranostic agents for intracellular gene delivery with spatiotemporal imaging. *NanoToday*, **8(1)**:21-38.
- Lehner R, Wang X, Marsch S and Hunziker P. 2013. Intelligent nanomaterials for medicine: carrier platforms and targeting strategies in the context of clinical application. *Nanomedicine: Nanotechnology, Biology and Medicine*, **9(6)**: 742-757.
- Li J Huang L. 2010. Targeted delivery of RNA: Therapeutics for cancer therapy. *Nanomedicine (London)*. **5(10)**:1483-1486.
- Lin HT, Otsu M and Nakauchi H. 2013. Stem cell therapy: an exercise in patience and prudence. *Phil. Trans. R. Soc. B.* doi:10.1098/rstb.2011.0334. 1471-2920.
- Liu C and Zhang N. 2011. Nanoparticles in gene therapy principles prospects and challenges. *Prog Mol Biol Transl Sci.*, **104**:509-562.
- Lyon DY, Adams LK, Falkner JC and Alvarez PJJ. 2006. Antibacterial activity of fullerene water suspensions: effects of preparation method and particle size. *Environ. Sci. Technol.*, **40(14)**:4360-4366.
- Mahasneh AM and Abbas MM. 2010. Probiotics and traditional fermented foods: the eternal connection. *Jordan J Biol Sci.*, **3(4)**:133-140.
- Mignani S, Elkazzonli S, Bousmina M and Majoral J. 2013. Expand classical drug administration ways by emerging routes

- using dendrimer drug delivery systems: A concise overview. *Advanced Drug Delivery Rev.*, **65**(10): 1316-1330.
- Min JS, Kim KS, Kim SW, Jung JH, Lamsal K, Kim SB, Jung MY and Lee YS 2009. Effects of colloidal silver nanoparticles on sclerotium-forming phytopathogenic fungi. *Plant Patho J.*, **25**(4):376-380.
- Mohanan D, Slutter M, Henriksen-Lacey M, Jiskoot W, Bouwstra JA and Perrie Y. 2010. Administration routes affect the quality of immune responses: a cross-sectional evaluation of particulate antigen-delivery systems. *J Control Release*, **147**(3):342-349.
- Mukherjee P, Bhattacharya R, Bone N, Lee YK, Parta CR and Wang S. 2007. Potential therapeutic application of gold nanoparticles in B-chronic lymphocytic leukemia (BCLL) enhancing apoptosis. *J Nanobiotechnol.*, **5**:4-11.
- Neves AR, Lucio M, Lima JL and Reis S. 2012. Resveratrol in medicinal chemistry: A critical review of its pharmacokinetics, drug delivery and membrane interactions. *Curr Med Chem.*, **19**(11):1663-1681.
- Park S, Cha K and Park J. 2010. Development of biomedical microbot for intravascular therapy. *Intern J A dvanced Robotic Systems*, **7**:21-27.
- Paulo CS, Vidal M and Ferreira LS. 2010 Antifungal nanoparticles and surfaces. *Biomacromolecules*, **11**(10):2810-2817.
- Pedro L, Soares SS and Ferreira GNM. 2008. Purification of bionanoparticles. *Chem Eng Technol.*, **31**(6):815-825.
- Pissuwan D, Niidome T and Cortie MB. 2011. The forthcoming applications of gold nanoparticles in drug and gene delivery systems. *J Controlled Release*, **149**:65-71.
- Ponarulselvam S, Panneerselvam C, Murugan K, Aarthi N, Kalimuthu K and Thangamani S. 2012. Synthesis of silver nanoparticles using leaves of *Catharanthus roseus* Linn. G. Don and their antiplasmodial activities. *Asian Pacific J Tropical Biomedicine*. **2**(7):574-580.
- Raja K, Saravanakumar A and Vijayakumar R. 2012. Efficient synthesis of silver nanoparticles from *Prosopis juliflora* leaf extract and its antimicrobial activity using sewage. *Spectrochimica Acta Part: Molecular and Biomolecular Spectroscopy*, **97**:490-494.
- Salvador-Morales C, Valencia PM, Thakkar AB, Swanson EW and Langer B. 2012. Recent developments in multifunctional hybrid nanoparticles: opportunities and challenges in cancer therapy. *Frontiers in Bioscience*, **4**:529-545.
- Sayed A and Ahmad A. 2013. Extracellular biosynthesis of CdTe quantum dots by the fungus *Fusarium oxysporum* and their antibacterial activity. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, **106**:41-47.
- Schneider GF, Subr V, Ulbrich K and Decher G. 2009. Multifunctional cytotoxic stealth nanoparticles: A model approach with potential for cancer therapy. *Nano Lett.*, **9**(2):636-642.
- Serpell C J, Cookson J, Ozkaya D and Beer PD. 2011. Core-shell bimetallic nanoparticle synthesis via anion coordination. *Nature Chem.*, **3**:48-483.
- Seth S, Johns R and Templin MV. 2012. Delivery and biodistribution of siRNA for cancer therapy: challenges and future prospects. *Ther Deliv.*, **3**(2):245-261.
- Shen H, Sun T and Ferrari M. 2012. Nanovector delivery of siRNA for cancer therapy. *Cancer Gene Ther.*, **19**:367-373.
- Sih J, Bansal SS, Filipini S, Ferrati S, Raghuwansi K, Zabre E, Nicolov E, Fine D, Ferrari M, Palapattu G and Grattoni A. 2013. Characterization of nanochannel delivery membrane systems for the sustained release of resveratrol and atorvastatin: new perspectives on promoting heart health. *Anal Bioanal Chem.*, **405**(5):1547-1557.
- Singh S. 2013. Nanomaterials as nonviral delivery agents for cancer therapy. *BioImpacts*. **3**(2):53-65.
- Singh S, Sharma A and Robertson G P. 2012. Realizing the clinical potential of cancer nanotechnology by minimizing toxicologic and targeted delivery concerns. *Cancer Res.*, **72**(22):5663-5668.
- Song JY and Kim BS 2009. Rapid biological synthesis of silver nanoparticles using plant leaf extracts. *Bioprocess Biosyst Eng.*, **32**(1):79-84.
- Suh WH, Suh YH and Stucky GD. 2009. Multifunctional nanosystems at the interface of physical and life sciences. *NanoToday*, **4**(1):27-36.
- Talib WH and Mahasneh AM. 2010a. Antimicrobial, cytotoxicity and phytochemical screening of Jordanian plants in traditional medicine. *Molecules*, **15**(3):1811-1824.
- Talib WH and Mahasneh AM. 2010b. Antiproliferative activity of plant extracts used against cancer in traditional medicine. *Scientia Pharmaceutica*, **78**(1):33-45.
- Talib WH and Mahasneh AM. 2012. Combination of *Ononis hirta* and *Bifidobacterium longum* decreases syngeneic mouse mammary tumor burden and enhances immune response. *J Cancer Res Therapeutics*, **8**(3):417-423.
- Talib WH, AbuZarga MH and Mahasneh AM. 2012. Antiproliferative, antimicrobial and apoptosis inducing effects of compounds isolated from *Inula visciosa*. *Molecules*, **17**(3):3291-3303.
- Tang D, Yuan R and Chai Y. 2006. Ligand-functionalized core/shell Ag-Au nanoparticles label-free amperometric immune-biosensor. *Biotechnol Bioeng.*, **94**(5):996-1004.
- Troiber C and Wagner E. 2011. Nucleic acid carriers based on precise polymer conjugates. *Bioconjugation Chem.*, **22**:1737-1752.
- Uchegbu IF and Siew A. 2013. Nanomedicines and nanodiagnostics come of age. *J Pharmaceutical Sci.*, **102**(2):305-310.
- Vauthier C and Bouchemal K. 2009. Methods for the preparation and manufacture of polymeric nanoparticles. *Pharmaceutical Res.*, **26**(5):1025-1058.
- Veerasamy R, Xin TZ, Gunasagaran S, Xiang TFW, Yang E F C, Jeyakumar N and Dhanaraj SA. 2011. Biosynthesis of silver nanoparticles using mangosteen leaf extract and evaluation of their antimicrobial activities. *J Saudi Chemical Soc.*, **15**(2):113-120.

Levels of Chromium and Copper in Liver and Muscle Tissues of the Round Sardinella *Sardinella aurita* (Valenciennes) from the Oran Coastline, Algeria

Nardjess Benamar and Boutiba Zitouni

Environmental Surveillance Laboratory, Department of Biology, University Oran ,Algeria

Received: December 24, 2012 Revised: February 9, 2013 Accepted: February 14, 2013

Abstract

The aim of the present study is to determine the levels of two heavy metals (Chromium and Copper) in the liver and muscle tissues of *Sardinella aurita* from the Oran coastline. Metal levels in fish samples were analyzed by using atomic absorption. 400 readings were carried out between May and October 2007. The order of heavy metal accumulation in the fish were in the order of magnitude as $Cu > Cr$. The average concentrations recorded in the fish, in this study, ranged, respectively, between 1.9 to 3.97 mg/kg for Copper, and between 0.07 to 0.12mg/kg for Chromium. The levels measured in liver tissue are higher than those reported for muscle tissue. The present study confirms that the round Sardinella *Sardinella aurita* (Valenciennes, 1847), from the Oran coastline, can bioaccumulate heavy metals from a polluted environment. The average concentrations of Cr (0.083 ± 0.01 mg/kg) and Cu (2.78 ± 2.92), recorded in our study, were above FAO limiting standards for food fish.

Key words. *Sardinella aurita*; Pollution, Heavy Metals, Chromium, Copper; Oran coastline.

1. Introduction

Heavy metal pollution has become a serious environmental and public health issue (Venkatramreddy *et al.*, 2009). Heavy metals are commonly found in natural waters and some are essential to living organisms. Yet they may become highly toxic when present in high concentrations (Ibok *et al.*, 1989). The toxicity for humans is mainly caused by their persistence in the environment. This situation is a result of the rapid growth of population, intense industrialization and other anthropogenic activities, like the exploration and exploitation of natural resources, the extension of irrigation and modern agricultural practices (FAO, 1992). Due to their bioaccumulative and non-biodegradable properties, heavy metals constitute a core group of aquatic pollutants.

The objective of the present study is to determine the level of two heavy metals: chromium and copper in organs of *Sardinella aurita* from gulf of Oran.

Sardinella aurita (Teleostei; Clupeidae) was selected for the present study. It is a small pelagic fish that lives in tropical and subtropical waters of the western and eastern Atlantic Ocean, the Pacific Ocean, the Mediterranean and, occasionally, the Black Sea (Sabate's, 2006). In the Algerian coasts; it is very common and very abundant (Benamar, 2011). It is

also considered, in Oran, the most consumed fish after sardine *Sardina pilchardus* and bug *Boops boops* (Benamar, 2011).

For the purposes of the present paper, two metals were detected: copper and chromium. It has been established that copper is one of the most abundant trace metals and for almost all organisms; it is an essential micronutrient (Duffus, 1980). The aquatic toxicology of Cr depends on both biotic and abiotic factors. The biotic factors include the type of species, age and developmental stage. When bioconcentrating in the food chain, heavy metals, including Cr, are potentially cytotoxic to aquatic biota. Therefore, an early detection and ecotoxicologic evaluation of a sensitive biomonitoring system comprising both in vivo and in vitro test systems is essential (Venkatramreddy *et al.*, 2009).

The Chromium was also shown to accumulate mainly in metabolically active organs such as liver, gill and kidneys at high concentrations. It was shown that fish go under some behavioral alterations such as suspending feeding, irregular swimming and accelerated operculum movement when first encountered with chromium (Svecevicius, 2009). Chromium also caused structural changes such as hypertrophy and hyperplasia at gill epithelium, degeneration in fin rays and weakening of immune system (Synder and Valle, 1991; Bennani *et al.*, 1996; Arunkumar *et al.*, 2000).

2. Materials and Methods

2.1. Study Area

The gulf of Oran, on the Algerian Mediterranean coast, is located between the industrial gulf of Arzew in the east, and the unspoiled Andalous Coast in the west (Figure 1). More than 90 million m³ of untreated wastewaters are discharged annually by the Oran metropolis and many industrial units. Generally, these industries shall carry out the evacuation of their waste without any processing. Chemical releases, discharges of hydrocarbons of petrochemical origin (industrial zone of Arzew), and dissolved mineral and metal waste are thrown more heavily in effluents from various industries. The sea constitutes a major receiving area for waste water in Oran, as is the case for the majority of Algerian coastal towns. The sea is also the direct discharge system for effluents from the town of Oran. Total waste-water discharge along the bay of Oran can be estimated for industrial plants (with water consumption of more than 1000 m³/year) and domestic discharge (SOGREAH Engineering, 1998).

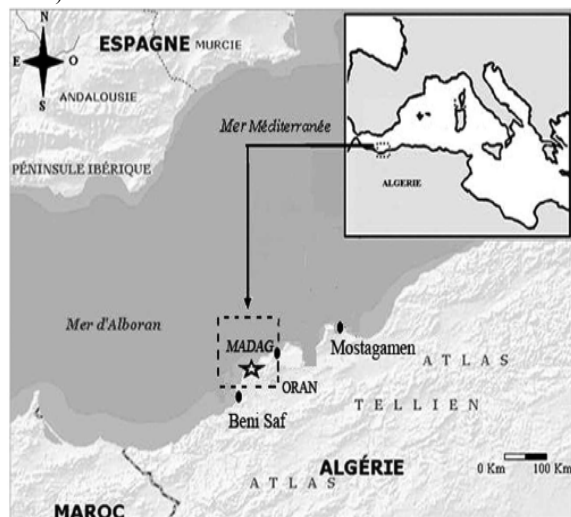


Figure 1. Geographical location of the site bay, Oran, Algeria

2.2. Sample Collection and Treatment

Samples of *Sardinella aurita* were collected between May 2007 and October 2007. Samples were placed in polyethylene bags and transported in a polystyrene ice-chest to the laboratory. The total lengths (cm) and weights (g) of the fish species were measured and after measurements, livers and muscles were removed, weighed and frozen until the time of their chemical analysis. Samples were prepared for trace metal analysis (UNEP, 1984a).

Wet mineralization of samples was performed according to the method of Amiard *et al.* (1987) using a mineralizer type VELP. One ml of nitric acid is added to 1g wet weight of fish sample and then adjusted to 4 ml of bidistilled water after one hour at 95°C. The trace metals were determined by flame atomization (UNEP, 1984b) using a Perkin Elmer, Analyst 100 Atomic Absorption Spectrophotometer. To ascertain the accuracy of the results, blanks were included in every batch of nine samples analyzed. Analytical quality control measures adopted by the International Atomic Energy Agency (IAEA-350) – the intercomparison run which uses tuna fish homogenate as a certified reference material was used. The statistical analyses were made by means of the software Statistica version 5.0 (Co Microsoft).

3. Results

In this study, the concentrations of metals copper and chromium were detected in all samples. The average concentration in mg/kg wet weight, in the liver and muscles of the round *Sardinella*, are summarized respectively in figure 2.

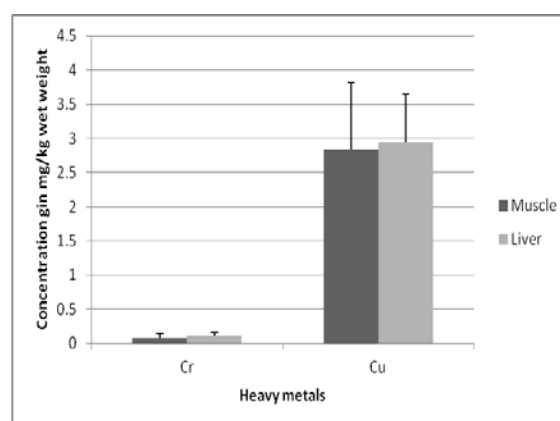


Figure 2. The average concentration of copper and chromium in organs of *Sardinella aurita* (mean \pm standard deviation).

The results showed that the values of copper were higher than that of chromium. The distribution patterns of Cr and Cu in the two organs of the *Sardinella* followed the order: liver > muscles.

The mean concentrations of copper in all examined tissues varied from 1.90 ± 0.4 to 3.96 ± 1.54 mg/kg (figure 3). The higher concentration level in the two organs was observed in August (figure 3). Meanwhile, the chromium concentration ranged from 0.07 ± 0.1 to 0.12 ± 0.04 mg/kg. The highest mean concentration level of chromium was observed between June and August, as shown in figure 4.

The mean concentrations of Cr and Cu in liver and muscle of the round *Sardinella* showed that the females accumulate more than the males but without statistical significance (figure 5).

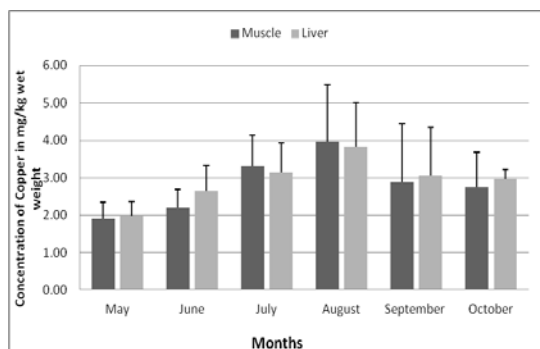


Figure 3. The monthly average concentration of copper in organs of *Sardinella aurita* (mean \pm standard deviation).

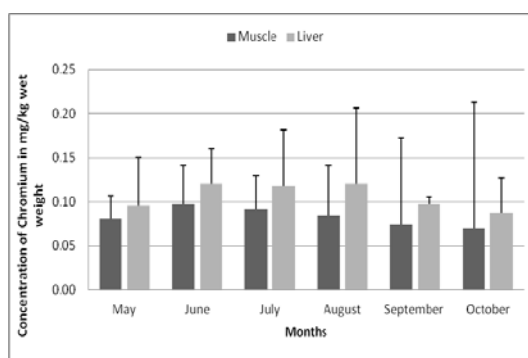


Figure 4. The monthly average concentration of chromium in organs of *Sardinella aurita* (mean \pm standard deviation).

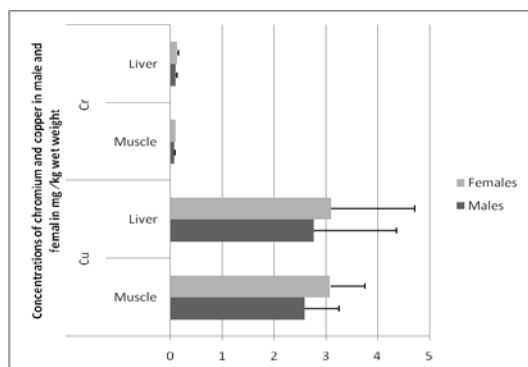


Figure 5. The average concentration between males and females of copper and chromium in organs of *Sardinella aurita* (mean \pm standard deviation).

4. Discussion

Concentrations of heavy metal, detected in the muscle and liver samples, showed different capacities for accumulating. The observed variability of heavy metal levels in *Sardinella aurita* depends on the level of exposure (water, food) and physiological factors (age, metabolic activity), together with factors related to environment such as temperature, physical-chemical parameters, presence of other metals (Kim *et al.*, 2004).

Metals accumulate in higher concentrations in the liver of *Sardinella aurita*, compared with muscle tissue. A similar result was found in the sardine *Sardina pichardus* fishing in the Bay of Oran (Merbouh, 1997). In fish, the most toxic metals tend to accumulate in the liver or kidneys (ICES, 1991). It has

been established that the liver is a target organ for the accumulation of heavy metals, and the higher levels of these metals in the liver, compared to other organs, may be attributed to the high coordination of metallothionein protein with the metals (Hogstrand and Haux, 1991). In addition, the liver is the principal organ responsible for the detoxification, transportation and storage of toxic substances and an active site of pathological effects induced by contamination. The muscle is not considered an active tissue in accumulating heavy metals (Yilmaz, 2003). However, it has been proven that all trace metals concentration in the muscle tissue of *Sardinella aurita* was positively correlated to lipid content in the muscle tissues (Wafica and Aboul Naga, 1996). As for regulation and guideline levels, the Food and Agriculture Organization (FAO) has set permissible limits for heavy metals in the muscle of fish (because it is the part consumed). The mean levels found in *Sardinella aurita* samples are 2,8 mg/kg for Copper and 0.08 mg/kg for Chromium.

The levels of (Cu) in the fish sampled were higher than those of (Cr). This is because it represents an essential homeostatically regulated metal in all living organisms (Goyer, 1996). Cu as a microelement is essential for the normal growth and metabolism of plants, animals and most microorganisms (Schroeder *et al.*, 1996). Additionally, it is the third largest trace element found in a living organism's body after iron and zinc. It is a component of many enzymes like cytochrome-c oxidase, superoxide dismutase, tyrosinase, dopamine beta hydroxylase ferroxidases and amine oxidase. Copper is involved in neurotransmitter regulation, nutrient metabolism, collagen synthesis, cellular respiration and immune function.

The copper contents in the samples were less than the FAO-permitted level of WHO (3.0mg/kg) (Onianwa *et al.*, 2001). Excessive intake of copper may lead to liver cirrhosis, dermatitis and neurological disorders (Onianwa *et al.*, 2001). Cu compounds, such as copper sulphate, are widely used as biocides to control macrophytes, freshwater snails that may harbour schistosomiasis or other disease pathogens, ectoparasites of fish and mammals, marine fouling organisms (Fisler, 1998).

On the other hand, Chromium does not normally accumulate in fish, and hence, low concentrations of Cr were reported even from different industrialized parts of the world (Moore and Ramamoorthy, 1984). Cr bioaccumulation in fish has been reported to cause impaired respiratory and osmoregulatory functions through structural damage to gill epithelium (Heath, 1991). The values of Cr recorded in *Sardinella aurita* in this study were above FAO limiting standards of 0.15mg/kg for food fish. The WHO has proposed that chromium (VI) is a human carcinogen. Several studies have shown that chromium (VI) compounds can increase in risk of lung cancer. Animal studies have also shown an increase in risk of cancer (Moore and Ramamoorthy, 1984).

In this study, higher concentrations of metals were observed in *Sardinella* collected between June and August, which coincide with the summer's months. During this period, peak temperature of the seawater is observed, in the bay of Oran. According to the Algerian National Office of Meteorology, the temperature attained between 26-27 °C (Benamar, 2011).

The relationship between metal accumulation and sex, found in this study, may be due to the difference in the metabolic activities between the males and the females. According to Mortet (1988), an ovarian sequence for *Sardinella aurita* begins in the Bay of Oran during summer. It is characterized by an intense vitellogenic activity. Vitellogenesis is accompanied by an accumulation of reserves for the growth of eggs, but at the same time, by the metallic pollutants found in the biota (Mortet, 1988).

When comparing the rate of metal traces found in the muscle of *Sardinella aurita* with those found in *Sardinella* from the Strait of Messina (Sicily, Italy) and *Sardinella* from Alexandria waters, we notice that our samples are the least contaminated by chromium. Contrary to the levels of copper which appears to be high in *Sardinella aurita* from Oran coastline (Table 1).

Table 1. Comparative concentrations of copper and chromium in muscles of *Sardinella aurita* from different areas.

Areas studies	Chromium	Copper	Authors
<i>Sardinella aurita</i> from the Strait of Messina (Sicily, Italy)	0,36 ± 0,07	0,373 ±0,012	Lo Turco et al., (2013)
<i>Sardinella aurita</i> from Alexandria waters	0,1± 0,04	0,98 ± 0,3	Wafica and Aboul-Naga (1996)
<i>Sardinella aurita</i> from Oran coastline	0,08± 0,06	2,83 ± 0,9	Present study

5. Conclusion

Compared to the muscle tissue, the liver tissue of *Sardinella aurita* from bay of Oran, contained elevated concentrations of the metals Cu and Cr. This capacity of metal sequestering highlights differences in elemental bioavailability between sites and thereby makes the liver more suitable for biomonitoring purposes than the muscle tissue.

This study shows that the concentration of heavy metals in different fish organs is still below the allowable limits. This means that the measured fish is healthy.

However, accumulation of heavy metal in tissues of fish species, that are consumed as protein sources, passed cumulatively along the food chain and can constitute, after a long time, a public health problem.

Acknowledgements

Our gratitude is expressed especially to Professor Boutiba Zitouni, Director of Laboratory network of environmental surveillance, University of Oran, for her invaluable help. Great thanks go to Professor Claude Gimenes, National Institute of Telecommunications, Paris and Madam Anne Chatellier, College of France, Paris for their precious help and advice.

References

- Amiar J.-C., Pineau A, Boiteau HL, Metayer C and Amiard-Triquet C. 1987. Application de la spectrométrie d'absorption atomique Zeeman aux dosages de huit éléments traces (Ag, Cd, Cr, Cu, Mn, Ni, Pb et Se) dans des matrices biologiques solides. *Water Res.*, **21**: 693-697.
- Arunkumar RI, Rajasekaran P and Michael RD. 2000. Differential effect of chromium compounds on the immune response of the African mouth breeder *Oreochromis mossambicus* (Peters). *Fish Selfish Immunol.*, **10**: 667-676.
- Benamar N. 2011. Study of the biology, exploitation and contamination by toxic heavy metals of round sardinella *Sardinella aurita* caught in the Bay of Oran. Doctorat thesis University of Oran, Faculty of Science, pp.153.
- Bennani N, Schmid-Alliana A and Lafaurie M. 1996. Immunotoxic effects of copper and cadmium in the sea bass *Dicentrarchus labrax*. *Immunopharmacol. Immunotoxicol.*, **18**: 129-144.
- Carvalho ML, Pimentel AC and Fernandes B. 2005. Study of heavy metals in wild edible mushrooms under different pollution conditions by X-ray fluorescence spectrometry. *Analytical Sci.*, **21**: 747-750.
- Duffus JH. 1980. **Environmental Toxicology**, Edward Arnold Publishers Ltd., London, pp.164.
- FAO. 1977 Central Fisheries Research Institute. Zambia. Interim report. Report prepared for the Government of Zambia by the FAO of the UN acting as executing agency for the UNDP. Rome, FAO/PNUD, FI:DP/ZAM/68/511:53 p.
- FAO. 1992. Wastewater Treatment and use in Agriculture. M.B. Pescod. FAO Irrigation and Drainage Paper 47, FAO, Rome. pp. 125.
- Heath AG. 1991. **Water Pollution and Fish Physiology**. Lewis Publishers. Boca Raton, Florida, USA. pp 359.
- Ibok UJ, Udosen ED and Udoidiong OM. 1989. Heavy metals in fishes from some streams in Ikot Ekpene area of Nigeria. *Nig. J. Tech. Res.*, **1**: 61-68.
- Kim S, Jee H and Kang C. 2004. Cadmium accumulation and elimination in tissues of juvenile olive flounder, *Paralichthys olivaceus* after sub-chronic cadmium exposure. *Environ Pollution*, **127**: 117-123.
- Lo Turco V, Di Bella G, Furci P, Cicero N, Pollicino G and Dugo G. 2013. Heavy metals content by ICP-OES in Sarda sarda, *Sardinella aurita* and *Lepidopus caudatus* from the Strait of Messina (Sicily, Italy). *Natural Product Res.*, **27** (6): 518-523.
- Onianwa P C, Adeyemo A O, Idowu O Eand Ogabiela E E. 2001. Copper and zinc contents of Nigerian foods and estimates of the adult dietary intakes. *Food Chem.*, **72**: 89-95.

- Oronsaye JAO and Obano EE. 2000. The uptake and loss of dissolved copper by *Clarias anguillaris* fingerlings. *Pak J Sci Ind Res.*, **43(6)**: 359-362.
- Oronsaye JAO. 1989. Historical changes in the kidneys and the gills of stickleback (*Gasterosteus aculeatus*) exposed to cadmium. *Ecotoxicol and Environ Safety*, **17**:279-290.
- Sabate's A, Paloma M, Josep L and Vanesa R.2006. Sea warming and fish distribution: the case of the small pelagic fish, *Sardinella aurita*, in the western Mediterranean. *Global Change Biol.*, **12**: 2209–2219.
- Simpkins J and Willaims J I. 1989. **Advanced Biology** (3rd Edition). English Language Book Society / Unwin Hyman. pp.760.
- Sogreah Engineering .1998. Etude de l'assainissement du groupement urbain d'Oran, Mission A : Actualisation du plan directeur d'assainissement PDAA, AGEF, Algérie
- Svecevicus G. 2009. Use of behavioral responses of rainbow trout *Oncorhynchus mykiss* in identifying sublethal exposure to hexavalent chromium. *Bull. Environ. Contam. Toxicol.*, **82**: 564-568.
- Snyder CA and Valle CD. 1991. Lymphocyte proliferation assays as potential biomarkers for toxicant exposures. *J Toxicol Environ Health*, **34(1)**:127-139.
- UNEP .1984a. Sampling of Selected Marine Organisms and Sample Preparation for Trace Metal Analysis. Reference Methods for Marine Pollution Studies No. 7 Rev. 2.
- UNEP. 1984b. Determination of Total Cadmium, Zinc, Lead, Copper in Selected Marine Organisms by Flameless Atomic Absorption Spectrophotometry. Reference Methods for Marine Pollution Studies No. 11, Rev. 1.
- Vassileva E, Dočekalová H, Baeten H, Vanhentenrijk S and Hoenig M. 2001. Revisitation of mineralization modes for arsenic and selenium determinations in environmental samples. *Talanta*, **54(1)**, : 187-196.
- Venkatramreddy V, Vutukuru SS, and Paul B T. 2009. Ecotoxicology of hexavalent chromium in freshwater fish: A Critical review. *Rev Environ Health*. **24(2)**: 129–145.
- Wafica M and Aboul Naga 1996. Comparative study of trace metals accumulated in the muscle tissues of the most common and marketable sea food in Alexandria waters *Inter J Environ Health Res.*, **6(4)** :289-300.
- Yilmaz AB. 2003. Levels of heavy metals (Fe, Cu, Ni, Cr, Pb, and Zn) in tissue of *Mugil cephalus* and *Trachurus mediterraneus* from Iskenderun Bay, Turkey. *Environ Res.*, **92**., 277–281.

Evaluation of Antioxidant Properties of *Morus nigra* L. Fruit Extracts [II]

Najlaa K. Issa^{1,*} and Rihan S. Abd-Aljabar²

¹Chemistry Department, Faculty of Science, Dohuk University;

²Chemistry Department, Faculty of Science, Zakho University, Kurdistan Region of Iraq, Iraq

Received: March 3, 2013

Revised: May 6, 2013

Accepted: May 24, 2013

Abstract

Antioxidant activity for *Morus nigra* L. fruit extracts ethanol (Eth. ext.), flavonoid (F. ext.) and anthocyanin (A. ext.) and the isolated pigment (Fig. a) was evaluated using different methods such as (β -carotene-linoleic acid assay, reducing power assay, scavenging of hydrogen peroxide and total antioxidant capacity). The results proved that Fig. (a) was the most potent antioxidant followed by A. ext. and Eth. ext. Black mulberry fruits contained the highest amount of total phenolic content followed by Eth. ext., whereas flavonoid ext. contained the highest amount of total flavonoid content followed by black mulberry fruits. The effect of pH, temperature and storage had been studied to evaluate the remaining antioxidant capacity of the prepared extracts and Fig. (a). The results showed that the antioxidant capacity was decreased with increasing the time of boiling, storage and extreme pH values.

Key words: *Morus nigra* Extracts, Antioxidant Activity, β -Carotene-linoleic acid, Reducing Power, Scavenging of H₂O₂, Total Antioxidant Capacity, Stability.

1. Introduction

Oxygen is an element obligatory for life where living systems have evolved to survive in the presence of molecular oxygen, which has double-edged properties, being essential for life; it can also aggravate the damage within the cell by oxidative events (Shinde *et al.*, 2006). Oxidative stress results when the balance between the production of ROS exceeds the antioxidant capability of the target cell (Ahmad *et al.*, 2009). The antioxidant defense system in most living cells is composed of two components: antioxidant enzymes (endogenous antioxidants), and small molecule antioxidants (exogenous antioxidant) (Mugwerua and Rusling, 2006). Diet plays a vital role in the production of the antioxidant defense system by providing essential nutrient antioxidants such as vitamin C, vitamin E, and β -carotene. Other antioxidant plant phenols are flavonoids and essential minerals form important antioxidant enzymes. For example, superoxide dismutase (SOD) contains zinc and glutathione peroxidase that contains selenium (Willcox *et al.*, 2004; Kumar *et al.*, 2008). Plants are considered as one of the most important and interesting subjects that should be

explored for the discovery and development of newer and safer drug candidates (Hamid *et al.*, 2011).

Under ideal circumstances the body would be in a steady state with free radicals produced and quenched by the endogenous antioxidants. When an endogenous and exogenous antioxidant system can't balance off the free radicals generated in human cells, oxidative stress occurs. Oxidative stress can cause oxidative damage to lipids, proteins and DNA (Jing, 2006). The antioxidants could attenuate this oxidative damage of a tissue indirectly by enhancing natural defenses of cell and/or directly by scavenging the free radical species (Goldberg, 2003).

Antioxidant activity of flavonoid polyphenols and other small molecules is dependent on bioavailability and redox potential which is closely related to its chemical structure (Pokorny *et al.*, 2001).

The antioxidant activity of anthocyanins is attributed to their ability to scavenge free radicals, and the binding with heavy metals such as iron, zinc, and copper. Anthocyanins are inducers of antioxidant enzymes such as glutathione-S-transferase (GST) and SOD (Hosseinian *et al.*, 2008). Besides their color features, anthocyanins have recently attracted even more interest due to their possible health attributes, such as a reducing risk of coronary diseases, reducing risk of stroke,

* Corresponding author. e-mail :saminmamr@yahoo.com.

anticarcinogen activity, anti-inflammatory effects and improved cognitive behavior (Zhang and Demain, 2005). Laleh *et al.* (2006) showed that increasing in pH, temperature or exposure to light was able to spoil the anthocyanin molecule. Hurtado *et al.* (2009) proved that the isolated anthocyanins had higher capacity to capture free radicals than ascorbic acid and the hydroxylation degree of the isolated rutinoides had great influence on the antioxidant capacity.

The mulberry belongs to the genus *Morus* of the family *Moraceae*. Mulberry trees have historically been used for leaf yield in sericulture. In addition, their fruit have been used in folk medicine (especially in Chinese medicine) to treat diabetes, hypertension, anemia and arthritis. Also, black mulberry fruits are used for treating mouth lesions in Turkey. Recently, red and black mulberries have gained an important position in the food industry due to the presence of anthocyanins (Ozgen *et al.*, 2009).

The literature survey revealed that no work has been done to assess the antioxidant activity of black mulberry fruits *Morus nigra* L. prevailed in Kurdistan region. Thereby, this study was conducted to determine the antioxidant activity. Moreover, it was deemed of interest to investigate the antioxidant activity of the prepared fruit extracts by comparing various methods including β -carotene-linoleic acid assay, reducing power assay and total antioxidant capacity, besides the determination of total phenolics and flavonoids contents. It was also important to evaluate the effect of heat treatment, pH and storage on the stability of antioxidant activity.

2. Materials and Methods

2.1. Collection of Plant Materials

Black mulberry fruits *Morus nigra* L. were collected in June, 2008 from Duhok-dam, Duhok city- Kurdistan Region of Iraq. The ripe fruits were kept in plastic containers at -10 °C until the time of use (Pantelidis *et al.*, 2007; Spada *et al.*, 2008). The plant was botanically authenticated in Agriculture College –Forestry Department, University of Duhok.

2.2. Chemicals

All chemicals used were of analytical grade. Tannic acid, ammonium molybdate tetrahydrate, potassium ferricyanide and Tween 20 were obtained from Fluka; quercetin dihydrate were obtained from ROTH; β -carotene from USP; Folin- Ciocalteu reagent from Ajax; linoleic acid from Merck; α -tocopherol from USD and others were obtained either from Fluka or Sigma chemicals.

2.3. Preparation of Plant Extracts

The ethanol extract (Eth. ext.) of black mulberry fruits was prepared according to the method of Laleh *et al.* (2006). The flavonoid extract (F. ext.) of black mulberry fruits was prepared according to the methods of (Peach *et al.*, 1955; Harborne, 1984; Andersen and Markham, 2006). Anthocyanin extract (A. ext.) was prepared according to the method of (Harborne, 1984; Schoofs, 2004; Andersen and Markham, 2006).

Chromatographic techniques were applied for purification and isolation of pigment (a) (Pig.(a)) from anthocyanin extract.

2.4. Antioxidant Activity of Black Mulberry Extracts and Pig. (a)

2.4.1. Total Phenolic Content (TPC)

Total phenolics content of black mulberry fruits and extracts were measured according to the methods that were involving Folin- Ciocalteu reagent and tannic acid as a standard (Slinkard and Singleton, 1997; Kaur and Kapoor, 2002). All samples were assayed in three replications. The same procedure was applied to all standard tannic acid solutions (0–2500 mg/50 ml), and standard curve was obtained. Results were expressed as milligrams of tannic acid equivalent per gram of extract (mg TAE/g).

2.4.2. Total Flavonoid Content (TFC)

Total flavonoid content was determined according to the method described by (Chang *et al.*, 2002; Hsu, 2006). All samples were assayed in three replications. The same procedure was applied to all standard quercetin solutions (12.5–100 μ g/ml) in methanol and standard curve was obtained. Results were expressed as milligrams of quercetin equivalent per gram of extract (mg QE/g).

2.4.3. Scavenging of Hydrogen Peroxide (H_2O_2)

The ability of the black mulberry fruits extracts and Pig. (a) to scavenge hydrogen peroxide was determined according to the method of (Ruch *et al.*, 1989; Oktaly *et al.*, 2003). All samples were assayed in three replications. Results were expressed as milligrams of ascorbic acid equivalent per gram of extracts (mg AAE/g).

2.4.4. Total Antioxidant Capacity

The phosphomolybdenum assay was performed according to the methods of (Prieto *et al.*, 1999; Delouee *et al.*, 2007). All samples were assayed in three replications. The antioxidant capacity of extracts was expressed as micrograms of α -tocopherol equivalent per milliliter of extracts using extinction coefficient of ($4 \times 10^3 \text{ M}^{-1} \text{ cm}^{-1}$) according to Beer's law.

2.4.5. β -Carotene-Linoleic Acid Assay

In β -carotene -linoleic acid assay, antioxidant activity of extracts and of some common antioxidants such as α -tocopherol was determined according to the method of (Marco, 1968; Barriere *et al.*, 2001) with slight modifications. Antioxidant activity (AA) was calculated as percentage of inhibition relative to the control (α -tocopherol sample).

2.4.6. Reducing Power Assay

The ability of extracts to reduce iron (III) was assessed by the method of (Oyaizu, 1986; Yildirim *et al.*, 2001; Tsasi *et al.*, 2006; Su *et al.*, 2009). The increase in absorbance of reaction mixture was interpreted as increase in reducing activity of the extract and the results were compared with ascorbic acid as (positive control). All samples were assayed in three replications.

2.5. Effect of Temperature, pH and Storage on the Antioxidant Capacity

The black mulberry fruits extracts and Fig. (a) were heated at 50 and 100 °C for (60 and 120 min) and the residual antioxidant capacity was determined using total antioxidant capacity method as previously described. For pH stability, the extracts were pre-incubated at different pH values (3, 5, 7, 9 and 11) and the residual antioxidant capacity was evaluated. The extracts were also stored in the dark at 5°C, and the antioxidant capacity was determined at intervals of 30 days over a period of 90 days. All samples were assayed in three replications. The effect of temperature, pH and storage on the antioxidant capacity of extracts was expressed as micrograms of α -tocopherol equivalent per milliliter of extracts using extinction coefficient of ($4 \times 10^3 \text{ M}^{-1} \text{ cm}^{-1}$) (Arabshahi *et al.*, 2007).

2.6. Statistical Analysis

The statistical analysis for all experiments was done by using (SAS, 2000). Means comparison was done by Duncan's Multiple Ranges Test under 1%, and figures were done by Graph Pad Prism 5 program.

3. Results and Discussion

3.1. Antioxidant Activity of Black Mulberry Extracts and Fig. (a)

3.1.1. Total Phenolic Content

Total phenolic content of black mulberry fruits, Eth., F. and A. ext. was determined using the calibration curve between concentration and absorbance at 760 nm of standard tannic acid as shown in Table 1 and Figure 1. Phenolic compounds are one class of antioxidant agents which considered as good proton donors resulting in free radical terminators and contributed to the antioxidant activities of plant (Garzon and Wroldstad, 2009). Mulberries are rich source of phenolics such as rutin, kaempferol, quercetin, isoquercetin and chlorogenic acid (Apak *et al.*, 2007; Turkoglu, *et al.*, 2007). A great variation in terms of total phenolics content was observed among black mulberry fruits and their extracts.

Table 1. Total phenolic content of black mulberry fruits, Eth., F. and A. ext.

Sample	Concentration (mg TAE/g)
Black mulberry fruits	20.19
Eth. ext.	14.26
F. ext.	5.36
A. ext.	3.38

The highest amount of total phenols was observed for black mulberry fruits (20.19 mg TAE/g) which was in agreement with the results that were obtained by Ercisli and Orhan (2007), where they reported that black mulberry fruits were rich in phenols. Also, Eth. extract was high in its phenols content (14.26 mg TAE/g).

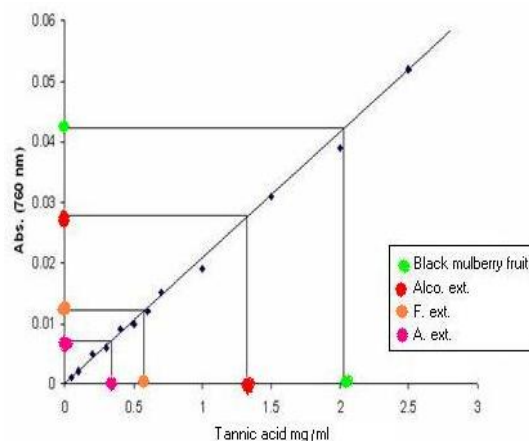


Figure 1. Total phenolic content of black mulberry fruits, Eth., F. and A. ext.

The high amount of phenols content in both black mulberry fruits and Eth. ext. might be due to their flavonoidal and non-flavonoidal phenolic compounds contents. Total phenols content of F.ext. had (5.36 mg TAE/g) due to the presence of flavonoid phenolic compounds only and A. ext. showed the least amount of total phenols content (3.38 mg TAE/g) due to the presence of anthocyanins only.

3.1.2. Total Flavonoid Content

Total flavonoid content of black mulberry fruits, Eth., F., A. ext. and Fig. (a) were determined using the calibration curve between concentration and absorbance at 415 nm of standard quercetin, as shown in Table 2 and Figure 2. Flavonoids are groups of polyphenolic compounds, which exhibit several biological effects such as antiinflammatory, antihepatotoxic, antiulcer, antiallergic, antiviral, and anticancer activities. They also act as inhibitors enzymes such as reducatase and xanthine oxidase. They are capable of effectively scavenging the reactive oxygen species because of their phenolic hydroxyl groups and are potent antioxidant (Chang *et al.*, 2002; Hsu, 2006).

Table 2. Total flavonoid content of black mulberry fruits, Eth., F., A. ext. and Fig. (a)

Sample	Concentration (mg QE/g)
Black mulberry fruits	1.11
Eth. ext.	1.17
F. ext.	1.26
A. ext.	0.94
Fig. (a)	0.26

The presence of high phenol and flavonoid contents in black mulberry had contributed directly to the antioxidant activity by neutralizing the free radicals (Umamaheswari and Chatterjee, 2008). Aluminum chloride colorimetric method was used for flavonoids determination. Figure 2 was revealed that F. ext. had the highest total flavonoid content (1.26 mg QE/g), followed by black mulberry fruits and Eth. ext. which contained approximately equal amounts of total flavonoids content (1.11 mg QE/g) and (1.17 mg QE/g), respectively.

This could be due to that ethanolic solvent that can extract most plant components especially phenolic compounds (Harborne, 1984). These results are in agreement with those obtained by Ercisli and Orhan (2007) who reported that total flavonoids content in black mulberry fruits was (2.76 mg QE/g) of extract. Total flavonoids content of A. ext. (0.94 mg QE/g) was lower than the forgoing ones indicating the presence of anthocyanins only. The least amount of total flavonoids content was found in Pig. (a) (0.26 mg QE/g) of extract containing one pigment only.

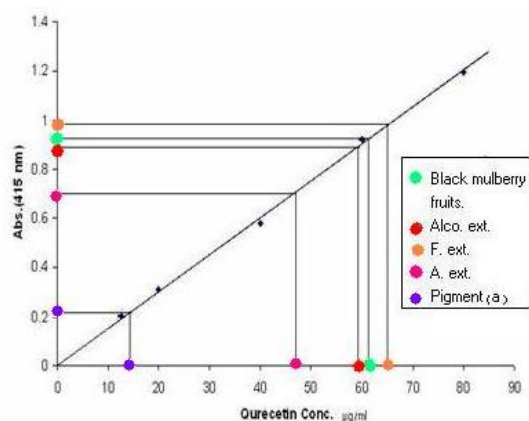


Figure 2. Total flavonoid content of black mulberry fruits, Eth., F., A. ext. and Pig. (a).

3.1.3. Scavenging of Hydrogen Peroxide

The ability of the Eth., F., A. ext. and Pig. (a) to scavenge H_2O_2 was determined using the calibration curve between concentration and absorbance at 230 nm of standard ascorbic acid, as shown in Table 3 and Figure 3. Since antioxidant compounds present in black mulberry extracts which are good electron donors, they may accelerate the conversion of H_2O_2 to H_2O (Shon *et al.*, 2007). Figure 3 illustrates that the strongest anti- H_2O_2 activity was observed for F. ext. (227.22 mg AAE/g) followed by A. ext. (208.37 mg AAE/g) which may be attributed to the presence of phenolic groups that could donate electrons to hydrogen peroxide and neutralizing it into water (Umamaheswari and Chatterjee, 2008). Eth. ext. and Pig. (a) exhibited weak scavenger H_2O_2 activity (199.10 mg AAE/g) and (191.90 mg AAE/g), respectively. The results showed that there was a strong correlation between total phenol content and scavenging of hydrogen peroxide.

Table 3. The ability of Eth., F., A. ext. and Pig. (a) to scavenge H_2O_2 .

Sample	Concentration (mg AAE/g)
Eth. ext.	199.10
F. ext.	227.22
A. ext.	208.37
Pig. (a)	191.90

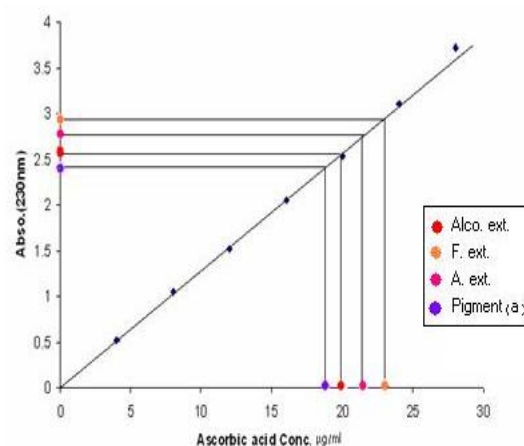
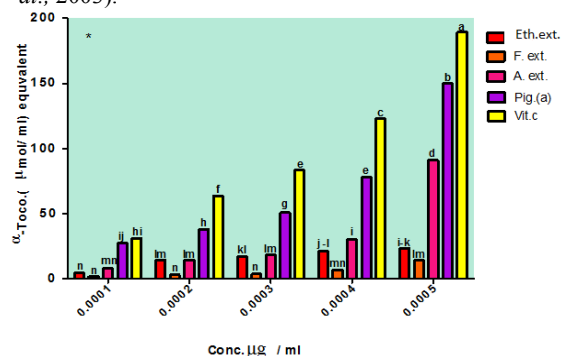


Figure 3. The ability of Eth., F., A. ext. and Pig. (a) to scavenge H_2O_2 .

3.1.4. Total antioxidant capacity

This assay is based on the reduction of Mo (VI) to Mo (V) by the samples of Eth., F., A. ext. and Pig. (a). The antioxidant capacity of the extracts was expressed as micromoles of α -tocopherol equivalent per milliliter of extracts using the calibration curve between micrograms of α -tocopherol equivalent and the concentration determined from Beer's law using extinction coefficient of ($4 \times 10^3 M^{-1} cm^{-1}$) at 760 nm, as shown in Figure 4. The extracts were demonstrated electron-donating capacity and, thus they may act as radical chain terminators, transforming reactive free radical species into more stable non-reactive products (Delouee *et al.*, 2007). Figure 4 illustrates that Pig. (a) was significantly exhibited the highest degree of activity at different concentrations especially at 0.0005 $\mu g/ml$ followed by A. ext. compared to ascorbic acid. This activity might be attributed to the presence of phytochemicals (antioxidant secondary metabolites) (Banso, 2009). Eth. ext. exhibited significant degree of antioxidant capacity followed by F. ext. due to the presence of phenolic compounds which had redox properties allowed them to act as reducing agents, hydrogen donors, and singlet oxygen quenchers. They may also had a metal chelating potential (Javanmardi *et al.*, 2003).



* Column followed by the same letter are not significantly different at 1% level based on Duncan's Multiple Rang Test.

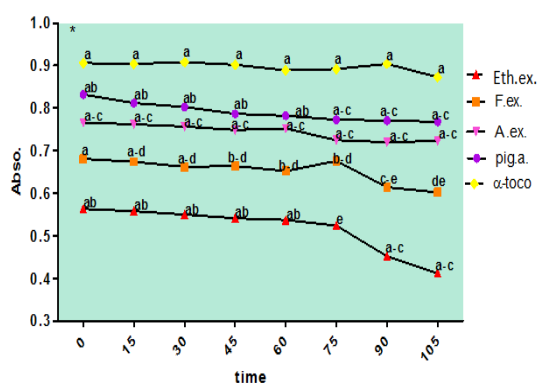
Figure 4. Total antioxidant capacity of Eth., F., A. ext. and Pig. (a).

3.1.5. β -Carotene-Linoleic Acid Assay

In β -carotene-linoleic acid assay, antioxidant activity (AA) of Eth., F., A. ext. and Pig. (a) was compared with antioxidant activity of some common antioxidants such as α -tocopherol when subjected to thermal auto oxidation at 50 °C. The AA was shown in Table 5 and Figure 5.

Table 5. Antioxidant activity of Eth., F., A. ext. and Pig.(a) as antioxidant compared to α -tocopherol.

Sample	A _i	A _t	A _i '	A _t '	AA%
Eth. ext.	0.496	0.412	0.379	0.207	51.20
F. ext.	0.681	0.602	0.379	0.207	54.10
A. ext.	0.766	0.723	0.379	0.207	75.00
Pig. (a)	0.832	0.768	0.379	0.207	62.8
α -tocopherol	0.906	0.873	0.379	0.207	80.9



*The same letters are not significantly different at 1% level based on Duncan's Multiple Rang Test.

Figure 5. Antioxidant activity of Eth., F., A. ext. and Pig. (a).

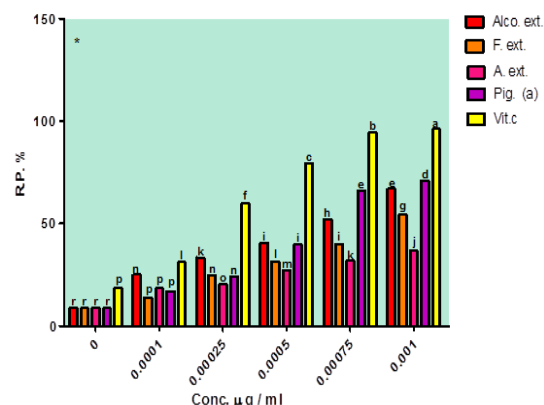
The results indicated that the major contributors among individual extracts to AA could be Pig. (a) (62.8%) followed by A. ext. (75 %). Regarding the structure of these compounds, they possess a high degree of hydroxylation and methoxylation on their aromatic rings which suggests their higher contribution to AA of black mulberry antioxidant (Lachman *et al.*, 2009).

Flavonoid ext. showed significant AA (54.10%) especially at zero time which, then decreased when temperature elevated. The obtained results were in agreement with the reports of Zhshen *et al.* (1999) who confirmed the presence of total flavonoids especially rutin, quercetin, isoquercetin and luteolin in mulberry plants. These compounds are the major phytochemicals responsible for antioxidant activity of mulberry plants. Eth. ext. showed a moderate activity (51.20%) due to the presence of polyphenolic compounds in black mulberry fruits especially chlorogenic acid (Lin and Tang, 2007). Positive correlation between total phenol and antioxidant activity was found by many researchers

(Brown *et al.*, 2005; Yang *et al.*, 2009; Garzon and Wrolstand, 2009).

3.1.6. Reducing Power Assay

In this assay, the ability of Eth., F., A. ext. and Pig. (a) to reduce iron (III) to iron (II) was determined and compared to that of ascorbic acid, which is known to be a strong reducing agent. The percentage reduction (R.P. %) of the samples was shown in Figure 6.



* Column followed by the same letters are not significantly different at 1% level based on Duncan's Multiple Rang Test.

Figure 6. Reducing power of Eth., F., A. ext. and Pig. (a) as a strong reducing agent compared to ascorbic acid.

Kaur and Arora (2008) determined reducing power of Eth., F., A. ext. and Pig. (a) and, compared to ascorbic acid, they considered it as a strong reducing agent. Data were shown that Eth., F., A. ext. and Pig. (a) possessed some degree of hydrogen donation capacity in a concentration dependent manner. The reducing power was increased with increasing concentration, but the capacities were inferior to that of ascorbic acid.

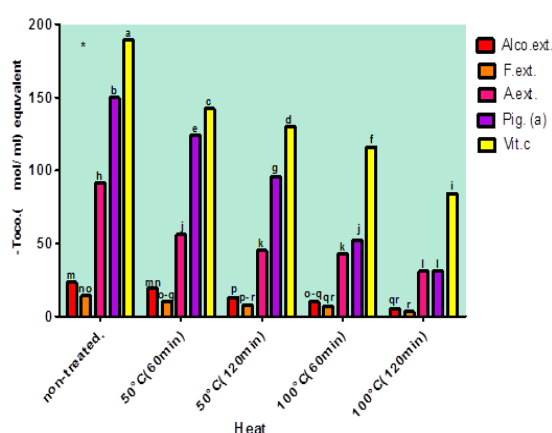
Pigment (a) was the most potent reducing agent followed by Eth. ext., F. and A. ext. due to the highest amount of total phenolics and phenolic acids. Phenols that have more number of hydrolysable groups (OH groups) attached to the ring acting as more powerful reducing agent considered as good proton donors resulting in the termination of free radicals chain reactions (Kaur and Arora, 2008). Hence, this may accelerate the conversion of H₂O₂ to H₂O (Ruch *et al.*, 1989). At high concentration, Pig. (a) showed a high potential for being a proton donor, wherever, A. ext. was the weakest in its activity due to the low content of total phenols. Similar relation between iron (III) reducing activity and total phenol content had been reported (Benzi and Szeto, 1999)

3.2. Effect of Temperature, pH and Storage on the Antioxidant Capacity

It is well known that many factors such as antioxidant concentration, temperature and pH of the media, processing treatment and storage are strongly influence the antioxidant capacity (Arabshahi *et al.*, 2007). In this study all extracts and Pig.(a) was subjected to thermal, pH and storage studies. Figure 7 shows the effect of temperature on the antioxidant stability of all extracts and Pig. (a).

The antioxidant capacity was decreased with increasing the time of boiling. This reduction was reached to 20.66% for Pig. (a) and to 34.06% for A. ext. when incubated at 100 °C for 120 min.. This observation indicated that anthocyanins were not stable at high temperature which reasoned to the speed destruction of anthocyanins at high temperatures (Arabshahi *et al.*, 2007).

Antioxidant capacity of Eth. ext. was reduced to 81.70% followed by F. ext. 72.18% when incubated at 50 °C for 60 min. The antioxidant capacity was decreased with increasing the time of boiling. The reduction activity was reduced for Eth. ext. and F. ext. when incubated at 100 °C for 120 min which might be explained to the loss of naturally occurring antioxidants or phytochemicals present in the extract (Arabshahi *et al.*, 2007).

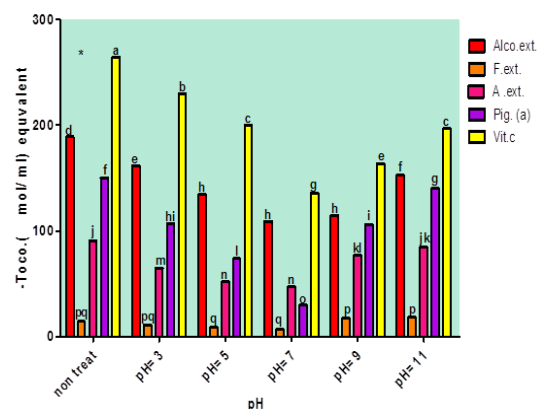


* Column followed by the same letters are not significantly different at 1% level based on Duncan's Multiple Rang Test.

Figure 7. Effect of temperature on the antioxidant capacity of Eth., F., A. ext. and Pig. (a).

The influence of pH on the antioxidant stability of all extracts and Pig. (a) is shown in Figure 8. The antioxidant capacity of all extracts and Pig. (a) gradually decreased with minimum value at pH 7 followed by continuous increase at alkaline pH, indicating strong dependence of antioxidant capacity of extract to the pH of the system. At pH 3 no significant effect was noticed in Pig. (a) as well as A. ext. comparing to vit. C. Anthocyanin can be found in different chemical forms which depend on the pH of the solution. At low pH 1, the flavylium cation (red color) was the predominant species and contributed to the purple and red colors. The antioxidant capacity of Pig. (a) and A. ext. was reduced at pH 5-7. At pH 7 the antioxidant capacity of Pig. (a) and A. ext. was reduced to 19.80% and 51.86%, respectively.

It had been shown that simple anthocyanins were unstable and quickly decolorized by hydration at the 2-position of the anthocyanidin skeleton. At alkaline pH 11, the antioxidant capacity of Pig. (a) and A. ext. was increased to 93.30% and 93.07%, respectively, due to the increased amount of anhydrous base at alkaline pH (Furtado *et al.*, 1993). The antioxidant capacity of Eth. ext. was least effected by pH (87.23%, at pH 3) followed by F. ext. (75.35%, at pH 3).

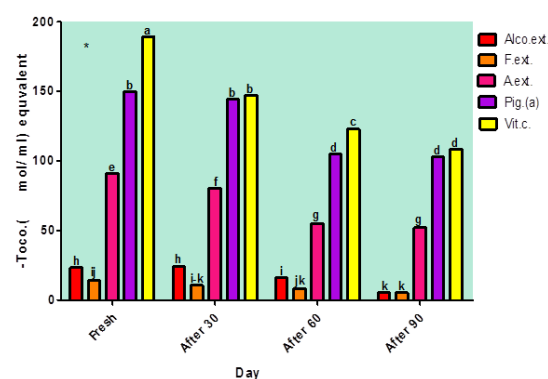


* Column followed by the same letters are not significantly different at 1% level based on Duncan's Multiple Rang Test.

Figure 8. Effect of pH on the antioxidant capacity of Eth., A. ext. and Pig. (a).

This observation could be attributed to the presence of antioxidant secondary metabolites other than anthocyanins like polyphenolic compounds in black mulberry fruits, especially chlorogenic acid (Lin and Tang, 2007).

The effect of storage on the antioxidant stability of all extracts and Pig. (a) was also studied up to 3 months at intervals of 30 days as shown in Figure 9. The antioxidant capacity of all extracts and Pig. (a) were gradually reduced with minimum value during 90 days period. The antioxidant capacity of Pig. (a) and A. ext. were unaffected by storage for 30 days and slightly reduced after 60 days only. Their antioxidant capacity after 90 days period were reduced to 68.66% and 57.14%, respectively, which was approximately similar to the antioxidant capacity of vit. C. with a reduction value of 57.25% after 90 days period storage.



* Column followed by the same letter are not significantly different at 1% level based on Duncan's Multiple Rang Test.

Figure 9. Effect of storage on the antioxidant capacity of Eth., F., A. ext. and Pig. (a).

Antioxidant capacity of Eth. and F. ext. was significantly affected by storage for 30 days and reduced after 60 days and their antioxidant capacity after 90 days period were reduced to 23.40% and 36.97%, respectively. These results were in agreement with the reports of Arabshahi *et al.*, (2007) who noticed that the

remaining activity of anthocyanin black mulberry extracts was about 65% after three months storage indicating that it could be still considered as a source of natural antioxidants.

4. Conclusions

Based on the obtained results, it may be concluded that *Morus nigra* fruits are a well recognized source of secondary metabolites like flavonoids and anthocyanins which considered as an excellent antioxidants. The antioxidant activities of *Morus nigra* fruits extracts varied with the test method. Pigment (a) showed a strong total antioxidant capacity, reducing power ability and inhibition of β -carotene bleaching. Flavonoid ext. exhibit a suitable H_2O_2 scavenging activity which may be correlated with the high amount of polyphenolic content especially flavonoids and anthocyanins. Increasing the time of boiling, storage, and extreme pH values decreased total antioxidant capacity of all extracts.

References

- Ahmad IM, Abdalla MY, Mustafa NH, Qnais EY and Abdulla FA. 2009. *Datura* aqueous leaf extract enhances cytotoxicity via metabolic oxidative stress on different human cancer cells. *Jordan J Biol Sci.*, **2**: 9-14.
- Andersen QM and Jordheim M. 2006. The anthocyanins. In: Andersen QM and Markham KR (Eds), **Flavonoids Chemistry, Biochemistry and Applications**. New York, pp. 471-553.
- Apak R, Guclu K, Demirata B, Ozyurek M, Celik SE, Bektasoglu B, Berker I and Ozyurt D. 2007. Comparative evaluation of various total antioxidant capacity assays applied to phenolic compounds with the CUPRAC assay. *J Molecules*, **12**: 1496-1547.
- Arabshahi DS, Devi DV and Urooj A. 2007. Evaluation of antioxidant activity of some plant extracts and their heat, pH and storage stability. *Food Chem.*, **100**: 1100-1105.
- Banso A. 2009. Phytochemical and antibacterial investigation of bark extracts of *Acacia nilotica*. *J Med Plants Res.*, **3**: 082-085.
- Barriere C, Centeno D, Lebert A, Leroy-Setrin S, Berdague, JL and Talon R. 2001. Roles of superoxide dismutase and catalase of *Staphylococcus xylosus* in the inhibition of linoleic acid oxidation. *Microbiol Lett.*, **201**: 181-185.
- Benzie IF and Fand Szeto YT. 1999. Total antioxidant capacity of tea by the ferric reducing antioxidant power assay. *J Agri Food Chem.*, **47**: 633-636.
- Brown C R. 2005. Antioxidant in potato. *J Amer Potato*, **82**: 163-172.
- Chang C, Yang M, Wen H and Chern J. 2002. Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *J Food Drug Anal.*, **10**: 178-182.
- Delouee SAW and Urooj A. 2007. Antioxidant properties of various solvent extracts of mulberry (*Morus indica* L.) leaves. *Food Chem.*, **102**: 1233-1240.
- Ercisli S and Orhan E. 2007. Chemical composition of white (*Morus alba*), red (*Morus rubra*) and black (*Morus nigra*) mulberry fruits. *Food Chem.*, **103**: 1380-1384.
- Furtado P, Figueiredo P, Neves, HC and Pina F. 1993. Photochemical and thermal degradation of anthocyanidins. *J Photochem Photobiol A: Chem.*, **75**: 113-118.
- Garzon GA and Wrolstad RE. 2009. Major anthocyanins and antioxidant activity of Nasturtium flowers (*Tropaeolum majus*). *Food Chem.*, **114**: 44-49.
- Goldberg G. 2003. **Plants: Diet and Health**. Blackwell Science, USA.
- Hamid K, Sultana S, Urmi KF, Obayed Ullah M, Zulfiker AM and Hossain A. 2011. *In vitro* Free radical scavenging and brine shrimp lethality bioassay of aqueous extract of *Ficus racemosa* seed. *Jordan J of Biol Sci.*, **4**: 51-54.
- Harborne JB. 1984. **Phytochemical Methods**. New York, Wiley, pp. 1-70.
- Hosseini FS, Li W and Beta T. 2008. Measurement of anthocyanins and other phytochemicals in purple wheat. *Food Chem.*, **109**: 916-924.
- Hsu C. 2006. Antioxidant activity of extract from *Polygonum aviculare* L. *Biol Res.*, **39**: 281-288.
- Hurtado NH, Morales AL, Gonzalez-Miret LG, Escudero-Gilete, ML and Heredia FJ. 2009. Color, pH stability and antioxidant activity of anthocyanin rutinosides isolated from tamarillo fruit (*Solanum betaceum* Cav.). *Food Chem.*, **117**: 88-93.
- Jayaprakasha GK, Singh RP and Sakariah KK. 2001. Antioxidant activity of grape seed (*Vitis vinifera*) extracts on peroxidation models *in vitro*. *Food Chem.*, **73**: 285-290.
- Jing P. 2006. Purple corn anthocyanins: chemical structure chemoprotective activity and structure/ function relationships. Ph.D. Thesis. Graduate School. Ohio State University, USA.
- Kaur C and Kapoor HC. 2002. Antioxidant activity and total phenolic content of some Asian vegetables. *Int J Food Sci Technol.*, **37**: 153-161.
- Kaur R and Arora S. 2008. Investigations of antioxidant activity of methanol extract of *Chukrasia tabularis* A. Juss. leaves. *J Clin Med.*, **3**: 200-205.
- Kumar V and Chauhan S. 2008. Mulberry: Life enhancer. *J Med Plants Res.*, **2**: 271-278.
- Lachman J, Hamouz K, Sulc M, Orsak M, Pivec V, Hejtmankova A, Dvorak P and Cepl J. 2009. Cultivar differences of total anthocyanins and anthocyanidins in red and purple-fleshed potatoes and their relation to antioxidant activity. *Food Chem.*, **114**: 836-843.
- Laleh GH, Frydoonfar H, Heidary R, Jameel R and Zare S. 2006. The effect of light, temperature, pH and species on stability of anthocyanin pigments in four *Berberis* species. *Pakistan J Nut.*, **5**: 90-92.
- Lin JY and Tang CY. 2007. Determination of total phenolics and flavonoid contents in selected fruits and vegetables, as well as their stimulatory effects on mouse splenocyte proliferation. *Food Chem.*, **101**: 140-147.
- Marco GJ. 1968. A rapid determination method for evaluation of antioxidants. *J Ame Oil Chem Soc.*, **45**: 594-598.
- Mugweru A and Rusling J. 2006. Studies of DNA damage inhibition by dietary antioxidants using metallopolypion / DNA sensors. *J Electro.*, **18**: 327 - 332.
- Oktay M, Gulcin I and Kufrevioglu U. 2003. Determination of *in vitro* antioxidant activity of fennel (*Foeniculum vulgare*) seed extracts. *Lebensm- Wiss. U.- Technol.*, **33**: 263-271.
- Oyaizu M. 1986. Antioxidative activity of browning products of glucosamine fractionated by organic solvent and thin-layer

chromatography. *Nippon Shokuhin Kogyo Gakkaishi*. **35**:771–775.

Ozgen M, Serc S and Kaya C. 2009. Phytochemical and antioxidant properties of anthocyanin-rich *Morus nigra* and *Morus rubra* fruits. *Scientia Horticulturae*, **119**: 275–279.

Pantelidis GE, Vasilakakis M, Manganaris GA and Diamantidis G. 2007. Antioxidant capacity, phenol, anthocyanin and ascorbic acid contents in raspberries, blackberries, red currants, gooseberries and cornelian cherries. *Food Chem.*, **102**:777–783.

Peach K and Tracey MV. 1955. **Modern Methods of Plant Analysis**. Springer-Verlag Bertin ,Germany.

Pokorny J, Yanishlieva N and Gordon M. 2001. **Antioxidants in Food: Practical Applications**, England, pp. 88.

Prieto P, Pineda M, and Aguilar M. 1999. Spectrophotometric quantization of antioxidant capacity through the formation of a phosphomolybdenum complex: specific application to the determination of vitamin E. *Anal Biochem.*, **269**: 337–341.

Ruch RJ, Cheng SJ and Klaunig JE. 1989. Prevention of cytotoxicity and inhibition of intracellular communication by antioxidant catechins isolated from Chinese green tea. *Carcinogen*. **10**:1003-1008.

Schofs B. 2004. Determination of pigments in vegetables. *J Chrom.*, **1045**: 217-226.

Shinde A, Abd Elmouttale AT, Ashraf T and Mostafa U . 2012. Effect of tomato and guava juices on oxidative stress in rats after strenuous exercise. *Jordan J of Bio Sci.*, **5**: 167 – 174.

Shon MY, Lee J and Choi JH. 2007. Antioxidant and free radical scavenging activity of methanol extract of chungkukjang. *J Food Composition and Analysis*, **20**: 113-118.

Slinkard K, and Singleton VL. 1997. Total phenol analysis: Automation and comparison with manual methods. *Amer J. Enol. Viticult.*, **28**: 49–55.

Spada PDS, Souza GGN, Bortolini GV, Henriques JAP and Salvador M. 2008. Antioxidant, mutagenic, and antimutagenic activity of frozen fruits. *J Med Food*, **1**: 144-151.

Su XS, Wang ZY and Liu JR. 2009. *In vitro* and *in vivo* antioxidant activity of *Pinus koraiensis* seed extract containing phenolic compounds. *Food Chem.*, **117**: 681–686.

Tsasi SY, Huang SJ and Mau JL. 2006. Antioxidant properties of hot water extracts from *Agrocybe cylindracea*. *Food Chem.*, **98**:670–677.

Turkoglu A, Duru ME and Mercan N. 2007. Antioxidant and antimicrobial activity of *Russula delica* Fr: An Edible wild mushroom. *Eurasian J of Anal Chem.*, **1**: 54-67.

Umamaheswari M and Chatterjee TK. 2008. *In vitro* antioxidant activities of the fractions of *Coccinia grandis* L. leaf extract. *Afr J Tradi.*, **5**:61-73.

Willcox J K, Ash SL. and Catignani GL. 2004. Antioxidants and prevention of chronic disease. *Critical Rev Food Sci Nut.*, **44**:275–295.

Yang J, Martinson TE and Liu RH. 2009. Phytochemical profiles and antioxidant activities of wine grapes. *Food Chem.*, **116**: 332–339.

Yildirim A, Mavi A and Kara AA. 2001. Determination of antioxidant and antimicrobial activities of *Rumex crispus* L. extracts. *J Agr Food Chem.*, **49**: 4083–4089.

Zhang L and Demain AL. 2007. **Natural Products: Drug Discovery and Therapeutic Medicine**, Springer,USA.

Zhishen J, Mengcheng T and Jianming W. 1999. The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chem.*, **64**: 555–559.

Antimicrobial Activity of Xerophytic Plant (*Cotula cinerea* Delile) Extracts Against Some Pathogenic Bacteria and Fungi

Djamel Bensizerara¹, Taha Menasria^{2,*}, Maimouna Melouka¹, Lamia Cheriet¹ and Haroun Chenchouni²

¹Department of Natural and Life Sciences, Faculty of Nature and Life Sciences and Sciences of Earth and Universe, University of Kasdi Merbah, Ouargla 30000, Algeria.

²Department of Natural and Life Sciences, Faculty of Exact Sciences and Natural and Life Sciences, University of Tebessa, Tebessa 12002, Algeria.

Received: February 13, 2013

Revised: May 9, 2013

Accepted: May 30, 2013

Abstract

In the present investigation, an Algerian commonly available plant namely *Cotula cinerea*, found throughout sandy desert grounds, was screening for antimicrobial activity against five different human pathogenic microbes namely, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Escherichia coli* and *Candida albicans*. The antimicrobial activity was evaluated using the agar disc diffusion method. Aerial parts of *C. cinerea* were subjected to extraction using four solvents of different polarity (70% ethanol, *n*-butanol, ethyl acetate and petroleum ether). Petroleum ether and *n*-butanol extracts had the most effective antimicrobial activity with Gram-negative *K. pneumoniae* demonstrating the highest susceptibility. Linear regression analysis was performed to find correlations between extract concentrations and inhibition activity. Results showed a significant increase in mean diameter of inhibition zone with increasing extract concentrations of all solvents except *n*-butanol. Two-way ANOVA test was used to compare the effect of *C. cinerea* extracts on the antimicrobial properties. All plant extracts have shown significant differences in their actions as antimicrobial agents. Indeed, the *n*-butanol extract at a low concentration of 0.25 mg mL⁻¹ indicated a potent antimicrobial activity of *C. cinerea* extracts.

Keywords: Antimicrobial activity, *Cotula cinerea*, Pathogenic microbes, Medicinal plant.

1. Introduction

Medicinal plants, which form the backbone of traditional medicine, have been in the last few decades the subject of very intense pharmacological studies. This has been brought by the acknowledgement of the value of medicinal plants as potential sources of new therapeutic compounds and drug development (Matu and van Staden, 2003). According to the World Health Organization, about 80% of the world's population living in developing countries rely mostly on plants for primary health care (McKay and Blumberg, 2007).

In recent years, pathogenic microorganisms have developed multiple drug resistance due to the indiscriminate use of commercial antimicrobial drugs commonly used in the treatment of infectious diseases (Katsumi *et al.*, 2005). An increase in the emergence of multidrug-resistant bacteria is worrying the world population. Infection rates have greatly increased and

antibiotics resistance has become an ever-increasing therapeutic problem (Shahid *et al.*, 2008). Therefore, there is a need to develop alternative antimicrobial drugs from various sources such as medicinal plants (Cordell, 2000).

Plants with antimicrobial activities have become more interesting because many people are aware of problems associated with the over-prescription and misuse of traditional antibiotics. However, only about 20% of the plants found in the world have been subjected to pharmacological or biological testing (Mothana and Lindequist, 2005). Plants in the environment are exposed to a range of abiotic stresses such as osmotic stress, salinity, and temperature variations. These, in-turn, affect their growth and the metabolic processes involved in the synthesis of a wide range of secondary metabolites, such as polyphenols, tannins, terpenoids, alkaloids etc, which may have remarkable antimicrobial properties (Cowan, 1999; Marzouk *et al.*, 2010; Daglia, 2012).

* Corresponding author e-mail: tahamenasria@hotmail.com

The Algerian flora plays a key role in supporting traditional medicine, which is widely practiced over the country. This flora holds a rich diversity of medicinal and endemic plants (Beloued, 2005). Many plants used in the Algerian traditional medicine have the potential to provide pharmacologically active natural products (Maiza *et al.*, 1993; Hammiche and Maiza, 2006). Ethnopharmacological interest in the sources of these compounds has increased nationally and worldwide, particularly in the search for drugs to counter multi-drug resistant microorganisms.

Cotula cinerea L., syn. *Brocchia cinerea* Del. (Asteraceae), is a xerophytic plant widely distributed in sandy and desert grounds (Markouk *et al.*, 1999a). This medicinal plant popularly known as (Gartoufa or Chouhiya), is commonly used in Algerian folk medicine, as well in the rest of the Maghreb region, as an anti-inflammatory, analgesic, antipyretic, antiseptic, and for treatment of various other diseases, including digestive problems (constipation and colic), rheumatism, and urinary and pulmonary infections. It is much appreciated in green tea or mixed with food to enhance the flavour (Maiza *et al.*, 1993; Markouk *et al.*, 1999a, 1999b; Larhsini *et al.*, 2002; Hammiche and Maiza, 2006). Several compounds have been isolated from *C. cinerea*, including flavonoids, sesquiterpene lactones, sesquiterpene coumarins and tannins (Ahmed *et al.*, 1987; Markouk *et al.*, 1999b). The objective of this work is to evaluate the antimicrobial activity of *C. cinerea* extracts, obtained using various solvents, against some pathogenic bacteria and fungi.

2. Materials and Methods

2.1. Plant Material

Cotula cinerea samples were collected from its natural range of distribution in El-Oued (Algerian Sahara Desert) (about 4 km Southeast of El-Oued city, 33°20'N to 33°19'N, 6°52'E to 6°53'E) in March 2011.

2.2. Extraction Protocol

2.2.1. Extracting Solvents

The extraction was carried out by using four solvents of increasing polarity: (i) petroleum ether (non-polar), (ii) ethanol, (iii) *n*-butanol, (iv) ethyl acetate (later three are moderately polar).

2.2.2. Preparation of Plant

The freshly picked aerial parts of the plant used in the screening were air-dried at room temperature for 2 weeks, with no direct sunlight. Once dry, plant was ground into fine powder and stored at 4 °C until time of extraction.

2.2.3. Preparation of the Extracts

The powdered plant material (20 g) was macerated for 24 h three times (3×24 h) in a mixture of ethanol/water (70:30; v/v) with frequent agitation at room temperature (25 ± 1 °C). Then the mixture was filtered using filter paper (Whatman No. 1) under the vacuum of a water pump and the ethanol was evaporated under low pressure using a rotary evaporator at 50 °C. The residue was taken as the hydro alcohol extract (Dall'Agnol *et al.*, 2003). The remaining aqueous

extract was fractioned with petroleum ether, ethyl acetate and *n*-butanol (3×100 mL for each solvent). These extracts were dried under reduced pressure using a rotatory evaporator at 40 °C. The residues were taken as the petroleum ether, ethyl acetate and *n*-butanol extracts of the plant (Bologn *et al.*, 2011).

2.3. Antimicrobial Activity

2.3.1. Microbial Strains and Growth Conditions

Five clinical isolates of microorganisms were used for assessing the plant antimicrobial properties, including the Gram-positive *Staphylococcus aureus*, the Gram-negative *Escherichia coli*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae*, and the yeast *Candida albicans*. Table 1 lists the strains and their resistance phenotypes. All strains were obtained from the Microbiology Laboratory of Hospital Benamor Djilani (El-Oued, Algeria) and were maintained at 4 °C on slants of Nutrient Agar (NA) for bacteria and Sabouraud Dextrose Agar (SDA) for the yeast. Active cultures were prepared by transferring a loop of cells from the agar slant to a test tube containing 5 mL of Nutrient Broth for bacteria and Sabouraud Dextrose broth for the yeast. They were then incubated overnight to reach the logarithmic phase of growth; for about 6–10 hours at 37 °C for bacteria and 12–16 h at 30°C for *C. albicans*.

Table 1. Resistance pattern of target organisms to antibiotics.

Strain	Phenotype of resistance	Phenotype of sensibility
<i>Staphylococcus aureus</i>	AMC, AMX, CE, CF, OX, PEN	CHL, CIP, GEN, PEF
<i>Escherichia coli</i>	AMC, AMX, CIP, PEF, SXT	CE, CF, CEF, C, GEN, IMI, STR
<i>Pseudomonas aeruginosa</i>	AMC, AMX, ATM, CF, CIP, PEF, SXT	GEN, IMI, PEF, PIP
<i>Klebsiella pneumoniae</i>	IMI	C, SXT
<i>Candida albicans</i>	ND	ND

(AMC: amoxicillin + acid clavulanic, AMX: amoxicillin, ATM: aztreonam, C: chloramphenicol, CE: cefatxin, CF: cefalotin, CEF: cefazolin, CIP: ciprofloxacin, GEN: gentamicin, IMI: imipenem, ND: not determined, OX: oxacillin, PEF: pefloxacin, PEN: penicillin, PIP: piperacillin, STR: streptomycin, SXT: trimethoprim/sulfamethoxazol)

2.3.2. Antimicrobial Assay (Disk Diffusion Assay)

The disc-diffusion assay (Qaralleh *et al.*, 2010) was used to determine growth inhibition caused by plant extracts. Inoculums, containing 10⁶–10⁸ CFU per milliliter, was spread on Mueller–Hinton (MH) agar plates for the four bacteria and 10⁴–10⁵ CFU per milliliter was poured over the base plates forming a homogenous top layer on SDA for the yeast. Using sterile forceps, Whatman's filter discs (Ø = 5 mm), impregnated with different dilutions of extracts (25, 50, 75 and 100%) from the initial concentration of 1 mg mL⁻¹, were placed on inoculated plates and left at 4 °C for 2 h before being incubated to allow the diffusion of the extract. Discs saturated with solvents (ethanol, petroleum ether, ethyl acetate, and *n*-butanol) were air-dried and used as negative controls. The plates were incubated at 37 °C for

24 h for bacteria and 48 h at 30 °C for the yeast, after which, inhibition zones around each disc (> 5 mm) were measured (disc diameter included).

2.4. Statistical Analysis

Linear regression analysis (LRA) was carried out to find statistically significant correlation between the different concentrations of each extract and their overall antimicrobial activity, assessed as diameter of inhibition with disregard to the tested strains. The data pertaining to antimicrobial activity of different *C. cinerea* extracts were also analysed with two-way ANOVAs, to test the effect of “tested strains” and “extract dilutions” on the levels of antimicrobial activity. Interaction between tested microbe-species and extract concentration was also included in the analysis for each plant extract. Besides the explanatory ability of LRA, it could be used, supported by ANOVA outputs, to detect potency “effectiveness” of the extract itself disregarding its concentration. Both LRA and ANOVA were considered statistically significant when P -value < 0.05.

3. Results

Hydro-alcohol extract of *C. cinerea* had a much higher extraction yield (w/w %) than the other extracts, whereas, the *n*-butanol extract had the lowest yield. Since extract yield increases with extracting solvent polarity. As a result, 70% ethanol, which was the most polar of all solvents and which was used for fractionation, has afforded the maximum yield (11.0%) compared to petroleum ether (1.0%), *n*-butanol (6.0%) and ethyl acetate (1.2%).

The inhibition zone, referring to antimicrobial activity of *C. cinerea* extracts, was measured after incubation of the plates. Each of the extracts was tested three times and the average (\pm SD) of three values was determined. Generally, the results showed that the inhibitory effect of extracts increased with increasing of concentrations (Figure 1).

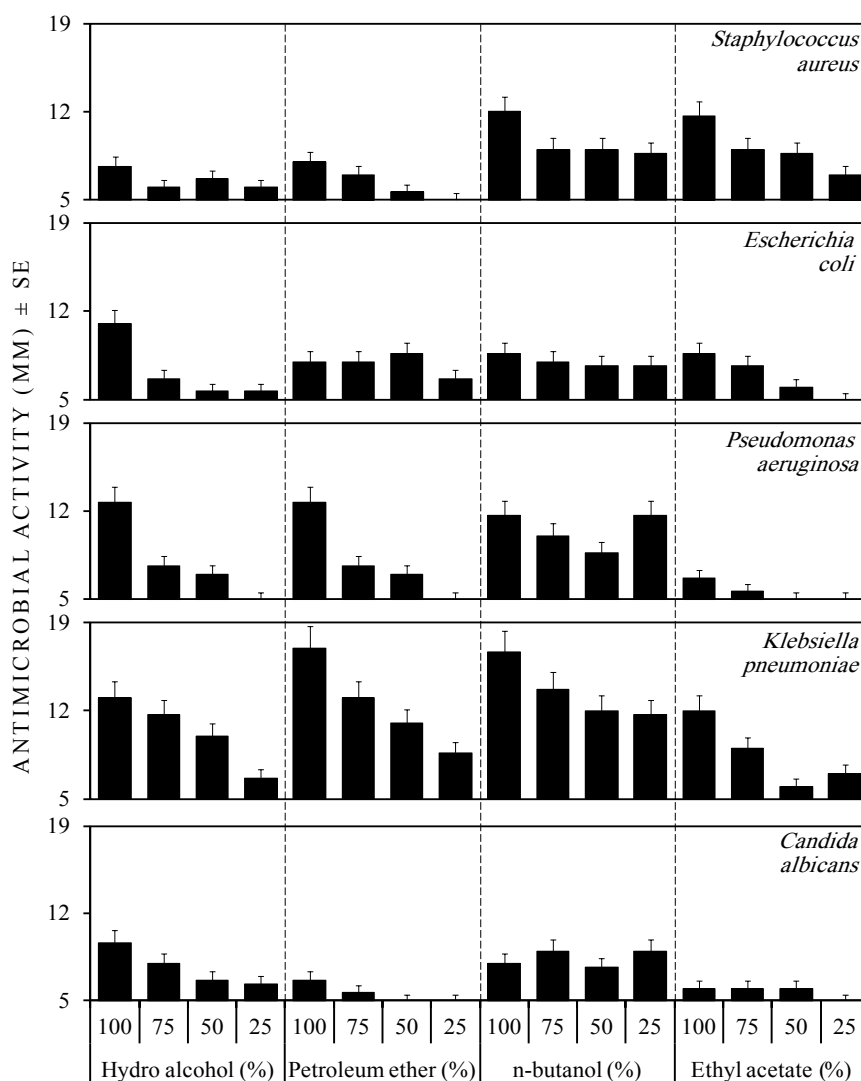


Figure 1. Antimicrobial activity of *Conula cinerea* extracts (Extract initial concentration = 1 mg mL⁻¹, disc diameter = 5 mm).

The antimicrobial activity assayed for *C. cinerea* extracts showed an overall inhibitory effect against *K. pneumoniae* (16.67 ± 5.77 mm) with the *n*-butanol extract, and 17 ± 1.73 mm with the petroleum ether extract. Little activity was observed against *E. coli*, *P. aeruginosa*, *S. aureus* and *C. albicans* at concentrations of 0.25 mg mL^{-1} . High activity against *S. aureus* was found with *n*-butanol and ethyl acetate extracts where inhibition zones equalled 12 ± 5.20 mm and 11.67 ± 3.79 mm, respectively. Moreover, the hydro alcohol extract was the most active extract against *E. coli*. Similarly, only the hydro alcohol extract of *C. cinerea* had antimicrobial activity against *C. albicans* (Figure 1). Comparing results of growth inhibition zones for the four extracts, it is evident that the petroleum ether extract possesses moderate antimicrobial properties as compared to the most active extract (*n*-butanol) and less active extracts (ethyl acetate and hydro alcohol).

The *n*-butanol extract at 25% of concentration demonstrated moderate to high antimicrobial activity against *K. pneumoniae*, *P. aeruginosa* and *S. aureus*, whereas *C. albicans*, *E. coli* and *P. aeruginosa* remained uninhibited at an equal concentration of ethyl acetate extract. The hydro alcohol extract at 25% of concentration also demonstrated low antimicrobial activity against all tested species.

The susceptibility of microbial species to crude *C. cinerea* extracts was in the following descending order: For hydro-alcohol extract, *K. pneumoniae* > *P. aeruginosa* > *E. coli* > *C. albicans* > *S. aureus*. For petroleum ether extract, *K. pneumoniae* > *P. aeruginosa* > *E. coli* > *S. aureus* > *C. albicans*. For *n*-butanol extract, *K. pneumoniae* > *S. aureus* > *P. aeruginosa* > *E. coli* > *C. albicans*. For ethyl acetate extract, *K. pneumoniae* > *S. aureus* > *E. coli* > *P. aeruginosa* > *C. albicans*.

LRA determined that the slopes were statistically significantly non-zero, i.e. one can assume that a relationship exists between inhibition activity and extract concentrations of hydro alcohol ($P < 0.001$), petroleum ether ($P < 0.001$), and ethyl acetate ($P < 0.001$). However, the linear regression slope for *n*-butanol was not statistically significantly different from zero ($P = 0.187$), indicating there was no significant trend between *n*-butanol extract concentrations and inhibition zone (Table 2). Additionally, comparison of regressions showed that slopes were not significantly different ($P = 0.232$) while the intercepts were highly significantly different ($P < 0.001$).

Table 2. Linear regression analysis applied for extract dilutions and antimicrobial activity.

Extracts	Slope	Intercept	R ²	S(C/I)	SS(I)	F	P
Hydro alcohol	6.16	4.13	0.304	28.88	584.98	25.34	<0.001
Petroleum ether	5.60	4.57	0.224	26.25	655.73	16.76	<0.001
<i>n</i> -butanol	2.37	8.53	0.030	11.13	886.98	1.78	0.187
Ethyl acetate	4.29	4.47	0.192	20.13	449.65	13.80	<0.001

(C/I: Concentrations/Inhibition, SS: Sum square).

Two-way ANOVA revealed that all plant extracts had significant differences in their actions as antimicrobial

agents with either tested strains or extract concentrations or even their interaction. Fisher-values in "Strain test" and "Extract dilutions" factors showed all highly or very highly significant effects for the four extracts (except for dilutions of *n*-butanol). Thus antimicrobial activity of *C. cinerea* varied significantly between tested strains (in particularly for the petroleum ether extract, $P < 0.001$) and according to dilutions (especially within the petroleum ether extract, $P < 0.001$). In general, the interaction effect of the two factors (Strain test * Extract concentration) had no statistical significance on the variation of antimicrobial activity in all extracts except that of petroleum ether ($P = 0.002$) (Table 3).

Table 3. Two-way analysis of variance (ANOVA).

Extracts	Effect	SS	Df	MS	F	P
Hydro alcohol	Strain test	97.6	4	24.4	4.1	0.007
	Extract dilutions	191.3	3	63.8	10.6	<0.001
	Interaction	56.2	12	4.7	0.8	0.667
	Residuals	240.0	40	6.0		
	Total	585.0	59	9.9		
Petroleum ether	Strain test	334.4	4	83.6	39.2	<0.001
	Extract dilutions	152.4	3	50.8	23.8	<0.001
	Interaction	83.6	12	7.0	3.3	0.002
	Residuals	85.3	40	2.1		
	Total	655.7	59	11.1		
<i>n</i> -butanol	Strain test	229.4	4	57.4	4.1	0.007
	Extract dilutions	45.5	3	15.2	1.1	0.370
	Interaction	48.7	12	4.1	0.3	0.988
	Residuals	563.3	40	14.1		
	Total	887.0	59	15.0		
Ethyl acetate	Strain test	120.9	4	30.2	6.0	<0.001
	Extract dilutions	90.2	3	30.1	6.0	0.002
	Interaction	38.6	12	3.2	0.6	0.793
	Residuals	200.0	40	5.0		
	Total	449.7	59	7.6		

(SS: Sum of squares, MS: Mean of the Sum of Squares, Df: Degrees of freedom).

4. Discussion

Infectious diseases represent a serious public health problem and remain the major cause of death throughout the world. Alternative natural products of plants could be of high interest to attenuate the increasing incidence of antibiotic resistance. Some phytochemicals and plant extracts are known to have antimicrobial properties, which could be of great importance in the therapy of microbial infections. Recently, various studies have been conducted over different countries, and have demonstrated the efficacy of this type of treatment (e.g. Coutinho *et al.*, 2008; Habbu *et al.*, 2009). Algeria has also recently increased research on traditional herbal medicines following scientific findings that verified their effectiveness in healing several health issues.

The present investigation explored the use of one such plant, *C. cinerea* Del., endemic in North Africa, for treating infectious diseases. The assay of antimicrobial activity of *C. cinerea* extracts showed that the hydro

alcohol extract was least active against the tested strains, and only the relatively polar fraction (*n*-butanol) had high activity against the strains. ANOVA analysis confirmed that *n*-butanol had significant variation in its antimicrobial activity against the tested strains, however there was no significant variation in activity for its tested concentrations, which was also demonstrated by LRA. These findings support the conclusion that the active antimicrobial compounds are highly concentrated in this fraction. The other extracts (hydro alcohol, petroleum ether and ethyl acetate) had the best antimicrobial activity when used at high concentrations. This was clearly revealed by both LRA and ANOVA. The wide range of antimicrobial activity shown by *C. cinerea* extracts might reflect the differences in chemical concentrations and composition obtained by each solvent. Indeed, the successful extraction of active botanical compounds from plant materials is dependent on the type of solvent used in the extraction procedure (Parekh *et al.*, 2005; Hayouni *et al.*, 2007).

The plant extracts were active against both Gram-positive and Gram-negative bacteria, though they were more active against the later. Concerning the antimicrobial activity against *C. albicans*, the present study revealed low to no activity by the plant extracts. In our study, the highest activity was recorded against the Gram-negative bacteria: *K. pneumoniae*, which was the most susceptible bacterium of all the tested strains. These results may be of great importance in infection therapy since *K. pneumoniae* can be commonly involved in urinary, intra-abdominal, and respiratory infections (Lavender *et al.*, 2005; Keynan and Rubinstein, 2007; Ahmad *et al.*, 2012).

Markouk *et al.* (1999a) reported that the acetate extract of *C. cinerea* collected from Zagora (Southern Morocco) exhibited an antibacterial effect with a minimum inhibitory concentration (MIC) of 200 µg/mL against all tested bacteria, and that the *n*-butanol extract was highly effective too, especially against *Pseudomonas fluorescens* and *Bacillus* sp., with an MIC of 12 µg/mL. In the same study, the ethyl ether extracts of *C. cinerea* were found to be inactive against all tested bacteria. These results are in agreement with ours, and confirm that bioactive components of any plant may differ in their solubility based on: (i) the extracting solvents (Hayouni *et al.*, 2007; Hassan *et al.*, 2009), (ii) the nature of biologically active components such as alkaloids, saponins, tannins, phenols, etc. (Hassan *et al.*, 2009; Marzouk *et al.*, 2010), and (iii) the geographical origin of the plant material (Seidel *et al.*, 2008), because ecological conditions in general (including abiotic factors "edaphic, climatic, water stress..." or biological interactions "such as intra and/or interspecific competitions...") may have a large impact on growth and fitness of vegetation species (Cordell, 2000; Seidel *et al.*, 2008), particularly by affecting their metabolism and secondary metabolites production (Cowan, 1999). Moreover, Ahmed *et al.* (1987) reported that *C. cinerea* is particularly rich with flavonic compounds besides sesquiterpene-lactone and sesquiterpene coumarins, which have been also isolated of this plant.

The results reported here can be considered as the first report on the antimicrobial properties of *Cotula cinerea*, an endemic species of the Algerian Saharan flora. Our findings also contribute to the knowledge of antimicrobial properties reported elsewhere for other *Cotula* species. Based on these antimicrobial results obtained using the disk diffusion method, it appears that this technique could not always be a reliable and sure method for screening the antimicrobial activity of plant extracts. As indicated by Moreno *et al.* (2006), the absence of an inhibition zone did not necessarily mean the compound was inactive, especially for the less polar compounds, which diffuse more slowly in the culture medium.

Consequently the analysis of the present results offers a simple scientific basis for traditional use of *C. cinerea* against microbial pathogen. However, *in vivo* studies on this medicinal plant are necessary and should seek to determine toxicity of different active compounds, their side effects, pharmacokinetic properties and reach their required minimum inhibitory concentration (MIC) in tissues and organs having the infection. The antimicrobial activities could be enhanced if active components are purified and adequate dosage determined for proper administration.

5. Conclusion

In the light of this study, *C. cinerea* is a prospective wild plant for the isolation of new antimicrobial substances. Although further investigations are clearly necessary to clarify and identify the bioactive constituents, we believe that our results presented herein represent a solid stepping-stone for other researchers in the field. Moreover, our antimicrobial assays results has justified and supported, at least in part, the Algerian common usage of the plant. The screening of some medicinal plant crude extracts has shown that some of those were potentially rich sources of antimicrobial agents. Finally, promoting human well-being deserves joining efforts in considering and valorising Saharan natural patrimony, as well as carrying out more scientific research on plants living in drylands by conducting chemical, biological, toxicological and pharmacological investigations as well as investigating therapeutic potential.

Acknowledgements

We gratefully acknowledge the staff of Laboratory of Biochemistry at Department of Natural and Life Sciences (University of Kasdi Merbah Ouargla, Algeria) for all facilities provided in carrying out this study.

References

- Ahmad TA, El-Sayed LH, Haroun M, Hussein AA and El-Ashry EH. 2012. Development of immunization trials against *Klebsiella pneumoniae*. *Vaccine*, **30**: 2411-2420.
- Ahmed AA, El-Sayed NH, El-Negoumy SI and Mabry TJ. 1987. Flavonoids of *Cotula cinerea*. *J Nat Prod*, **50**: 519-520.
- Beloued A. 2005. **Les Plantes Médicinales d'Algérie (Medicinal Plants of Algeria)**. Ed Office des Publications Universitaires, Algiers, 284 p.
- Boligon AA, Agertt V, Janovik V, Cruz RC, Campos MMA, Guillaume D, Athayde ML and dos Santos ARS. 2011.

- Antimycobacterial activity of the fractions and compounds from *Scutia buxifolia*. *Rev Bras Farmacogn*, **22**: 45-52.
- Cordell GA. 2000. Biodiversity and drug discovery - a symbiotic relationship. *Phytochem*, **55**: 463-480.
- Coutinho HDM, Costa JGM, Siqueira-Junior JP and Lima EO. 2008. *In vitro* anti staphylococcal activity of *Hyptis martiusii* Benth against methicillin-resistant *Staphylococcus aureus*-MRSA strains. *Rev Bras Farmacogn*, **18**: 670-675.
- Cowan MM. 1999. Plant products as antimicrobial agents. *Clin Microbiol Rev*, **12**: 564-582.
- Daglia M. 2012. Polyphenols as antimicrobial agents. *Curr Opin Biotech*, **23**: 174-181.
- Dall'Agnol R, Ferraz A, Bernardi AP, Albring D, Nör C, Sarmiento L, Lamb L, Hass M, von Poser G and Schapoval EES. 2003. Antimicrobial activity of some *Hypericum* species. *Phytomed*, **10**: 511-516.
- Habbu PV, Mahadevan KM, Shastry RA and Manjunatha H. 2009. Antimicrobial activity of flavonoid sulphates and other fractions of *Argyrea speciosa* (Burm.f) Boj. *Indian J Exp Biol*, **47**: 121-128.
- Hammiche V and Maiza K. 2006. Traditional medicine in Central Sahara: Pharmacopoeia of Tassili N'ajjer. *J Ethnopharmacol*, **105**: 358-367.
- Hassan A, Rahman S, Deebe F and Mahmud S. 2009. Antimicrobial activity of some plant extracts having hepatoprotective effects. *J Med Plant Res*, **3**: 20-23.
- Hayouni EA, Abedrabba M, Bouix M and Hamdi M. 2007. The effects of solvents and extraction method on the phenolic contents and biological activities in vitro of Tunisian *Quercus coccifera* L. and *Juniperus phoenicea* L. fruit extracts. *Food Chem*, **105**: 1126-1134.
- Katsumi S, Kazshi T and Hiroshi O. 2005. Pathogen occurrence and antimicrobial susceptibility of urinary tract infection cases during a 20-year period (1983-2002) at a single institution in Japan. *Jpn J Infect Dis*, **58**: 303-308.
- Keynan Y and Rubinstein E. 2007. The changing face of *Klebsiella pneumoniae* infections in the community. *Inter J Antimicrob Agents*, **30**: 385-389.
- Larhsini M, Markouk M, Jaouhari JT, Bekkouche K, Lazrek HB and Jana M. 2002. The antipyretic activity of some Moroccan medicinal plants. *Phytother Res*, **16**: 97-98.
- Lavender H, Jennifer J and Clegg JS. 2005. *Klebsiella pneumoniae* type 3 fimbria-mediated immunity to infection in the murine model of respiratory disease. *Inter J Med Microbiol*, **295**: 153-159.
- Maiza K, Brac de la Perrière RA and Hammiche V. 1993. Pharmacopée traditionnelle saharienne: Sahara septentrional. *2nd Proc of European Conf on Ethnopharmacology & 11th Int Conf of Ethnomedicine*, Heidelberg France, pp 169-171.
- Markouk M, Redwane A, Lazrek HB, Jana M and Benjama A. 1999a. Antibacterial activity of *Cotula cinerea* extracts. *Fitoterapia*, **70**: 314-316.
- Markouk M, Lazrek HB and Jana M. 1999b. Analgesic Effect of Extracts from *Cotula cinerea* (L). *Phytother Res*, **13**: 229-230.
- Marzouk B, Marzouk Z, Décor R, Mhadhebi L, Fenina N and Aouni M. 2010. Antibacterial and antifungal activities of several populations of Tunisian *Citrullus colocynthis* Schrad. immature fruits and seeds. *J Med Mycol*, **20**: 179-184.
- Matu EN and van Staden J. 2003. Antibacterial and anti-inflammatory activities of some plants used for medicinal purposes in Kenya. *J Ethnopharmacol*, **87**: 35-41.
- McKay DL and Blumberg JB. 2007. A review of the bioactivity of South African herbal teas: Roobos (*Aspalathus linearis*) and Honey comb (*Cyclopia intermedia*). *Phytother Res*, **21**: 1-16.
- Moreno S, Scheyer T, Romano CS and Vojnov AA. 2006. Antioxidant and antimicrobial activities of rosemary extracts linked to their polyphenol composition. *Free Radic Res*, **40**: 223-231.
- Mothana RA and Lindequist U. 2005. Antimicrobial activity of some medicinal plants of the island Soqatra. *J Ethnopharmacol*, **96**: 177-181.
- Parekh J, Jadeja D and Chanda S. 2005. Efficacy of aqueous and methanol extracts of some medicinal plants for potential antibacterial activity. *Turkish J Biol*, **29**: 203-210.
- Qaralleh H, Iddid S, Saad S, Susanti D, Taher M and Khleifat K. 2010. Antifungal and antibacterial activities of four Malaysian sponge species (Petrosiidae). *J Med Mycol*, **20**: 315-320.
- Seidel V, Peyfoon E, Watson DG and Fearnley J. 2008. Comparative study of the antibacterial activity of propolis from different geographical and climatic zones. *Phytother Res*, **22**: 1256-1263.
- Shahid M, Malik A, Akram M, Agrawal LM, Khan AU and Agrawal M. 2008. Prevalent phenotypes and antibiotic resistance in *Escherichia coli* and *Klebsiella pneumoniae* at an Indian tertiary care hospital: plasmid-mediated Cefoxitin resistance. *Inter J Infect Dis*, **12**: 256-264.

Correlation Between Numerical Profiles Generated for Soil Spore Forming Bacilli and Their Inhibitory Potential Against *Staphylococcus aureus* ATCC 6538

Qasem M. Abu Shaqra*

Department of Allied Medical Sciences, Zarqa University College, Al- Balqa Applied University- P.O. Box 5341,

Code number: 13111, Zarqa- Jordan

Received: May 9, 2013

Revised: June 18, 2013

Accepted June 21, 2013

Abstract

Numerical profiling using conventional tests for studying bacterial diversity of soil have probably never been employed. The main objective of this work was to describe the heterogeneity of spore forming bacilli in a vegetative soil using numerical profiling and to establish the possible application of this approach for the prediction of the antimicrobial potentials of soil dwelling *Bacillus* species. A total of 100 spore forming Gram positive bacilli were recovered from soil samples and their characters were determined using 9 conventional tests. A 3 digit numerical profile was constructed for each isolate. This labelling system resulted in the generation of 12 different profiles which indicated the diversity of soil *Bacillus* isolates. The highest frequency of occurrence was detected among the isolates labelled with the profile 4.1.5 and the lowest among those labelled with 4.7.5; 4.6.6 and 1.0.3 profiles. Out of 13 isolates labelled with 4.4.3, the cell free extract of 12 exhibited no antimicrobial activity against *Staphylococcus aureus* ATCC 6538 whereas, extracts obtained from 15 out of 18 isolates labelled with the profile 4.1.5 were inhibitory to the test organism. It is concluded that the numerical profiling system is a significant tool for the establishment of diversity among soil spore forming bacilli which can also be used for the prediction of the potentials of isolates to produce antimicrobial metabolites.

Keywords: *Bacillus* species, Vegetative soil, Diversity, Antimicrobial effect, Numerical profiles.

1. Introduction

Bacillus species are soil dwelling organisms that produce metabolites with diverse chemical structures and potentials to inhibit a wide spectrum of bacteria (Morsi and Atef, 2010). Antibiotics produced by *Bacillus* species include bacitracin, colistin, circulin, gramycidin, polymyxin, and tyrotricidin. However, these antibiotics were discovered many decades ago, they are still used in medical practice (Fickers, 2012).

The isolation of *Bacillus* species from soil is usually carried out by traditional techniques which involve the treatment of soil with heat to inhibit vegetative microorganisms and to allow the selection of spore forming bacteria (Kuta *et al*, 2009). Isolates may be characterized by a range of methods. Morphological and physiological characteristics were used for classification and identification. However, such techniques can provide the basis for bacterial, and

Bacillus taxonomy, additional tests were found to be necessary for confirmation and for fine-scale resolution (Maughan and Van der Auwera, 2011).

Identification of spore forming bacteria is a difficult task, it was suggested that for the accurate identification of these Bacilli, several standard schemes should be used together for the establishment of different phenotypic characteristics of all known species (Morsi *et al*, 2010). In recent years, molecular techniques were introduced for the characterization of isolates at a genetic level (Kadyan *et al*, 2013). Although, this characterization approach is valuable in discriminating and grouping *Bacillus* strains, it provides little ecological information. For example, 16S rRNA gene based methods provides good phylogenetic information to the genus level, but in themselves, give little information on function (Mandic-Mulec and Prosser, 2011; Maughan and Van der Auwera, 2011). On the other hand, traditional approaches that group isolates on the basis of common metabolic properties

* Corresponding author. e-mail: qabushaqra@hotmail.com.

may be limited in terms of phylogenetic power but provide clues to environmental factors and can be strong indicators of potential ecosystem function (Koeppel *et al.*, 2008). Kim *et al.* (2003) indicated that for the accurate identification and classification of spore forming Bacilli from soil, a multiphasic approach may be required.

Antimicrobial activity is usually established by screening cell free extract derived from soil isolates against a panel of bacteria using the agar diffusion method (Manga and Oyeleke, 2008). Therefore, it is evident that identification procedures in addition to preliminary screening are tedious and time consuming. If a system or a model can be devised to label the isolates and simultaneously predict their potential to produce antimicrobial substances, then definitely, time, efforts and money can be saved. The effective labeling system should be capable of grouping isolates according to their biochemical properties and should detect variations among isolates. These stipulations were taken into considerations by (Griffiths and Lovitt, 1980), who suggested that numerical profiling can be used as a significant tool for the study of bacterial diversity in different ecosystems. The advantage of this system over traditional and modern identifications methods is that all isolates are labeled and none is left unidentified. Depending on the tests included in the characterization of isolates, it can provide an idea about the function of the isolate from the respective environment. The objectives of this communication were to establish the diversity of endospore forming *Bacillus* species recovered from rich vegetative Jordanian soil using the numerical profiling system and to investigate the possible application of this system for the prediction of antimicrobial potential of these isolates.

2. Materials and Methods

2.1. Collection of Soil Samples

A total of 10 soil samples were collected from highly vegetative locations in Ajloun Mountains (near the castle). Samples were derived from the depth of 5 cm below the surface, placed in sterile containers and processed in the laboratory within 2 days of collection.

2.2. Isolation of *Bacillus* Species

One gram of each soil sample was separately suspended in a sterile tube containing 9 ml of sterile distilled water and shaken vigorously for 2 minutes using a vortex mixer. The samples were heat treated at 80 °C for 10 minutes in a water bath. Upon cooling to room temperature, each soil suspension was 10 fold serially diluted in sterile distilled water. Aliquots of 0.1 ml of the appropriate dilution were plated on Nutrient Agar Medium (Difco- USA) using the spread plate technique. All plates were then incubated at 37 °C for 2 days. From plates that contained counts between 30 to 250 colonies, 10 colonies with apparently different colonial morphology were isolated and further purified by repeated subculture.

2.3. Preparation of Cell Free Extracts

Each purified isolate was inoculated into a sterile tube containing 10 ml of Nutrient Broth (Difco- USA) and then incubated at 30 °C for 72 hours. The tubes were centrifuged and supernatant was then filtered through 0.22 µm bacteriological filters and stored at 4°C until used as described in the next experiment.

2.4. Antimicrobial Activity Test

Cell suspension of *Staphylococcus aureus* ATCC 6538 was prepared in accordance with the method described by Abu Shaqra and Al Groom (2012) using 0.5 McFarland standard. The suspension prepared, contained 10⁶ Colony Forming Unit /ml and was employed to streak plates of Mueller Hinton Agar (Difco- USA). After drying, wells were bored using sterilized cork borer of 6 mm diameter and each well received 50 µl of the extract of a single isolate. Plates were incubated for 24-48 h at 37 °C before measuring the diameter of inhibition zones developed.

2.5. Numerical Profiles

Tests used to characterize each soil isolate are presented in table 1. These tests were performed as described by Baron *et al.* (1994). Results were arranged in groups of 3 and scores of 1, 2, or 4 were assigned to label the isolates according to their position in the table. In this context, attention should be drawn to two important points; first, all tests carried equal weights as they were randomly positioned in the table and second, scoring numbers were chosen to avoid overlap between tests in case of positive results when the total score is calculated. For example, if the first digit was 3, this would indicate that the organism was xylose fermentor, grew in 10% concentration of NaCl and was unable to grow under anaerobic conditions. If the score of the third test in the profile was 3, then one would be confused whether the organism was positive for the first two tests or merely positive for the third test.

The percentage diversity of the *Bacillus* isolates in relation to the profiles generated was calculated according to the equation given by Abu Shaqra and Mashni (2006) as follows:

$$\frac{\text{Number of profiles}}{\text{Total number of isolates}} \times 100$$

The work was concluded by relating the antimicrobial activity of each isolate to its generated numerical profile to determine if the profiles constructed can provide indications regarding the inhibitory activity of the cell free extract derived from the isolates.

Table 1. Characters used to generate numerical profiles for spore forming bacilli recovered from soil samples.

Score	First digit	Second digit	Third digit
0 or 1	Xylose fermentation	Mannose fermentation	Nitrate reduction
0 or 2	Growth in 10% NaCl	Raffinose fermentation	Hipurate hydrolysis
0 or 4	Anaerobic growth	Voges Proskauer	Starch hydrolysis

2.6. Tabulation of Results

Each numerical profile generated has taken a separate row in a table and the numbers of isolates labelled with the respective profile were tabulated next to it. Upon measurement of the antibacterial effect against *Staphylococcus aureus* ATCC 6538, zones of inhibitions were categorized into 3 groups; no zone of inhibition, ≤ 10 mm and > 11 mm. Isolates that belonged to a specific profile generated were grouped according to the size of inhibitory zone determined and were also placed in the table next to their profiles (Table 2). Therefore, it was possible to correlate the number of isolates in each profile with their antimicrobial activity and this is discussed in more details in the results section.

2.7. Statistical Analysis

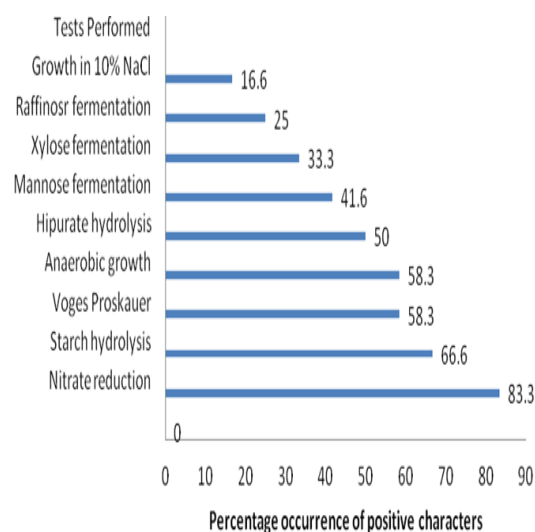
The system of Microsirir statistical analysis and data management was employed. Student t test was performed to establish significance of difference between the number of isolates grouped in each numerical profile and the zone of inhibition measured for the cell free extract derived from each isolate against *S. aureus* ATCC 6538. The test was performed to profiles that contained equal or more than 7 isolates. The difference was considered as significant when Probability (*p*) value was less than 0.05.

3. Results

A total of 100 pure isolates of bacteria were obtained from 10 soil samples collected from mountainous and highly vegetative region in Jordan. All recovered organisms were spore forming, Gram positive bacilli in pairs or longer chains. Most of the isolated colonies were white, opaque or translucent and they were rough, granular, or wispy. These characteristics were considered as presumptive for the identification of the isolates as *Bacillus* species.

Using the numerical profiling system and the 9 identification tests given in table 1, it was possible to generate 12 different profiles and this indicated the diversity of the *Bacillus* species recovered (12% diversity). Figure 1 illustrates the percentage occurrence of positive characters which constituted the back bone of this diversity. It is evident that 83% of the isolates were capable of nitrate reduction while 67% were starch hydrolysers.

Table 2 demonstrates the frequency of occurrence of each profile among the isolates. This table clearly shows that profile 4.1.5 was the label for the most common isolates whereas; 4.6.6; 4.7.5 and 1.0.3 were the least encountered.

**Figure 1.** Percentage occurrence of positive characters among spore forming isolates recovered from soil samples.

The diameter of the inhibition zone produced by cell free extract derived from each isolate was taken as a criterion to mark its antimicrobial activity. Table 2 also shows the number of isolates in each profile generated in relation to the zone of inhibition measured. Two examples are given below to explain the content of this table; out of 9 isolates labeled with the profile 2.5.2, 5 produced zones of inhibition > 11 mm, while 3 exhibited zones ≤ 10 mm and only 1 isolate was none inhibitory. On the other hand, 13 isolates were labelled with the profile 4.4.3, 12 of these isolates were inhibitory and none showed the formation of any inhibitory zone.

According to results presented in table 2, 43% of isolates demonstrated no inhibitory zone, while 21% and 36% of the isolates showed inhibitory zone equal or less than 10 and > 11 mm in diameter, respectively. It is worth noting that the majority of *Bacillus* isolates labeled with profiles such as 4.1.5 and 3.4.5 were effective inhibitors to the test organism whereas, the majority of species labelled with 2.1.7, 4.4.3 and 0.0.1 were none inhibitors. Although it was not the intention of this work to identify isolates to species level, it was possible using the few tests performed and the identification tables given by (Barrow and Feltham, 1993) to predict closely the *Bacillus* species to which many of the isolates belonged. For instance, profile 4.1.5 fitted *B. licheniformis* while profiles 4.4.3, 2.1.7 and 3.7.7 most probably denoted *B. laterosporus*, *B. firmus* and *B. subtilis*, respectively.

Table 2. Frequency of numerical profiles occurrence in relation to the inhibition zone measured for their cell free extract against *S. aureus* ATCC 6538

Profiles generated	occurrence Frequency of each profile	No. of isolates in relation to inhibition zone diameter		
		None	≤ 10 mm.	> 11 mm.
4.1.5	18	3	5	10
3.7.7	6	0	2	4
2.5.2	9	1	3	5
4.4.3	13	12	1	0
2.1.7	12	10	2	0
4.4.5	9	6	1	2
5.3.4	7	4	2	1
0.0.1	7	7	0	0
3.4.5	7	0	2	5
4.7.5	4	0	0	4
4.6.6	4	0	1	3
1.0.3	4	0	2	2
Total	100	43	21	36

4. Discussion

Numerical profiling is not a new approach for the study of bacterial diversity. Griffiths and Lovitt (1980) were the first to use this system while investigating the bacterial diversity of oil storage tanks water bottom. These authors suggested that the system could be used to study bacterial diversity in different ecosystems. Abu Shaqra and Mashni (2006) used the numerical profiling for studying the bacterial diversity of hard gelatine capsules contaminants and proposed the system as a significant tool for establishing the diversity of contaminants in cosmetic and pharmaceutical products. However, methods for studying bacterial diversity in soil are available (Kirk *et al.*, 2004), as far as is known, numerical profiling as a mean for establishing the diversity of spore forming bacilli in soil was never used.

Tests used for the characterization of isolates were chosen on the basis of their convenience and ability to establish diversity. For instance, spore formation test and reaction to Gram stain though were carried out, they were excluded because they were of no significance in determining the diversity of *Bacillus* as the vast majority of species gave positive reaction to Gram stain and by most produced endospores (Sneath, 1986).

The best test to be selected for the detection of bacterial diversity in a particular habitat is the one which can differentiate between isolates on equal bases. For clarification, the ideal test should give positive results for 50% of isolates and negative test results for the remaining 50% (Griffiths and Lovitt, 1980). Figure 1 demonstrates the percentage occurrence of positive characters among our isolates and indicates that none of the tests used was ideal as no single test was capable of 50% differentiation. It is evident that this method relies

on stressing the characters of the organism rather than focussing on their names, although in occasions as the case is in this investigation, assigning names was possible.

The diversity of *Bacillus* species in soil varies considerably in terms of numbers and types of isolates. Nishijima *et al.*, (2005) have studied soil samples collected from different locations in Japan and found that number of spore forming bacteria did not differ a lot from one place to another but the types of isolates did. They were able to recover up to 18 different species from one location whereas; only one species was isolated from another. In this work 12 different numerical profiles were generated for the isolates. Had more tests been used in the generation of profiles, diversity could have been higher. In fact addition of tests for the construction of the numerical profiling system merely requires the increase in the number of digits; instead of 3 digits, 4 or 5 digits could be constructed.

The frequency of occurrence of each profile generated is illustrated in table 2. It is clear from this table that profile 4.1.5 was the most frequently encountered, followed by 4.4.3 while isolates labeled with profiles 5.7.5, 4.6.6 and 1.0.3 were the least encountered. These observations are consistent with those made by Kuta *et al.*, (2009) who found that frequency of occurrence among their *Bacillus* soil isolates (identified to species level by conventional methods) varied between 1.9% and 30.8%. The close similarity of results reported herein and those of Kuta *et al.*, (2009) indicated the effectiveness of the numerical profiling system in dealing with soil bacterial diversity as compared to traditional approaches.

Soil is a well known source of microorganisms capable of producing a variety of biologically active metabolites including antibiotics. Table 2 shows that 36% of *Bacillus* isolates recovered from Ajloun soil exhibited effective antagonistic activity against *S. aureus* ATCC 6538. This percentage is not far away from that determined by Kuta *et al.*, (2009) who found that 41.7% of *Bacillus* species isolated from soil samples obtained from 16 refuse dump sites in Minna – Niger were inhibitory against a strain of *S. aureus*.

It is of profound importance to note that isolates with similar profiles did not give 100% exact results in the antimicrobial activity test. This particular observation is of dual significance. First, had isolates with the same profile been with absolute phylogenetic relation and this is clearly not the case, all would have given the same inhibitory activity against the test organism. The lack of this absolute relatedness was expected as tests used were dependent on the biochemical activity and no tests were performed to identify the genotype of isolates. However, simple and limited number of tests was used, numerical profiling system proved to be as good as any conventional approach employed for the study of bacterial diversity. Second, not all isolates with similar profiles gave the same spectrum of activity against *S. aureus*. Out of 18 isolates labeled with the profile 4.1.5, 10 were effective inhibitors while, 5 and 3 isolates were with limited or no antimicrobial activity, respectively.

Statistical significance was found to exist ($p < 0.05$) between each profile and the number of isolates grouped in that profile with respect to the zone of inhibition measured. Table 2 shows that isolates with profiles 4.4.3 and 2.1.7 catered for 13 as well as 12 isolates, respectively, with the majority of these isolates being none inhibitory to the test organism.

Based on the outcome of this work, future study may exempt isolates labeled with profiles that were found to be of poor antimicrobial effect from being screened for inhibitory activity as they are unlikely to exhibit antimicrobial property. It might be thought that the use of a single bacterial species in the antimicrobial study is a draw back in the set up of this investigation, the fact is, this work was not designed to isolate a strain or strains of *Bacillus* with broad spectrum of antimicrobial activity but to demonstrate the usefulness of the profiling system in predicting the antimicrobial potentials of isolates and this was achieved.

5. Conclusion

This investigation has shown for the first time that numerical profiling is a useful technique for studying the diversity of soil spore forming bacteria. It also points out to the possibility of exploiting the same system in the prediction of antibacterial activity of isolates recovered from this extremely diverse habitat.

References

- Abdulkadir M and Waliyu S. 2012. Screening and isolation of the soil bacteria for ability to produce antibiotics. *Eur J Appl Sci.*, **4** (5): 211-215.
- Abu Shaqra QM and Al Groom RM. 2012. Microbiological quality of hair and skin care cosmetics manufactured in Jordan. *Int Biodeter Biodegr.*, **69**: 69-72
- Abu Shaqra QM and Mashni YI. 2006. Bacterial diversity of the contaminants of hard gelatine capsules using numerical profiles and conventional methods. *Eur J Parenter Pharm Sci.*, **11**(3): 71- 74.
- Baron EJO, Peterson LR and Finegold SM 9 (eds). 1994. **Bailey and Scott's Diagnostic Microbiology**. Mosby, Missouri, USA.
- Barrow G and Feltham R. 1993. **Cowan and Steels Manual for the Identification of Medical Bacteria**. 3rd ed. Cambridge University Press, Cambridge.
- Fickers P. 2012. Antibiotic Compounds from *Bacillus*: Why are they so amazing? *Am J Biochem Biotechnol.*, **8** (1): 40-46,
- Griffiths AJ and Lovitt R. 1980. Use of numerical profiles for studying bacterial diversity. *Microb Ecol.*, **6**: 35- 43.
- Kadyan S, Panghal M, Kumar S, Singh K and Yadav JP. 2013. Assessment of functional and genetic diversity of aerobic endospore forming Bacilli from rhizospheric soil of *Phyllanthusamarus* L. *World J Microbiol Biotechnol.*, Mar 23. [Epub ahead of print]
- Kim JS, Kwon SW, Jordan F and Ryu JC. 2003. Analysis of bacterial community structure in bulk soil, rhizosphere soil, and root samples of hot pepper plants using FAME and 16S rDNA clone libraries. *J Microbiol Biotechnol.*, **13**:236-242
- Kirk JL, Beadette LA, Hart M, Moutogliss P, Klironomos JN, Lee H and Trevors JT. 2004. Methods of studying soil microbial diversity. *J Microbiol Methods.*, **58** (2): 169- 188.
- Koeppel A, Perry EB, Sikorski J, Krizanc D, Warner A, Ward DM, Rooney AP, Brambilla E, Connor N, Ratcliff RM, Nevo E and Cohan FM. 2008. Identifying the fundamental units of bacterial diversity: a paradigm shift to incorporate ecology into bacterial systematics. *Proc Natl Acad Sci, U S A*, **105**: 2504-2509.
- Kuta FA, Nimzing L and Orka'a Y. 2009. Screening of *Bacillus* species with potentials of antibiotics production. *Appl Informatics J.*, **24** (1-2): 42 – 46.
- Mandic-Mulec I and Prosser JI. 2011. Diversity of endospore-forming bacteria in soil: Characterization and driving mechanisms. In: Logan NA and De Vos P (Eds), **Endospore-forming Soil Bacteria, Soil Biology** 27, Springer-Verlag Berlin Heidelberg, pp. 31-59.
- Manga BS and Oyeleke SB. 2008. **Essentials of Laboratory Practical's in Microbiology**. 1st ed. Tobes Publishers, pp. 56-76.
- Maughan H and Van der Auwera G. 2011. *Bacillus* taxonomy in the genomic era finds phenotypes to be essential though often misleading Infection, *Genetics and Evolution*, **11**: 789-797.
- Morsi NM, Atef NM and El-Hendawy H. 2010. Screening for some *Bacillus* spp. inhabiting Egyptian soil for the biosynthesis of biologically active metabolites. *J Food Agric Environ*, **8** (2): 1166-1173.
- Nishijima T, Toyota K and Mochizuki M. 2005. Predominant culturable *Bacillus* species in Japan arable soil and their potential as biocontrol agents. *Microbes Environ.*, **20** (1): 61-68.
- Sneath, PHA. 1986. In **Bergey's Manual of Systematic Bacteriology**. Bergey DH, Sneath PHA and Holt JG (Eds.) Williams & Wilkins, Baltimore Vol. 2: pp 1104-1139.

Nitrate Reductase Assay Using Sodium Nitrate for Rapid Drug Susceptibility Testing of *Mycobacterium tuberculosis* Directly on Sputum Samples

Mohammed Abdul- Imam Almazini*

Department of Biology, College of Science, University of Basrah, Iraq

Received: May 1, 2013 Revised: June 14, 2013 Accepted: June 22, 2013

Abstract

Multidrug-resistant tuberculosis is an increasing public health concern in many parts of the world, especially in low-income countries, where most cases occur. Traditional drug susceptibility testing is either time-consuming, such the proportion method on solid media, or expensive, such as the BACTEC 960 System. The aim of this study was to evaluate a nitrate reductase assay (NRA) using sodium nitrate (NaNO_3) on smear- positive sputum for the detection of multidrug-resistant *Mycobacterium tuberculosis* (MDR-TB) and compared it with the nitrate reductase assay using potassium nitrate (KNO_3) and Proportion Method (PM) or Direct Proportion Method (DPM). The NRA- NaNO_3 results were compared with other methods for 91 sputum samples for which comparable results were available. The sensitivity (ability to detect true drug resistance) and specificity (ability to detect true drug susceptibility) of the NRA- NaNO_3 , were 100% and 96%, 93% and 100%, 85% and 98%, and 76% and 97% for Rifampin, Isoniazid, Streptomycin and Ethambutol, respectively. The results were in most cases available in 10 days. NRA- NaNO_3 is simple to perform and provides a rapid, accurate, especially in low-income countries and might become alternative to traditional methods.

Keyword: Nitrate reductase assay, Sodium nitrate, *Mycobacterium tuberculosis*.

1. Introduction

Tuberculosis (TB) remains one of the major causes of morbidity and mortality from infection in humans. The World Health Organization (WHO) estimates that one third of the world population is infected with *Mycobacterium tuberculosis*, 9.4 million new cases of tuberculosis and 1.3 million deaths from tuberculosis occurring worldwide. The worldwide incidence is 140 cases per 100,000 population (WHO, 2010).

The emergence of multidrug-resistant (MDR) tuberculosis, defined as tuberculosis caused by strains resistant to the two first-line drugs (Isoniazid and Rifampin), and extensively drug-resistant (XDR) tuberculosis, defined as tuberculosis caused by strains resistant to the two above mentioned drugs, to at least one fluoroquinolone, and to at least one of three injectable second-line drugs (Amikacin, Kanamycin and Capreomycin) (Bwanga *et al.*, 2009).

Data from more than 100 countries collected during the last decade show that 5% of all TB cases have MDR-TB. There were an estimated 500,000 new MDR-TB cases in 2007. Twenty- seven countries accounted for 85% of all MDR-TB cases. The top five countries with the largest number of MDR-TB cases are India, China, the Russian Federation, South Africa and

Bangladesh, while XDR-TB has been found only in 58 countries to date (WHO, 2010).

Tuberculosis is one of the most important health problems worldwide. For this reason, the rapid diagnosis of TB drug resistance is a priority to avoid the spread of resistant strains (Palomino, 2005). There are different methods for detection of TB drug-resistance.

The BACTEC radiometric system has the advantage of being more rapid (5-10 days), but requires the use of radio-isotopes and can be costly to be performed routinely. Commercial tests (MGIT, E-Test) and molecular tools (INNO –LIPA) have been proposed, but are expensive and also impractical for routine use (Lemus *et al.*, 2004; Palomino, 2005).

For developing countries, it would be useful to have a simple and inexpensive test that could rapidly detect drug-resistant *M. tuberculosis* strains. Several methods have been reported, including colorimetric methods that use redox indicators (MTT and resazurin) and phage amplification technology (Martin *et al.*, 2003; Simboli *et al.*, 2005).

Conventional tests for the detection of drug resistance require several weeks to yield results. Recently, alternative rapid methods have been developed (Solis *et al.*, 2010). Among them, the colorimetric nitrate reductase assays (NRA), based on the ability of *M. tuberculosis* to reduce nitrate to nitrite,

* Corresponding author. e-mail: almazini_engineer@yahoo.com.

has been successfully applied on solid medium. This indirect method result in less than 14 days but requires an initial 3 to 4 weeks for cultivation of the isolate (Coban *et al.*, 2004). Another conventional method is proportion method (PM) or Direct Proportion Method (DST) for mycobacterial drug susceptibility testing requires several weeks of incubation to give results (Canetti *et al.*, 1989).

The aim of the present study was to comprise performance of a direct NRA, PM and using sodium nitrate (NaNO_3) with clinical sputum samples instead of bacterial isolates in determining the susceptibilities to rifampin (RIF), isoniazid (INH), streptomycin (STR), and ethambutol (EMB) of *M. tuberculosis* strains in microscopy-positive clinical samples from patients with pulmonary tuberculosis.

2. Materials and Methods

2.1. Specimen Processing

From February to August 2012, a total of 100 smear-positive sputum samples from new and treated patients, with positive score of 1+ or more (>1 acid-fast bacillus-AFB) per field (WHO, 1998), were collected at the tuberculosis chest disease clinic in Basrah city. The samples (one per patient) were processed using the Modified Petroff Digestion Decontamination (WHO, 1999). The sediment was re-suspended in 1ml of sterile distilled water, and portions were plated onto NRA drug susceptibility testing medium and into a Lowenstein Jensen (LJ) tube without nitrate, which was later used for the Indirect Proportion Method (IPM)

2.2. Direct NRA Drug Susceptibility test (by using KNO_3)

The NRA was performed as described previously by Musa *et al.* (2005). We used standard LJ medium with 1.000 μg of KNO_3/ml and with or without Rifampin (RIF). For LJ medium with RIF, the critical concentration of 40 $\mu\text{g}/\text{ml}$ was used. Before NRA, part of the decontaminated suspension was diluted 1:10 in sterile distilled water. For each specimen, 0.2 ml of the undiluted suspension was inoculated into LJ medium containing KNO_3 and RIF, and 0.2 ml of the 1:10 dilution was inoculated into four drug-free LJ medium tubes containing KNO_3 . The tubes were incubated at 37°C.

The assay was performed as described previously by Angeby *et al.* (2002). After 10 days of incubation, 0.5 ml of freshly prepared reagent mixture (1 part 50% concentrated hydrochloric acid, 2 parts 0.2% sulfanilamide, and 2 parts 0.1% n-1-naphtyl-ethylenediamine dihydrochloride) was added to one drug-free tube. If any color appeared, the tube with RMP was developed with the reagent mixture. Otherwise, the other tubes were re-incubated, and the procedure was repeated at day 14, day 18, and finally at day 28. An isolate was considered to be resistant if there was a color change in the RMP tube equal or greater than that in the 1:10-diluted growth control. An isolate was considered to be susceptible if there was no color change or a color change less than that in the 1:10-

diluted growth control. NRA was considered to be invalid if the nitrate reaction was negative in the drug-free medium at day 28 despite the presence of colonies.

2.3. Direct Proportion Method (DST) or Proportion Method (PM)

The technique was carried out on normal LJ medium according to the laboratory standard procedure (Canetti, 1993). The medium was prepared in 7-ml portions in 150-by-155 mm glass tubes with rubber plugs, with or without antimicrobial agents incorporated. Critical concentrations of antituberculosis drugs were the same as were used for NRA. The critical proportion values were 10% for RIF and STR and 1% for INH and EMB. For each strain, part of the suspension was diluted 1:100, and 0.2ml of the dilution was inoculated into two tubes of LJ medium without antibiotics. Then, 0.2ml of the undiluted suspension was inoculated into the tubes containing LJ medium with antibiotics. The tubes were incubated at 37°C. Final susceptibility results were reported after 40 days following the laboratory standard procedure, but preliminary results could be reported earlier for resistant strains, sometimes as early as after 20 days.

2.4. Direct NRA by using Sodium Nitrate (NaNO_3)

The method is similar of direct NRA drug susceptibility test but using sodium nitrate (NaNO_3) in replacement of potassium nitrate (KNO_3) (Maira *et al.*, 2012).

2.5. Quality

Internal quality control was done using the fully susceptible *M. tuberculosis* H37Rv and Known MDR *M. tuberculosis* isolate.

2.6. Statistical Analysis

In the present study, the term sensitivity reflects the ability to detect a true drug resistance in a strain, whereas specificity reflects the ability to detect a true drug susceptibility. Statistical analysis of data was carried out by using SPSS analysis (Moore, 2000).

3. Results

One hundred sputum samples of *M. tuberculosis* were analyzed by the Direct NRA- KNO_3 , Direct NRA- NaNO_3 and DST methods. Table 1 shows the results obtained with Direct NRA- KNO_3 compared to DST method using sputum samples. The smear results for AFB were positive with more than 10 AFB per field (+++). Of the 100 smear microscopy-positive results, 9 had negative growth control as determined by the NRA method and could thus not be used in the comparison. Then, 91 sputum samples could be used for the comparison between three methods.

In table 1, for RIF, 60 isolates were found resistant and 24 susceptible by both methods. For INH, 64 isolates were resistant and 22 susceptible by both methods; four strain gave a discordant result being susceptible by DST method. For STR, 67 isolates were resistant and 20 susceptible by both methods; three isolates were susceptible by DST but resistant by NRA- KNO_3 . In other hand, for EMB, 67 isolates were

resistant and 18 susceptible by both methods; 4 isolates were susceptible by DST but 2 isolates susceptible by NRA-KNO₃. The results were available in 10 days for 11 samples, in 14 days for 45 samples, and in 18 days for 35 samples.

Table 2 shows the sensitivity and specificity obtained with NRA using NaNO₃ and KNO₃ compared to the DST method. Drug susceptibility testing for RIF showed a sensitivity of 93% with KNO₃ and 100% with NaNO₃, but specificity was 96% for both nitrate

sources. For INH the sensitivity was 90% with KNO₃ and 93% with NaNO₃ while the specificity was 97% and 100%. For STR the sensitivity was 80% with KNO₃ and 85% with NaNO₃ while the specificity was 94% and 98%. In addition, for EMB the sensitivity was 71% with KNO₃ and 76% with NaNO₃ while the specificity was 90% and 97%.

Figure 1 shows the comparison of three methods are NRA- NaNO₃, NRA-KNO₃ and DST.

Table 1. Comparison of the susceptibility results, sensitivity and specificity to the Direct NRA method by using KNO₃, NaNO₃ and DST for *M. tuberculosis* in sputum samples.

Drug	Direct proportion method (DST) determination	Direct NRA by using KNO ₃			
		NO.		%	
		R	S	Sensitivity	Specificity
RIF	R	60	5	93	-
	S	2	24	-	96
INH	R	64	4	90	-
	S	1	22	-	97
STR	R	67	3	80	-
	S	1	20	-	94
EMB	R	67	4	71	-
	S	2	18	-	90
Total	R	258	16	80	-
	S	6	84	-	97

R= Resistant ; S= Susceptible

Sensitivity = reflects the ability to detect (true resistant).

Specificity = reflects the ability to detect (true susceptibility).

Table 2. Sensitivity and specificity of the NRA using KNO₃ and NaNO₃ compared to the DST method for *M. tuberculosis* in sputum samples.

Drug	Direct proportion method (DST) determination	NRA – KNO ₃				NRA- NaNO ₃			
		NO		%		NO		%	
		R	S	Sensitivity	Specificity	R	S	Sensitivity	Specificity
RIF	R	60	5	93	-	61	3	100	-
	S	2	24	-	96	2	25	-	96
INH	R	64	4	90	-	65	5	93	-
	S	1	22	-	97	1	20	-	100
STR	R	67	3	80	-	63	4	85	-
	S	1	20	-	94	2	22	-	98
EMB	R	67	4	71	-	60	6	76	-
	S	2	18	-	90	1	24	-	97
Total	R	258	16	80	-	250	18	86	-
	S	6	84	-	97	6	91	-	98

R = Resistant ; S = Susceptible

Sensitivity = reflects the ability to detect true resistant.

Specificity = reflects the ability to detect true susceptibility.

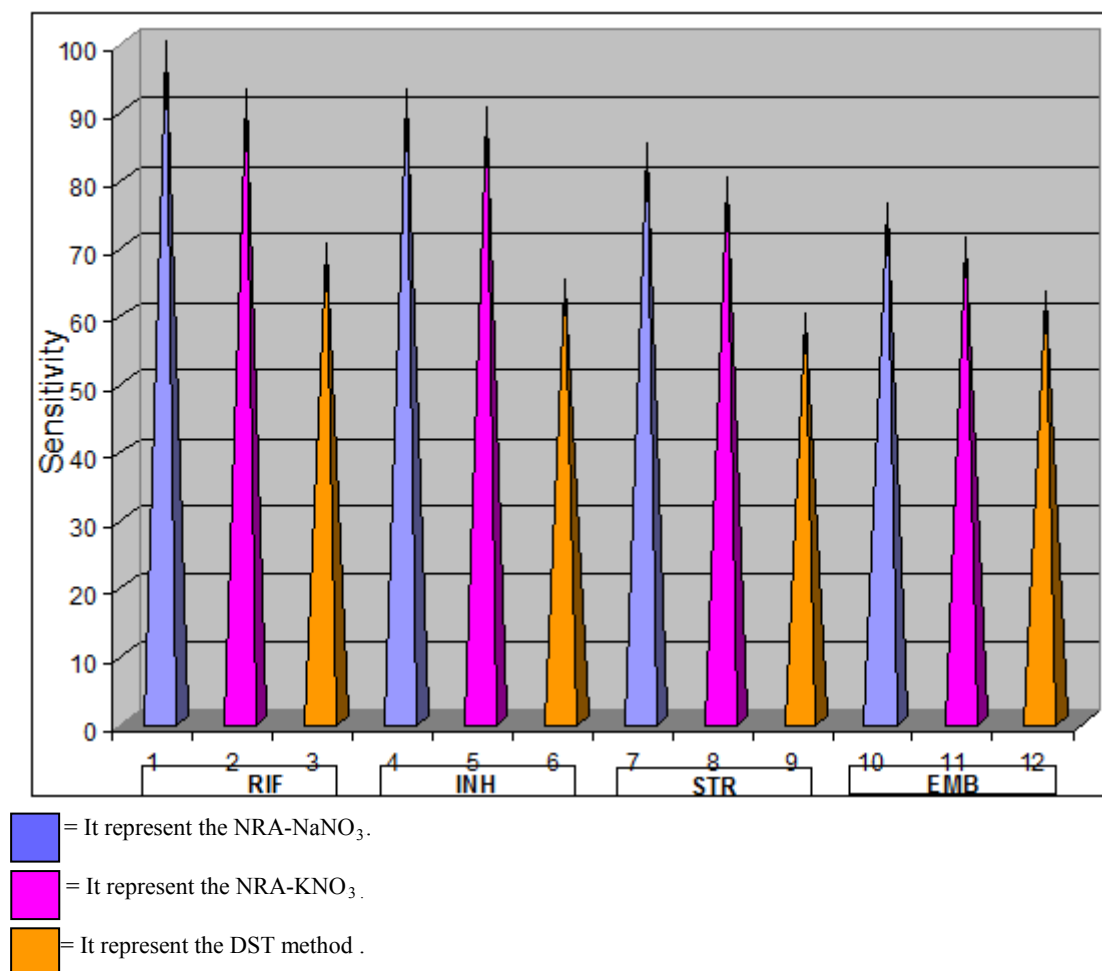


Figure 1. Results the sensitivity of NRA using KNO₃, NaNO₃ compared to DST method.

4. Discussion

To our knowledge, this is the first evaluation of the direct NRA in Iraq. The most worrisome trend during recent years is an increase in multidrug-resistant MDR (for example resistant to RIF and INH) TB strains. Rapid detection of MDR strains is very important to restrict their spread in the population. Current method for DST of MDR-TB are either costly or very slow. So, a cost-effective and rapid drug-susceptibility method is required to guide the treatment of TB (Coban *et al.*, 2004 ; Mishra *et al.*, 2009). Complete agreement between the results of the direct NRA and DST method was found for RIF, which is important since RIF, together with INH, is the most important antituberculosis drug. Resistance to RIF is also almost always associated with multidrug resistance (Vareldzis *et al.*, 1994) and can thus serve as a marker of MDR of *M. tuberculosis* strains if resources are limited.

The direct NRA was comparable to the direct DST method regarding susceptibility testing of INH (sensitivity to detect resistant was 90% with specificity was 97%). In addition, the sensitivity to detect resistance to STR and EMB were low to be acceptable (80% and 71%), but the specificity were 94% and 90% respectively. Results for RIF and INH susceptibility were similar to indirect NRA method (Sethi *et al.*, 2004 ; Musa *et al.*, 2005). These results may be need to adjusting the critical drug

concentrations used in the NRA test, although the susceptibility of *M. tuberculosis* to STR and EMB is more complicated to determine the antibiotic sensitivity (Maire *et al.*, 2012) .

The NRA method utilizes the detection of nitrate reduction as an indication of growth, and therefore, results can be obtained faster than by visual detection of colonies. The ability to reduce nitrate is typical for *M. tuberculosis*, although some other mycobacterial species, like *Mycobacterium kansasii*, and most rapid growers share this characteristic, nitrate reductase-negative strains of *M. tuberculosis* are rare (< 1%)(Rosales *et al.*, 2009).

In the other hand, this study showed that the NRA gave similar results using KNO₃ or NaNO₃ as nitrate source. NRA using NaNO₃ showed high sensitivity and specificity for RIF (100% and 96%) and INH (93% and 100%). These results are in agreement with previous studies presented in a meta-analysis that evaluated the accuracy of the NRA for the detection of MDR .

According to that meta-analysis; the sensitivity and specificity were more than 94% and 92% for RIF and INH (Maire *et al.*, 2012). Another important finding in this study was that 97% of the isolates showed results in 10 days with NRA using NaNO₃ whereas 88% of the isolates gave results in 10 days with NRA using KNO₃ in the previous studies (Coban *et al.*, 2004). Our study suggests the use of NaNO₃ as the source of nitrate for NRA.

References

- Angeby K, Klintz L and Hoffner E. 2002. Rapid and inexpensive drug susceptibility testing of *Mycobacterium tuberculosis* with a nitrate reductase assay. *J Clin Microbiol.*, **40**:553-555.
- Bwanga F, Hoffer S, Haile M and Joloba M. 2009. Direct susceptibility testing for multi-drug resistant tuberculosis: a meta- analysis. *BMC Infect Dis.*, **9**:67-75.
- Canetti G, Fox W, Khownenko A and Grosset J. 1989. Advances in techniques of testing mycobacterial drug sensitivity and the use of sensitivity tests in tuberculosis control programmes. *Bulletin of the World Health Organization*, **41**:21-43.
- Canetti G, Froman F and Grosset J. 1993. Mycobacteria: laboratory methods for testing drug sensitivity and resistance. *Bulletin of the World Health Organization*, **29**: 565-578.
- Coban A, Birinci B, Ekinci A and Durupinar B. 2004. Drug susceptibility testing of *Mycobacterium tuberculosis* with nitrate reductase assay. *Int J Antimicrob Agents*, **24**:304-306.
- Lemus D, Martin A, Montoro E, Portaels F and Palomino J. 2004. Rapid alternative methods for detection of rifampicin resistance in *Mycobacterium tuberculosis*. *J Antimicrob Chemother.*, **54**, 130-133.
- Maira B, Andrea V, Krista F, Juan C, Pedro E and Almeida D. 2012. Nitrate reductase assay using sodium nitrate for rapid detection of multidrug resistance tuberculosis. *Brazilian J Microbiol.*, **43**(3):981-983.
- Martin A, Camacho M, Portaels F and Palomino J. 2003. Resazurin microtiter assay plate testing of *Mycobacterium tuberculosis* susceptibilities to second-line drugs: rapid, simple, and inexpensive method. *Antimicrob Agents Chemother.*, **47**: 3616-3619.
- Mishra B, Muralidharan S and Srinivasa H. 2009. Direct drug susceptibility testing of *Mycobacterium tuberculosis* to primary anti-tubercular drugs by nitrate reductase assay. *Indian J Pathol Microbiol.*, **52**:343-352.
- Moore D. 2000. **The Basic Practice Of Statistics** (2nd ed.). New York : W.H. Freeman and Company.
- Musa H, Ambroggi M, Souto A and Angeby A. 2005. Drug susceptibility testing of *Mycobacterium tuberculosis* by a nitrate reductase assay applied directly on microscopy- positive sputum samples. *J Clin Microbiol.*, **43**:3159-3561.
- Palomino J. 2005. Nonconventional and new methods in the diagnosis of tuberculosis : feasibility and applicability in the field. *Eur Respir J.*, **26**:339-350.
- Rosales S, Pineda L, Andino N, Almendarez N and Hoffner S. 2009. Evaluation of the nitrate reductase assay for rapid detection of extensively drug- resistant tuberculosis. *Int J Tuber Lung Dis.*, **13**:1542-1549.
- Sethi S, Sharma S, Lee K, Meharwai S and Jinadl M. 2004. Drug susceptibility of *Mycobacterium tuberculosis* to primary antitubercular drugs by nitrate reductase assay. *Indian J Med Res.*, **120**: 468-471.
- Simboli N, Takiff H, Mc Nerney R, Lopez B, Martin M and Palomino J. 2005. In-house phage amplification assay is a sound alternative for detecting rifampin- resistant *Mycobacterium tuberculosis* in low- resource settings. *Antimicrob Agents Chemother.*, **49**: 425-427.
- Solis L, Shin S, Han F, Lianos M and Sloutsky A. 2010. Validation of a rapid method for detection of *M. tuberculosis* to isoniazid and rifampin in Lima, Peru. *Int J Tubere Lung Dis.*, **9**:760-764.
- Vareldzis B, Grosset J, Dekantor J, Crofton A, Laszio M and Felten M. 1994. Drug- resistant tuberculosis : laboratory issues. World Health Organization recommendation. *Tuber Lung Dis.*, **75**:1-7.
- World Health Organization. 1998. Laboratory services in tuberculosis control. Part II . Microscopy. WHO, Geneva, Switzerland.
- World Health Organization. 2010 a. Multidrug and Extensively Drug- Resistant TB(M/XDR –TB) 2010 Global Report on surveillance and response. Geneva : World Health Organization.
- World Health Organization. 2010 b. The WHO / IUATLD global project on anti-tuberculosis drug resistance surveillance. Anti-tuberculosis drug resistance in the world, report no 2: prevalence and trends. World Health Organization, Geneva, Switzerland.

Protective Effects of *Enantia chlorantha* Stem Bark Extracts on Acetaminophen Induced Liver Damage in Rats

Olamide E. Adebisi* and Mathew O. Abatan

Department of Veterinary Physiology, Biochemistry and Pharmacology, University of Ibadan,

P.O. Box 28112, Agodi, Ibadan, Oyo State, Nigeria

Received: April 23, 2013

Revised: June 4, 2013

Accepted: July 10, 2013

Abstract

The study was designed to evaluate the hepatoprotective activity of different solvent extracts (hexane, chloroform, ethyl acetate and methanol) of *Enantia chlorantha* stem bark in acute experimental liver injury induced by acetaminophen. The effects observed were compared with a known hepatoprotective agent, silymarin (100 mg/kg p.o.). Preliminary phytochemical tests and acute toxicity study were done. The degree of hepatoprotection was measured using serum transaminases (AST and ALT), alkaline phosphatase, bilirubin, albumin, and total protein levels. In the acute liver damage induced by acetaminophen, *E. chlorantha* stem bark extracts (200 mg/kg, p.o.) significantly reduced the elevated serum levels of aspartate aminotransferase, alanine aminotransferase, alkaline phosphatase and bilirubin in acetaminophen induced hepatotoxicity. The total serum protein was significantly increased ($P < 0.05$) by all the solvent extracts. Histological examination of the liver tissues supported the hepatoprotection. Our findings suggested that *E. chlorantha* stem bark extracts possessed hepatoprotective activity; the hexane extract of stem bark of *E. chlorantha* plant possesses better hepatoprotective activity compared to other extracts.

Keywords: Hepatoprotection; Acetaminophen; Liver; Stem Bark Extracts; *Enantia chlorantha*

1. Introduction

The liver is the most important organ in the body. The liver plays a pivotal role in regulating various physiological processes (Rajib *et al.*, 2009). It is the centre of metabolism of nutrients such as carbohydrates, proteins and lipids. It is also involved in the metabolism and excretion of waste metabolites, drugs and other xenobiotics from the body thereby providing protection against foreign substances by detoxifying and eliminating them (Mohamed *et al.*, 2010). As a result of this, the liver is exposed to all types of toxic abuse from both endogenous and exogenous substances which may produce liver degeneration.

Liver diseases have become one of the major causes of morbidity and mortality in man and animals and hepatotoxicity due to drugs appears to be the most common contributing factor (Russmann *et al.*, 2009). For instance, drug-induced liver injury accounts for at least 13% of acute liver failure cases in the United States (Au *et al.*, 2011). The manifestations of drug-induced hepatotoxicity are highly variable, ranging from asymptomatic elevation of liver enzymes to fulminant

hepatic failure. Acetaminophen also known as paracetamol, taken in overdose can cause severe hepatotoxicity and nephrotoxicity (Yakubu *et al.*, 2008). In spite of the tremendous advances in modern medicine, there is no effective drug available that stimulates liver function, offers protection to the liver from damage or helps to regenerate hepatic cells (Chaudhary, 2010).

Medicinal plants play a key role in human and animal health care. About 80% of the world population relies on the use of traditional medicine, which is predominantly based on plant material (WHO, 1993). Despite the significant popularity of several herbal medicines in general, and for liver diseases in particular, they are still unacceptable treatment modalities for liver diseases due to lack of standardization of the herbal drugs, lack of identification of active ingredient(s)/principles(s), lack of randomized controlled clinical trials (RCTs) and lack of toxicological evaluation (Radha & Yogesh, 2005). Therefore, due importance has been given globally to develop plant-based hepatoprotective drugs effective against a variety of liver disorders.

There are numerous plants and traditional formulations available for the treatment of liver diseases. *Silybum marianum*, *Orthosiphon stamineus* and *Foeniculum*

* Corresponding author. e-mail: olamideadebiyi24@gmail.com.

vulgare are amongst natural products whose hepatoprotective effects have been investigated and documented (Hanefi *et al.*, 2004; Pradhan and Girish, 2006; Subramanian *et al.*, 2006).

Enantia chlorantha Oliv (family-Annonaceae) common name-African Yellow Wood is widely distributed along the coasts of West and Central Africa and also very common in the forest regions of Nigeria (Adesokan *et al.*, 2007).

Several studies have shown that the stem bark of *E. chlorantha* possesses wide spectrum antimicrobial and antimalarial (Adesokan *et al.*, 2007; Odugbemi *et al.*, 2007) activities. In Cameroon, the stem bark extract is also used to treat jaundice, urinary tract infections (Adjanooun *et al.*, 1996), hypoglycaemia, typhoid fever (FAO, 2001). The stem bark is also used for treating leprosy spots, as haemostatic agent and uterus stimulant (Gill, 1992). An anti-sickling compound has also been isolated from the ethanolic extract of the plant (Ejele *et al.*, 2012).

Despite its numerous medicinal uses and importance in the treatment of many illness and diseases in Africa, to our knowledge, no concrete scientific study has been reported to prove the folklore claim of the utility of *E. chlorantha* in the treatment of liver diseases and hence one of the objectives of the present study was to correlate the ethnobotanical evidence with scientific study. Further, the study also attempts to evaluate *in vivo* hepatoprotective and curative effects of stem bark extracts of *E. chlorantha* on acetaminophen induced hepatotoxicity models in rats using solvents of various polarities.

2. Materials and Methods

2.1. Plant Material and Authentication

The plant samples were collected from local region between September and October (rainy season), 2012. The plant was identified and authenticated at the Forestry Research Institute of Nigeria, Ibadan and voucher specimen (FHI. 109950) was preserved at the herbarium.

2.2. Plant Materials Extraction

Stem barks of *E. chlorantha* were dried under shade for 7 days until a constant weight was obtained. This was ground into powder using an electric blender (Blender/Miller III, model MS-223, Taiwan, China). The powder was packed into Soxhlet column and extracted with hexane. The same material was successively extracted with chloroform, ethyl acetate and methanol. The solvents were filtered, squeezed off and evaporated off under reduced pressure in a rotary evaporator to obtain the crude extract. After concentrated preparation, the dried powder extract was stored at 4°C (Prakash *et al.*, 2008).

2.3. Experimental Animals

Thirty five wistar albino rats (*Rattus norvegicus*) consisting of both male and female with average weight of 150-200 g were obtained from the Animal Holding Unit of the Department of Veterinary Physiology, Biochemistry and Pharmacology, Faculty of Veterinary Medicine, University of Ibadan, Nigeria. The animals

were allowed free access to feed and fresh water *ad libitum*. All the animals were acclimatized to laboratory conditions for two weeks before commencement of the experiment. The study was approved by the Animal Ethics Committee of the Faculty of Veterinary Medicine, University of Ibadan.

2.4. Drugs and Chemicals

Silymarin (Micro labs, Tamilnadu, India), ethanol, hexane, chloroform, ethyl acetate, methanol and the rest of the chemicals utilized were of analytical grade and were prepared in all glass distilled water. Acetaminophen (Emzor Paracetamol[®]) was purchased over the counter.

2.5. Phytochemical Screening

The qualitative methods already established to test for classes of compounds in plant extracts by Ciulei (1964) and Chitravadivu *et al.* (2009) were used. The substances that were tested for included: phenolics, alkaloids, steroids, tannins, flavonoids, saponins, glycosides and phlebotanins. The hexane, chloroform, ethyl acetate and methanol extracts of *Enantia chlorantha* stem bark were used to determine the compounds.

2.6. Acute Toxicity Study

This study was conducted according to the Organisation for Economic Cooperation and Development's (OECD) revised up and down procedure for acute toxicity testing (OECD, 2001). Animals were divided into eight groups of five rats each. The control group received distilled water (10 ml/kg) groups II-VIII received 100, 200, 400, 800, 1000, 2000 or 3000 mg/kg of ethanol extract of *Enantia chlorantha* stem bark orally in a single dose. Immediately after dosing, the rats were observed for mortality and clinical signs for the first hour, then hourly for three hours and then periodically for 72 hours and then kept for up to 14 days post-treatment in order to observe for any toxic symptoms and mortality.

2.7. Experimental Design

Animals were randomly divided into seven groups (I-VII) of five animals per group. Group I (normal control) received neither the plant extract nor acetaminophen for 8 days. Group II (negative control group) Induction of hepatotoxicity using acetaminophen: The animals received distilled water for 7 days and were administered acetaminophen (500 mg/kg) orally on day 8. Group III (positive control group) – pre-treatment with silymarin (100 mg/kg) for 7 days (p.o) followed by a single dose of acetaminophen on day 8. Groups IV, V, VI and VII – Pre-treatment with hexane, chloroform, ethyl acetate and methanol extract of *Enantia chlorantha* stem bark respectively at 200 mg/kg for 7 days (p.o) followed by a single dose of acetaminophen on day 8.

During the period of drug treatment the rats were fed *ad libitum* with standard pellet diet and had free access to water. The biochemical parameters were estimated after 24 hours following the administration of acetaminophen.

2.7.1. Serum Biochemical Analyses

Blood was obtained from all animals by puncturing retro-orbital plexus. The blood samples were allowed to coagulate and then serum was separated by centrifuging at 3000 rpm for 20 min, collected into sterilized tubes and

stored at -20 °C. Serum biochemical parameters were analyzed: Aspartate aminotransferase (AST) (Reitman and Frankel, 1957), alanine aminotransferase (ALT) (Reitman and Frankel, 1957), alkaline phosphatase (ALP) (Kind and King, 1954), serum bilirubin (Mallay and Evelyn, 1937), total protein, albumin, blood urea nitrogen (BUN) using RANDOX[®] laboratory reagent kits (RANDOX[®] Laboratories Ltd., Ardmore, United Kingdom).

2.7.2. Histopathological Examination

After collection of blood samples the rats in different groups were sacrificed. 3-5 mm samples of the liver tissue were collected and placed in 10% formaldehyde solution for histopathological study. The pieces of liver were processed and embedded in paraffin wax and sections were made about 4-6 µm in thickness. After staining with haematoxylin and eosin (H&E), slides were examined under microscope (Olympus, Japan) for histopathological changes and photographed.

2.8. Statistical Analysis

All data were expressed as mean ± standard error of mean (SEM), comparison was by the student t test using Graphpad Prism version 4.00 for Windows, Graphpad Software. Significance was reported at $P < 0.05$.

3. Results

Phytochemical screening of the extracts of *E. chlorantha* stem bark revealed the presence of phenolics, flavonoids, alkaloids, glycosides and saponins (Table 1).

All four extracts of *E. chlorantha* stem bark at a dose of 3000 mg/kg *p.o.*, did not produce any mortality in the rats during the pilot acute toxicity study.

There was a significant ($P < 0.05$) increase in the level of serum total protein (TP) in the test groups when compared with the negative control/ untreated group (distilled water). Rats pre-treated with HX, CH, EE and ME extracts of *E. chlorantha* stem bark showed an insignificant increase ($P > 0.05$) in the serum albumin contents when compared with the untreated group. Groups pretreated with HX, CH, EE and ME extracts of

E. chlorantha stem bark showed a significant increase in the serum globulin contents when compared with the negative control/ untreated group.

The test groups had a significant decrease ($P < 0.05$) in activities of AST, ALT and ALP when compared with the negative control/ untreated group. The extracts (HX, CH, EE and ME) also decreased significantly the activities of ALT and ALP relative to silymarin (positive control group) (Table 2).

The results reported in table 2 also showed that groups treated with HX, CH, EE and ME extracts of *E. chlorantha* stem bark had significantly decreased ($P < 0.05$) levels of serum bilirubin when compared with the negative control/ untreated group.

Liver sections from acetaminophen treated rats showed vacuolar degeneration and different stages of necrotic alterations in the hepatocytes surrounding the central veins. Focal mononuclear leucocytes inflammatory cells infiltration was observed in between the degenerated and necrotic hepatocytes, as well as in the portal area. There was marked congestion of portal vessels and central vein (Figure 1). Liver sections from rats pre-treated with silymarin showed diffused proliferation of Kupffer cells between the hepatocytes associated with dilatation in the portal vein and inflammatory cells infiltration in the portal area (Figure 2).

Liver of rats pre-treated with hexane extract of *E. chlorantha* prior to acetaminophen administration showed diffused proliferation of the Kupffer cells between the hepatocytes, associated with dilatation in the central vein, and focal inflammatory cells infiltration in the hepatic parenchyma (Figure 3). The liver of rats given chloroform extract of *E. chlorantha* before administration of acetaminophen showed dilatation in the central and portal veins with newly formed bile ductules, oedema and inflammatory cells infiltration in the portal area. Focal haemorrhage was noticed in the hepatic parenchyma (Figure 4). The liver of rats dosed with ethyl acetate and methanol extract of *E. chlorantha* stem bark showed no visible histopathological lesion (Figure 5 and 6, respectively).

Table 1. Phytochemical constituents of hexane, chloroform, ethyl acetate and methanol extracts of *Enantia chlorantha* stem bark.

Phytochemical	Hexane	Chloroform	Ethyl acetate	Methanol
Phenolics	-	+	++	++
Flavonoids	+	+	+	+
Alkaloids	++	++	++	++
Glycosides	±	+	±	+
Saponins	++	++	++	++
Tannins	-	-	-	-
Phlebotanins	-	-	-	-
Steroids	-	-	-	-

++ = Strongly positive, + = positive, ± = weakly positive, and - = not detected

Table 2. Serum biochemical values of rats administered with hexane, chloroform, ethyl acetate, methanol extracts of *E. Chlorantha* stem bark and the control groups

	Group I	Group II	Group III	Group IV	Group V	Group VI	Group VII
TP (g/dL)	2.75 ± 0.15 ^a	1.28 ± 0.09 ^b	1.88 ± 0.23 ^c	2.65 ± 0.22 ^a	1.84 ± 0.02 ^c	1.46 ± 0.04 ^b	1.98 ± 0.19 ^a
ALB (g/dL)	0.85 ± 0.07 ^a	0.43 ± 0.09 ^b	0.72 ± 0.09 ^a	0.45 ± 0.05 ^b	0.80 ± 0.20 ^a	0.44 ± 0.08 ^b	0.88 ± 0.03 ^a
GLB (g/dL)	1.90 ± 0.18 ^a	0.85 ± 0.05 ^b	1.16 ± 0.09 ^c	2.20 ± 0.17 ^a	1.04 ± 0.02 ^c	1.02 ± 0.10 ^c	1.10 ± 0.08 ^c
AST (U/L)	35.46 ± 2.81 ^a	56.00 ± 1.33 ^b	38.50 ± 2.26 ^a	34.50 ± 0.63 ^a	43.04 ± 0.60 ^c	35.20 ± 4.50 ^a	41.50 ± 1.71 ^c
ALT (U/L)	41.39 ± 2.61 ^a	75.25 ± 1.80 ^b	59.75 ± 2.40 ^c	42.25 ± 1.03 ^a	55.00 ± 1.41 ^c	46.00 ± 7.48 ^a	55.25 ± 3.35 ^c
ALP (U/L)	35.61 ± 3.11 ^a	72.25 ± 0.85 ^b	46.25 ± 6.76 ^b	34.00 ± 2.94 ^a	30.00 ± 2.97 ^a	38.40 ± 4.35 ^a	46.75 ± 9.16 ^b
BIL(mg/dl)	6.56 ± 0.34 ^a	9.92 ± 0.17 ^b	8.57 ± 0.06 ^b	6.52 ± 0.36 ^a	9.05 ± 0.15 ^b	6.96 ± 0.94 ^a	7.95 ± 0.20 ^c

Values are expressed as mean ± SEM (n= 5 mice/ group).

TP- total protein

ALB- albumin

GLB- globulin

AST- aspartate aminotransferase

ALT- alanine aminotransferase

ALP- alkaline phosphatase

BIL- bilirubin

Means with different superscripts within rows are significantly different at $P < 0.05$

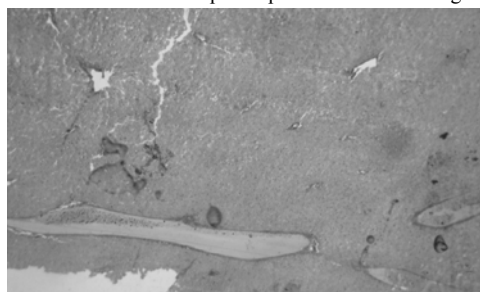


Figure 1. Shows vacuolar degeneration, necrotic hepatocytes, marked congestion of portal vessels and central vein. H & E stain (x100)

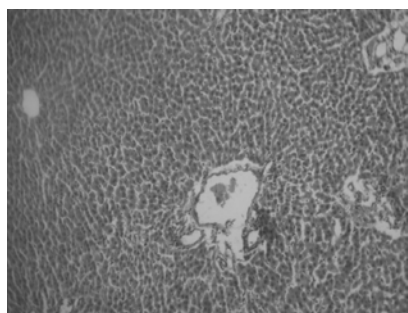


Figure 2. Shows diffused proliferation of Kupffer cells between the hepatocytes inflammatory cells infiltration. H & E stain (x100)

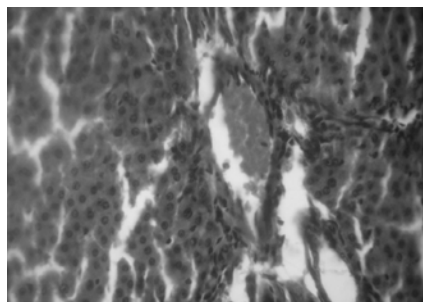


Figure 3. Shows diffused proliferation of Kupffer cells between the hepatocytes dilatation in the central vein and inflammatory cells infiltration in the hepatic parenchyma. H & E stain (x100).

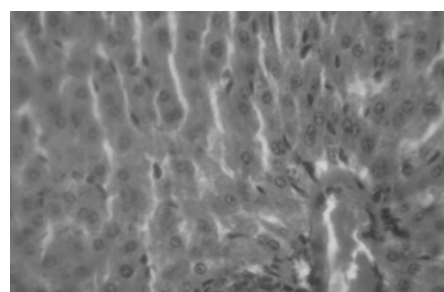


Figure 4. Dilatation in the central and portal veins with newly formed bile ductules, oedema and inflammatory cells infiltration in the portal area. Focal haemorrhage was noticed in the hepatic parenchyma (H & E stain(x100).

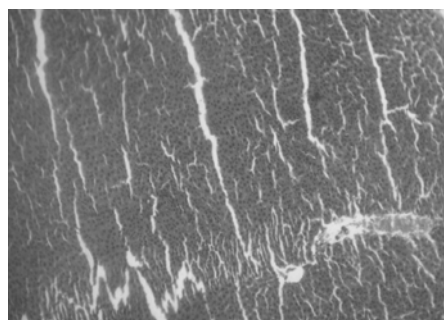


Figure 5. Shows no visible histopathological lesion. (H & E stain) x 100.

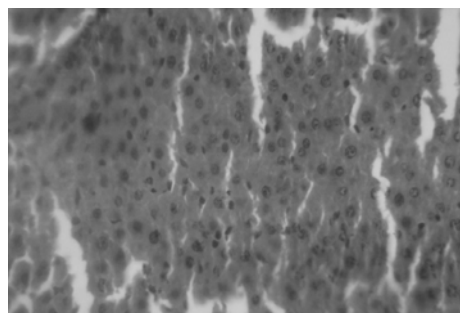


Figure 6. Shows no visible histopathological lesion. H & E stain (100x).

4. Discussion

The stem bark extracts of *E. chlorantha* was found to contain phenolics, flavonoids, alkaloids, glycosides and saponins. These are secondary metabolites which have been reported to cure a lot of diseases (Dongmo *et al*, 2007; Suman *et al*, 2011). The fact that methanol exhibited the strongest reactions; thus being able to extract more phytochemicals, could mean that there are more non-polar phytochemicals in the stem bark.

Acetaminophen is a common antipyretic agent, which is safe in therapeutic doses but can produce fatal hepatic necrosis in man, rats and mice with toxic doses (Dash *et al*, 2007). Protection against acetaminophen-induced toxicity has been used as a test for potential hepatoprotective activity by several investigators (Sabir & Rocha, 2008; Parmar *et al*, 2010).

Proteins are important organic constituents of the animal cells playing a vital role in the process of interactions between intra and extra cellular media (Waqar *et al*, 2004). In the present study, the untreated/ negative control group administered with acetaminophen showed a decrease in the level of serum total protein (TP) while pre-treatment with solvent extracts of *E. chlorantha* (500mg/kg) showed an increase in the level of serum protein. Being a part of cell membrane and as an enzyme, protein helps to balance sub cellular fractions. Protein and amino acids are also very important nutrients and they play a major role in the synthesis of microsomal detoxifying enzymes which help detoxify toxicants that enter into the animal's body (Abubakar *et al*, 2010). The reduction in the protein levels in the untreated/ negative control group might thus be as a result of their metabolism to liberate energy during acetaminophen toxicity. The liver is also an important site for the synthesis of many serum proteins (Ahsan *et al*, 2009). The reduction in serum total protein observed in the acetaminophen group may also be associated with decrease in the number of hepatocytes which consequently results in decreased hepatic capacity to synthesize protein. Pre-treatment with HX, CH, EE and ME extracts of *E. chlorantha* stem bark significantly increased TP indicating the hepatoprotective activity of the extracts most probably through hepatic cell regeneration (Olorunnisola *et al*, 2011). These results are in line with the report by Manokaran *et al*, (2008) that oral administration of *Aerva lanata* to acetaminophen treated rats showed increased serum protein level when compared to acetaminophen alone treated rats. Similarly oral administration of hydro-ethanolic extract (70%) of *Calotropis procera* flowers to acetaminophen treated rats showed significantly increased serum protein level (Setty *et al*, 2007). The highest increase in serum total protein content was noticed in the rats treated with the HX extract (this group had more than a two-fold increase) of *E. chlorantha* stem bark and this increase was statistically significant when compared with the acetaminophen treated rats. This also explains the corresponding reduction and increases in albumin in the untreated group and the test groups as about 60% of total serum protein is albumin (Musa *et al*, 2005).

It has been reported in several studies that liver enzymes are liberated into the blood whenever liver cells are damaged and enzyme activity in the plasma is increased (Chang, 2009). Thus ALT, AST and ALP activity and serum bilirubin level are largely used as most common biochemical markers to evaluate liver injury (Ajayi *et al*, 2009). Elevation of these liver enzymes is also associated with cell necrosis of many tissues especially the liver (Adedapo *et al*, 2004). The current study also confirmed these effects of acetaminophen overdose toxicity, as indicated by marked increases in serum hepatic enzymes in the control/ untreated group. This is in consonance with the findings of Vadivu *et al*, (2008) who stated that acetaminophen causes liver damage in rats and significantly ($P < 0.05$) increased the AST and ALT levels in serum when compared with silymarin which has a remarkable protection of serum AST and ALT levels towards acetaminophen induced hepatotoxicity. The significant ($P < 0.05$) decrease in activities of these enzymes by the extracts may indicate that the plant extracts did not have necrotic effect on the liver. This may be due to the fact that the extracts offer protection and maintain the functional integrity of hepatic cells. The protective effect may be the result of stabilization of plasma membrane thereby preserving the structural integrity of cell as well as the repair of hepatic tissue damage caused by acetaminophen (Murugaian *et al*, 2008). The increased ALP concentration following acetaminophen administration is in line with existing literature that ALP synthesis is increased by cells lining bile canaliculi usually in response to cholestasis and increased biliary pressure (Gaw *et al*, 1999). Increased level was obtained after acetaminophen administration and it was brought to near normal level by *E. chlorantha* treatment. This further signifies the curative nature of the extract against acetaminophen toxicity.

Serum bilirubin is considered an index for the assessment of hepatic function and any abnormal increase indicates hepatobiliary disease and severe disturbance of hepatocellular architecture (Martin and Friedman, 1992). Acetaminophen administration resulted in increased serum bilirubin level, (Table 2) thereby suggesting severe hepatic injury and confirming the hepatotoxic nature of acetaminophen. Treatment with *E. chlorantha* stem bark extracts significantly decreased the elevated level of total bilirubin in serum towards normalcy indicating its hepatoprotective efficacy. The hexane extracts demonstrated the highest potency in this regard.

Liver of rats administered with acetaminophen showed severe necrosis, with disappearance of nuclei. This could be due to the formation of highly reactive radicals because of oxidative threat caused by acetaminophen (Shardul, 2010). Histopathological changes of the group pre-treated with the extracts showed significant improvement in architecture. Pretreatment with the ethyl acetate and methanol extracts restored the hepatic architecture and protected the liver tissue from fatty and degenerative changes, by preventing the toxic chemical reaction. Although, necrotic changes were still evident in the liver of rats

pre-treated with the hexane and chloroform extracts, the severity of the damage was less intense significantly. The various phytoconstituents of the stem bark extracts of *E. chlorantha* might be helpful in the changes in the membrane, in the mitochondria or at the ionic level like calcium (Rang *et al.*, 2003). The extracts of *E. chlorantha* stem bark may have a role in the process of regeneration and prevention of fibrosis. However, our study has shown the centrilobular necrosis by acetaminophen and prevention of such changes and restoration to normalcy in the centrilobular area by extracts of *E. chlorantha* stem bark.

The possible mechanism responsible for the protection of the acetaminophen induced liver damage by the extract of *E. chlorantha* maybe a result of the extract acting as a free radical scavenger by intercepting the radicals involved in acetaminophen metabolism by microsomal enzymes or the phytochemicals constituents of the plant because a number of scientific reports indicate the role of certain flavonoids and steroids in hepatoprotection against hepatotoxins. The presence of these compounds in *E. chlorantha* may be responsible for the protective effect on acetaminophen induced liver damage in rats.

5. Conclusion

Based on the above results, it could be concluded that hexane, chloroform, ethyl acetate and methanol extracts of *Enantia chlorantha* stem bark exert significant hepatoprotection against acetaminophen-induced toxicity.

The hexane extract of *E. chlorantha* showed better hepatoprotective activity in acetaminophen induced liver damage compared to chloroform, ethyl acetate and methanolic extract(s) as indicated by maximum prevention of increased serum biochemical parameters.

The stem bark extract of *E. chlorantha* extract may be hepatoprotective.

References

- Abubakar MG, Lawal A and Usman MR. 2010. Hepatotoxicity studies of sub-chronic administration of aqueous stem bark of *Khaya senegalensis* in albino rats. *Bayero J Pure and Applied Sci.*, **3**(1): 26 – 28.
- Adedapo AA, Abatan MO and Olorunsogo OO. 2004. Toxic effects of some plants in the genus *Euphorbia* on haematological and biochemical parameters of rats. *Veterinary Archives*, **74**(1): 53-62.
- Adesokan AA, Akanji MA and Yakubu MT. 2007. Antibacterial potentials of aqueous extract of *Enantia chlorantha* stem bark. *Afr J Biotechnol.*, **6**(22): 2502-2505.
- Adjanohoun JE, Aboobakar N and Dramane K. 1996. Traditional Medicine and Pharmacopoeia: Contribution to Ethnobotanical and Floristic Studies in Cameroon, Porto Novo, Benin. Organisation of African Unity Scientific Technical and Research Commission. Centre National de Production de Mannels Scolaires.
- Ajayi GO, Adeniyi TT and Babayemi DO. 2009. Hepatoprotective and some haematological effects of *Allium sativum* and vitamin C in lead exposed wistar rats. *Inter J Medicine and Medical Sci.*, **1**(3): 064-067.
- Ahsan MR, Islam KM and Bulbul IJ. 2009. Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride-induced hepatotoxicity in rats. *Europ JScientific Res.*, **37**(2): 302-310.
- Au JS, Navarro VJ and Rossi S. 2011. Drug-induced liver injury- its pathophysiology and evolving diagnostic tools. *Alimentary Pharmacol Therapeutics*, **34**(1): 11-20.
- Chang S. 2009. Liver Enzymes [online]. Available from URL: <http://www.LiverEnzymesonRightHealth.html> (cited on 11th Feb 2013)
- Chaudhary GD, Kamboj P, Singh I and Kalia AN. 2010. Herbs as liver savers- A review. *Indian J Natural Products and Resources*, **1**(4): 397-408.
- Chitravadivu C, Manian S and Kalachelvi K. 2009. Qualitative analysis of selected medicinal plants, Tamilnadu, India. *Middle East J Sci Res.*, **4**: 144-146.
- Ciulei I. 1964. **Practical Manuals on the Industrial Utilization of Medicinal and Aromatic Plants**, University of Bucharest, Romania.
- Dash DK, Yeligar VC, Nayak SS, Ghosh T, Rajalingam D and Sengupta P. 2007. Evaluation of hepatoprotective and antioxidant activity of *Ichnocarpus frutescens* (Linn.) on paracetamol-induced hepatotoxicity in rats. *Tropical J Pharmaceutical Res.*, **6**: 755-765.
- Dongmo F, Julius EO, Claudia ENM and Damaris EM. 2007. Phytochemical constituents and antioxidant potential of some Cameroonian medicinal plants *Pharmacol online*, **2**: 436-452.
- Ejele AE, Akpan IO, Ogukwe CE, Onyechoa VO and Ukiwe LN. 2012. Bioassay-guided isolation and partial characterization of an antisickling compound from *Enantia chlorantha*. *Inter Res J BiochemBioinformatics*, **2**(7): 149-154.
- Food and Agriculture Organization (FAO) 2001. Collecte et analyse de données pour l'aménagement durable des forêts - joindre les efforts nationaux et internationaux. Programme de partenariat CE-FAO (1998-2001). Données statistiques des produits forestiers non-ligneux du Cameroun. pp. 36
- Gaw A, Cowan, RA, O'Reilly DSJ, Stewart MJ and Shepherd J. 1999. **Clinical Biochemistry – An Illustrated Color Text**. 1st ed. New York: Churchill Livingstone; pp. 51-53
- Gill LS. 1992. **Ethnomedical Uses of Plants in Nigeria**. Uniben Press Benin Nigeria. pp.143.
- Gupta A, Chitme H, Dass SK and Misra N. 2006. Hepatoprotective activity of *Rauwolfia serpentina* rhizome in paracetamol intoxicated rats. *J Pharmacol Toxicol.*, **1**: 82-88.
- Hanefi O, Serdar U, Irfan, Ismail U, Ender E, Abdurrahman O and Zübeyir H. 2004. Hepatoprotective effect of *Foeniculum vulgare* essential oil: A carbon-tetrachloride induced liver fibrosis model in rats. *Scand J Lab Anim Sci.*, **31**: 9-17.
- Kind PRN and King E. 1954. Estimation of plasma phosphatase by determination of hydrolysed phenol with amino antipyrine. *J Clin Pathol.*, **7**: 322.
- Maheswari C, Maryammal R and Venkatanarayanan R. 2008. Hepatoprotective activity of "*Orthosiphon stamineus*" on liver damage caused by paracetamol in rats. *Jordan J Biol Sc.*, **1**(3): 105 -108
- Mallay HT and Evelyn KA. 1937. Estimation of serum bilirubin level with the photoelectric colorimeter. *J Biological Chem.*, **119**: 481-484.

- Manokaran S, Jaswanth A, Sengottuvelu S, Nandhakumar J, Duraisamy R, Karthikeyan D and Mallegaswari R. 2008. Hepatoprotective activity of *Aerva lanata* Linn. against paracetamol induced hepatotoxicity in rats. *Res J Pharmacy and Technol.*, **1**: 398-400.
- Martin P and Friedman LS. 1992. Assessment of liver function and diagnostic studies. In: Freidman L S and Keefe E B (Eds.), **Hand Book of Liver Disease**. Churchill Livingstone, Philadelphia, pp.1-14.
- Mohamed TS, Madhusudhana C, Ramkanth S, Rajan VST, Mahesh K and Gauthaman K. 2010. Hepatoprotective Herbs. *Inter J Res Pharmaceutical Sci.*, **1**(1): 1-5.
- Murugaian P, Ramamurthy V and Karmegam N. 2008. Hepatoprotective Activity of *Wedelia calendulacea* L. against Acute Hepatotoxicity in Rats. *Res J Agriculture and Biol Sci.*, **4**(6): 685-87.
- Musa TY, Adebayo OJ, Egwim EC and Owoyele VB. 2005. Increased liver alkaline phosphatase and amino transferases activities following administration of ethanolic extract of *Khaya senegalensis* stem bark to rats. *Biochem.*, **17**(1): 27-32.
- OECD. 2001. OECD Guideline 425: Acute Oral Toxicity—Up-and-Down Procedure. In: *OECD Guidelines for the Testing of Chemicals* Vol.2 Organization for Economic Cooperation and Development. Paris, France.
- Odugbemi TO, Odunayo R, Akinsulire I, Aibinu E and Fabeku O. 2007. Medicinal plants useful for malaria therapy In Okeigbo, Ondo State, Southwest Nigeria. *Afr J Traditional, Complementary and Alternative Med.*, **4**(2): 191- 198.
- Olorunnisola OS, Bradley G and Afolayan AJ. 2011. Antioxidant properties and cytotoxicity evaluation of methanolic extract of dried and fresh rhizomes of *Tulbaghia violacea*. *Afr J Pharmacy and Pharmacol.*, **5**: 2490–2497.
- Parmar SR, Patel HV and Kiran K. 2010. Hepatoprotective activity of some plants extract against paracetamol induced hepatotoxicity in rats. *J Herbal Med and Toxicol.*, **4**(2): 101-106
- Pradhan SC and Girish C. 2006. Hepatoprotective herbal drug, silymarin from experimental pharmacology to clinical medicine. *Indian J Med Res.*, **124**: 491-504
- Prakash T, Snehal DF, Uday RS, Surendra V, Divakar G, Perfect S and Kotresha D. 2008. Hepatoprotective activity of leaves of *Rhododendron arboreum* in CCl₄ induced hepatotoxicity in rats. *J Med Plants Res.*, **2**(11): 315-320.
- Radha KD and Yogesh KC. 2005. Herbal medicines for liver diseases. *Digestive Dis Sci.*, **50**(10): 1807–1812.
- Rajib A, Islam KM, Musaddik A and Haque E. 2009. Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride induced hepatotoxicity in albino rats. *Global J Pharmacol.*, **3**(3): 116-122.
- Rang HP, Dale MM, Ritter JM and Moore PK. 2003. Harmful effects of drugs. In: **Pharmacology**, 5th ed. Churchill Livingstone. Edinburgh. pp. 724-37.
- Reitman S and Frankel SA. 1957. Colorimetric method for the determination of serum glutamic Oxaloacetic and glutamic pyruvic transaminase. *American J Clin Pathol.*, **28**: 56 – 63.
- Russmann S, Gerd A and Grattagliano I. 2009. Current concepts of mechanism in drug induced hepatotoxicity. *Curr Med Chem.*, **16**: 3041-3053.
- Sabir SM and Rocha JBT. 2008. Water-extractable phytochemicals from *Phyllanthus niruri* exhibit distinct in vitro antioxidant and in vivo hepatoprotective activity against paracetamol-induced liver damage in mice. *Food Chem.*, **111**(4): 845-851.
- Setty SR, Quereshi AA, Viswanath Swamy AHM, Patil T, Prakash T, Prabhu K and Veeran GA. 2007. Hepatoprotective activity of *Calotropis procera* flowers against paracetamol induced hepatic injury in rats. *Fitoterapia*. **78**: 451- 454.
- Shardul SW. 2010. Antioxidant and hepatoprotective activity of *Tridax procumbens* Linn, against paracetamol induced hepatotoxicity in male albino rats. *Advanced Studies in Biol.*, **2**(3): 105 – 112.
- Subramanian K, Nalini R, Palani G, Elango V and Carani VA. 2006. Fenugreek (*Trigonella foenum graecum*) seed extract prevents ethanol-induced toxicity and apoptosis in change liver cells. *Alcohol & Alcoholism* **41**(3): 267–273
- Suman P, Siva SN, Durga PP, Subas CD, Vikas S and Amol J. 2011. Hepatoprotective activity of crude flavonoids extract of *Cajanus scarabaeoides* (L) in paracetamol intoxicated albino rats. *Asian J Pharmaceutical and Biol Res.*, **1**(1): 22-27
- Vadivu R, Krithika A, Biplab C, Dedeepya P, Shoeb N and Lakshmi KS. 2008. Evaluation of hepatoprotective activity of the fruits of *Coccinia grandis* Linn. *Inter J Health Res.*, **1**(3): 163-168.
- Waqar A, Saad A, Khalid A, Haleema S, Tariq R and Luqman M. 2004. Diagnostic significance of serum protein electrophoresis. *Biomedica*, **20**: 40-44.
- World Health Organisation (WHO) 1993. Regional Office for Western Pacific, research guidelines for evaluating the safety and efficacy of herbal medicines, Manila.
- Yakubu MT, Akanji MA and Oladiji AT. 2008. Alterations in serum lipid profile of male rats by oral administration of aqueous extract of *Fadogia argrestis* stem. *Res J Medicinal Plant*, **2**: 66-73.

Odonata of Wadi Al Mujib Catchment with Notes on the Impact of Wadi Al Mujib Dam, Jordan (Insecta: Odonata)

Zuhair S. Amr^{1,*}, Loay S. Al Azzam², Ahmad Katbeh-Bader³ and Ehab K. Eid²

¹ Department of Biology, Jordan University of Science and Technology, P. O. Box 3030, Irbid,

² The Royal Society for the Conservation of Nature, Amman,

³ Faculty of Agriculture, Department of Plant Protection, University of Jordan, Amman, Jordan

Received: May 18, 2013

Revised: July 16, 2013

Accepted: July 22, 2013

Abstract

A total of 14 species pertaining to five families (Platycnemididae, Coenagrionidae, Gomphidae, Aeshnidae and Libellulidae) were identified along eight sites in the Wadi Al Mujib catchment. Collected species varied along the eight sampling sites, ranging from a single species from Al Mujib dam to a maximum of seven species in the waterfalls to the bridge site. In the present study, eight species are considered as new records to the Odonata of the Wadi Al Mujib catchment (*Ischnura elegans*, *I. evansi*, *I. fountaineae*, *I. senegalensis*, *Anax parthenope*, *Orthetrum sabina*, *Sympetrum fonscolombii* and *Zygonyx torridus*). Such changes over the past 35 years reflect the dynamics of dragonflies' spatial movement within their distribution range. Results shown in this study strongly indicates the negative impact of the Wadi Al Mujib dam on the dragonfly fauna, as a single species was recovered from the dam proper. This is mainly due to the sharp cliffs and water level fluctuation and the limited breeding areas. Few flat areas with scarce vegetation were identified around the dam. Water level fluctuation does not allow steady vegetation growth around the edges of the dam, yet, the abrupt water depth is not suitable for development of the larval stages of dragonflies. Sharp edges are not suitable for breeding and perching of these insects.

Key words. Dragonflies, Wadi Al Mujib, Jordan, Biodiversity.

1. Introduction

The Wadi Al-Mujib basin drains a large part of central Jordan. Its principal wadis form a deep canyon complex that developed through the Moab plateau, the eastern uplifted shoulder of the Dead Sea rift. The drainage development of the Wadi Al Mujib basin since the Late Oligocene regression of the Tethys has been submitted to the combined effects of different environmental changes: rift activity, neotectonic history and climate changes (De Jaeger and De Dapper, 2002). The catchment is fed by several tributaries extending from the east near the main spring in Um Al Rasas, wadi systems around Karak then join Wadi Al Hidan near Al Mujib Nature Reserve.

Two major biogeographical zones of Jordan are represented in the reserve, namely the Irano-Turanian and the Saharo-Arabian zones. These characteristics and variations enhance the formation of five vegetation types, which, as a result, enrich the diversity of both faunal and floral elements in the reserve. These major vegetation types are the Mediterranean non-forest, saline, water,

tropical and steppe, with a total size area of 2.95, 10.38, 30.28, 39.39 and 97.42 km² respectively. The reserve is characterized by three major wadis with continuous water flow (Wadi Mujib, Wadi Hidan and Wadi Zarqa Ma'in).

Dragonflies and damselflies are among the most ancient insects that inhabited the earth. These remarkable species are considered as one of the most sensitive species of freshwater ecosystems together with other groups such as Ephemeroptera and Trichoptera. Their existence is dependent on the water quality and its availability as well as river structure. Both larvae and adults are predators feeding on aquatic and flying insects. They are often used as indicators for environmental health and conservation management (Corbet, 1999). Dragonflies are used in both basic and applied research due to the relative ease them (Kalkman *et al.*, 2009).

The Odonata of Jordan received attention during the late seventies and early eighties by Dumont (1973, 1975). As a part of his dissertation, W. Schneider studied the dragonfly fauna of the Levant, including Jordan (Schneider 1981a,b, 1982a,b, 1985, 1986). Ever since, few studies were undertaken to explore this insect group.

* Corresponding author. e-mail: amrz@just.edu.jo.

Amr *et al.* (1997) studied the dragonflies of the Azraq oasis and later Katbeh-Bader *et al.* (2002) examined the Odonata collections deposited in the Insect Museum at the Jordan University and at the Jordan University of Science and Technology. They reported a total of 47 species of dragonflies from Jordan, of which 29 were formerly known in the country. A review article with emphasis on conservation of dragonflies and their habitats in Jordan was published by Katbeh-Bader *et al.* (2004). Recently, Kunz *et al.* (2006) published a comprehensive study on the distribution of *Zygonyx torridus* in the Palaearctic, including new localities from Jordan.

The most important contribution on the dragonflies of Wadi Al Mujib catchment was undertaken by Schneider (1986). In his dissertation, he listed a total of nine species met along the Al Mujib catchment from 1981 to 1983. Three species were recorded from the upper reaches of Wadi Al Mujib, five from the current dam location and four along the wadi until it reaches the Dead Sea.

Dumont (1991) listed seven species from Wadi Al Mujib without referring to specific localities. Some of these species were already reported by Schneider (1986).

The objectives of this study are: to know the Odonata of Wadi Al Mujib catchment, identify key species that can be used as indicators and to define the effects of the Al Mujib dam on dragonflies populations.

2. Materials and Methods

2.1. Sampling Sites

Eight sampling sites were selected within the Wadi Al Mujib catchment in order to represent various habitats (Figure 1 and 2). The study was conducted during July 2011.

These sites were selected based on the overall habitat type and their accessibility. Some areas are very difficult to access due to the sharp ridges around the main flow of the water course. Additionally, the upper reaches of Wadi Al Mujib beyond Um Al Rasas are drained out due to water pumping and the main water course disappears before it reaches the main dam.

2.2. Description of the Sites Studied

Site 1: Um Al Rasas (E 35°53'46" N 31°23'59")

This is the main spring that feeds the Al Mujib river. This spring is surrounded by relatively thick vegetation of *Typha domingensis* and *Nerium oleander* and by a flat rocky area. Plastic water pipes to pump water for irrigation are scattered along the spring and the main stream. The area is not polluted by animal excrement or plastic.

Site 2: Al Mujib dam lake (35°49'26", N 31°25'48")

The lake created by the Wadi Al Mujib dam is located within the heart of the Wadi Al Mujib catchment. The elevation is about 208 m asl. The dam collects water from springs and main floods during winter. Maximum depth is 60 m and the total capacity reaches 32 million m³. The periphery of the lake consists of sharp rocky cliffs that were formed during the construction phase (Figure 2B). Very limited open areas are available around the water. Vegetation is very scarce due to the continuous flooding

and water retraction. Only small patches of *Juncus* sp. are present in some areas.

Small scaled agriculture occurs along the margins of the lake and on the relief around. Water is pumped from the dam by means of pumping engines. Insecticides are used by farmer to spray their vegetables in an unorganized fashion.

Site 3: Under the dam (E 35°48'58", N 31°26'47")

This site is characterized by the presence of side pools with a dense vegetation of *Typha* sp. and *Phragmites* near the main stream. The latter is narrow and is bordered by rocky edges. Scattered *Tamarix* trees and *Nerium oleander* are in close vicinity to the main water course. Common frog (*Rana ridibunda*) and the Tree frog (*Hyla savignyi*) are not uncommon. Several species of freshwater fishes were identified.

Site 4: Al Sdeer (E 35°46'56", N 31°27'05")

This site is about 5 km west to the dam. It is located within a deep gorge with relatively fast running water. Sides are rocky with a vegetation cover of *Typha* and *Phragmites* and small patches of *Juncus*, scattered *Tamarix* sp. and *Nerium oleander* (Figure 2C). Water depth varies from shallow to small ponds within the main water course with depth reaching up to 1 meter.

Site 5: Al Malaqee (E 35°36'11", N 31°26'48")

This site represents the junction between Wadi Al Mujib with Wadi Al Hidan tributary. Its name in Arabic, "Al Malaqee", means the meeting point. The upper junction of Wadi Al Mujib is densely vegetated and cannot be walked through. The area is characterized by a dense vegetation of *Typha* on both sides of the water course, while *Tamarix* and *Nerium oleander* grow more on the dry edges (Figure 2D). Water temperature is relatively higher than in other sections of the water catchment, due to mixing of hot springs feeding into Wadi Al Hidan. This area is quite rich in freshwater fishes that can be seen swimming against the water current. Few frogs, if any, were seen.

Site 6: Between Al Malaqee and Al Mujib Gorge (E 35°35'15", N 31°27'20")

This site extends from the beginning of the gorge leading to the waterfalls to Al Malaqee site. It extends for over four km from the beginning of the gorge until it reaches Al Malaqee. Water depth varies from shallow to up to 40 cm with variable dense vegetation of *Typha*, *Phragmites* and *Tamarix* (Figure 2E). In some instance, the water course is wide, reaching up to 3 meters.

Site 7: From the waterfalls to the bridge (E 35°34'59", N 31°27'40")

This area lies immediately after the gorge and extends to the Mujib Bridge. The water is shallow over a gravel bed. A few scattered vegetation is found, with mainly *Tamarix* and few *Juncus* communities. Edges are narrow with a gravel cover (Figure 2F).

Site 8: Mouth of Wadi Al Mujib to the Dead Sea (E 35°34'38", N 31°27'58")

This area is located near the Dead Sea. No open water flow exists as the river flows underground. The vegetation consists of *Tamarix*.

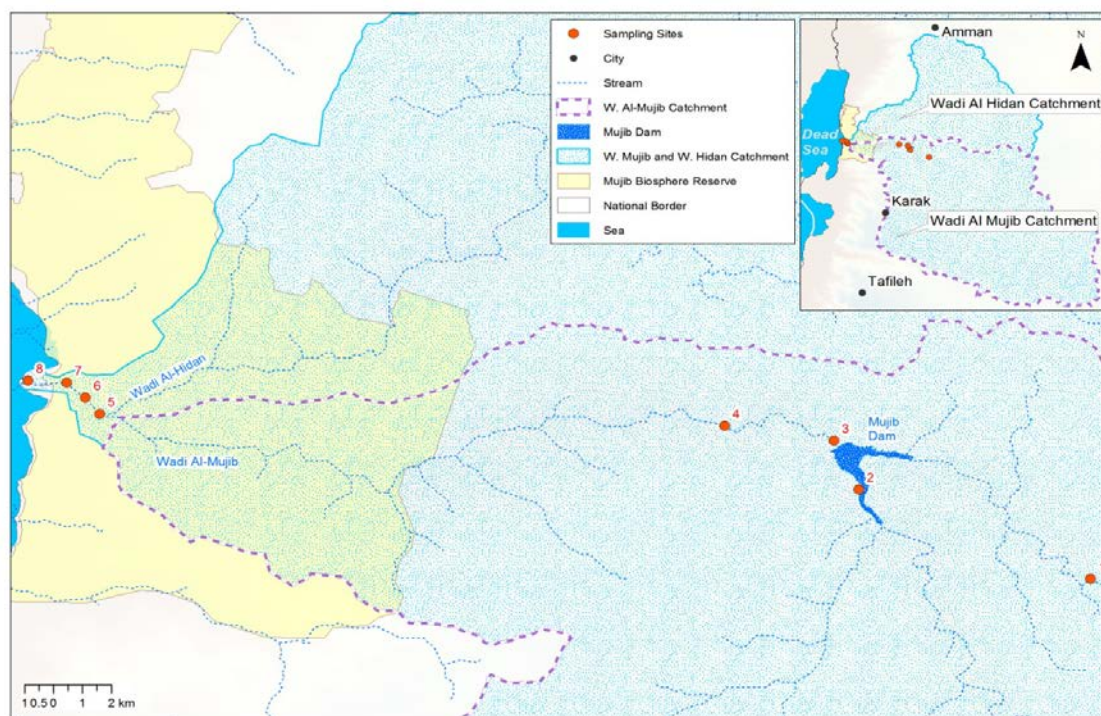


Figure 1. Study sites: 1. Um Al Rasas. 2. Wadi Al Mujib dam. 3. Under Wadi Al Mujib dam. 4. Al Sdeer. 5. Al Malaqee. 6. Between Al Malaqee and Al Mujib gorge. 7. From the waterfalls to the bridge. 8. Mouth of Wadi Al Mujib to the Dead Sea.



Figure 2. Habitats along Wadi Al Mujib Catchment. A. Um Al Rasas. B. Wadi Al Mujib dam. C. Al Sdeer. D. Between Al Malaqee and Al Mujib gorge. E. Al Malaqee. F. From the waterfalls to the bridge.

2.3. Sampling and Collecting Dragonflies

The vegetation (shrubs, low vegetation, trees, etc.) along Wadi Al Mujib was checked for the presence of dragonflies. In some instance, the vegetation was shaken to check for the presence of resting and perching adults. A handled aerial net with a wide opening was used to capture adult dragonflies.

In each site, a total of at least five hours was spent to check for the presence of Odonata. Transects along water edges were carefully examined for the presence of damselflies at rest on the low vegetation. Dragonflies usually occur on shrubs, trees and rocks in shaded areas. The team consists of two persons for the first three sites, and then additional persons joined the team for the rest of the sites. Both edges of the water courses were examined with a maximum distance from the water course not exceeding five meters.

Captured Odonata were kept in a plastic container and killed in a killing jar. Identification was followed according to Schneider (1986) and Dumont (1991). Both male and female genitalia were examined under the microscope to confirm their identification. The collected material was deposited in the University of Jordan Insects Museum (UJIM), was identified by Ahmad Katbeh-Bader and was later revised by Christian Monnerat (Switzerland).

3. Results

3.1. Species Diversity in the Sites Studied

A total of 14 species representing five families (Platynemidae, Coenagrionidae, Gomphidae, Aeshnidae and Libellulidae) were identified during this work. The number of species collected varied from a locality to another and ranged from a single one at the Al Mujib dam to seven at Al Malaqee (Table 1). Among the families, the Libellulidae included the highest number of species with seven species.

Site 1: Um Al Rasas

Materials collected and examined: *Platynemis dealbata*, 3 ♂♂ and 3 ♀♀, 20.7.2011; *Orthetrum chrysostigma*, 7 ♂♂ and 1 ♀, 20.7.2011; *Orthetrum brunneum*, 3 ♂♂ and 1 ♀, 20.7.2011; *Trithemis arteriosa*, 2 ♂♂ and 2 ♀♀, 20.7.2011. L. Al Azzam and R. Al Omari leg. throughout.

Remarks: Four species were collected and observed along the main spring of Um Al Rasas. *Platynemis dealbata* was very common and was found among the lower part of thick *Typha belts*. Libellulidae species were also common with varying relative abundance.

Site 2: Wadi Al Mujib dam lake

Materials collected and examined: *Sympetrum fonscolombii*, 1 ♂, 10.7.2011, Z. Amr and O. Abed leg.

Remarks: During extensive survey for over eight hours, two specimens of *S. fonscolombii* were observed and only one specimen was collected. Both specimens were found perching on dry plants.

Site 3: Under the dam

Materials collected and examined: *Ischnura elegans*, 3 ♀♀, 11.7.2011; *Platynemis dealbata*, 3 ♂♂ and 3 ♀♀, 11.7.2011; *Crocothemis erythraea*, 1 ♂ and 1 ♀,

11.7.2011; *Orthetrum chrysostigma*, 1 ♂, 11.7.2011. Z. Amr and O. Abed leg. throughout. Observed: *Anax parthenope*, 11.7.2011.

Remarks: Five species were collected and observed in this site. *Anax parthenope*, was observed among the thick vegetation near an open pool. Both *Ischnura elegans* and *Platynemis dealbata* were common within the shady vegetation near the pools and also among the vegetation in the vicinity of the main water course. Perching males of *Crocothemis erythraea* and *Orthetrum chrysostigma* were less common.

Site 4: Al Sedeer

Materials collected and examined: *Platynemis dealbata*, 3 ♀♀ and 1 ♂, 12.7.2011; *Paragomphus genei*, 1 ♀, 12.7.2011; *Trithemis arteriosa*, 2 ♂♂, 12.7.2011; *Orthetrum chrysostigma*, 1 ♀, 12.7.2011. Z. Amr and O. Abed leg. throughout.

Remarks: Four species of dragonflies were collected from this site. Most notably was the presence of *Paragomphus genei*. This species prefers open water such as open streams, rivers or pools.

Site 5: Al Malaqee

Materials collected and examined: *Platynemis dealbata*, 1 ♂ and 1 ♀, 20.7.2011; *Orthetrum chrysostigma*, 3 ♂, 20.7.2011; *Trithemis arteriosa*, 3 ♂, 20.7.2011. L. Al Azzam and R. Al Omari leg. throughout.

Remarks: Three species were collected from this site. They were common, particularly *P. dealbata*. Males of *O. chrysostigma* were the most common Anisoptera observed. They were perching on stones, rocks and among vegetation.

Site 6: Between Al Malaqee and the gorge

Materials collected and examined: *Platynemis dealbata*, 1 ♂ and 1 ♀, 20.7.2011; *Paragomphus genei*, 1 ♂, 20.7.2011; *Crocothemis erythraea*, 1 ♀, 20.7.2011; *Orthetrum chrysostigma*, 3 ♂♂ and 1 ♀, 20.7.2011; *Trithemis arteriosa*, 5 ♀♀, 20.7.2011; *Zygonyx torridus*, 1 ♀, 20.7.2011. L. Al Azzam and R. Al Omari leg. throughout.

Remarks: This site exhibited high species diversity (6 species). This is mainly due to the wide sides of the water course in addition to the relatively thick vegetation. *Orthetrum chrysostigma*, *C. erythraea* and *T. arteriosa* were the most common species. Males of *O. chrysostigma* and *C. erythraea* were very distinctive, perching on vegetation and rocky areas.

Site 7: From the Waterfalls to the bridge

Materials collected and examined: *Ischnura senegalensis*, 1 ♂, 21.7.2011; *I. fountaineae*, 1 ♂, 21.7.2011; *I. evansi*, 1 ♀, 21.7.2011; *Platynemis dealbata*, 1 ♀, 21.7.2011; *Paragomphus genei*, 1 ♀, 21.7.2011; *Orthetrum sabina*, 1 ♂, 21.7.2011; *Trithemis arteriosa*, 2 ♂♂, 21.7.2011. L. Al Azzam and R. Al Omari leg. throughout.

Remarks: This site exhibited the highest species diversity. Seven species were either collected or observed in this site.

Site 8: Mouth of Wadi Al Mujib to the Dead Sea

Materials collected and examined: *Ischnura fountaineae*, 1 ♂, 21.7.2011; *Trithemis arteriosa*, 2 ♂♂, 21.7.2011. L. Al Azzam & R. Al Omari leg. throughout.

Remarks: Two species were collected from this site (Table 1). Number of individuals was low compared to other sites.

Table 1. Odonata collected and observed along Wadi Al Mujib Catchment

		Collection sites							
Species	Common name	1	2	3	4	5	6	7	8
Suborder Zygoptera									
Family Platycnemididae									
<i>Platycnemis dealbata</i>	Ivory Feather leg	•		•	•	•	•	•	
Family Coenagrionidae									
<i>Ischnura elegans</i>	Blue-tailed Damselfly			•				•	
<i>Ischnura fountaineae</i>									•
<i>Ischnura evansi</i>	Evans' Blue tail Damselfly								•
<i>Ischnura senegalensis</i>									•
Suborder Anisoptera									
Family Gomphidae									
<i>Paragomphus genei</i>	Green Hooktail					•		•	•
Family Aeshnidae									
<i>Anax parthenope</i>	Lesser Emperor			•					
Family Libellulidae									
<i>Crocothemis erythraea</i>	Common Scarlet-darter			•				•	
<i>Orthetrum brunneum</i>	Brown Skimmer	•							
<i>Orthetrum chrysostigma</i>	Epaulet Skimmer	•		•	•	•	•		
<i>Orthetrum sabina</i>	Green Skimmer								•
<i>Sympetrum fonscolombii</i>	Red-veined Darter		•						
<i>Trithemis annulata</i>	Violet Dropwing								
<i>Trithemis arteriosa</i>	Red-veined Dropwing	•			•	•	•	•	•
<i>Zygonyx torridus</i>	Ringed Cascader							•	
Total		4	1	5	4	3	6	5	4

Sites: **1.** Um Al Rasas. **2.** Wadi Al Mujib Dam. **3.** Under Wadi Al Mujib Dam. **4.** Al Sdeer. **5.** Al Malaqee. **6.** Between Al Malaqee and Al Mujib gorge. **7.** After the water falls to the bridge. **8.** Entrance of Wadi Al Mujib to the Dead Sea.

4. Discussion

4.1. Species Diversity of Odonata in the Al Mujib Catchment: past and present

The present Odonata fauna of the Wadi Al Mujib catchment includes 14 species belonging to five families. This represents about 30% of the total known species occurring in Jordan. Previously, Schneider (1986) reported eight species; four were not found during the present study (*Crocothemis sanguinolenta*, *Diplacodes lefebvrii*, *Orthetrum abbotti* and *Trithemis annulata*). Also, five species reported by Dumont (1991) were not found again (*Calopteryx syriaca*, *Crocothemis*

sanguinolenta, *Epallage fatime*, *Onychogomphus flexuosus*, *Orthetrum abbotti* and *Trithemis annulata*) (Table 2). Schneider (1986) doubted the occurrence of *Calopteryx syriaca* from old reports.

Table 2. Comparison between the Odonata fauna in the present study with previous studies.

Species	Schneider (1986)	Dumont (1991)	Present study
<i>Calopteryx syriaca</i>	?	•	
<i>Epallage fatime</i>		•	
<i>Ischnura elegans</i>			•
<i>Ischnura evansi</i>			•
<i>Ischnura fountaineae</i>			•
<i>Ischnura senegalensis</i>			•
<i>Paragomphus genei</i>	•		•
<i>Onychogomphus flexuosus</i>		•	
<i>Platycnemis dealbata</i>		•	•
<i>Anax parthenope</i>			•
<i>Crocothemis erythraea</i>	•		•
<i>Crocothemis sanguinolenta</i>	•	•	
<i>Diplacodes lefebvrii</i>	•		
<i>Orthetrum abbotti</i>	•	•	
<i>Orthetrum brunneum</i>	•		•
<i>Orthetrum chrysostigma</i>	•		•
<i>Orthetrum sabina</i>			•
<i>Sympetrum fonscolombii</i>			•
<i>Trithemis annulata</i>	•	•	•
<i>Trithemis arteriosa</i>	•		•
<i>Zygonyx torridus</i>			•
Total	9	7	15

Six species were shared between the present and past studies, namely *P. genei*, *P. dealbata*, *C. erythraea*, *O. brunneum*, *O. chrysostigma*, and *T. arteriosa*.

Eight species are considered as new for the Wadi Al Mujib catchment (*I. elegans*, *I. evansi*, *A. parthenope*, *O. sabina*, *S. fonscolombii* and *Z. torridus*). Such changes over the past 35 years reflect the dynamics of dragonflies' spatial movement within their distribution range. This pattern of faunal changes was documented by Moore (2001), which reported radical changes in dragonfly communities in pond in the United Kingdom over the period 1962-1988. Such changes were attributed to changes in the aquatic flora, vegetation cover, presence of predators etc (Wellborn et al., 1996).

Within the Dead Sea basin, Furth (1983) reported 13 species of dragonflies. Among these, *A. parthenope*, *C. erythraea*, *O. brunneum*, *O. chrysostigma*, *S. fonscolombii*, *T. arteriosa*, *T. annulata* and *Z. torridus* were also recorded from the western side of the Dead Sea, that means they could easily colonize other localities in this area, to which Wadi Al Mujib pertains.

4.2. Conservation Status of Dragonflies in Jordan

Jordan is a small country with very limited water resources. Over the past sixty years, the human population increased enormously, that led to increased pressure on water resources. Dam constructions, altering natural surface waters increased in number to meet water demand for both agriculture and domestic use. Such activities and development took its toll on freshwater habitats and its biological communities.

Schneider (2004) revised the status of the Odonata of the Levant (Table 3). In his treatment, *C. syriaca* was considered as endangered and its status is now even worse. *Orthetrum abbotti* is considered as an Afrotropical

relict in Jordan. Such species requires further evaluation in Jordan.

Table 3. Conservation status of some levantine Odonata in Jordan, according to Schneider (2004).

Species	IUCN 2003	Suggested status 2004	Remarks
<i>Calopteryx syriaca</i>	EN	EN	Endemic
<i>P. sublacteum mortoni</i>		VU	Endemic
<i>Pseudagrion syriacum</i>			Endemic
<i>Gomphus davidi</i>			Endemic
<i>Onychogomphus macrodon</i>	VU	VU, DD	Endemic
<i>Crocothemis sanguinolenta</i>		VU (?), DD	Relict
<i>Libellula pontica</i>		DD	Endemic
<i>Orthetrum abboti</i>		DD	Relict

The diversity of Odonata in the Levant is considered high and including a number of endemic and threatened species. At present, Jordan is witnessing increasing demand for water. In combination with the unpredictable effects of climate change, this suggests strongly that the conservation status of many species will deteriorate in the near future (Riservato *et al.*, 2009), at such a point that when a new examination of their conservation status will be done, species like *Calopteryx hyalina* and *Onychogomphus macrodon* will certainly join the strongest categories of threat (CR and EN or CR, respectively).

Within the Mediterranean region, habitat loss and degradation caused by human activities is the major threat for both threatened and non-threatened dragonfly species. It is estimated that these threats are currently affecting 110 dragonfly species, including 30 of the 31 threatened species (Riservato *et al.*, 2009).

4.3. Impact of the Wadi Al Mujib Dam on Dragonflies Diversity

Results shown in this study strongly indicate the negative impact of the Wadi Al Mujib dam on the dragonfly fauna. Only a single species (*S. fonscolombii*) was recovered from the dam proper. This is mainly due to the sharp cliffs and water level fluctuation. Few flat areas with scarce vegetation were found around the lake. Water level fluctuation does not allow a steady vegetation growth around the banks. Moreover, the abrupt water depth is not suitable for the development of the larvae of the dragonflies. Sharp edges are not suitable for breeding and perching for these insects. So, *S. fonscolombii* may represent vagrant or migrant individuals from nearby areas or from other countries, and certainly not a breeding population given the depth of the dam and the absence of

suitable breeding areas. Previously, five species (*C. erythraea*, *Diplacodes lefebvrei*, *O. brunneum*, *O. chrysostigma* and *P. genei*) were recorded at the present location of the Al Mujib dam (Schneider, 1986).

In addition, several species of alien freshwater fishes were found breeding in the dam (*Clarias gariepinus*, *Cyprinus carpio*, *Oreochromis aureus* and *Tilapia zillii*). They may also have a negative impact on the larval stages of dragonflies.

Globally, many dragonfly species have shown a dramatic decline in their distribution and abundance patterns since the second half of the 20th century (Westfall and May, 1996; Sahlen *et al.*, 2004; Inoue, 2004). This decline is mainly due to habitat destruction, eutrophication, acidification, pollution and water management (channelization, dams construction, modifications of the structure of rivers). Not all anthropogenic disturbances are harmful to dragonflies. In some instances, small dams play an important role in increasing the overall density of many lotic species (Samways, 1999).

Wilcove *et al.* (1998) gave a comprehensive review on endangered species due to habitat loss and occurrence of alien species. Thirty three species of Odonata were considered threatened due to these causes. The sensitivity of dragonflies to habitat structure and their amphibious habits make them well suited for use in evaluating long and short environmental changes (Kalkman *et al.*, 2008).

On the other hand, shallow farm dams with a relatively dense helophyte vegetation offer suitable habitats for Odonata communities. In southern Africa, farm dams, which are a necessary and characteristic feature of the agricultural landscape, attract dragonflies, each functioning as a small nature reserve (Samways, 1999).

4.4. Key species that can be used as indicators

Within the Wadi Al Mujib catchment, at least four species can be regarded as key species (Table 4). Although *Onychogomphus lefebvrei* and *Onychogomphus flexuosus* were not recorded during this study, the former species was reported by Dumont (1991). These species are very sensitive to changes in vegetation cover as well as to water quality. These species were selected since they were the most common species in the stretches exhibiting relatively high species diversity. Also, these species are easy to identify on the spot without the need for a close-up examination. These key species can provide information on changes in vegetation cover over seasons or due to man induced changes in a particular site. Also, they bring some information about water velocity and quality.

Table 4. Some selected species that can be considered as indicator species.

Species	Habitat Requirement	Remarks
<i>O. lefebvri</i>	Thick vegetation of <i>Typha</i> and slow running water	Found on medium sized vegetation
<i>P. genei</i>	Thick vegetation with trees and large shrubs in shaded areas with fast running water	Observed on shrubs, rocks and trees.
<i>O. flexuosus</i>	Thick vegetation with trees and large shrubs in shaded areas with fast running water	Observed on shrubs, rocks and trees.
<i>Z. torridus</i>	Thick vegetation with trees and large shrubs in shaded areas with fast running water	Observed on shrubs, rocks and trees.

Perhaps *Orthetrum abboti* and *Crocothemis sanguinolenta* should be also considered as a major key species to the study area, since they are known only from the Dead Sea rift within the Levant. During the present study, we failed to record them. Additional effort is required during different seasons to confirm their occurrence in the Wadi Al Mujib catchment. Both were found in the past in this catchment (Dumont, 1977) and Monnerat and Hoess (2011) mapped them in several locations of the Dead Sea rift.

Several species of the dragonflies of Jordan disappeared or are on the verge of extinction (i.e. *C. syriaca*). Indeed, Schneider (1982b) pointed out earlier on such changes and their impact on the regional dragonfly fauna. For example, *E. fatime* was common all over the Jordan Valley and currently is very scarce. Similarly, *C. syriaca* populations are diminishing in previously known breeding sites (Katbeh *et al.*, 2004). This species is very sensitive to pollution and habitat structure degradation and could be eradicated easily from its natural habitats. Schneider (1982b) pointed out that the endemic *Pseudagrion syriacum* is threatened due to the overwhelming changes in freshwater ecosystems in the Jordan valley.

Acknowledgements

This project was funded by the Canadian International Development Agency (CIDA) through the capacity building program of Wetlands International (WI). We are heartily thankful to the Royal Society for the Conservation of Nature for its ongoing support to scientific research and for the excellent coordination of the field trips and logistic support.

We would like to thank Christian Monnerat, Swiss Biological Switzerland for his help in confirming the identification of collected specimens. Also, we would like to thank Dr. Nashat Hamidan for his outstanding knowledge on the freshwater biology of Wadi Al Mujib, and for his company during the fieldwork. Our thanks are also extended to Rami Al Omari, Omar Abed, Thabit Al Share, Tariq Qaneer and Aqeel Shora for their help during field work. Thanks are also due to Mrs. Natalia Bolad for the map preparation.

Special thanks are extended to the Jordan Valley Authority, particularly Engineer Nidal Shenanat, Mujib dam Manager, for providing facilities, accommodation and arranging boat tour in Wadi Al Mujib dam. Our

gratitude's are extended to Mujib Nature Reserve rangers for their hospitality and help during our mission, Mr. Raed Karadsheh, Enforcement Section, for his help to identify the Um Al Rasas site.

References

- Amr ZS, Al-Melhim WN, Katbeh-Bader A and Schneider W. 1997. On the common Insecta of Al Azraq, Jordan. *Entomologist's Gazette*, **48**: 55-66.
- Corbet PS. 1999. **Dragonflies. Behaviour and Ecology of Odonata**. Harley Books, Essex, England.
- De Jaeger C and De Dapper M. 2002. Tectonic control in the geomorphologic development of the Wadi el-Mujib canyon (Jordan). *EGU Stephan Mueller Special Publication Series*, **2**: 83-94.
- Dumont HJ. 1977. *Orthetrum abboti* Calvert, 1892, a new Ethiopian representative in the Palaearctic fauna (Anisoptera: Libellulidae). *Odonatol.*, **6**: 199-203.
- Dumont HJ. 1973. The genus *Pseudagrion* Selys in Israel and Egypt, with a key to the regional species (Insecta: Odonata). *Israel J Zoology*, **22**: 169-195.
- Dumont HJ. 1975. Endemic dragonflies of the Pleistocene age of the Hula Lake area (northern Israel), with notes of the Calopterygidae of the River Jordan (Israel, Jordan) and Litani (The Lebanon) and description of *Urothemis edwardsi hulae* subpec. nov. (Libellulidae). *Odonatol.*, **4** (1):11-19.
- Dumont HJ. 1991. **Odonata of the Levant**. Israel Academy of Sciences and Humanities, Jerusalem.
- Furth DG. 1983. Aquatic entomofauna of a Dead Sea oasis. *Hydrobiol.*, **102**: 3-25.
- Inoue K. 2004. Critical species of Odonata in Japan. In: Clausnitzer V and Jödicke R (Eds), **Guardians of the Watershed. Global Status of Dragonflies: Critical Species, Threat and Conservation**. *Inter J Odonatol.*, **7**: 311-324.
- Kalkman VJ, Clausnitzer V, Dijkstra K-DB, Orr AG, Paulson DR and Van Tol J. 2008. Global diversity of dragonflies (Odonata) in freshwater. *Hydrobiol.*, **595**: 351-363.
- Katbeh-Bader A, Amr ZS, Abu Baker M and Mahasneh A. 2004. The dragonflies (Insecta: Odonata) of Jordan. *Denisia*, **14**: 309-317.
- Katbeh-Bader A, Amr Z and Schneider W. 2002. Odonata of Jordan. *Fragmenta Entomol.*, **34**: 147-170.
- Kunz B, Ober S and Jödicke R. 2006. The distribution of *Zygonyx torridus* in the Palaearctic (Odonata: Libellulidae). *Libellula*, **25**: 89-108.
- Monnerat C and Hoess R. 2011. Libellen aus Jordanien, dem Westjordanland und dem Libanon, gesammelt von Johann Friedrich Klapperich zwischen 1956 und 1969 (Odonata). *Libellula*, **30**: 77-88.
- Moore NW. 2001. Changes in the dragonfly communities at the twenty ponds at Woodwalton Fen, Cambridgeshire, United Kingdom, since the study of 1962-1988. *Odonatol.*, **30**: 289-298.
- Riservato E, Boudot J-P, Ferreira S, Jović M, Kalkman VJ, Schneider W, Samraoui, B and Cuttelod A. 2009. **The Status and Distribution of Dragonflies of the Mediterranean Basin**. Gland, Switzerland and Malaga, Spain: IUCN. vii + 33 pp.
- Sahlen G, Bernard R, Rivera AC, Ketelaar R and Suhling F. 2004. Critical species of Odonata in Europe. In: Clausnitzer V and Jödicke R (eds), **Guardians of the Watershed. Global Status of Dragonflies: Critical Species, Threat and Conservation**. *Inter J Odonatol.*, **7**: 385-398.

- Samways MJ. 1999. Diversity and conservation status of South African dragonflies (Odonata). *Odonatol*, **28**: 13-62.
- Schneider W. 1981a. Eine Massenwanderung von *Selysiotthemis nigra* (Vander Linden, 1825) (Odonata: Macrodiplactidae) und *Lindenia tetraphylla* (Vander Linden, 1825) (Odonata: Gomphidae) in Südjudanien. *Entomologische Zeitschrift*, **91**: 97-102.
- Schneider W. 1981b. On a dragonfly collection from Syria. *Odonatol*, **10**: 131-145.
- Schneider W. 1982a. *Crocothemis sanguinolenta arabica* n. subsp. (Odonata: Anisoptera: Libellulidae), ein afrikanisches Relikt in der südlichen Levante. *Entomologische Zeitschrift*, **92**: 25-31.
- Schneider W. 1982b. Man-induced changes in the dragonfly fauna of the Jordan Valley. *Advances in Odonatol*, **1**: 243-249.
- Schneider W. 1985. Die Gattung *Crocothemis* Bauer 1868 im Nahen Osten (Insecta: Odonata: Libellulidae). *Senckenbergiana Biologica*, **66** (1-3): 79-88.
- Schneider W. 1986. Systematic und Zoogeographie der Odonata der Levante unter besonderer Berücksichtigung der Zygoptera. Ph.D. Dissertation, 3 Volumes, University of Mainz.
- Schneider W. 2004. Critical species of Odonata in the Levant. *Inter J Odonatol*, **7**: 399-407.
- Wellborn GA, Skelly DK and Werne EE. 1996. Mechanisms creating community structure across a freshwater habitat gradient. *Annual Rev Ecol Systems*, **27**: 337-63.
- Westfall MJ. and May ML. 1996. **Damselflies of North America**. Scientific Publishers, Gainesville.
- Wilcove DS, Rothstein D, Dubow J, Phillips A and Losos E. 1998. Quantifying threats to imperiled species in the United States. *Bio Sci.*, **48**: 607-615.

Comparative Studies on Anti-hyperglycemic Effects of Ethyl Acetate and Methanol Extract of *Albizzia lucida* Benth Bark in Alloxan Induced Diabetic Rats

Arumugam S. Kumar^{1,*}, Subramanian Kavimani² and Korlakunta N. Jayaveera³

¹Department of Pharmacology, Jawaharlal Nehru Technological University, Anantapur, Andhra Pradesh, 515002, India.

²Department of Pharmacology, Mother Theresa Post Graduate and Research Institute of Health Sciences, Puducherry, 605 006, India.

³Department of Chemistry, Jawaharlal Nehru Engineering College, Jawaharlal Nehru Technological University, Anantapur, Andhra Pradesh, 515002, India.

Received: July 16, 2013

Revised: August 5, 2013

Accepted: August 8, 2013

Abstract

The decoction of the bark of *Albizzia lucida* Benth was used in ayurvedic system for management of diabetes mellitus. To confirm the traditional claim, the comparative anti-hyperglycemic efficiency of ethyl acetate extract of bark of *Albizzia lucida* Benth. (EEAL) and methanol extract of bark of *Albizzia lucida* Benth. (MEAL) was done in alloxan induced diabetic rats. Acute and sub-acute effect of oral administration of EEAL and MEAL (200 & 400 mg/kg, b.wt) on blood glucose was observed in different time interval for 10 days. By the end of the study, the collected blood sample and pancreas was used for estimation of serum lipid profile and histological studies. EEAL and MEAL showed that the serum blood glucose and serum lipids (total cholesterol, triglyceride, VLDL-cholesterol, LDL-cholesterol) was reduced significantly ($p < 0.01$) in dose dependent manner when compare to control group. At the same time MEAL was showed potent effect than EEAL. EEAL and MEAL does not affect the body weight and alter the histopathology of pancreas similar to that of normal. It was concluded that EEAL extract of bark of *Albizzia lucida* Benth and MEAL extract of bark of *Albizzia lucida* Benth. possess anti-hyperglycemic activity in alloxan-induced diabetic model rats.

Keywords: *Albizzia lucida* Benth., Ayurveda, Diabetes, Alloxan, Serum Lipid Profile, Histopathology, Anti-hyperglycemic.

1. Introduction

Diabetes is chronic metabolic disorder characterized by hyperglycemia due to absence or insufficient of circulating insulin levels (Holmann and Turner, 1991). Though insulin and various anti-diabetic drugs are available but insulin could not be used by oral and continuous use of synthetic agents leads severe side effects (Valiathan, 1998). Herbal medicines are prescribed universally even though their biological active constituents are unknown due to their potency with minimum side effects and relatively low cost (Verspohl, 2002; Villasenor and Lamadrid, 2006).

In folk medicine, the decoction of bark of *Albizzia lucida* Benth. (Family: Mimosaceae) is considered useful in diabetes, pregnancy and stomachache. It is also used as a medicine for water buffalo when given with salt (Bulusu Sitaram and Chuneekar, 2006). Therefore, the present research was undertaken to verify the anti-hyperglycemic potential of ethyl acetate extract of bark of *Albizzia lucida*

Benth. (EEAL) and methanol extract of bark of *Albizzia lucida* Benth. (MEAL) in alloxan induced diabetic rats.

2. Materials and Methods

2.1. Plant Material and Preparation of Extract

The bark of *Albizzia lucida* Benth. was collected from Tirumala hills, Chittoor district of Andhra Pradesh, India in March 2011. The plant was authenticated Dr. K. Madhava Chetty, Department of Botany, Sri Venkateswara University, Tirupati. The bark of *Albizzia lucida* Benth. was dried in shade and pulverized in the grinder-mixer to obtain a coarse powder, then passed through the 40 mesh sieve. A weighed quantity (100gm) of powder was subjected to continuous hot extraction with ethyl acetate and methanol in soxhlet apparatus for 48 hours. Then the extracts were evaporated at reduced pressure using rotary evaporator until all the solvent has been removed to give an extract sample. The percentage yield of ethyl acetate and methanol extract of *Albizzia*

* Corresponding author. e-mail: sarganjune1@gmail.com.

lucida Benth. was found to be 7.40 & 9.85%w/w respectively.

2.2. Preliminary Phytochemical Investigation of EEAL and MEAL

The preliminary phytochemical investigation was performed by using several standard phytochemical tests for the qualitative estimation of presence of various phytochemicals in ethyl acetate (EEAL) and methanol extract (MEAL) of bark of *Albizzia lucida* Benth. (Harbone, 1973).

2.3. Animals

Male albino Wistar rats (200-250gm) were obtained from the animal house in Sree Vidyanikethan College of Pharmacy, Tirupati, Andhra Pradesh. The animals were maintained in a well-ventilated room with 12:12 hour light/dark cycle and water was given ad libitum. The experiments were performed after approval (Approval no: SVCP/IAEC/I-009/2011-12) of the protocol by the Institutional Animal Ethics Committee (IAEC) and were carried out in accordance with the current guidelines on the care of laboratory animals.

2.4. Acute Toxicity Study

The acute toxicity of EEAL and MEAL was determined as per the OECD guideline no. 423 (Acute toxic class method). Albino wistar rats (n=6) either sex selected and kept fasting for overnight providing only water. EEAL and MEAL was administered orally at the dose level of 2000mg/kg by oral needle and observed for 14 days. Hence, 1/10th (200 mg/kg) and 1/5th (400 mg/kg) of the doses were selected for further study (OECD, 2002).

2.5. Anti-hyperglycemic Efficiency of *Albizzia lucida* Benth. Extracts

2.5.1. Induction of diabetes

Male wistar albino normoglycemic rats were injected intraperitoneally with alloxan monohydrate dissolved in normal saline at the dose of 120 mg/kg b.wt (Lachin and Reza, 2012; Chaudhary *et al.*, 2012). After 3 days, the fasting blood glucose was checked and above 250 mg/dl of blood glucose reached rats was considered as diabetic rats. These diabetic rats were segregated into seven groups of six rats each group. Group I (Normal control): Vehicle 1% w/v CMC; 5ml/kg, b.w. p.o); Group II (Alloxan induced Diabetic control) received only vehicle (1%CMC; 5ml/kg, b.w. p.o); Group III & IV - Alloxan induced diabetic rats received the EEAL 200 & 400 mg/kg/day p.o suspended in 1% w/v CMC; Group V & VI - Alloxan induced diabetic rats received the MEAL 200 & 400 mg/kg/day p.o suspended in 1% w/v CMC; Group VII (Standard) Alloxan induced diabetic rats received Glibenclamide (2.5 mg/kg p.o) suspended in 1% w/v CMC, respectively.

2.5.2. Acute experimental study

Blood glucose level of all group rats were checked after 0, 1, 3, 6 & 9h of oral administration of single dose of test drugs.

2.5.3. Sub-acute experimental study

The same treatments continued with same dose once daily for 10 days. The blood samples collected from tail

vein and measured the glucose level using commercially available glucose strips (Accu-Chek) using one-touch glucometer (Johnson-Johnson, India) on initial day, 3, 7, 10th day respectively. After terminate the study, the animals were sacrificed by cervical dislocation. The blood was collected without anticoagulant and serum was separated by centrifugation at 6000 rpm for 5min. the pancreas organ of rats also separated carefully for histological study.

2.5.4. Estimation of serum lipid profile

Serum total cholesterol (TC), triglycerides (TG), LDL-C, VLDL, HDL-C were measured for all animals (n=6) from each group by commercially available diagnostic kits (Span Diagnostics, India). The serum low-density lipoprotein (LDL) and very low-density lipoprotein (VLDL) levels were calculated by Friedewald formula (Friedewald *et al.*, 1972): VLDL = TG / 5; LDL = TC - (HDL + VLDL).

2.6 Histological Assessment of Pancreas

The separated pancreas was kept in 10% formaldehyde solution for histopathological studies and standard procedure followed for fixation. This study was done at pathology department of Sri Venkateswara University, Tirupati, Andhra Pradesh.

2.7 Statistical Analysis

The present research observations were signified as Mean \pm Standard Error Mean. Statistical significance of dissimilarities amid the groups was evaluated by one way and multiple way analysis of variance (ANOVA) followed by Turkey test. *P* values less than 0.05 were deliberated as significance.

3. Results

3.1. Preliminary Phytochemical Investigation of EEAL & MEAL

Ethyl acetate and methanol extracts of bark of *Albizzia lucida* Benth. were showed proteins, steroids, alkaloids, flavonoids, terpenoids, tannins and phenolic compounds and saponins.

3.2. After single dose of EEAL & MEAL on serum blood glucose level in diabetic rats

EEAL and MEAL extracts were significantly ($P < 0.01$) decreased the serum blood glucose level. In acute experimental study, single dose response of EEAL and MEAL in alloxan induced diabetic rats at 0, 1, 3, 6 & 9h. Table 1 and Figure 1 showed that the methanolic extract of bark of *Albizzia lucida* Benth. (MEAL 200 and 400mg/kg, p.o) effectively ($P < 0.01$) reduced blood glucose level of diabetic rats (47.59% and 53.34%, respectively) than ethyl acetate extract of bark of *Albizzia lucida* Benth. (EEAL 200 and 400mg/kg, p.o) (26.34% and 37.49%, respectively). These reduced values were very close to that of glibenclamide (2.5mg/kg p.o) (61.74%). After oral administration of MEAL, acute reduction ($P < 0.01$) of glucose level in time dependent manner when compared to EEAL.

3.3. After Multidose of EEAL & MEAL on serum blood glucose level in diabetic rats

In sub-acute experimental study, the blood glucose was determined on initial day, 3, 7, 10th day after oral administration of EEAL and MEAL (200 & 400mg/kg, p.o) (Table 2 and Figure 2). Both doses of MEAL (200 and 400mg/kg, p.o) showed significant ($P<0.01$) chronic reduction of serum blood glucose level (54.59% and 57.73%, respectively) when compared to diabetic rats. MEAL effect was near to that of ($P<0.01$) of glibenclamide (2.5mg/kg p.o) (67.06%). Anti-hyperglycemic effect of MEAL was greater effect ($P<0.01$) than EEAL 200 and 400mg/kg, p.o (40.89% & 48.96%, respectively).

3.4. After Multidose of EEAL & MEAL on serum lipid profile and body weight in diabetic rats

The serum blood glucose was raised ($P<0.01$) accompanied by an increase in serum lipid profile such as total cholesterol (TC), triglycerides (TG), LDL-C, VLDL & HDL-C in alloxan-induced diabetic rats (Table 3 and Figure 3). MEAL (200 and 400mg) afforded greater reduction of TC, TG, LDL-C & VLDL, whereas HDL-C level was significantly ($P<0.01$) increased. Anti-

hyperlipidemic effect of MEAL was great efficient than EEAL. Standard group rats showed significant ($P<0.01$) anti-hyperlipidemic effect with glibenclamide (2.5mg/kg p.o).

Table 4 and Figure 4 showed that the alloxan induced diabetic rats showed loss of body weight when compared to normal control and EEAL and MEAL (200 and 400mg/kg, p.o) treated rats. These results apparently recorded that treatment with EEAL and MEAL extracts showed better control in the loss of body weight.

3.5. Histopathological studies on islets of pancreas

Pancreatic beta cells of normal rats showed adorned islets with well defined border and normal allocation of islets of langerhans. The histopathological studies of alloxan induced diabetic rat pancreas illustrates that atrophy, suppress the number of beta cells of islets of langerhans was noted when compared to control group. Necrosis and damage of islets was caused by alloxan. The oral administration of EEAL and MEAL to groups III, IV, V & VI of experimental rats showed in the recovery of beta cells necrosis and the restoration of the islets similar to that of control and standard groups (glibenclamide 2.5mg/kg p.o) treated animals (Figure 5).

Table 1. After single dose of EEAL & MEAL on serum blood glucose in alloxan induced diabetic rats

Groups	Serum Blood Glucose (mg/dl) (% of glucose reduction)				
	0 h	1 h	3 h	6 h	9 h
I - Normal control (1% w/v CMC)	86.67±1.856**a	87.00±2.221**a	84.00±1.00**a	87.17±1.424**a	84.83±1.276**a
II - Alloxan induced Diabetic control	258.00±2.113	263.50±3.490	268.17±3.807	282.67±2.418	288.00±2.517
III - Alloxan + EEAL (200 mg/kg, p.o)	264.50±2.513 ^b	256.17±1.600 ^b (3.15)	243.50±1.147 ^{tb} (7.94)	223.33±1.453 ^{tb} (15.56)	194.83±1.400 ^{tb} (26.34)
IV - Alloxan + EEAL (400 mg/kg, p.o)	265.83±2.600 ^b	252.67±1.978 ^b (4.95)	234.00±2.129 ^{tb} (11.97)	193.17±1.400 ^{tb} (27.33)	166.17±1.424 ^{tb} (37.49)
V - Alloxan + MEAL (200 mg/kg, p.o)	267.17±4.102 ^b	254.17±1.400 ^{tb} (4.87)	237.33±0.667 ^{tb} (11.17)	197.83±2.227 ^{tb} (25.95)	140.00±0.931 ^{tb} (47.59)
VI - Alloxan + MEAL (400 mg/kg, p.o)	261.83±2.857 ^b	238.67±1.282 ^{tb} (8.85)	221.50±1.727 ^{tb} (15.40)	161.67±2.186 ^{tb} (38.25)	122.17±1.621 ^{tb} (53.34)
VII - Alloxan + Glibenclamide (2.5mg/kg, p.o)	266.17±3.380 ^b	233.83±2.372 ^{tb} (12.15)	208.00±1.033 ^{tb} (21.85)	135.67±1.229 ^{tb} (49.03)	101.83±2.167 ^{tb} (61.74)

All values expressed in mean ± SEM for six animals in each group (n=6);

All values compared with diabetic control groups. * $p < 0.05$; ** $p < 0.01$;

a – Normal control group I /s diabetic control group II; b – Groups III, IV, V, VI & VII /s diabetic control group II.

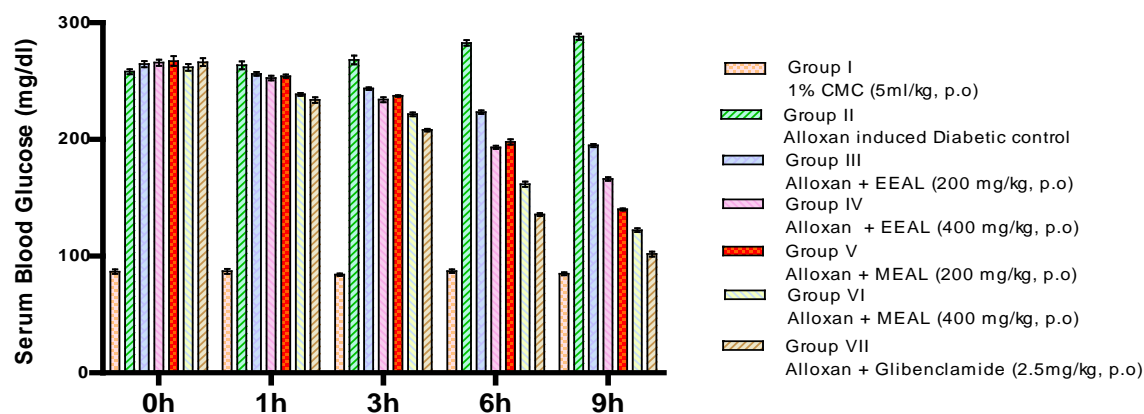


Figure 1. After single dose of EEAL & MEAL on serum blood glucose in alloxan induced diabetic rats

All values expressed in mean \pm SEM for six animals in each group (n=6);

All values compared with diabetic control groups. * $p < 0.05$; ** $p < 0.01$;

a – Normal control group I /s diabetic control group II; b – Groups III, IV, V, VI & VII /s diabetic control group II.

Table 2. After Multidose of EEAL & MEAL on serum blood glucose in alloxan induced diabetic rats

Groups	Serum Blood Glucose (mg/dl) (% of glucose reduction)			
	0 day	3 day	7 day	10 day
I - Normal control (1% w/v CMC)	86.67 \pm 1.856 ^{**a}	87.50 \pm 2.802 ^{**a}	86.00 \pm 0.730 ^{**a}	83.83 \pm 0.946 ^{**a}
II - Alloxan induced Diabetic control	258.00 \pm 2.113	297.83 \pm 1.990	290.50 \pm 2.078	286.50 \pm 3.074
III - Alloxan + EEAL (200 mg/kg, p.o.)	264.50 \pm 2.513 ^b	207.00 \pm 0.966 ^{**b} (21.74)	183.67 \pm 0.919 ^{**b} (30.56)	156.33 \pm 1.333 ^{**b} (40.89)
IV - Alloxan + EEAL (400 mg/kg, p.o.)	265.83 \pm 2.600 ^b	178.33 \pm 1.333 ^{**b} (32.92)	149.00 \pm 1.528 ^{**b} (43.94)	135.67 \pm 0.715 ^{**b} (48.96)
V - Alloxan + MEAL (200 mg/kg, p.o.)	267.17 \pm 4.102 ^b	176.33 \pm 1.726 ^{**b} (34.00)	145.17 \pm 0.946 ^{**b} (45.66)	121.33 \pm 1.145 ^{**b} (54.59)
VI - Alloxan + MEAL (400 mg/kg, p.o.)	261.83 \pm 2.857 ^b	154.00 \pm 1.238 ^{**b} (41.18)	120.00 \pm 1.732 ^{**b} (54.17)	110.67 \pm 1.542 ^{**b} (57.73)
VII - Alloxan + Glibenclamide (2.5mg/kg, p.o.)	266.17 \pm 3.380 ^b	96.67 \pm 0.919 ^{**b} (63.68)	93.67 \pm 1.174 ^{**b} (64.81)	87.67 \pm 2.459 ^{**b} (67.06)

All values expressed in mean \pm SEM for six animals in each group (n=6);

All values compared with diabetic control groups. * $p < 0.05$; ** $p < 0.01$;

a – Normal control group I /s diabetic control group II; b – Groups III, IV, V, VI & VII /s diabetic control group II.

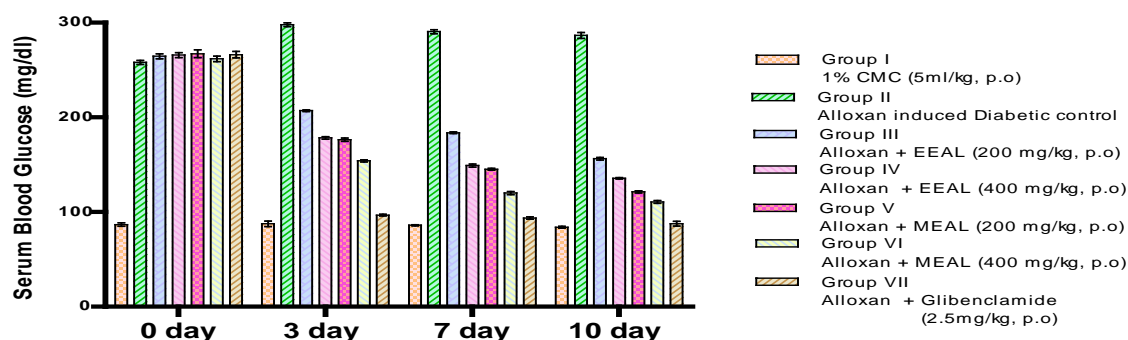


Figure 2. After Multidose of EEAL & MEAL on serum blood glucose in alloxan induced diabetic rats

All values expressed in mean \pm SEM for six animals in each group (n=6);

All values compared with diabetic control groups. * $p < 0.05$; ** $p < 0.01$;

a – Normal control group I Vs diabetic control group II; b – Groups III, IV, V, VI & VII Vs diabetic control group II.

Table 3. After Multidose of EEAL & MEAL on serum lipid profile in alloxan induced diabetic rats

Groups	Serum Lipid Profile (mg/dl)				
	Total Cholesterol (mg/dl)	Triglycerides (mg/dl)	LDL- C (mg/dl)	VLDL- C (mg/dl)	HDL- C (mg/dl)
I - Normal control (1% w/v CMC)	119.67 \pm 1.282**a	115.83 \pm 1.493**a	40.33 \pm 1.833**a	23.17 \pm 0.299**a	56.17 \pm 0.833**a
II - Alloxan induced Diabetic control	233.33 \pm 2.951	163.33 \pm 1.667	168.00 \pm 2.266	32.67 \pm 0.333	29.67 \pm 0.494
III - Alloxan + EEAL (200 mg/kg, p.o)	173.83 \pm 1.014**b	150.00 \pm 1.238**b	106.33 \pm 1.077**b	30.00 \pm 0.248**b	37.50 \pm 0.563**b
IV - Alloxan + EEAL (400 mg/kg, p.o)	147.83 \pm 1.579**b	146.83 \pm 0.703**b	80.80 \pm 1.838**b	28.37 \pm 0.141**b	37.67 \pm 0.803**b
V - Alloxan + MEAL (200 mg/kg, p.o)	139.83 \pm 0.543**b	141.33 \pm 1.116**b	69.90 \pm 0.834**b	28.27 \pm 0.223**b	41.67 \pm 0.422**b
VI - Alloxan + MEAL (400 mg/kg, p.o)	123.83 \pm 1.014**b	126.50 \pm 1.118**b	52.20 \pm 1.108**b	25.30 \pm 0.224**b	46.17 \pm 0.401**b
VII - Alloxan + Glibenclamide (2.5mg/kg, p.o)	116.67 \pm 1.282**b	115.00 \pm 0.730**b	42.83 \pm 1.237**b	23.00 \pm 0.146**b	50.83 \pm 0.909**b

All values expressed in mean \pm SEM for six animals in each group (n=6);

All values compared with diabetic control groups. * $p < 0.05$; ** $p < 0.01$;

a – Normal control group I Vs diabetic control group II; b – Groups III, IV, V, VI & VII Vs diabetic control group II.

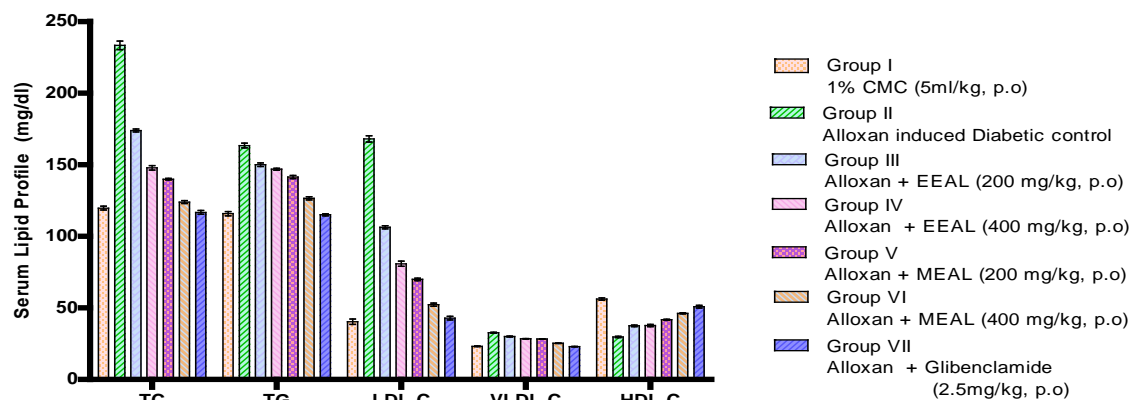


Figure 3. After Multidose of EEAL & MEAL on serum lipid profile in alloxan induced diabetic rats

All values expressed in mean \pm SEM for six animals in each group (n=6);

All values compared with diabetic control groups. * $p < 0.05$; ** $p < 0.01$;

a – Normal control group I /s diabetic control group II; b – Groups III, IV, V, VI & VII /s diabetic control group II.

Table 4. Efficiency of EEAL & MEAL on body weight in alloxan induced diabetic rats

Groups	Body weight (g)	
	Initial	Final
I - Normal control (1% w/v CMC)	164.83 \pm 1.621 ^a	194.17 \pm 0.703 ^{**a}
II - Alloxan induced Diabetic control	163.50 \pm 1.586	158.67 \pm 1.202
III - Alloxan + EEAL (200 mg/kg, p.o)	164.33 \pm 1.229 ^b	171.00 \pm 1.033 ^{**b}
IV - Alloxan + EEAL (400 mg/kg, p.o)	163.83 \pm 0.601 ^b	177.17 \pm 1.493 ^{**b}
V - Alloxan + MEAL (200 mg/kg, p.o)	164.00 \pm 1.528 ^b	173.33 \pm 1.308 ^{**b}
VI - Alloxan + MEAL (400 mg/kg, p.o)	164.33 \pm 1.174 ^b	185.17 \pm 1.138 ^{**b}
VII - Alloxan + Glibenclamide (2.5mg/kg, p.o)	167.50 \pm 1.607 ^b	187.33 \pm 0.843 ^{**b}

All values expressed in mean \pm SEM for six animals in each group (n=6);

All values compared with diabetic control groups. * $p < 0.05$; ** $p < 0.01$.

a – Normal control group I /s diabetic control group II.

b – Groups III, IV, V, VI & VII /s diabetic control group II.

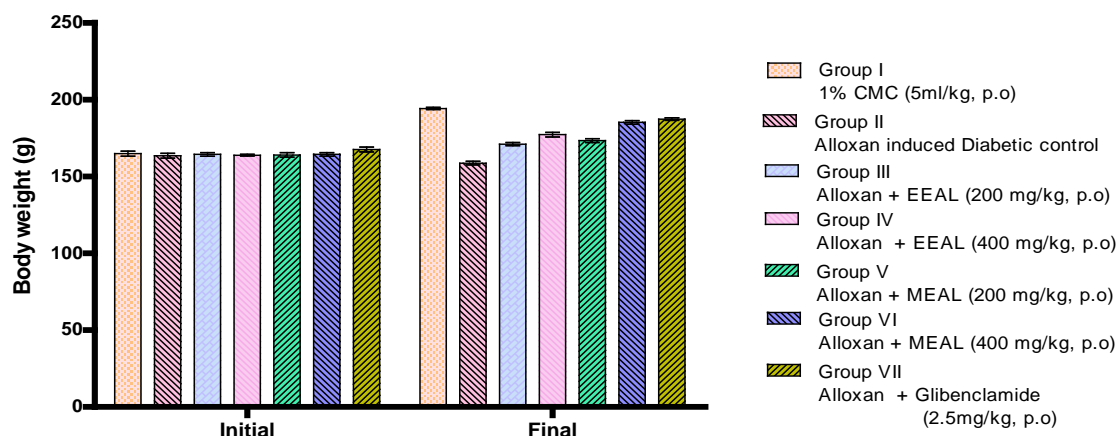


Figure 4. Efficiency of EEAL & MEAL on body weight in alloxan induced diabetic rats

All values expressed in mean \pm SEM for six animals in each group (n=6);

All values compared with diabetic control groups. * $p < 0.05$; ** $p < 0.01$.

a – Normal control group I /s diabetic control group II.

b – Groups III, IV, V, VI & VII /s diabetic control group II.

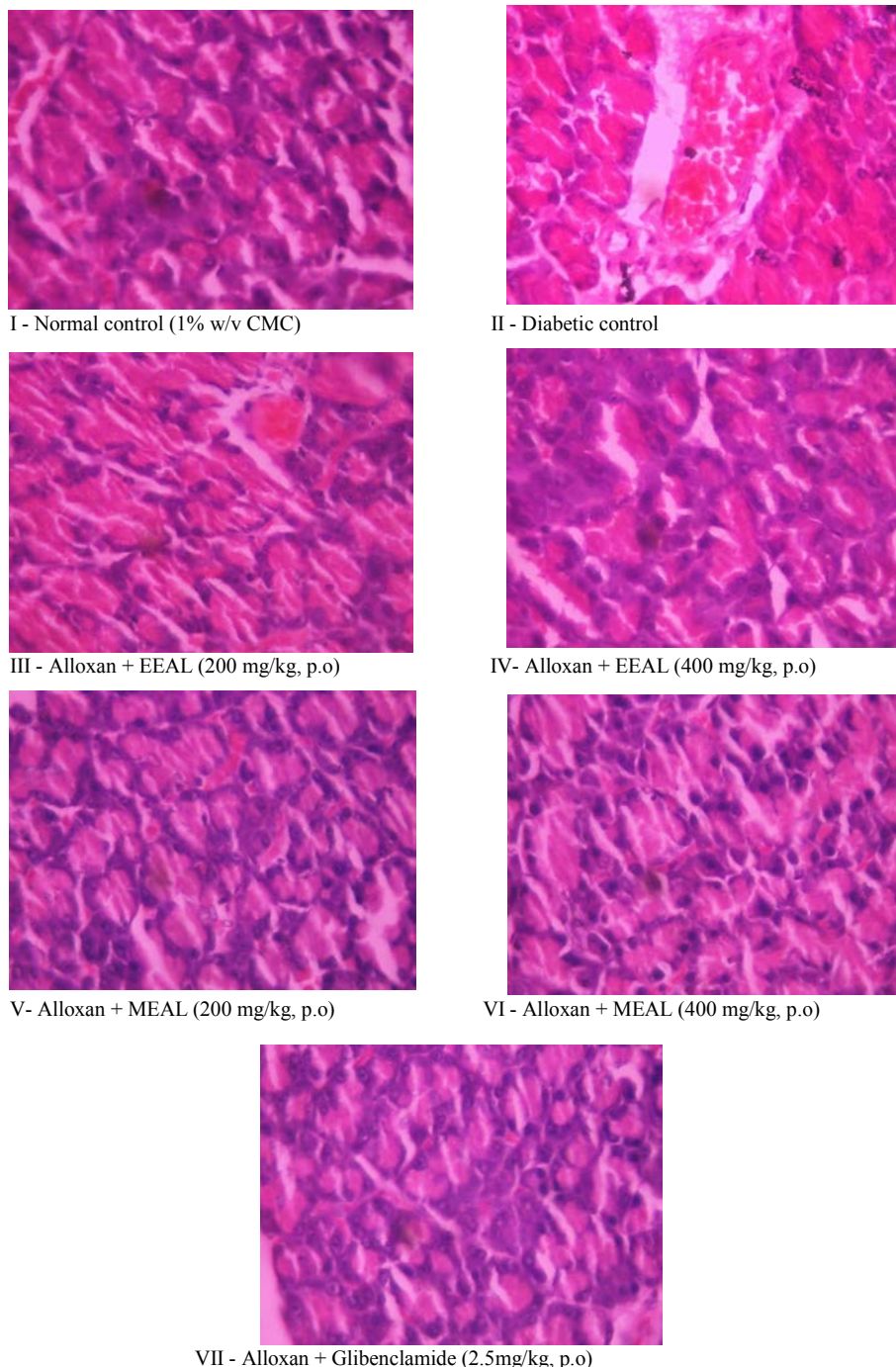


Figure 5. Histopathological studies on pancreas

4. Discussion

Ethyl acetate extract of bark of *Albizzia lucida* Benth. (EEAL) and Methanol Extract of bark of *Albizzia lucida* Benth. (MEAL) was screened for discover the scientific evidence of decoction of this bark of *Albizzia lucida* Benth. used for treatment of diabetes in Ayurveda (Sitaram and Chuneekar, 2006). Nevertheless, presence of triterpenoid saponins which possess anti-hyperglycemic, anti-hyperlipidemic and anti-angiogenic properties has been reported in previous literature (Tan *et al.*, 2008; Joseph and Jin, 2013).

The intraperitoneal injection of alloxan monohydrate (β -Cytotoxin) induces chemical diabetes in albino rats causes hyperglycemia, hyperlipidemia due to impaired insulin release and insulin resistance. The acute and sub-acute administration, the effect of EEAL and MEAL was reduced blood glucose significantly. Glibenclamide (sulphonylureas) produce hypoglycemia by enhancing the release of insulin and these drugs are very potent in alloxanized diabetic rats. Hence our results displayed that the acute and sub-acute anti-hyperglycemic effects of EEAL and MEAL might be potentiating the insulin secretion, it was similar to that of Glibenclamide (Meliani *et al.*, 2011).

In alloxanized diabetes also correlated with hyperlipidemia and hypertriglyceridemia. Observation of hyperlipidemia, the total cholesterol (TC), LDL-C, VLDL was increased & fall of HDL-C due to underutilization of glucose leads to excess mobilization of fat from adipose tissue. At the same time, oral administration of EEAL and MEAL suppresses the mobilization effect by increased glucose utilization. Moreover, EEAL and MEAL showed significantly decreases the triglycerides (TG) by activating or releasing the lipoprotein lipase resulting in regulates the metabolism of lipids (Nammi *et al.*, 2003).

Additionally, EEAL and MEAL treated animals body weight was not affected and it is comparable to that of normal control animals. In histological studies on pancreas, there is no necrosis and damage of islets of langerhans in oral administration of EEAL and MEAL treated groups. It was accepted the potency and safety of EEAL and MEAL.

In conclusion, our observations of the present research clearly demonstrated that Ethyl acetate extract of bark of *Albizia lucida* Benth. (EEAL) and Methanol Extract of bark of *Albizia lucida* Benth. (MEAL) exerts markable anti-hyperglycemic and anti-hyperlipidemic efficiency due to the presence of biologically active constituents with possible multiple mechanism involving both pancreatic and extra pancreatic effects. But still more research is needed for find out its specific mechanism of action and long term effects in diabetes mellitus.

5. Conflict of Interest

The authors declare that there are no conflicts of interest.

Acknowledgements

The authors are grateful to Padmashree Dr. M. Mohan Babu, Chairman, Sree Vidyanikethan Educational Trust, Tirupati, India for providing the necessary facilities to carry out this work.

References

- Chaudhary P, Goel B and Ghosh AK. 2012. Antidiabetic activity of *Adina cordifolia* (Roxb) leaves in alloxan induced diabetic rats. *Asian Pac J Trop Biomedicine*, S1630-S1632.
- Friedewald WT, Levy RJ and Fredrickson DS. 1972. Estimation of VLDL and LDL-cholesterol in plasma, without use of the preparative ultracentrifuge. *Clin Chem*, **18**: 499–502.
- Harbone JP. 1973. **Phytochemical Methods, A Guide To Modern Technique Of Plant Analysis**. Chapman and Hall, London, pp. 1-271.
- Holmann RR and Turner RC. 1991. **Textbook of Diabetes**. Oxford: Blackwell.
- Joseph B and D Jin D. 2013. Antidiabetic effects of *Momordica charantia* (bitter melon) and its medicinal potency. *Asian Pac J Trop Dis*, **3**(2): 93-102.
- Krishnakumar K, August KT and Vijayammal PL. 2000. Hypolipidemic effect of *Salacia oblonga* Wall. Root bark in Streptozotocin diabetic rats. *Med Sci Res.*, **28**; 65-67.
- Lachin T and Reza H. 2012. Anti-diabetic effect of cherries in alloxan induced diabetic rats. *Recent Pat Endocr Metab Immune Drug Discov*, **6**(1): 67-72.
- Meliani N, Dib M E-A, Allali H and Boufeldja T B. 2011. Hypoglycaemic effect of *Berberis vulgaris* L. in normal and streptozotocin-induced diabetic rats. *Asian Pac J Trop Biomedicine*, 468-471.
- Nammi S, Boini MK, Lodagala SD and Behara RBS. 2003. The juice of fresh leaves of *Catharanthus roseus* Linn. reduces blood glucose in normal and alloxan diabetic rabbits. *BMC Complementary and Alternative Medicine*, **3**; 1–4.
- OECD, 2002. **Acute oral toxicity**. Acute oral toxic class method guideline 423 adopted 23.03.1996. In: Eleventh Addendum to the, OECD, guidelines for the testing of chemicals organisation for economical co-operation and development, Paris, June, 2000.
- Sitaram B and Chunekar KC. 2006. **Bhavaprakasa of Bhavamisra** - Original Text Along with Commentary and Translated Including Nighantu Portion, Volume 1, Chaukhambha Orientalia Publication Varanasi, **6**; 355.
- Tan MJ, Ye JM, Turner N, Hohnen-Behrens C, Ke CQ, Tang CP, Chen T, Weiss HC, Gesing ER, Rowland A, James DE and Ye Y. 2008. Antidiabetic activities of triterpenoids isolated from bitter melon associated with activation of the AMPK pathway. *Chemistry & Biology*, **15**(3): 263-273.
- Taskimen MR. 1987. Lipoprotein lipase in diabetes. *Diabetes Metab Rev.*, **3**: 551-570.
- Valiathan MS. 1998. Healing Plants. *Current Sci.*, **75**: 1122–1126.
- Verspohl EJ. 2002. Recommended testing in diabetes research. *Planta Medica*, **68**: 581–590.
- Villasenor IM and Lamadrid MRA. 2006. Comparative anti-hyperglycemic potentials of medicinal plants. *J Ethnopharmacol.*, **104**: 129–131.

Bacteriological and Mycological Assessment for Water Quality of Duhok Reservoir, Iraq

Yahya A. Shekha¹, Hero M. Ismael² and Akhter A. Ahmed^{2,*}

¹Environmental Science Department,

²Biology Department, College of Science- Salahaddin University- Erbil, Iraq.

Received June 24, 2013

Revised: August 6, 2013

Accepted: August 21, 2013

Abstract

Duhok dam reservoir is situated in Duhok city, Iraq. It is an artificial reservoir which supplies water for crops land and orchard around the canal throughout its path. The objective of this investigation is to assess physical, chemical and microbiological aspects of aquatic ecosystem in the lake. The water quality variables (water temperature, pH, EC, total dissolved nitrogen and phosphate, SO₂, BOD₅ and microorganisms) were being measured seasonally during 2011. The results reported that high conductivity and sulphate concentrations were recorded during different seasons. Heterotrophic plate count and faecal coliform exceed Iraqi and WHO standards for drinking purposes. Statistically, no differences were found between studied sites for all variables. Microbiological isolates, counts, total occurrence and diversity index were more in sediment than in water samples. The occurrence of mycobiota was surveyed by three isolation methods 19 fungal species assigned to 14 genera were isolated. The most frequent species in order were: *Aspergillus* spp., *Penicillium* spp. and *Eurotium* spp. The occurrence of keratinophilic fungi was detected in the sediment by hair baiting method. The most frequent genera isolated in this study were *Chrysosporium* spp., *Trichophyton* spp. and *Microsporum* spp.

Keyword: Water Quality, Duhok Reservoir, Bacterial Count, Fungal isolation, Biodiversity.

1. Introduction

Water quality performs an important role in the health of human beings, animals, and plants. Surface water quality is an essential component of the natural environment and a matter of serious concern today (Liu *et al.*, 2011). Rivers and reservoirs play a major role in drinking water, agricultural use, fishery, and electricity production, so protection of water quality is a very important issue and it should be kept at acceptable levels (Venkatesharaju *et al.*, 2010). The variation of water quality is the essential combination of both anthropogenic (such as urban, industrial, agricultural activities and the human exploitation of water sources) and natural contributions (such as precipitation rate, weathering processes and soil erosion) (Pejman *et al.*, 2009). Deterioration of lake and river water quality is common in many aquatic systems and potential causes are usually various including point and non- point sources of pollution (Pisinaras *et al.*, 2007).

In a well- balanced aquatic ecosystem, the quality of water plays a critical role between, the organisms and environment which is also extremely important for the health of the ecosystem (Akbulut *et al.*, 2010). In water quality assessment the microbial community has special significance, especially in terms of protecting public health.

Coliform bacteria, normally present in intestinal tract of humans and worm- blooded animals, can secondary be found on plants, in the soil and in waters. Although the occurrence of primarily non- pathogenic refers to the presence of disease- causing organisms, they reach natural waters mainly during rainfall, through runoff from agricultural and urban lands, as well as through drainage (Radojevic *et al.*, 2012). Total coliform (TC) is used as a parameter giving basic information on microbiological quality of surface waters (WHO, 2008). For more than a century the presence of coliform bacteria in drinking and recreational waters has been taken as an indication of fecal contamination, and thus of a health hazard. Total coliform and thermotolerant (fecal) coliform (FC) indicator tests are common public health tests of the safety of water and wastewater which might be contaminated with sewage or fecal material (APHA, 1998).

Historically, water has played a significant role in the transmission of human disease. Typhoid fever, cholera, infectious hepatitis, bacillary and amoebic dysenteries and many varieties of gastrointestinal diseases can all be transmitted by water (Rompere *et al.*, 2002).

The qualitative and quantitative composition of fungi in water sediments depend on the origin and composition of waste water sediments, stabilization degree of their organic

* Corresponding author. e-mail: akhter_micro@uni-sci.org.

matter, hydration degree and structure. It was postulated that keratinophilic fungi may be utilized as microbiological indices for the transformations of organic matter of waste water sediments as well as of the degree of their deactivation from the sanitary standpoint (Ulfig and Korcz, 1991).

Duhok dam is a high earth fill dam with central clay core and gravel shell. The main aim of the dam was irrigation of the agricultural areas inside Duhok city and areas around it till Summel city through a tunnel, now the reservoir area of the dam is used for supplying Duhok city with water beside it became a touristic area (Mustafa and Noori, 2013). It is 60 m (197 ft) tall and can withhold 52,000,000 m³ (42,157 acre.ft) of water. The dam has a bell-mouth spillway with a maximum discharge of 81 m³/s (2,860 cu ft/s) (Wikipedia, 2008).

The main objectives of this study were to assess the microbial water quality by detecting the presence of coliforms and fungi, as well as to determine the spatial and temporal pattern in the community structure of biota richness and relations to some physico- chemical parameters, in order to know the water quality of Duhok dam and their suitability to using it for different purposes.

2. Materials and Methods

2.1. Study Area

Duhok dam reservoir was established in 1988 on Duhok rubar (River) forming an impoundment of surface area around 256 hectares, coming third in Iraqi Kurdistan region, northern part of Iraq (Figure 1) after Dokan and Derbandikan reservoirs. Duhok impoundment is located about 2 Km to the north of Duhok city center of latitude 36° 50' 49" and longitude 43° 00' 33". It is an artificial reservoir with maximum depth exceeding 60 meter having an area of 6.8 Km² (1.7 Km width, 4 Km length) (Al-Nakhabandi, 2002).

The reservoir is situated on a hilly plane surrounded from the south by a mountain Chai Spii, whereas from the north by a chain of Zahio mountain. Most parts of the area are formed from the slopes and steep mountains crossed by numerous valleys all of which generally direct the water flow of rain, and snow melts through different creeks and canals to the main Duhok rubar, because of the steep slopes toward the center of the reservoir. This reservoir (rubar Duhok canal) supplies water for crops land and orchard around the canal throughout its path (Al-Ganabi, 1985). The establishment of the reservoir extended the irrigation nowadays to about 4600 hectares on the west of Duhok town, extending to approach the main international road Zakho. The source of the water to the dam is mainly rain, snowmelt and the main tributaries Sundor and Gurmava that on their joining made up Duhok rubar.

The impoundment has almost a fan- like shape with an elongated part to the north west, where the forest vegetation and orchards have been permanently under the water, the west and east shores of the impoundment are arable lands, therefore, the use of the fertilizers and pesticides are common, whereas in Spring and Summer seasons views of cattle and sheep for grazing were common in the area. The outflow of the impoundment is through the spillway discharge.



Figure 1. Maps of: Duhok city, Iraq, Duhok Dam Lake and studied sites.

2.2. Sample Collection

Water samples were collected at surface (0- 20cm) from three sites, while sediment samples were collected from two sites at depth 1m near banks of the reservoir (sites 1 and 3) in four different seasons during 2011. All samples were kept in a 2L sterilized plastic bottles, and stored in insulated cooler containing ice and delivered on the same day to laboratory and all samples were kept at 4 °C until processing and analysis.

2.3. Analytical Methods

2.3.1. Physico- chemical analysis

Water quality parameters includes: water temperature was measured by using a thermometer (accurate to nearest 0.1 °C), pH using pH- meter (Philips, 4014, UK), electrical conductivity using (EC meter, Philips, 4025, UK). The BOD₅ by the Winkler azid method, Sulfate by titrimetric method (APHA, 1998). Total nitrogen wet mineralized by using potassium persulphate as described by (MacKareth *et al.*, 1978), persulphate digestion method was used for total dissolved phosphate as described by (Lind, 1979).

2.3.2. Microbiological analysis

For the bacteriological analysis of water samples, Coliform test was performed by the most probable number (MPN) technique (Benson, 1998) and heterotrophic plate count (aerobic) by Pour Plate method (Sugita *et al.*, 1993). Standard MPN technique was applied using glucose azide broth for isolation of fecal streptococci (APHA, 1998).

Detection of *Salmonella* spp was done by the enrichment of water samples on Selenite F broth, followed by isolation of the typical organism on selective medium, Xylose Lysine Deoxycholate Agar (XLD). Detection of *Vibrio cholerae* was done by enriching the samples in 1% alkaline peptone water for 6 to 8 hours followed by isolation on Thiosulphate Citrate Bile salt sucrose (TCBS) agar medium (Collee *et al.*, 1996). For *Pseudomonas aeruginosa* both MacConkey agar, Nutrient agar were used as presumptive cultures and Mannitol salt agar and Blood agar were used to isolate *Staphylococcus aureus* (Benson, 1998). All colonies with different characteristics on their selective media were identified on the basis of their colonial, morphological and biochemical properties following Bergey's Manual of Determinative Bacteriology (Holt *et al.*, 1994).

The fungi were isolated from the water samples seasonally by using two methods: The direct plate and the dilution plate, two types of growth media were used for isolation of fungi Potato dextrose agar (PDA) and Sabouraud's dextrose agar (SDA) supplemented with chloramphenicol (50 mg/l) and cycloheximide (500 mg/l). While for the isolation of fungi from soil sediment two methods were used: A dilution of 10⁻³ was chosen for the estimation of the fungal total count and the hair bait technique of Vanbreuseghem (1952) was used to isolate keratinophilic fungi. For this purpose, sterile Petri dishes were half filled with the soil samples and moistened with water and baited with burying sterile human hair in the soil. These dishes were incubated at room temperature (20 ± 1 °C) and examined for fungal growth over a period of four weeks. After observing the growth under a stereoscopic binocular microscope it was cultured on SDA supplemented with chloramphenicol (50 mg/l) and cycloheximide (500 mg/l) (Deshmukh and Verekar 2006).

2.4. Statistical Analysis

Statistical analysis was conducted for the data using software program (SPSS version 20). One way ANOVA (Analysis of variance). Post hoc test (Duncan) was applied to determine significant differences in spatial and in temporal variation. All data are expressed as mean ± SE. A

P value of 0.05 was considered as the limit for statistical significance.

3. Results and Discussion

Seasonal variation of physico-chemical and bacteriological characteristics are given in Table 1. The maximum temperature was 28 °C recorded during summer and minimum during winter 9 °C. The fluctuation in water temperature usually depends on the season, geographic location and sampling time.

Table 1. Seasonal variation of physico- chemical and biological characteristics of water for Duhok dam. (data represented as mean± SE).

Variables	Winter	Spring	Summer	Autumn
Water temperature (°C)	9.0± 0.0 ^a	28± 0.0 ^b	27.1± 0.16 ^c	22± 0.0 ^d
pH	7.7± 0.057 ^a	8.3± 0.081 ^b	8.0± 0.043 ^c	7.9± 0.052 ^c
EC (µs.cm ⁻¹)	753± 0.88 ^a	817± 4.72 ^b	1237± 6.22 ^c	1038± 0.66 ^d
Total Nitrogen (µg.l ⁻¹)	18.6± 6.21 ^a	117.9± 37.7 ^b	98.6± 30.0 ^b	5.8± 2.26 ^a
Total phosphate (µg.l ⁻¹)	1.5± 0.31 ^a	7.98± 4.52 ^a	7.4± 3.80 ^a	0.73± 0.18 ^a
Sulphate (mg.l ⁻¹)	548.8± 9.93 ^a	518± 39.3 ^a	486± 39.5 ^a	579± 13.8 ^a
BOD ₅ (mg.l ⁻¹)	1.26± 0.14 ^a	1.16± 0.16 ^a	2.2± 0.15 ^b	0.93± 0.07 ^a
Heterotrophic plate count (CFU.ml ⁻¹)	291± 18 ^a	348000± 326 ^a	51600± 351 ^a	1296± 266 ^a
Fecal coliform MPN.100m ⁻¹)	1.0± 0.1 ^a	8.3± 1.3 ^a	10.6± 1.5 ^a	0.0± 0.0 ^a
Fungi (CFU.ml ⁻¹)	1.0± 0.05 ^a	7.6± 0.28 ^b	1.6± 0.07 ^a	2.6± 0.08 ^{ab}

Note: Values in each row with different letters are significantly different at *P*≤0.05 according to Duncan test. Values in rows with same letters are not significantly different.

Highest microorganisms counting recorded during spring and summer seasons. Statistically significant differences (*P*≤0.05) were observed between seasons regarding to microorganisms counts. The minimum microorganisms count in winter might be due to cold climate condition, which is not been supportive for bacterial and fungal duplication in a greater extent (Venkateshharaju *et al.*, 2010).

The pH value was more than 7 and the maximum value was recorded during spring season (pH max. 8.3). High photosynthesis rate during spring adversely affect the pH value. These results came in agreement with the results of Al- Nakshabandi (2002) who also worked on Duhok reservoir. Conductivity values varied from 753 to 1237 µs. cm⁻¹ for winter and summer seasons respectively. The high salt concentration of water shows that significant dissolution and/ or precipitation reactions are taking place in the lake depending upon the solubility constants of different minerals present in the lake (Millero, 2001). The minimum value observed in rainy season is due to dilution with the rain water and maximum owing to evaporation and high water temperature (Aqel, 2012). The maximum limit of conductivity value exceeds the desirable limits for drinking water (WHO, 1984). While, water quality classified as high salinity type C3 for irrigation purposes (Ayers and Wescot, 1994). Al- Nakshabandi (2002)

recorded higher electrical conductivity value for the same impoundment.

Total dissolved nitrogen and total dissolved phosphorus determination is important in assessing the potential biological productivity of surface waters, increasing concentration of phosphorus and nitrogen compounds in lakes and reservoirs leads to eutrophication (Welch and Tindell, 1992). Maximum concentration observed during Spring 117.9 and 7.98 $\mu\text{g.l}^{-1}$ for total nitrogen and total phosphorus respectively with minimum concentration recorded during autumn season.

The sulphate concentration of natural water is an important factor in determining their suitability to using it for different purposes (Sawyer and McCarty, 1978). High sulphate content recorded during studied period and never fall below 486 mg.l^{-1} . It exceeds maximum permissible range of Iraqi standard (1986) and WHO (2011) drinking water quality standard. Statistically significant difference ($P \leq 0.05$) was observed with conductivity value. Similar results were obtained by Al-Nakshbandi (2002) and Duhoki (1997) at the same reservoir.

BOD₅ represents the amount of oxygen that microbes need to stabilize biologically oxidizable matter. Higher BOD₅ value was recorded during Summer season (2.2 mg.l^{-1}). This may be due to high water temperature and bacterial counts. Statistically significant differences ($P \leq 0.05$) was found between different seasons.

High heterotrophic plate counts during all seasons were recorded with maximum number 348×10^3 and 51.6×10^3 CFU during Spring and Summer seasons respectively. The maximum permissible level of Iraqi drinking water standard must not exceed 50 CFU.ml⁻¹. It may reveal the human activity (Tourist and waste disposal) as bacterial population was estimated in higher concentration from water samples collected from the bank of the lake (Shafiq *et al.*, 2011).

Faecal coliforms in the present investigation exhibits more counts during summer followed by spring seasons. During both seasons it exceeds Iraqi standards (1986) for drinking purposes. The presence of FC suggests that water may have been contaminated with faeces either of human or animal origin (Omezuruike *et al.*, 2008).

Total fungal count was high during spring season (7.6 CFU.ml⁻¹), this may be related to human activities and waste disposal around lake bank.

Statistically, there are no significant differences ($P \leq 0.05$) between all studied sites for physico-chemical and biological water characteristics of Duhok reservoir (Table 2).

Table 2. Physico-chemical and biological characteristics of water for Duhok dam (data represented as mean \pm SE).

Locations	Seasons	Heterotrophic plate count (CFU.gm ⁻¹)	Fecal coliform (MPN. 100m ⁻¹)	Fungi x 10 ³ (CFU.gm ⁻¹)
Site 1	Winter	3.1×10^2	0	21
	Spring	7×10^3	4	12
	Summer	5×10^5	4	4
	Autumn	5×10^2	0	10
Site 3	Winter	2.3×10^2	21	9
	Spring	5.2×10^5	4	13
	Summer	8×10^4	21	9
	Autumn	9×10^2	0	8

As noticed in Table 3, the results showed higher microbial count for all groups in sediment samples compared with water samples. The highest heterotrophic plate count and faecal coliform count were recorded in sediments of site 3. This site characterized by the presence of some vegetative plants near the bank. The presence of coliforms group in collected samples generally suggests that a certain selection of water and sediment samples may have been contaminated with faeces either of human and animal origin and other more dangerous microorganisms could be present (Omezuruike *et al.*, 2008). Bano (2006) reported that presence of bushes, shrubs or plants makes likely possible that smaller mammals may have been coming around these water bodies to drink water, thereby passing out faeces into the water. In addition to, tourist activities nears this site that exposed to more pollutant sources.

Table 3. Seasonal variation of microbial count in sediment of Duhok dam.

Variables	Site1	Site2	Site3
Water temperature (°C)	21.5 \pm 4.3 ^a	21.5 \pm 4.3 ^a	21.6 \pm 4.4 ^a
pH	8.02 \pm 0.11 ^a	7.96 \pm 0.17 ^a	7.99 \pm 0.09 ^a
EC ($\mu\text{S.cm}^{-1}$)	967 \pm 111 ^a	960 \pm 109 ^a	957 \pm 109 ^a
Total Nitrogen (mg.l ⁻¹)	80 \pm 43.5 ^a	73.5 \pm 32 ^a	27.1 \pm 9 ^a
Total phosphate (mg.l ⁻¹)	2.55 \pm 0.9 ^a	8.63 \pm 4.2 ^a	2 \pm 0.5 ^a
Sulphate (mg.l ⁻¹)	491 \pm 36 ^a	561 \pm 22.8 ^a	546.8 \pm 12.6 ^a
BOD ₅ (mg.l ⁻¹)	1.5 \pm 0.35 ^a	1.35 \pm 0.25 ^a	1.32 \pm 0.26 ^a
Heterotrophic plate count ((CFU.ml ⁻¹))	18.2 \pm 1.03 ^a	2.21 \pm 0.81 ^a	280 \pm 24.1 ^a
Fecal coliform MPN .100m ⁻¹)	3.5 \pm 2 ^a	1.75 \pm 1.03 ^a	9.75 \pm 4.95 ^a
Fungi (CFU.ml ⁻¹)	3.75 \pm 0.24 ^a	2 \pm 0.2 ^a	4 \pm 0.52 ^a

As shown in Table 4, *E. coli* was found in water sample almost during Winter and Summer seasons. In sediments, it was more abundant and recorded in all seasons except Autumn season. *E. coli* is the most widely adopted indicator of faecal pollution and they can also be isolated and identified simply (Kumar *et al.*, 2010). *E. coli* has frequently been reported to be the causative agent of traveler's diarrhoea, urinary tract infection, hemorrhagic colitis, and haemolytic uraemic syndrome (Al-Otaibi, 2009). *Streptococcus faecalis* with *E. coli* are good indicators of gastrointestinal diseases.

The presence of such bacteria indicates the possible presence of faecal material (Leclerc *et al.*, 1996). *Pseudomonas aeruginosa* has been isolated in both water and sediment samples. *Pseudomonas* can, in rare circumstances, cause community acquired pneumonias as well as ventilator associated pneumonias, being one of the most common agents isolated in several studies (Radojevic *et al.*, 2012; Fine *et al.*, 1996). Karfistan and Arik-Colagolu (2005) enter *Pseudomonas* bacteria as indicators of microbiological water quality during their study on Manyas lake. *Staphylococcus aureus* regarded as important indicators of the whole aquatic ecosystem health, including fish, and birds via the food web (Kumar *et al.*, 2010). Most of these isolated bacteria species from Duhok water dam have been isolated in different water bodies in other studies (Bano, 1996; Omezurike *et al.*, 2008; Uzoigwe and Agwa, 2012). No detection of *Vibrio cholerae* was found in water and sediment samples of Duhok water dam.

Table 5. A total of 5 fungal genera (9 species) were identified in water samples and 9 genera (12 species) were identified in sediment sample. The most dominant species includes *Aspergillus niger* (28%), *A. flavus* (16%), *A. ochraceus* and *Penicillium* spp. (10%) in water samples, while in sediment samples the most common species were *A. niger* (22%), *Rhodotorulla* spp. (19%), *A. flavus* (16%) and *Rhizopus* spp. (10%).

Higher diversity index ($H' = 2.136$) was accounted in sediment rather than water samples ($H' = 1.956$).

The fungal communities were identified at the all four seasons in water samples. The highest number of taxa, 6 species were collected in Spring season followed by 4 species in Autumn and 3 species in Summer and Winter.

In sediment samples, highest number of taxa, 8 species were observed during Autumn season, 5 species collected during Spring and Winter seasons and 4 species were identified during Summer. Seasonal changes in the water temperature have been shown distinct effects on occurrence and percentage of fungal communities and compositions. The reason of low fungal isolation genera in Duhok water dam may be due to absent of plants growing along its banks, with exception of few macrophytes near site 3. Luo *et al.*, (2004) commented that riparian vegetation had been regarded as an important factor influencing freshwater fungal communities through availability of detritus for these organisms. Higher species isolation, occurrence and diversity index (H') and counting in sediments than in water, may be related to availability of organic detritus on sediments that supply a good sources of food and habitat for these decomposers (El-Dohlob and Ali, 1981). It seems quite clear that the availability of organic matter, pH and water temperature play important role in the existence and propagation of aquatic fungi in lakes (Mahmoud and Abou-Zeid, 2002).

Table 4. Isolation of some bacteria in water and sediment of Duhok dam.

Samples	Bacterial isolation	Seasons												Total occurrence
		Winter			Spring			Summer			Autumn			
		Site 1	Site 2	Site3	Site 1	Site 2	Site3	Site 1	Site 2	Site3	Site 1	Site 2	Site3	
Water	<i>E. coli</i>			+				+	+	+				4
	<i>Salmonella spp.</i>							+	+	+	+			4
	<i>Pseudomonas aeruginosa</i>	+		+	+									3
	<i>Streptococcus faecalis</i>				+			+						2
	<i>Staphylococcus aureus (MRSA)</i>			+										1
	<i>Vibrio cholera</i>													
Total		1		3	2		2	2	2	2				14
Sediment	<i>E. coli</i>	+		+	+			+	+		+			6
	<i>Salmonella spp.</i>							+	+		+			3
	<i>Pseudomonas aeruginosa</i>	+		+										2
	<i>Streptococcus faecalis</i>										+			1
	<i>Staphylococcus aureus (MRSA)</i>	+		+							+			3
	<i>Vibrio cholera</i>													
Total		3		3	1		2	2		4				15

Table 5. The occurrence of fungi at different seasons in water and sediment of Duhok dam

Samples	Fungal isolation	Seasons												Total occurrence
		Winter			Spring			Summer			Autumn			
		Site 1	Site 2	Site3	Site 1	Site 2	Site3	Site 1	Site 2	Site3	Site 1	Site 2	Site3	
Water (CFU.ml ⁻¹)	<i>Aspergillus candidus</i>	1						1						2
	<i>Aspergillus flavus</i>				5		4							9
	<i>Aspergillus niger</i>				2	1	3		1	1	1		2	11
	<i>Aspergillus ochraceous</i>				4									4
	<i>Emericella</i> spp.	1										2		3
	<i>Penicillium</i> spp.		1			1						2		4
	<i>Rhodotorulla glutins</i>						2							2
	<i>Cladosporium</i> spp.						1							1
	<i>Eurotium</i> sp.									2			1	3
Total		2	1		11	2	10	1	1	3	1	4	3	39
Sediment x 10 ³ (CFU.ml ⁻¹)	<i>Aspergillus flavus</i>	2		2	1		4	2		2			1	14
	<i>Aspergillus fumigates</i>				2									2
	<i>Aspergillus niger</i>			3	5		5			3			3	19
	<i>Aspergillus ochraceous</i>				4									4
	<i>Cladosporium</i> sp.										5			5
	<i>Emericella</i> sp.										2			2
	<i>Mucor</i> sp.										1			1
	<i>Neosartoria</i> sp.	2						2						4
	<i>Rhizopus</i> sp.			4						4	1			9
	<i>Rhodotorulla</i> sp.	17												17
	<i>Alternaria</i> sp.										1			1
	<i>Penicillium citrinum</i>						4						4	8
Total		21		9	12		13	4		9	10		8	86

Microsporum gypseum, *Chrysosporium* spp., *Trichophyton* spp. and *Absidia* spp. were isolated by hair-baiting technique (Table 6). Among the isolated species, *M. gypseum* and *Trichophyton* spp. were common agent of dermatophytosis (tinea) in human and animals that can cause many problems for human health (Hedayati and Mirzakhani, 2009). In addition, *Chrysosporium* spp. and *Absidia* spp. keratinophilic and saprophytic fungi, were isolated by this technique. These results are partially agree with that found by (Hedayati and Mirzakhani, 2009).

Table 6. Keratinophilic fungi isolated in sediment samples from different sites of water Sari city, by the hair-baiting technique.

Fungal genera	Spring	Summer	Autumn	Winter
<i>Absidia</i> spp.			+	
<i>Chrysosporium</i> spp.	+	+	+	+
<i>Microsporum</i> spp.	+			
<i>Trichophyton</i> spp.			+	

4. Conclusion

High water temperature was 28 °C recorded during Summer. All tested water samples were alkaline in their nature. Water quality characterized by high salt concentration according to electrical conductivity values, exceeded guideline standards for drinking and irrigation purposes. Sulphate content exceeded Iraqi standard for drinking water quality and may be attributed to some sulphur springs discharged into Duhok water dam. Fecal coliform group was detected in all studied sites. Higher microbial count, isolation and diversity index were found in sediment rather than water samples. This may be related to organic detritus which used as microhabitat and food sources for microbial growth. Isolation of some pathogenic microorganisms (bacteria and fungi) in water and sediment samples, may be due to non point sources of pollutant discharged into Duhok water dam, can cause problems for human health as used for drinking and swimming activities. Fourteen fungal genera were isolated during this investigation.

References

- Akbulut M, Kaya H, Celik ES, Odabasi DA, Odabasi SS and Selvi K. 2010. Assessment of surface water quality in Atikhisar reservoir and Saricay creek (Canakkale, Turkey). *Ekoloji*. 19, **74**: 139- 149.
- Al- Gabani HA. 1985. Duhok city , a study in urban geography. MSc. Thesis- Mosul University-Iraq. pp.102.
- Al- Nakshbandi IZR. 2002. A phyecologicalstudy on Duhok impoundment its main watershed. PhD Dissertation- Duhok University-Iraq. pp.143.
- Al-Otaibi EL. 2009. Bacteriological assessment of urban water resources in KhamisMushait governorate, southwestern Saudi Arabia. *Inter J Health Geographics*. **8**:16.
- American Public Health Association (A.P.H.A.) 1998. **Standard methods for the examination of water and wastewater**. Twentieth ed. A.P.H.A.,1015 Fifteenth Street, NW, Washington, DC. 20005-2605.
- Aql H. 2012. Preliminary investigation on the chemical, physical and microbiological properties of Dumat lake in Al- Jouf region, Saudi Arabia. *Eur J Biological Sci.*, **4**(1): 5- 12.
- Ayers R S and Westcot D. W. 1994. Water Quality for Agriculture, FAO Irrigation and Drainage Paper 29, revision 1, Food and Agriculture Organization of United Nations, Rome, Italy.
- Bano K. 2006. Nutrient load and pollution study of some selected station along Ogunpa river in Ibadan, Nigeria. M.Sc Dissertation. University of Ibadan, Nigeria.
- Benson HJ. 1998. **Microbiological applications: Laboratory manual in general Microbiology**, seventh edition, pp. 208-211.
- Collee JG, Frasher AG, Marmion BP and Simmons A. 1996. **Mackie and McCartney Practical Medical Microbiology**. Fourteenth Edition, Churchill Living Stone
- Deshmukh S K and Verekar S A. 2006. The occurrence of dermatophytes and other keratinophilic fungi from the soils of Himachal Pradesh (India). *Czech Mycol.*, **58**(1-2): 117-124.
- Dohuki MSS. 1997. Classification of some wells and springs water in Duhok governorate for irrigation purposes. M.Sc. Thesis. University of Duhok, Iraq.
- El-Dohlob M and Ali BZ. 1981. Fungal population inhabiting polluted water of the river Shatt Al-Arab and its creek at Basrah, Iraq. *J. of Univ. of Kuwait*: 235- 241.
- Fine MJ, Smith MA and Carson CA. 1996. Prognosis outcome of patient with community acquired pneumonia. *JAMA*. **275** (2): 134- 141.
- Hedayati MT and Mirzakhani M. 2009. Survey of keratinophilic fungi in sewage sludge from wastewater treatment plants of Mazandaran, Islamic Republic of Iran. *Eastern Mediterranean Health J.*, **15**(2): 451-454.
- Holt JG, Krieg NR, Senath PHA, Staley JT and Williams ST 1994. **Bergey's Manual of Determinative Bacteriology**. Ninth Edition. Baltimore Md.,Willaims and Wilkins.
- Iraqi Standards. 1986. Environmental legislation. Iraqi Directorate for Environment Protection and Improvement. Ministry of Health. Baghdad-Iraq.
- Karafistan A and Arik- Colakoglu F. 2005. Physical, chemical and microbiological water quality of the Manyas lake, Turkey. *Mitigation and adaptation Strategies for Global Change*. **10**:127 -143.
- Kumar A, Bisht BS, JoshiVD, Singh AK and Talwer A. 2010. Physical, chemical and bacteriological study of water from river of Uttarakhand. *J Human Ecol.* **32**(3): 169- 173.
- Leclerc H, Devriese LA and Mossel DAA. 1996. Taxonomical changes in intestinal (faecal) enterococci and streptococci: Consequences on their us as indicators of faecal contamination in drinking water. *J. Appl. Bacteriol.*, **81**:459- 466.
- Lind OT. 1979. **Handbook of common methods in limnology**. C.V. Company. USA. pp 197.
- Liu W, Yu H and Chung C. 2011. Assessment of water quality in a subtropical Alpine lake using multivariate statistical techniques and geostatistical mapping: A case study. *Inter J Environ Res Public Health*. **8**: 1126-1140.
- Luo J, Yin J, Cai L, Zhang K and Hyde KD. 2004. Freshwater fungi in lakeDianchi, a heavily polluted lake in Yunnan, China. *Fungal Diversity*. **16** : 93-112.
- Mackereth FJ.H, Heron J and Talling JF. 1978. **Water Analysis: Some Revised Methods For Limnologists Science Publication**. No. 36. Freshwater Biol. Assoc., Titus Wilson and SonsLtd. UK. pp 121.
- Mahmoud Y A G and Abou- Zeid A M. 2002. Zoo sporic fungi isolated from four Egyptian lakes and the uptake of radioactive waste. *Mycology*. **30**(2):76- 81.
- Millero FJ. 2001. **The Physical Chemistry of Natural Waters**. Wiley- Interscience, New York.
- Mustafa YT and Noori MJ. 2013. Satellite remote sensing and geographic information systems (GIS) to assess changes in the water level in the Duhok dam. *Inter J Water Resources and Environ Eng.*, **5**(6), 351-359.
- Omezuruike OI, Damilola AO, Adeola OT, Fajobi EA and Shittu OB. 2008. Microbiological and physico- chemical analysis of different water samples used for domestic purposes in Abeokuta and ojota, Lagos state, Nigeria. *Afr J Biotechnol.* **7**(5): 617- 621.
- Pejman AH, NabiBidhendi GR, Karbassi AR, Mehrdadi N and Esmaeili Bidhendi M. 2009. Evaluation of spatial and seasonal variations in surface water quality using multivariate statistical techniques. *Inter. J. of Environ. Sci Technol.*, **6**(3): 467- 476.
- Pisinaris V, Petulas C, Gemitzi A and Tsihrintz VA. 2007. Water quantity and quality monitoring of Kosynthosriver, norh- eastern Greece. *Global Nest J.*, **9**(3): 259- 268.

- Radojevic ID, Stefanovic DM, Comic LR, Ostojic AM, Topuzovic MD and Strfanovic ND. 2012. Total coliforms and data mining as a tool in water quality monitoring. *Afr J Microbiol Res.*, **6(10)**: 2346- 2356.
- Rompere A, Servias P, Baudart J, de-Roubin M and Laurent P. 2002. Detection and enumeration of coliforms in drinking water. Current methods and emerging approaches. *J Microbiol Methods*, **49**: 31-54.
- Sawyer CN and McCarthy PL. 1978. **Chemistry for Environmental Engineering**. 3rd Ed. McGraw-Hill Book Company. Singapore. pp. 532.
- Shafiq HB, Ajaz M and Rasool SA. 2011. Bacterial and toxic pollutants in lake of river Indus. *Pak J of Botany*, **43(3)**: 1765-1772.
- Sugita H, Okamoto N and Nakamura T. 1993. Characterization of microaerophilic bacteria isolated from the coasted waters of Tokyo Bay, Japan. *FEMS Microbiol Ecol.*, **13**: 37-46.
- Ulfing K and Korcz M G. 1991. keratynofilnewosadachsciekowych [Keratinophilic fungi in wastewater sediments]. *Roczniki Panstwowego Zakladu Higieny*, **42**: 309-15.
- Uzoigwe CI and Agwa OK. 2012. Microbiological quality of water collected from boreholes sited near refuse dumpsites in Port Harcourt, Nigeria. *Afr J Biotechnol.*, **11(13)**: 3135- 3139.
- Vanbreuseghem R. 1952. Technique biologique pour l'isolement des dermatophytes du sol. – *Ann. Soc. Belge. Med. Trop.* **32**: 173-178.
- Venkatesharaju K, Ravikumar P, Somashekar R K and Prakash KL. 2010. Physico- chemical and bacteriological investigation on the river Cauvery of Kollegal stretch in Karnataka. *Kathmandu University J Sci, Eng Technol.*, **6 (1)**: 50- 59.
- Welch E.B. and Lindell T. 1992. **Ecological Effects of Wastewater: Applied Limnology and Pollutant Effects**. 2nd Ed., Taylor and Francis Group LLC. pp.419.
- WHO. 1984. Guidelines for drinking water quality recommendation. *World Health Organization*. Geneva. **1**: 130.
- WHO. 2008. Guidelines for drinking water quality (3rd Ed.). Incorporating first and second addenda. World Health Organization Press, Switzerland. **1**: 281- 294.
- WHO. 2011. Guidelines for drinking water quality. 4th. Ed. NML. Classification: WA **675**: 541.
- Wikipedia. 2008. "Iraqi Dam Assessments". Iraq: United States Army, Corps of Engineers. 6 June 2003. Retrieved 27 February 2012. http://en.wikipedia.org/wiki/Dohuk_Dam

Effects of *Theileria lestoquardi* Infection on Haematological and Biochemical Parameters in Experimentally Infected Desert Ewes

Aisha A. Elsadig, Yousif H. Abdalla Elmansoury*, Husna M. Elbasheir, Amna E. Babiker, Aza A. Adam, Tahani O. Abdelmageed and Sabri Hussein

Department of Radioisotopes, Veterinary Research Institute, Sudan

Received: May 2, 2013

Revised: June 15, 2013

Accepted: August 22, 2013

Abstract

In an experimental infection of sheep with *Theileria lestoquardi* some biochemical and haematological parameters were studied in ten Sudanese desert ewes together with 10 clinically healthy ewes. The obtained results showed a significant ($P<0.05$) decrease in haemoglobin, packed cell volume and white blood cells counts compared to the control group. The decrease in Hb concentrations and PCV% was observed 7-10 weeks after tick application whereas; the decrease in WBCs occurred at week 5 and week 6 after tick application. There was also a significant decrease ($P<0.05$) in serum total proteins, and serum globulins values of the infected group at week 3- week7 after tick application. A significant increase ($P<0.05$) in serum urea and serum creatinine values in the infected group was noticed during the first 7-8 weeks in the infected group compared to the control one's.

Key words: *Theileria lestoquardi*, Sheep, Haematological, Serobiochemical, Sudan

1. Introduction

Tick- borne diseases of small ruminant are of highly economic importance in many countries. Malignant theileriosis of sheep and goats caused by *Theileria lestoquardi* is considered among the most important small ruminant diseases and constituted an obstacle to the industry in countries like the Sudan (Bakheit *et al.*, 2006). When the infested sheep develop theileriosis, *Theileria* schizonts are demonstrated in liver, spleen, lung, kidney lymph node and peripheral blood (Hong *et al.*, 2003).

Theileria lestoquardi, was shown to be widely distributed in main sheep grazing areas in Sudan where 16.3% of sheep surveyed showed reactive antibodies in IFAT (Salih *et al.*, 2003). Biochemical investigations were very important to understand the host parasite relationship and to study the metabolism of the parasite, which may be helpful in the diagnosis and therapeutic processes (Yadav *et al.*, 1986).

2. Materials and Methods

2.1. Experimental Animals

This study was carried out at the Department of Radioisotopes, Central Veterinary Research Laboratories, Khartoum, Sudan. It was conducted on Sudanese desert ewes. The ewes were clinically healthy, free of ticks, parasitic infections including theileriosis and *Brucellosis*.

Before the commencement of the experiments, the animals were kept in the premises of the Veterinary Research Administration for five weeks to adapt themselves to the new environment and to be examined for any clinical detectable diseases such as brucellosis, trichomoniasis, theileriosis and any reproductive disorders or abnormalities. All animals were dosed with broad spectrum antihelminthic and anti coccidial. The animals were fed on green forage and supplemented with a commercial pelleted diet. Ewes were randomly divided into two groups (A and C) of ten each. Each group was housed separately at the Department of radioisotopes barns of 20 square meters each. They were provided with metal feeding troughs and plastic containers for water. Ewes of group A were experimentally infected with *Theileria lestoquardi*, whereas; ewes in group C remained as the uninfected control.

2.2. Infection with *Theileria lestoquardi*

A ram which was naturally infected with *Theileria lestoquardi* was brought from Atbara, North Sudan by the aid of the regional veterinary research laboratory in Atbara. This ram was particularly brought as it was proved to carry 10% parasitaemia the parasitaemia was further confirmed in the department of ticks at Soba. This sheep was used as the source of infection.

Flat nymphs of *Hyalomma anatolicum* were applied on this ram to pick up the infection using ear bags according to

* Corresponding author. e-mail: yousifelmansoury@hotmail.com.

the method described by Bailey (1960). Engorged nymphs collected and kept in the laboratory to the second stage (flat adult). Infected flat adults emerging from the nymphs were allowed to feed again on the experimental ewes.

Establishment of infection was monitored daily by measuring rectal temperature. Thin blood smears from the ear vein were prepared daily, fixed with absolute methanol and then stained with fresh Giemsa stain.

2.3. Collection of Samples

Weekly Blood samples were collected from the jugular vein into plain and heparinized vacutainer tubes from each ewe throughout the study. Sera were separated and kept at -20 °C until analyzed. All the haematological parameters were estimated by the methods described by Schalm (1965).

Total protein was determined using Biuret reagent as described by King and Wooton (1965), Albumin was determined according to Bartholmew and Delany (1966) while, Urea level was determined according to the method outlined by Evans (1968) using Randox commercial kits (Diamond Road, Crumlin, Co. Antrim, United Kingdom, BT29 4QY).

2.4. Statistical Analysis

Using (SPSS) version 10.0 data were analyzed using student T-test analysis, differences were considered significant at $P < 0.05$ level.

3. Results

3.1. Haematological Parameters

3.1.1. Haemoglobin concentration

The weekly mean Hb concentration, PCV and total white blood counts were monitored for 10 weeks in both groups of sheep post tick application. The starting concentrations of Hb at the first week was >8.0 g/dL then gradual decrease was noticed in the infected group which showed lower Hb values compared to the control group (Table 1), however, the difference was significant ($P < 0.05$) in weeks 7-10 reaching lowest concentration at week 10 (< 7.0 g/dL). While in control group the mean Hb concentration remained as high as (7.6- 8.9g/dL) throughout the study period.

3.1.2. PCV

The mean values of PCV in the infected group were found to be lower than that of the control group ranging between (22-28%) and (26-29%), respectively (Table 1). However, the differences were statistically significant ($P < 0.05$) at weeks 8,9 and 10 representing the lowest value ($< 22\%$).

3.1.3. White Blood Cells (WBCs)

The mean WBCs counts were (8.1 ± 2.12 and 8.3 ± 2.24) in the control and infected group, respectively. As shown in Table (1) there was a gradual decrease in WBCs counts in the infected group and at week 5 and week 6 the counts dropped to significant levels ($P < 0.05$). However, at week 7 to week 10 the counts increased to the starting level (8-11) in both groups.

Table 1. Haematological values (mean \pm SD) in *Theileria lestoquardi*-infected Sudanese desert ewes

Weeks post tick application	Hb	PCV	WBCs
1	8.27 ± 1.20^a 8.10 ± 1.35^a	25.11 ± 4.7^a 24.11 ± 5.62^a	8.12 ± 2.12^a 8.36 ± 2.24^a
2	7.65 ± 1.12^a 6.91 ± 1.49^a	28.72 ± 8.3^a 25.56 ± 6.44^a	7.96 ± 3.23^a 7.52 ± 3.25^a
3	8.40 ± 0.95^a 7.44 ± 1.04^a	27.34 ± 5.16^a 23.00 ± 5.17^a	7.02 ± 1.82^a 6.69 ± 1.66^a
4	8.00 ± 0.61^a 7.88 ± 0.79^a	29.47 ± 4.71^a 27.88 ± 4.58^a	6.20 ± 2.56^a 6.10 ± 1.93^a
5	7.97 ± 0.78^a 7.91 ± 1.04^a	28.79 ± 4.9^a 28.11 ± 5.25^a	6.20 ± 2.84^a 4.96 ± 1.41^b
6	8.38 ± 0.64^a 8.26 ± 0.64^a	25.87 ± 4.73^a 23.00 ± 4.44^b	7.20 ± 2.25^a 6.31 ± 4.65^b
7	7.67 ± 1.13^a 6.78 ± 1.10^b	26.00 ± 5.04^a 25.43 ± 4.65^a	6.92 ± 2.11^a 6.78 ± 2.23^a
8	8.35 ± 1.82^a 4.20 ± 0.01^b	27.93 ± 5.16^a 21.89 ± 6.39^b	9.13 ± 2.16^a 9.33 ± 2.29^a
9	8.55 ± 0.33^a 6.92 ± 1.7^b	26.82 ± 5.56^a 22.15 ± 6.41^b	11.0 ± 3.24^a 8.92 ± 1.09^a
10	8.90 ± 1.13^a 6.98 ± 1.27^b	27.64 ± 5.39^a 22.30 ± 4.00^b	8.45 ± 1.75^a 8.92 ± 1.09^a

Values with different small superscripts within the same column were significantly different at $P < 0.05$

3.2. Biochemical Parameters

3.2.1. Serum Total Proteins

Decreases in the weekly serum total protein concentrations was observed for up to 10 weeks post tick application in the infected animals compared to the controls (Table 2). However, the difference was significant ($P < 0.05$) at weeks 3 – week 7, whereas, in week 8- 10, the mean values of serum total proteins showed insignificant difference ($P > 0.05$) between the two groups.

3.2.2. Serum albumin

Although the mean values of serum albumin during the sampling period of 10 weeks, were slightly lower in the infected group compared to the control group, yet the differences were insignificant ($P > 0.05$).

3.2.3. Serum globulins

A gradual decrease in the mean concentration of serum globulin was noticed in the infected group from (> 4.0 g/dl) at week 3 to (< 3.0 g/dl) at week 7 post infection (Table 2), however, at week 8 the values began to increase to the control level and by week 10 there were no significant difference in globulin concentration between both group.

3.2.4. Serum creatinine

The mean serum creatinine concentrations (mg/dl) were significantly higher in the infected group of sheep compared to the control one's (Table 2). These differences were observed during the first 7 weeks reaching the highest value (22.05 ± 14.4 mg/dl) at week 5. Then it started to drop again to the baseline level. Later, at week 10 the

creatinine levels reached its lowest values in both groups (4- 4.3 mg/dl).

3.2.5. Serum urea

The weekly serum urea concentrations varied between (30 - 34mg/dl) in the infected group which is significantly higher ($P<0.05$) than in the control group (28-31mg/dl) during the first eight weeks, whereas, during weeks 9-10, there were no significant differences in urea concentration of both groups where they showed relatively constant value (32.4-32.9mg/dl) as presented in Table 2.

Table 2. Serobiochemical values (mean± SD) in *Theileria lestoquardi*-infected Sudanese desert ewes

Weeks post tick application	Total Proteins (g/dl)	Albumins (g/dl)	Globulins (g/dl)	Creatinine (mg/dl)	Urea (mg/dl)
1	7.31±0.32 ^a	3.32±0.33 ^a	3.98±0.54 ^a	5.32±5.86 ^a	34.76±5.58 ^a
	7.34±2.05 ^a	3.31±0.82 ^a	4.03±1.37 ^a	7.59±5.03 ^b	33.86±3.19 ^b
2	7.45±0.49 ^a	3.38±0.32 ^a	4.07±0.76 ^a	5.46±0.87 ^a	32.84±3.26 ^a
	7.73±2.62 ^a	3.41±0.57 ^a	4.32±2.01 ^a	10.14±3.85 ^b	34.24±2.45 ^b
3	7.35±0.78 ^a	3.00±0.27 ^a	4.34±0.88 ^a	4.26±3.74 ^a	31.38±2.89 ^a
	5.56±1.11 ^b	2.84±0.97 ^a	2.72±1.03 ^b	8.99±5.99 ^b	32.89±7.19 ^b
4	7.27±0.37 ^a	3.12±0.32 ^a	4.15±0.51 ^a	6.83±3.59 ^a	28.51±4.10 ^a
	6.11±0.15 ^b	2.99±0.61 ^a	2.87±0.78 ^b	11.34±2.55 ^b	30.02±3.19 ^b
5	7.19±0.51 ^a	3.26±0.58 ^a	3.93±0.81 ^a	7.00±3.07 ^a	30.71±3.62 ^a
	5.73±69 ^b	3.14±0.27 ^a	2.83±0.68 ^b	22.05±14.7 ^b	33.71±6.63 ^b
6	7.24±0.40 ^a	3.24±0.24 ^a	3.97±0.51 ^a	6.16±5.51 ^a	31.52±4.92 ^a
	5.98±1.36 ^b	3.09±0.43 ^a	2.84±1.12 ^b	10.27±4.17 ^b	32.88±6.24 ^b
7	7.37±0.44 ^a	3.21±0.17 ^a	4.16±0.54 ^a	4.29±2.20 ^a	31.39±2.39 ^a
	5.83±1.25 ^b	3.07±0.22 ^a	2.98±1.21 ^b	9.21±3.59 ^b	34.78±3.90 ^b
8	7.19±0.38 ^a	3.31±0.33 ^a	3.89±0.60 ^a	10.26±1.21 ^a	31.70±4.06 ^a
	7.15±1.96 ^a	3.12±0.35 ^a	3.79±1.59 ^a	12.74±4.12 ^a	33.36±3.32 ^b
8	7.17±0.36 ^a	3.13±0.29 ^a	4.04±0.49 ^a	7.38±3.79 ^a	32.40±3.24 ^a
	7.63±1.69 ^a	3.12±0.52 ^a	4.25±0.91 ^a	7.12±4.38 ^a	32.95±11.34 ^a
10	7.19±0.59 ^a	3.15±0.31 ^a	4.15±0.52 ^a	4.30±2.81 ^a	32.31±4.25 ^a
	7.54±0.82 ^a	3.11±0.42 ^a	4.16±0.29 ^a	4.13±3.25 ^a	32.59±5.68 ^a

Values with different small superscripts within the same column were significantly different at $P<0.05$

4. Discussion

Attempts to explain the haematological changes on pathophysiological basis, revealed that in infection with *T. lestoquardi*, there was a marked decrease in haemoglobin, packed cell volume and white cell counts. This decrease fluctuated in weeks following tick application, after which, the values returned back to normal. Our findings were similar to the findings of other worker (Ahmed, 2004; Mehta *et al.*, 1988; Rayules and Hafeez, 1995; Sandhu *et al.*, 1998; Singh *et al.*, 2001). Nazifi *et al.* (2010) also reported that, as the parasitaemia increased, a significant decrease was observed in RBCs, PCV and Hb. In contrast, with an increase in the parasitaemia rate, a significant increase in the mean corpuscular volume, haptoglobin (Hp), serum amyloid A (SAA), ceruloplasmin and fibrinogen was evident. The decline of the above mentioned values might probably be attributed to the destruction of erythrocytes by macrophages in the lymph nodes, spleen and other organs of the reticuloendothelial system as previously suggested. Singh *et al.* (2001) and Omer *et al.* (2002) reported that these changes in blood parameters pictured by the decrease in haemoglobin, packed cell volume and white blood cell counts may finally lead to the occurrence of severe anemia.

With respect to the biochemical changes as a result of *T. lestoquardi* infection, the study demonstrated that there was an apparent decrease in the concentrations of serum total protein, serum albumin and serum globulins. This decrease fluctuated in weeks following the tick application, after which, these values returned back to normal. This finding is in line with that of Ahmed (2004), Singh *et al.* (2001), Ramazan and Uguruslu (2007), and Yadav and Sharma (1986). However, Sandhu *et al.* (1998) reported an insignificant decrease in these parameters. Stockham *et al.* (2000) attributed the decrease in the concentrations of the serum protein and the serum albumin to the extra vascular accumulation of proteinaceous fluids resulting from affected lymph nodes. On the other hand, Singh *et al.* (2001) and Omer *et al.* (2003) attributed the decrease in serum proteins to hypoalbuminaemia and hypoglobulinaemia arising from liver failure.

Our findings indicated that ewes infected with *Theileria lestoquardi* had higher concentrations of serum creatinine and serum urea. This increase fluctuated in week3-7 following tick application, after which, they returned to normal levels. These findings were in agreement with the findings of Ramazan and Uguruslu (2007), Ahmed (2004), and Yerham *et al.* (1998) who attributed the increase in creatinine to damage observed in the liver and kidney in babesiosis in sheep. The increase in urea level was similar to that reported by Singh *et al.* (2001) and Sandhu *et al.* (1998). However, our observation contradicts that of Omer *et al.* (2003) who showed significant decrease in urea and creatinine in cattle naturally infected with *Theileria annulata*.

5. Conclusions and Recommendations

In conclusion it could be stated that infection of desert sheep with *Theileria lestoquardi* had significantly and adversely affected the haematological and biochemical parameters under this study.

Further investigations were needed to trace the causes of such fluctuations in serum and blood parameters.

Acknowledgments

The authors are highly indebted to the general director of the Veterinary Research Institute for financial support of this study.

References

- Ahmed Y A E. 2004. Effect of bovine theileriosis on haematology and serum constituents in calves. M.Sc. dissertation. University of Khartoum.
- Bailey K P. 1960. Notes on the rearing of *Rhipicephalus appendiculatus* and their infection with *Theileria parva* for experimental transmission. *Bull. Epiz. Dis. Afri.*, **8**: 33-43.
- Bakheit M A, Seitzer U and Ahmed J S. 2006. A new recombinant protein-based ELISA for the diagnosis of malignant theileriosis of sheep and goats. *Parasitol. Res.*, **98**: 145-149.
- Bartholmew RJ and Delany AM. 1966. Determination of serum albumin. *Proc Aust Assoc Clinical Biochem.*, **1**: 214-218.
- Evans RT. 1968. Manual and automated methods for measuring urea based on a modification of its reaction with diacetyl monoxime and thiosemicarbazide. *J Clin Pathol.*, **21**(4): 527-529.
- King EJ and Wootton TDP. 1965. Micro-analysis. In: **Medical Biochemistry. Determination of Total Protein in Plasma or Serum**. pp138, Churchill, LTD, London.
- Kolmer J A, Spaulding E H and Robinson H W. 1951. **Approved Laboratory Technique**. 5th edition, Appleton Century crofts. New York. U.S.A.
- Mehta H K, Sisodia R S and Misraula R S. 1988. Clinical and haematological observation in experimentally induced cases of bovine theileriosis. *Indian J Clin Invest.*, **26**: 636.
- Nazifi SI, Razavi SM, Reiszadeh M, Zahra E and Maryam A. 2010. Diagnostic values of acute phase proteins in Iranian indigenous cattle infected with *Theileria annulata*. *Veterinarski Arhiv.*, **80** (2): 205-214.
- Omer O H, El-Malik K H, Mahmoud O M, Haroun E M, Hawas A, Sweeney D and Magzoub M. 2002. Haematological profiles in pure bred cattle naturally infected with *Theileria annulata* in Saudi Arabia. *Veterinary Parasitol.*, **107**: 161-168.
- Omer O H, El-Malik K H, Magzoub M, Mahmoud O M, Haroun E M, Hawas A and Omar HM. 2003. Biochemical profiles in Friesian cattle naturally infected with *Theileria annulata* in Saudi Arabia. *Veterinary Res Communications*, **27**(1): 15-25.
- Ramazan C and Ugur U. 2007. Changes in selected serum components in cattle naturally infected with *Theileria annulata*. *Bull Vet. Inst Pulawy*, **51**: 15-18.
- Rayulu V C and Hafeez M D. 1995. Haematological values in cattle infected with *Theileria annulata*. *J Animal Sci.*, **65**: 1202-1203.
- Salih D A, ElHusseini A M, Hayat M and Taha K M. 2003. Survey of *Theileria lestoquardi* antibodies among Sudanese sheep. *Vet. Parasitol.*, **111** (4): 361-370.
- Sandhu G S, Grewal A S, Singh A, Kondal J K, Singh J and Brar R S. 1998. Haematological and biochemical studies on experimental *Theileria annulata* infection in crossbred calves. *Vet Res Commun*, **22**: 347-354.
- Schalm O W. 1965. **The Determination of Complete Haematological Picture**. London, Bailliere, Tindall and Cassell, Ltd. pp. 65-100.
- Singh A, Singh J, Grewal A S and Brar R S. 2001. Studies on some blood parameters of crossbred calves with experimental *Theileria annulata* infections. *Vet Res Communications*, **25**(4): 289-300.
- Stockham S L, Kjemtrup A M, Conrad P A, Schmidt D A, Scott M A, Robinson T W, Tyler J W, Jonson G C, Carson C A and Cuddihoe P. 2000. Theileriosis in a Missouri beef herd caused by *Theileria buffeli*. Case report: herd investigation, ultrastructure, phylogenetic analysis, and experimental transmission. *Vet. Pathol.*, **37**: 11.
- Yadav C L and Sharma N N. 1986. Changes in blood chemical components during experimentally induced *Theileria annulata* infection in cattle. *Vet. Parasitol.*, **21**: 91-98.
- Yerham I, Hadani A and Galker F. 1998. Some epizootical and clinical aspects of ovine babesiosis caused by *Babesia ovis*- a review. *Vet Parasitol.*, **74**: 153-163.
- Yin H, Liu G, Luo J, Guan G, Ma M, Ahmed J and Bai Q. 2003. Observation on the schizont stage of an unidentified *Theileria* sp. in experimentally infected sheep. *Parasitol Res.*, **91**(1): 43-49.

Computational Prediction of Binding of Methyl Carbamate, Sarin, Deltamethrin and Endosulfan Pesticides on Human Oxyhaemoglobin

Padma Saxena *

Department of Zoology, D.A-V College, Civil Lines, Kanpur 208001 Uttar Pradesh, India Address -51/7 Vijay Nagar Kanpur, U.P. India

Received: June 29, 2013

Revised: August 20, 2013

Accepted: August 26, 2013

Abstract

Pesticides are used to control insects and pests. However, their use has become so frequent that they may create problems for non target animal species. Methyl carbamate, sarin, deltamethrin and endosulfan pesticides are used to control insects and pests. In the current study computational prediction of binding of these pesticides on human oxyhaemoglobin using Molegro Virtual Docker (MVD) and evaluating the comparative Mole Dock Score, Root-mean square deviation (RMSD), affinity, interacting residues of human oxyhaemoglobin, number of Hydrogen bond interaction, docking score, Protein steric interaction energy (Protein EvdW) and interacting interaction of residues. In the present study the energy bound conformation with lower value of selected ligands shows hydrogen bonding and electrostatic interactions. The binding affinity of selected pesticides is found to be in decreasing order i.e., sarin > methyl carbamate > deltamethrin > endosulfan. All the pesticides bind with the serine 133. Frequent alterations in the expression of serine (amino acid) due to pesticide interaction with oxyhemoglobin may lead to produce carcinogenic cells in human beings.

Key words: Pesticide, Human oxyhaemoglobin, MVD, RMSD, Protein steric interaction energy, Serine.

1. Introduction

Due to increase in population scarcity of food is likely to create immense pressure to full fill the demand of food production. Nearly 29% of food produced is destroyed by insects, pests, birds, rats, etc. With the increasing pressure for food production, there is a great increase in the use of insecticides and pesticides to reduce the loss due to insects and pests (Tomlin, 2006). Though their use reduces the loss from insects and pests, on the other hand, they have adverse effects on crops nutrient value and leads to hazardous results on non target animals as well as human beings (Dich *et al.*, 1997). Methyl carbamate, sarin, deltamethrin and endosulfan are widely used pesticides in homes, gardens and agriculture (Bradberry *et al.*, 2005, Ayaz *et al.*, 2013). They are used in commercial crops, recreational uses and control variety of pests and inhibit cholinesterase enzyme activity, thus they have similar symptomatology during exposure of acute and chronic toxicity (Lifshitz *et al.*, 1997, Burr & Ray, 2004). They can produce a variety of acute health problems and also carries several ecological risks (Lifshitz *et al.*, 1994, Goswamy *et al.*, 1994, Saxena and Saxena, 1997). Haemogrammic

studies in albino rat after pesticide intoxication were revealed by Shakoori *et al.* (1992), Saxena and Tomer (2003), Shah *et al.* (2007) and Saxena and Saxena (2010). They reported that total erythrocyte count, hemoglobin concentration, packed cell volume, mean corpuscular volume were decreased after pesticide toxicity. It becomes necessary to carry out hematological examination to evaluate the normal and abnormal physiological states of the body. Our cells require regular supply of fuel and oxygen. Blood has capability to fulfill these requirements for proper functioning of cells resulting in maintaining good health. Low hemoglobin is the main cause of anemia. Further, low hemoglobin indicates lower levels of oxygen in the blood, which often causes shortness of breath (Shakoori *et al.*, 1988; Saxena *et al.*, 2009). Low hemoglobin level may also exasperate extant heart problems (Villarini *et al.*, 1998). In this work, we report computational prediction of binding of methyl carbamate, sarin, deltamethrin and endosulfan pesticides on human oxyhaemoglobin and evaluate the comparative Mole Dock Score, RMSD, binding affinity, interacting residues of receptor human oxyhaemoglobin, number of H-bond interaction, docking score, protein EvdW and interacting Interaction of residues using MVD.

* Corresponding author. e-mail: padmasaxenadav@gmail.com.

2. Materials and Methods

Three dimensional X ray crystallized structure of human oxyhaemoglobin (PDB: 1HHO, 2.1 Å resolution) was downloaded from the protein data bank (Shaanan, 1983). The downloaded protein have the two chain hemoglobin A (oxy) (alpha chain) with 141 residues and hemoglobin A(oxy) (beta chain) with 146 residues. The protein was taken as receptor protein and most suitable site was predicted by using q sitefinder ligand binding site prediction (<http://www.modelling.leeds.ac.uk>). Because of priority of site, oxyhaemoglobin has been selected for docking with ligand methyl carbamate (CID_11722), sarin (CID_7871), deltamethrin (CID_40585), and endosulfan (CID_6434141) pesticides, recently use for plant protection and control pests in homes & gardens. The selected ligands (pesticides) were downloaded from Pub Chem Compound (<http://www.ncbi.nlm.nih.gov>). Docking study was done with Molegro Virtual Docker (MVD). It is automated docking software with fast processing. The binding site cavity detection was performed by q site finder ligand binding site prediction tool (Figure 1). The docking simulation was performed by using docking software, namely MVD for the selected pesticides (ligands) and oxyhaemoglobin (protein). It shows mole dock score, RMSD, affinity (the estimated binding affinity in kJ/mol), docking score, protein EvdW and interacting interaction (the interaction energy among the pose and the cofactor), number of H-bond and interaction between interacting residues of receptor human oxyhaemoglobin, which indicates towards the formation of stable complex among ligand and receptor molecule (Thomsen and Christensen, 2006). MVD visualizer is used for interaction site prediction.

3. Results and Discussion

The comparative results obtained (using MVD) from docking simulation are given in Table-1. The interaction analysis for binding of human oxyhaemoglobin with methyl carbamate, sarin, deltamethrin and endosulfan have been done to find out the residues that are involved in binding site residues and number of hydrogen bonds are involved in interaction among selected pesticides, Table - 2. The energy bound conformation with lower value of selected ligands shows hydrogen bonding and electrostatic interactions are given in Figures 2a, b, 3a, b, 4a, b and 5a, b, for methyl carbamate, sarin, deltamethrin and endosulfan respectively.

Docking energy for most of pesticides were found favorable for hemoglobin A (oxy) (alpha chain) rather than hemoglobin A (oxy) (beta chain) which shows that these compounds can get stuck into hemoglobin A (oxy) (alpha chain) due to positive interaction (Singh, 2012). The methyl carbamate shows very high affinity to bind with human oxyhaemoglobin and it interacts with Phe 98, Ser133,

Val132, Ser102, Leu 120 residues of human oxyhaemoglobin. All these residues involved in binding belong to the cavity-1. The methyl carbamate forms 5 hydrogen bonds with Phe 98, Ser133, Val132, Ser102, Leu 120 (Table 2 and Figure 2 (a, b)). The hydrogen bonding is very significant for interaction of biomolecules. Sarin forms low binding energy complex as compare to methyl carbamate which shows binding affinity with Ser 133, Thr 134, Phe 98, Lys 99, Ala 130, Thr137 and forms three hydrogen bond with Ser 133, Thr 134 (Table 2 and Figure 3 (a,b)). The deltamethrin forms low binding energy complex as compare to sarin, it shows binding affinity with Ser133, Ser102, Tyr 35, Ala 130, Asp 126, Phe 98, Lys 99, Leu 105 residues of human oxyhaemoglobin. Whereas, deltamethrin formed three hydrogen bond with Ser133, Ser102, Tyr 35 residues (Table 2 and Figure 4 (a,b)). The Endosulfan forms low binding energy complex as compare to methyl carbamate shows binding affinity with Ser133, Asp 126, Leu 129, Ser102, Phe 98, Lys 99, Ala 130, Tyr35 and forms one hydrogen bond with Ser133 residue of human oxyhaemoglobin [Table 2 and Figure 5 (a, b)]. The binding affinity of selected pesticides methyl carbamate, sarin, deltamethrin and endosulfan at the active site of human oxyhaemoglobin using MVD is in decreasing order i.e. sarin > methyl carbamate > deltamethrin > endosulfan. The binding of pesticide with oxyhaemoglobin also supported by *in vitro* studies. Mourad, 2005 reported a fall in hemoglobin contents after insecticide intoxication in farm workers. Further, Shakoori *et al.* (1992), Khan and Ali (1993), Saxena *et al.* (2009), and Saxena and Saxena (2010) also observed a significant reduction in hemoglobin concentration after pesticide toxicity.

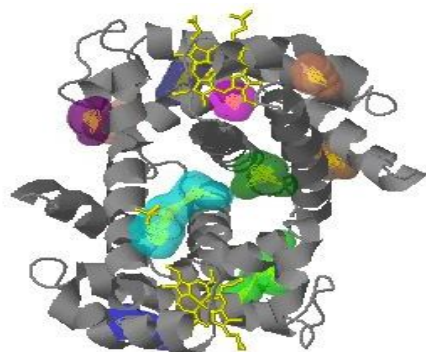
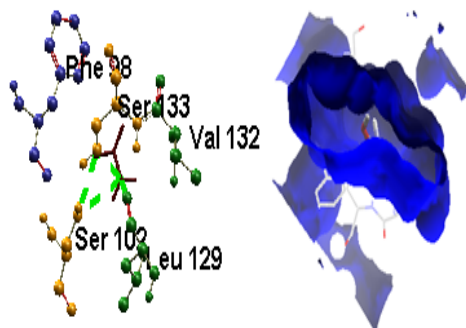
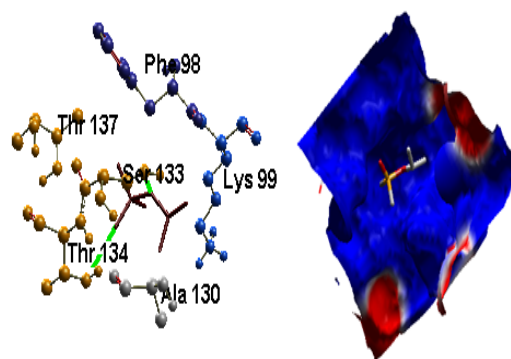
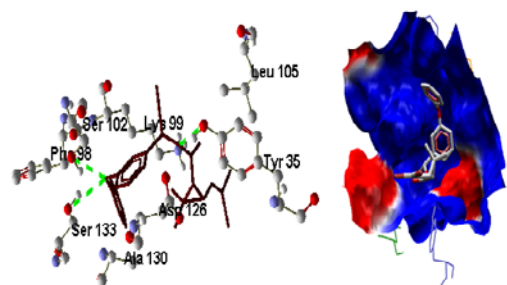
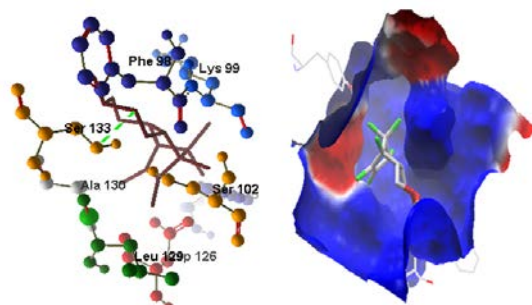
Even today our understanding of the relationship between pesticides and human health is limited due to numerous methodological problems faced in laboratory studies. Computational methods can turn out very useful for comparing *in vitro* results for new hypotheses. Docking study shows that amino acid residue i.e., Ser133 is common in binding for all the pesticides. Serine is a polar amino acid and important for catalytic function of many enzymes (Serine, 2013). Further, the OH group of serine phosphorylates and form kinase enzyme. Serine kinases also play an important role in cellular homeostasis due to phosphorylate transcription factors, regulate cell cycle, and arrange properly cytoplasmic and nuclear effectors (Blume-Jensen and Hunter, 2001; Weichenthal *et al.*, 2012). Capra *et al.* (2006) revealed that some serine kinase might play a tumor suppressor role and have a causal role in certain malignancies. The misregulation of kinases enzyme stimulate tumor growth, metastasis and poor clinical outcome (Warner *et al.*, 2003; Freeman and Whartenby, 2004; Eckerdt *et al.*, 2005). It has been also reported that frequent alterations in the expression of serine kinases causes cancer in human being (Edelman *et al.*, 1987).

Table 1. Comparative Docking Simulation Result of Selected Pesticides Methyl Carbamate, Sarin, Deltamethrin and Endosulfan with Human Oxyhaemoglobin Protein from X-ray Crystallized Data of Protein Data Bank (PDB: 1HHO) using MVD.

S. No.	Ligands	MoleDock Score	RMSD	Affinity	Protein EvdW	Intracting	Docking Score
1.	Methyl Carbamate	-45.0674	0.118821	-26.473	-34.8891	-47.1982	-48.0108
2.	Sarin	-51.4733	1.51773	-24.1619	-49.6729	-53.5958	-52.1534
3.	Deltamethrin	-124.791	1.76639	-36.9168	-129.268	-134.458	-123.268
4.	Endosulfan	-80.0703	3.52641	-42.6401	-75.845	-76.882	-80.3116

Table 2. Human Oxyhaemoglobin from X-ray Crystallized data of Protein Data Bank (PDB: 1HHO), protein residues interact with selected pesticides Methyl Carbamate, Sarin, Deltamethrin and Endosulfan using MVD (highlighted residues are involved in H-bonding interaction with ligands).

S. No.	ligands	Interacting residues of receptor Human oxyhaemoglobin	No. of H-bond interaction
1.	Methyl Carbamate	Ser133, Ser102, Leu 120 Phe 98, Val132 Ser 133, Thr 134,	05
2.	Sarin	Phe 98, Lys 99, Ala 130, Thr137, Ser133, Ser102, Tyr 35, Ala 130, Asp 126, Phe 98, Lys 99, Leu 105,	02
3.	Deltamethrin	Ser133, Asp 126, Leu 129, Ser102, Phe 98, Lys 99, Ala 130, Tyr35,	03
4.	Endosulfan		01

**Figure1.** Binding site for oxyhaemoglobin.**Figure2.** (a) Docked conformation of hydrogen bonding view and (b) with Electrostatic interaction of Methyl Carbamate with interacting Human Oxyhaemoglobin protein at the active site cavity.**Figure3.** (a) Docked conformation of hydrogen bonding view and (b) with Electrostatic interaction of Sarin with interacting Human Oxyhaemoglobin protein at the active site cavity.**Figure4.** (a) Docked conformation of hydrogen bonding view and (b) with Electrostatic interaction of Deltamethrin with interacting Human Oxyhaemoglobin protein at the active site cavity.**Figure5.** (a) Docked conformation of hydrogen bonding view and (b) with Electrostatic interaction of Endosulfan with interacting Human Oxyhaemoglobin protein at the active site cavity.

4. Conclusions

We predict that pesticides bind human oxyhaemoglobin with varying affinities, and all tested pesticides bind to serine 133 which might lead to cancer. Thus, pesticides have both short term and long term

hazardous effects and, therefore, their use on crops & plants should be limited to a certain extent.

Acknowledgement

I am thankful to C.S.J.M. University, Kanpur for providing lab facility.

References

- Ayaz AK, Shah MA and Rahman SU. 2013 Occupational exposure to pesticides and its effects on health status of workers in Swat, Khyber Pakhtunkhwa, *Pak J Biol Life Sci*, **4**(2): 43-55
- Blume-Jensen P and Hunter T. 2001. Oncogenic kinase signaling. *Nature*, **411**: 355–65
- Bradberry SM, Cage SA, Proudfoot AT, Vale JA. 2005. Poisoning due to Pyrethroids. *Toxicol Rev*, **24** (2): 93-106.
- Burr SA and Ray DE. 2004. Structure-activity and interaction effects of 14 different pyrethroids on voltage-gated chloride ion channels. *Toxicol Sci*, **77**: 341-346.
- Capra M., Nuciforo P, Confalonieri GS, Quarto MM, Bianchi, Nebuloni M, Boldorini R, Pallotti F, Viale G, Gishizky ML, Draetta GF and Fiore PPD. 2006. Frequent alterations in the expression of serine/threonine kinases in human cancers. *Can Res*, **66**:8147-8154.
- Dich J, Zahm SH, Hanberg A and Adami HO. 1997. Pesticides and cancer. *Can C Cont*, **8**(3):420-43
- Eckardt F, Yuan J and Strebhardt K. 2005. Polo-like kinases and oncogenesis. *Oncogene*, **24**: 267–76.
- Edelman AM, Blumenthal DK and Krebs EG. 1987. Protein serine/threonine kinases. *Annu Rev Biochem*, **56**: 567–613.
- Freeman SM and Whartenby KA. 2004. The role of the mitogen-activated protein kinase cellular signaling pathway in tumor cell survival and apoptosis. *Drug News Perspect*, **17**: 237–42.
- Goswamy R, Chaudhuri A and Mahashur AA. 1994. Study of respiratory failure in organophosphate and carbamate poisoning. *Heart Lung*, **23**:466-72.
- Khan SA and Ali SA. 1993. Assessment of certain haematological responses of factory workers exposed to pesticides. *Bull Environ Contam Toxicol*, **51**: 740-747.
- Lifshitz M, Rotenberg M, Sofer S, Tamiri T, Shahak E and Almog S. 1994. Carbamate poisoning and oxime treatment in children: A clinical and laboratory study. *Pediatrics*, **93**: 652-655
- Lifshitz M, Shahak E, Bolotin A and Sofer S. 1997. Carbamate poisoning in early childhood and in adults. *Clin Toxicol*, **35**:25-27.
- Mourad TA. 2005. Adverse impact of insecticidal on the health of Palestinian farm workers in the Gaza strip: A haematologic biomarker study. *Int J Occup Environ Health*, **11**:144–149.
- Saxena P, Saxena VL and Saxena AK. 2009. Cypermethrin induced toxicity in the blood of *Rattus norvegicus*. *Trends in Life Sci*, **24** (1): 59-63
- Saxena PN and Saxena P. 1997. Haemogrammic studies in albino rat after Cybil intoxication. *J Environ Biol*, **18**(4): 425-428
- Saxena PN and Tomer V. 2003. Assessment of comparative Hemotoxicity of Cybil and Fenvalerate in *Rattus norvegicus*. *Bull Environ Contam Toxicol*, **70**:839-846.
- Saxena P and Saxena AK. 2010. Cypermethrin Induced Biochemical Alterations in the Blood of Albino Rats. *Jordan J Biol Sci*, **3**(3): 111-114.
- Serine. 2013 . **The Columbia Encyclopedia**, 6th ed. Encyclopedia.com. 25Jun.2013<<http://www.encyclopedia.com>>.
- Shaanan B. 1983. Structure of human oxyhaemoglobin at 2.1 Å resolution. *J Mol Biol*, **171**: 31-59
- Shah MK, Khan A, Rizvi F, Siddique M and Rehman SU. 2007. Effect of cypermethrin on clinico-haematological parameters in rabbits. *Pak Vet J*, **27** (4):171-175.
- Shakoori AR, Ali SS and Saleem MA. 1988. Effect of six months feeding of cypermethrin on the blood and liver of albino rat. *J Biochem Toxicol*, **3**: 59-71.
- Shakoori AR, Aslam F and Sabir M. 1992. Effect of prolonged administration of insecticide (Cyhalothrin/Karate) on the blood and liver of rabbit. *Folia Biol*, **40**: 91-99.
- Singh DV, Agarwal S, Kesharwani RK and Misra K. 2012. Molecular modeling and computational simulation of the photosystem-II reaction center to address isoproturon resistance in *Phalaris minor*. *J Mol Model*, **18**:3903–3913
- Thomsen R and Christensen MH. 2006. MolDock: a new technique for high-accuracy molecular docking. *J Med Chem*, **49**(11): 3315–3321.
- Tomlin C D S. 2006. **The Pesticide Manual: A World Compendium**, 14th ed.; British Crop Protection Council: Farnham, UK : 286-287
- Villarini M, Moretti M, Pasquini R, Scassellati-Sforzolini G, Fatigoni C, Marcarelli M, Monarca S and Rodríguez AV. 1998. In vitro genotoxic effects of the insecticide deltamethrin in human peripheral blood leukocytes: DNA damage ('comet' assay) in relation to the induction of sister-chromatid exchanges and micronuclei. *Toxicol*, **130** (2-3):129-39.
- Warner SL, Bearss DJ, Han H and Von Hoff DD. 2003. Targeting Aurora-2 kinase in cancer. *Mol Can Ter*, **2**: 589–95.
- Weichenthal S, Moase C and Chan P. 2012. A review of pesticide exposure and cancer incidence in the agricultural health study cohort. *Cien Saude Colet*, **17**(1):255-70.

New Records of Arthropod Ectoparasites of Bats from North-Eastern Algeria

Mohamed Lamine Bendjeddou¹, Idir Bitam¹, Awatef Abiadh², Zihad Bouslama¹ and Zuhair S. Amr^{3,*}

¹Ecology of Terrestrial and Aquatic Systems (EcoSTaQ). University of Badji Mokhtar, BP12, Annaba 23000;

²Unité de Recherche 05/ UR/ 09-10. Biodiversité & Biologie des Populations. Faculté des Sciences de Tunis. Campus Universitaire, El Manar II, 1092, Algeria;

³Department of Biology, Jordan University of Science and Technology, P. O. Box 3030, Irbid, Jordan.

Received: July 21, 2013 Revised: August 29, 2013 Accepted: September 3, 2013

Abstract

Eight species of ectoparasites were recovered from nine bat species in north-eastern Algeria. Seven species belong to class Insecta and two species belong to class Arachnida. Insecta is represented by three orders namely, Diptera, Hemiptera and Siphonaptera. Two species of dipteran ectoparasites identified as *Phthiridium biarticulata* and *Brachytarsina falvipinnis*, belongs to Streblidae and Nycteribiidae, respectively. Order Hemiptera was represented by two species (*Cimex pipistrellus* and *Cimex lectularius*). One species of Siphonaptera was identified as *Ischnopsyllus octactenus*. Two species of arachnids were identified as *Rhipicephalus sanguineus* and *Spinturnix myoti*.

Key Words: Bats, Acarina, Flies, Hemiptera, Siphonaptera, Northeast Algeria.

1. Introduction

Our knowledge on the ectoparasites of bats of Algeria is very limited. Anciaux de Faveaux (1976) provided a list of insect parasites on Algerian bats based on previous records including Falcoz (1923), Ségué (1933) and Aellen (1955) for Nycteribiidae and Streblidae. Amr and Qumsiyeh (1993) reported five species of Nycteribiidae and Streblidae. Other reports included records of Spinturnicidae from Algeria such as Deunff (1977) and Uchikawa *et al.* (1994), and for Cimicidae (Usinger, 1966).

In the course of extensive collection of bats in north-eastern Algeria, nine bat species were collected. The present study documents the ectoparasites associated with bats collected from north eastern Algeria.

2. Materials and Methods

A total of 283 bats representing nine species were captured using mist-nets and hand-nets. Hand-nets were used to capture bats of cave roots, while mist-nets were used in open habitats. Bats were collected from eight localities within north-eastern Algeria during August,

November and December 2012. Bats were identified according to Dietz and von Helversen (2010).

Each bat was individually examined and their ectoparasites were stored and labelled in a vial containing 75% alcohol. "N" and "NP" designate the number of bats examined and the number of recovered parasites respectively. Collected ectoparasites were identified according to Hopkins and Rothschild (1956), Rudnick (1960), Dusbabek (1962), Usinger (1966), Radovsky (1967), Theodor (1967), Beaucoirn (1961), and Hürka (1982).

Collected parasites were cleared and mounted using Hoyer's medium for mites and Canada balsam for ticks and the other insects on microscopic slides. Specimens were deposited in the Pasteur Institute in Algeria (L'Institut Pasteur d'Algérie).

3. Results and Discussion

Bat species

A total of 283 bats representing nine species (Family Vespertilionidae: *Eptesicus serotinus* (N=26), *Hypsugo savii* (N=17), *Myotis capaccinii* (N=15), *Myotis punicus* (N=32), *Pipistrellus kuhlii* (N=42), *Pipistrellus pipistrellus* (N=83); Family Rhinolophidae: *Rhinolophus*

* Corresponding author. e-mail: amrz@just.edu.jo.

ferrumequinum (N=22), *Rhinolophus hipposideros* (N=18); Family Miniopteridae: *Miniopterus schreibersii* (N=28) were collected from eight localities within north-eastern Algeria (Table 1).

Table 1. Localities and bat species collected

Localities	Coordinates		Bat Species collected
	E	N	
Benmhidi	36.77	7.87	<i>M. capaccinii</i>
El Chatt	36.83	7.82	<i>M. capaccini</i>
El Hadjar	36.80	7.74	<i>P. pipistrellus</i> and <i>P. kuhlii</i>
Grotte	36.92	8.52	<i>Rh. hipposideros</i> and <i>M. schreibersii</i>
Kehf Lagareb	36.90	8.54	<i>Rh. ferrumequinum</i> and <i>M. punicus</i>
Kehf Nasser	36.92	8.55	<i>P. kuhlii</i> and <i>H. savii</i>
Sidi Amar	36.75	7.70	<i>P. pipistrellus</i>
Sidi Kassi	36.75	7.97	<i>E. serotinus</i>

Ectoparasites

Eight species of ectoparasites belonging to four orders were recovered from nine bat species from north-eastern Algeria. Three species of bat flies (Insecta: Diptera: Streblidae) *Brachytarsina falvipennis* and (Insecta: Diptera: Nycteribiidae) *Phthiridium biarticulatum* and *Penicillidia dufouri* were recovered. Order Hemiptera was represented by two species (*Cimex pipistrellus* and *Cimex lectularius*), while order Siphonaptera was represented by a single species (*Ischnopsyllus octactenus*). One ixodid tick (*Rhipicephalus sanguineus*) and one species of mesostigmatid mites (*Spinturnix myoti*) were collected (Table 2).

Class Insecta

Order Diptera

Family Streblidae

Brachytarsina flavipennis Macquart 1851

Material examined: ex. *Rh. ferrumequinum*, (N=22, NP=10), Kehf Lagareb, 10-11.VIII.2012.

Remarks: From Algeria, it was recovered from *M. schreibersii*, *R. mehelyi* and *Rh. hipposideros* (Anciaux de Faveaux, 1976) and *Myotis blythi* (Amr and Qumsiyeh, 1993). This is the only streblid known within the Mediterranean area associated with horseshow bats such as *Rh. blasi*, *Rh. ferrumequinum*, *Rh. hipposideros* and *Rh. mehelyi* from Cyprus, Jordan and Syria (Amr and Qumsiyeh, 1993; Walter and Ebenau, 1997; Ševčík *et al.*, 2013).

Family: Nycteribiidae

Phthiridium biarticulatum Hermann, 1804

Material examined: ex. *P. kuhlii*, (N=, 42, NP=16), Kehf Nasser, 8.XI.2012; ex. *Rh. ferrumequinum*, (N=22, NP=7), Kehf Lagareb, 8.XII.2012; ex. *Rh. hipposideros*, (N=18, NP=13), Kehf Lagareb, 8.XII.2012; ex. *E. serotinus*, (N=26, NP=10), Sidi Kassi, 9.XII.2012; ex. *M. punicus*, (N=32, NP=19), Kehf Lagareb, 8.XII.2012.

Table 2. Summary for ectoparasites recovered from nine species of bats from north-eastern Algeria

Bat species	Ectoparasites recovered
Family Vespertilionidae	
<i>E. serotinus</i> (Schreber, 1774)	<i>Phthiridium biarticulatum</i>
<i>H. savii</i> (Bonaparte, 1837)	<i>Rhipicephalus sanguineus</i>
<i>M. capaccinii</i> (Bonaparte, 1837)	<i>Cimex lectularius</i>
	<i>Rhipicephalus sanguineus</i>
<i>M. punicus</i> Felten, 1977	<i>Phthiridium biarticulatum</i>
	<i>Penicillidia dufouri</i>
	<i>Spinturnix myoti</i>
	<i>Rhipicephalus sanguineus</i>
<i>P. kuhlii</i> (Kuhl, 1817)	<i>Phthiridium biarticulatum</i>
	<i>Cimex pipistrelli</i>
<i>P. pipistrellus</i> (Schreber, 1774)	<i>Ischnopsyllus octactenus</i>
	<i>Cimex pipistrelli</i>
	<i>Ischnopsyllus octactenus</i>
Family Rhinolophidae	
<i>Rh. ferrumequinum</i> (Schreber, 1774)	<i>Brachytarsina flavipennis</i>
	<i>Phthiridium biarticulatum</i>
	<i>Penicillidia dufouri</i>
<i>Rh. hipposideros</i> (Bechstein, 1800)	<i>Phthiridium biarticulatum</i>
	<i>Cimex lectularius</i>
Family Miniopteridae	
<i>M. schreibersii</i> (Kuhl, 1817)	<i>Rhipicephalus sanguineus</i>

Remarks: It was previously recovered from *M. blythi*, *M. mehelyi* and *M. schreibersii* from Algeria (Amr and Qumsiyeh, 1993). This bat fly is distributed within the circum-Mediterranean and associated with cave dwelling bats such as *Rh. ferrumequinum* (Ševčík *et al.*, 2013).

Penicillidia dufouri (Westwood, 1825)

Material examined: ex. *Rh. ferrumequinum*, (N=22, NP=24), Kehf Lagareb, 9.XI.2012 and 8.XII.2012; ex. *M. punicus*, (N=32, NP=2), Kehf Lagareb, 8.XI.2012 and 8.XII.2012.

Remarks: It was previously recorded from four Algerian bat species: *Myotis blythi* and *Rh. hipposideros* (Theodor, 1967) and from *M. blythi*, *M. schreibersii* and *Rh. euryale* (Amr and Qumsiyeh, 1993). This is a bat ectoparasite with a wide range of distribution extended from Europe, North Africa and Asia, reaching to India (Krištofik and Danko, 2012). It is usually associated with bats of the genus *Myotis*, however, it was recovered from a wide range of species including *M. schreibersii* and *Rh. euryale* (Hürka, 1980). In the present study, *Rh. ferrumequinum* is an additional new host for this nycteribiid fly. It was found among 17 *Rh. ferrumequinum* individuals.

Order Hemiptera

Family Cimicidae

Cimex lectularius (Linnaeus, 1758)

Material examined: ex. *M. capaccinii*, (N=15, NP=10), Benmhidi (El-Tref), 6-7.XI.2012; ex. *Rh. hipposideros*, (N=18, NP=14), Grottem 7-8.XI.2012.

Remarks: *C. lectularius* was collected from maternity roosts of *M. myotis* (Morkel, 1999).

Cimex pipistrelli Jenyns, 1839

Material examined: ex. *P. kuhlii*, (N=42, NP=4), El Hadjar, 7.XI.2012; ex. *P. pipistrellus*, (N=83, NP=18), Sidi Amar, 6-7.XI.2012.

Remarks: The species was reported mainly among bats of the genus *Pipistrellus* within the Palearctic region (Balvín, 2008). It was recovered from other species in Europe including *Myotis myotis* and *Nyctalus noctula* (Krištofik and Danko, 2012) and from maternity roosts of *M. myotis* (Morkel, 1999).

Order Siphonaptera

Family Ischnopsyllidae

Ischnopsyllus octactenus (Kolenati, 1856)

Material examined: ex. *P. kuhlii*, (N=42, NP=4), El Hadjar, 7.XI.2012; ex. *P. pipistrellus*, (N=83, NP=16), El Hadjar, 7.XI.2012.

Remarks: This is a western Palearctic species with distribution extending from Morocco, Spain and Great Britain to Middle Asia (Hürka, 1997). This flea was reported from *P. kuhlii* in Tunisia (Beaucournu and Kock, 1996). It is mostly associated with bats of the genus *Pipistrellus* (Beaucournu and Launay, 1990).

Class Arachnida

Order Mesostigmata

Family Spinturnicidae

Spinturnix myoti (Kolenati, 1856)

Material examined: ex. *M. punicus*, (N=32, NP=55), Kehf Lagareb, 8.XI.2012 and 8.XII.2012.

Remarks: This bat mite has a wide range of distribution through Europe, North Africa and Asia (Krištofik and Danko, 2012). It is mostly associated with bats of the genus *Myotis*, but reported from other genera as well (Stanyukovich, 1997).

Order Ixodida

Family Ixodidae

Rhipicephalus sanguineus (Latreille, 1806)

Material examined: ex. *M. capaccinii*, Benmhidi, 6-7.XII.2012; ex. *M. punicus*, Kehf Lagareb, 8-19.XII.2012; ex. *M. schreibersii*, Grotte, 7-8.XII.2012; ex. *H. savii*, Kehf Nasser, 8.XI.2012 and 8.XII.2012.

Remarks: Other ixodid ticks were reported from a variety of bats such as *Ixodes simplex* and *Ixodes vespertilionis* (Krištofik and Danko, 2012). *Ixodes vespertilionis* are the most common known ectoparasites of bats in Europe (Beaucournu, 1961). Most of the recovered ticks were females, and about 20% were nymphs.

Acknowledgments

We gratefully acknowledge A. Belabed, H. Soualah-Alila, P. Christe, S. Aulagnier, J-C. Beaucournu for their advice and help. We also thank F. Vrankenne for the materials support, A. Zenoun and S. Hamami, C. Merah and B. Boudjatit for indispensable help assistance in capturing bats, and A. Beneldjouzi for the confirming the identification of ectoparasites. This study is a part of an international program of research (Eurobats and EcoSTaQ) on the management and conservation of Algerian Bats.

References

- Aellen V 1955. *Rhinolophus blasii* Peters, 1866. Chauve-scuris nouvelle pour l'Afrique du Nord. *Mammalia*, **19**:361-366.
- Amr ZS and Qumsiyeh M 1993. Records of bat flies from Jordan, Libya and Algeria. *Entomol News*, **104**:43-46.
- Anciaux de Favaux M. 1976. Distribution des Chiroptères en Algérie, avec notes écologiques et parasitologiques. *Le Bulletin de la Société d'histoire naturelle d'Afrique du Nord*, **67**:69-80.
- Balvín O. 2008. Revision of the West Palearctic *Cimex* species. Preliminary report. *Bulletin of Insectology*, **61**:129-130.
- Beaucournu J-C. 1961. Ectoparasites des Chiroptères de l'Ouest de la France. La partie: Ixodoides-Cimicidae et Nycteribiidae. *Bulletin de la Société Scientifique de Bretagne*, **36**:315-338.
- Beaucournu J-C. and Kock D. 1996. Notes on the Ischnopsyllinae of the African continent, III. Additions to the distribution of species (Insecta: Siphonaptera: Ischnopsyllidae). *Senckenbergiana Biologica*, **75**:163-169.
- Beaucournu J-C. and Launay H. 1990. Les puces (Siphonaptera) de France et du Bassin méditerranéen occidental. Faune de France 76. Fédération Française des Sociétés de Sciences Naturelles, Paris, 548 pp.
- Dietz C. and von Helversen O. (2010). Illustrated identification key to the bats of Europe. Electronic publication Version 1.0. First released 15.12.2004. Tuebingen & Erlangen (Germany)
- Deunff J. 1977. Observations sur les Spinturnicidae de la région paléarctique occidentale (Acarina, Mesostigmata) spécificité, répartition et morphologie. *Acarologia*, **8**: 602-617.
- Dusbábek F. 1962. Parasitische Fledermausmilben der Tschechoslowakei I. Fam. Spinturnicidae Oudms., 1901 (Acarina, Gamasides). *Časopis Československé Společnosti Entomologické*, **59**:357-380.
- Falcoz L. 1923. Biospeologica, No. 49. Diptera Pupipara, *Arch. Zool. Exp. Gén.*, **61**:521-552.
- Hopkins GHE. and Rothschild M. 1956. An Illustrated Catalogue of Rothschild Collection of Fleas (Siphonaptera) in the British Museum (NH). Volume II. Cotopsyllidae, Vermipsyllidae, Stephanocircidae, Ischnopsyllidae, Hypsophthalmidae, and Xiphopsyllidae. London: British Museum (Natural History), 445 pp.
- Hürka K. 1980. Čeled' Nycteribiidae – Muchulovití [Family Nycteribiidae – bat flies]. Pp.: 479-509. In: Chvála M., Hürka K., Chalupský J., Knoz J., Minář J. & Országh I. (eds.): Krevsajci mouchy a střechci – Diptera. Fauna ČSSR svazek 22 [Hematophagous Flies and Bat Flies – Diptera. Fauna of Czechoslovakia. Volume 22]. Academia, Praha, 538pp (in Czech, with summaries in Russian and English).
- Hürka K. 1982. On the insect bat ectoparasites of coastal Libya (Cimicidae, Nycteribiidae, Streblidae, Ischnopsyllidae). *Věstník Československé Společnosti Zoologické*, **46**: 85-91.
- Hürka K. 1997. New data on taxonomy and distribution of Palearctic, Oriental and Neotropical Ischnopsyllidae (Siphonaptera), Nycteribiidae and Streblidae (Diptera). *Acta Societatis Zoologicae Bohemicae*, **61**: 23-33.
- Krištofik J. and Danko S. 2012. Arthropod ectoparasites (Acarina, Heteroptera, Diptera, Siphonaptera) of bats in Slovakia. *Vespertilio*, **16**:167-189.
- Morkel C. 1999. Zum Vorkommen von an Fledermausen (Chiroptera) parasitierenden Bettwanzen der Gattung *Cimex* Linnaeus 1758 (Heteroptera: Cimicidae) in Hessen. *Hessische Faunistisch Rundbriefe (Darmstadt)*, **18**:38-48

Radovsky FJ. 1967. **The Macronyssidae and Laelapidae (Acarina: Mesostigmata) Parasitic on Bats**. Berkeley & Los Angeles: University of California Press, viii+288 pp

Rudnick A. 1960. A revision of the mites of the family Spinturnicidae (Acarina). *University of California Publications in Entomology*, **17**: 157–284

Séguy E. 1933. Mission saharienne (Augieras-Draper, 1927-1928). Insectes Dipteres. *Bull. Mus. Natl. Hist. Nat. Paris*, **5**:70-76 and 122-127.

Stanyukovich MK. 1997. Keys to the gamasid mites (Acari, Parasitiformes, Mesostigmata, Macronyssoidea et Laelaptoidea) parasiting bats (Mammalia, Chiroptera) from Russia and adjacent countries. *Rudolstädter Naturhistorische Schriften*, **7**: 13–46.

Ševčík M., Benda P. and Lučan RK. 2013. Diptera Pupipara from bats of two large eastern Mediterranean islands, Crete and Cyprus. *Turkish J Zool*, **37**:31-37.

Theodor O. 1967. An illustrated catalogue of the Rothschild Collection of Nycteribiidae (Diptera) in the British Museum (Natural History), with keys and short descriptions for the identification of subfamilies, genera, species and subspecies with an Introduction by Miriam Rothschild. British Museum (Natural History), London, viii+506 pp.

Uchikawa K., Meng-Yu Z., O'Connor BM and Klompen H. 1994. Contribution to the taxonomy of genus *Spinturnix* (Acari: Spinturnicidae), with the erection of a new genus, *Emballonuria*. *Folia Parasitologica*, **41**: 283-304.

Usinger RL. 1966. *Monograph of Cimicidae*. College Park: Entomological Society of America, xi+585 pp.

Walter G. and Ebenau C. 1997. Nachweise von Fledermaus fliegen aus Syrien (Diptera: Streblidae, Nycteribiidae). *Zool Middle East*, **14**:115-119.

Evaluation of Immunomodulatory Effects of Antiepileptic Drug Phenytoin

Mohammad A. Al- Fararjeh^{*}, Mohammad H. Jaber and Yaseen S. Abdelrahman

Department of Medical Laboratory Sciences, Faculty of Allied Health Sciences,

The Hashemite University, B.O.Box :13115, Zarqa, Jordan.

Received: June 30, 2013 Revised: September 2, 2013 Accepted: September 6, 2013

Abstract

The immunomodulatory effects of phenytoin (PHN), used as an antiepileptic agent were investigated *in vivo* using female Balb/c mice. The main aim of this study was to investigate the immunotoxicity of PHN. Animals were divided into six groups, eight animals per group. Group I, served as a control group, received only the vehicle. Groups (II-VI) were received a daily intraperitoneal dose of PHN (2.5, 5, 10, 15 and 25 mg/kg, respectively) over a period of 21 days. PHN has shown a significant decrease in the animal body weight. The relative weights of animal's spleens were also decreased significantly at doses of (10, 15, and 25 mg/kg). PHN showed a significant decrease in the percentage of circulating neutrophils and lymphocytes and an increase in the percentage of circulating eosinophils. The result showed a marked suppression in antibody production capacity as a humoral immune response and a suppression in the Delayed type hypersensitivity response as a cell mediated immune response in PHN-treated mice compared to the control group. Detectable changes have also been noticed in the histology of the footpad tissue and spleen.

Key words: Phenytoin, Immunotoxicity, Balb/c mice, DTH, Humoral Immune Response, Hemagglutination Titer Assay.

1. Introduction

The Immune system is a collection of organs, cells and tissues that work together to protect our body from potentially harmful infectious agents and certain tumor cells (Ponce *et al.*, 2009). Immunotoxicology is an important portion of the safety evaluation of drugs and chemicals (Descotes, 2006). Immunotoxicology studies focuses on the modulation of the immune system following exposure to environmental chemicals and drugs. The modulation may include immunosuppression (non- specific), immunostimulation, hypersensitivity, or autoimmunity (Veraldi, 2006; Descotes, 2005). There is increasing evidence that many toxic effects on the immune system components and their functions takes place as a result of drug treatments or chemical exposure (Rooney *et al.*, 2008). Therefore, it is very important to identify and evaluate the potential effects of chemical compounds that produce immunotoxicity during the processes of drug development (Spanhaak, 2006). Determining mechanism of immunotoxicity is an important issue in understanding the clinical relevance of the observed adverse effects. For instance, changes in blood cellular elements could suggest immunosuppression (Schulte, 2006).

Phenytoin (PHN) is an antiepileptic drug which can be useful in the treatment of epilepsy. This drug acts to suppress abnormal brain activity that has been seen in seizure by reducing electrical conductance among brain cells by stabilizing the inactive state of voltage-gated sodium channel, PHN known as (5,5-diphenylimidazolidine-2,4-dione). Molecular formula: $C_{15}H_{12}N_2O_2$. It is white crystalline powder or granule with melting point 296 °C, insoluble in water, while it is soluble in ethanol, acetone and ether (Bernaskova *et al.*, 2010).

The usual human therapeutic dosage of PHN is (1 g) orally divided in 3 doses (400 mg, 300 mg, 300 mg) given at 2 hour intervals (Ratanakorn *et al.*, 1997). Due to risk of serious side effects of PHN, it must be titrated gradually over several weeks to reach this target dose range (Keppel, 1998) PHN is associated with adverse side effects, depending on dosage, side effects occurred in 1/3 of the patients, especially those whose serum concentration was more than 20 µg/ml (Beier *et al.*, 1978), increasing with higher serum concentrations and in combination therapy, including diplopia, nystagmus. Allergic skin rash, change in blood counts such as leukopenia, and impairment of hepatic function are seen and may return to normal after dose reduction (Walia, 2004). In some cases, serious allergic reaction, such as

^{*} Corresponding author. e-mail: alqudslaboratory@yahoo.com.

skin inflammation with large area flaking (exfoliative dermatitis) (Pelekanos *et al.*, 1991). Hematological side effects of antiepileptic drugs occur infrequently but remain a potential cause of severe toxicity. It is recommended that patients receiving PHN are necessary to have a complete blood count (CBC) every two weeks. According to the international monitoring guidelines, total white blood cells count (WBCs) and differential WBC count must be performed, with PHN treatment. Other hematological effects of PHN include eosinophilia also has been reported in 34% of PHN treated patients (Yang *et al.*, 2011; Bjornsson *et al.*, 2007; and Fararjeh *et al.*, 2008). The aim of the present study was to investigate the toxic effect of PHN on immune system cells and tissues in mice by assessing humoral and cell mediated immunity. Routine hematological parameters, screening for changes in organ and body weight, hemagglutination titer (HA), and delayed type hypersensitivity (DTH) in PHN-treated animal groups were tested. Histological examination of the spleen and inflamed tissue was also evaluated for each animal (Putman *et al.*, 2003).

2. Materials and Methods

2.1. Animals

Adult female balb/c mice (6-8 weeks old; 18-20 g weight) were obtained from The Hashimate University animal house, Zarqa', Jordan. Animals were housed in plastic cages containing saw-dust bedding and adapted for 1 week in the lab prior to usage. The animal's room was maintained at a temperature of ($23 \pm 2^\circ\text{C}$) with relative humidity ($50 \pm 20\%$) under a 12 hour light/dark cycle (lights on at 07:00). All animals were maintained at standard laboratory food and tap water ad libitum. Animals were cared for in accordance with the guide to the care and use of experimental animals. All procedures involving animals were reviewed and approved by the institutional review board (IRB) issued by The Hashemite University.

2.2. Chemicals

One hundred milligram of phenytoin sodium (Goedecke AG/Germany) was purchased from local drug store (Jordan). Phosphate buffered saline (PBS), RBC lysing buffer, Sheep- RBCs (SRBC) and RPMI-1640 medium were purchased from Sigma (USA).

2.3. Phenyton Administration

Phenyton (One hundred mg) was dissolved in 8 ml of absolute ethanol (12.5 mg/ml). PHN was made up in a concentrated stock solution and diluted to an appropriate dilution with PBS for intraperitoneal (i.p.) injection each day of experiment (Tomson *et al.*, 2007; Wyllie *et al.*, 1991).

2.4. Dose and Exposure Schedules

Mice were randomly divided into six groups (I-VI), each of 8 animals. Group I (control group) received Phosphate buffer saline (PBS) and the same percentage of drug solvent (ethanol). Groups II-VI (treatment groups) received the corresponding doses of PHN. Animals were treated with PHN in PBS intraperitoneally for 21 consecutive days. Phenyton was administered at a volume of 10 ml/kg. The drug doses were 0.5,

1.0, 2.0, 3.0 and 5.0 X times of the human therapeutic dose, and were equal to 2.5, 5, 10, 15, and 25 mg/kg respectively. Mice were sacrificed by cervical dislocation 4 hours after the administration of the last drug dose.

2.5. Organ and Body Weight Ratio

The weights of animals were recorded at the beginning of the study and 4 hours after the last day of treatment (day twenty one). On the day of experiment, animals were sacrificed by cervical dislocation and body organs such as spleen, liver, kidneys and thymus were removed, and weighed. Connective and adipose tissue were removed from these organs before weighing. The relative weight of the organs of each mouse was calculated as organ weight (mg)/ body weight (g).

2.6. Determination of the Hematological Parameters

Blood was collected from the retro-orbital plexus of each mouse before being sacrificed using heparinized capillary tubes. 0.2 ml of blood were collected in a sterile ($\text{K}_2\text{-EDTA}$) anticoagulated tubes (Minicollect®, Impromini, China) so that the blood to anticoagulant ratio was 1:0.075 v/v. Routine hematological parameters were assessed including hemoglobin content, packed cell volume percent (PCV%), red blood cell (RBC) count, and WBC count. Blood films were prepared and stained with Gimsa stain for each treated animal for differential WBCs counts, slides were observed under light microscope (Nikon, China). Blood samples analyses were confirmed by Mindray BC 2800 hematology analyzer (Jiangsu, China) at Hamdan institution for medical equipment.

2.6.1. Serum Antibody Titer: Hemagglutination (HA) Titer Assay

Seven days before ending the treatments, mice were immunized by i.p injection of $10^8/50\ \mu\text{l}$ Sheep-RBCs (SRBCs) in PBS (Lee, 2004). Phenyton treatments were continued up to 21 days. At the end of experimentation (day 21), sera were prepared from peripheral blood samples, then 25 μl of twofold diluted sera in PBS were placed in the wells of a U-shape 96-microtiter plates. All samples were challenged with 25 μl of 5% v/v SRBCs suspension and mixed according to (Fararjeh *et al.*, 2008). The plates were incubated at 37°C for 1 h. Hemagglutination was then observed according to (Riahi *et al.*, 2010).

2.7. Delayed-type Hypersensitivity Response (DTH)

Delayed-type hypersensitivity response (DTH) was determined for all groups using a modified method of Bin-hafeez *et al.*, 2003. On the 14th day of the treatment, animals were immunized with $10^8/50\ \mu\text{l}$ SRBCs. After seven days of immunization (day twenty of the experiment), all animals were again challenged with a booster dose of $10^8/50\ \mu\text{l}$ SRBCs in the right hind footpad according to (Bin-hafeez *et al.*, 2003; and Fararjeh *et al.*, 2008). The left hind footpad was injected with an equal volume of PBS to serve as trauma control for nonspecific swelling. Increased volume of footpad was measured 24 h after the last challenge with SRBCs using digital plethysmometer LE 7500 (Harvard, UK) and the differences between right and left hind footpad volumes were calculated (Dietert and Holsapple, 2007).

2.8. Histological Examination

The collected tissues of right footpad and spleen of each treatment group of mice were fixed in 10% formalin and sectioned using Shandon rotary microtome (Egenolf, 2011). Various sections (4-5 μ m thickness) were prepared and stained with Hematoxylin-Eosin (H-E). Histological changes in these organs were examined under light microscope (Nikon, china) by an experienced pathologist and scored according to the degrees of changes in cellular infiltration and architectural distortion as described by (Kugelberg *et al.*, 2005; and Fararjeh *et al.*, 2008).

Scoring for the presence of the mononuclear and polymorphonuclear leukocytes in the dermal layer was considered as follow; 0, neither mononuclear nor polymorphonuclear leukocytes were present; 1, (1-3 cells per field); 2, (5-7 cells per field); and 3, abundant occurrence of mononuclear and polymorphonuclear leukocytes (more than 7 cells per field) (Kugelberg *et al.*, 2005). The presence of the mononuclear and polymorphonuclear leukocytes in the muscular layer was also scored in a similar fashion. Histological changes in spleen were scored according to the white pulp, red pulp, and trabecular changes. Negative sign (-) indicates no changes observed; +, minimal changes; and ++, readily detectable changes.

2.9. Statistical Analysis

Data are presented as mean \pm standard error of the mean (SEM) of eight independent experiments. Statistical analysis was performed using a one-way analysis of variance (ANOVA) followed by Dunnett's multiple comparison post-test using Graphpad Prism version 5 software package. In the figures, asterisks represent a statistically significant, increase or decrease, compared to control, where * = $p < 0.05$, ** = $p < 0.01$ and *** = $p < 0.001$.

3. Results

3.1. Effect of PHN on Organ and Body Weight

At the beginning of the study, all animals have almost comparable body weights since none of the groups showed any significant difference between groups as calculated by multiple comparison test. None of the used doses of PHN caused mortality in the treated animal groups. Animals treated with PHN showed a significant decrease in their body weight at the highest used doses (15 and 25 mg/kg) when compared to the control group. However, neither the subtherapeutic nor therapeutic doses of PHN (2.5, 5 and 10 mg/kg, respectively) showed any significant difference to the control group. On the other hand, PHN at the doses of (10, 15 and 25 mg/kg) caused a decrease in the relative (% body weight) of spleen. However, only (15 and 25 mg/kg) PHN doses showed a significant decrease in relative weight of thymus. No effects were observed in liver and kidney relative weights at any given doses when compared to the control group (Table 1).

Table 1. Effects of PHN on Red blood cells (RBCs) parameters of different treatment groups of Balb/c mice females.

PHN (mg/kg)	RBCs parameters		
	RBCs count ($\times 10^6/\text{mm}^3$)	PCV (%)	Hemoglobin (g/dl)
Control	8.70 \pm 1.28	45.99 \pm 3.82	14.66 \pm 1.27
2.5	8.69 \pm 1.19	43.54 \pm 6.66	14.51 \pm 2.22
5	8.56 \pm 0.89	47.55 \pm 4.35	15.85 \pm 1.45
10	8.67 \pm 0.93	45.00 \pm 5.33	15.00 \pm 1.77
15	7.05 \pm 1.60	44.63 \pm 4.43	14.88 \pm 1.47
25	7.03 \pm 1.53*	45.64 \pm 5.99	15.21 \pm 1.99

Data are means \pm SEM of eight animals. (PHN; phenytoin, PCV; packed cell volume, RBC; Red blood cells). * $P < 0.05$ when compared to control.

3.2. Effect of PHN on Hematological Parameters

Hematological tests of the peripheral blood revealed that mice treated with PHN caused a significant decrease in erythrocytes count at the highest dose 25 mg/kg (Table 1). Also, PHN caused a significant decrease in leukocytes count at the doses (15, 25 mg/kg), while doses (2.5, 5, 10 mg/kg) of PHN did not show any significant change compared to the control group (Table 2). Moreover, PHN caused a significant decrease in the percentage of circulating peripheral blood neutrophils and lymphocytes at 15, 25 mg/kg doses. However, there is no significant difference in the percentage of peripheral blood leukocytes among the lower three doses. Interestingly as shown in Table 2, PHN caused a significant dose dependent increase in the percentage of eosinophils. A significant increase was also found between 5, 10, 15 and 25 mg/kg doses. While none of the PHN doses used appears to have any effect on other hematological parameters (Table 1 and Table 2).

Table 2. Effects of PHN on White blood cells (WBCs) parameters of different treatment groups of female Balb/c mice

PHN (mg/kg)	WBCs parameters				
	WBCs count ($\times 10^3/\text{mm}^3$)	Neutrophils (%)	Lymphocytes (%)	Monocytes (%)	Eosinophils (%)
Control	7.21 \pm 0.97	23.33 \pm 2.54	65.81 \pm 2.86	10.09 \pm 1.69	0.75 \pm 0.71
2.5	7.22 \pm 1.99	22.48 \pm 5.71	62.16 \pm 9.71	9.34 \pm 1.85	1.25 \pm 0.46
5	5.89 \pm 1.17	22.44 \pm 2.70	61.9 \pm 14.06	9.66 \pm 0.79	2.5 \pm 1.19*
10	5.94 \pm 1.21	22.69 \pm 3.17	56.13 \pm 17.41	10.15 \pm 1.48	2.38 \pm 1.51*
15	5.03 \pm 1.41**	19.21 \pm 2.17*	43.3 \pm 12.9**	10.18 \pm 1.83	3.5 \pm 0.9***
25	3.7 \pm 0.9***	16.4 \pm 2.2***	41.8 \pm 8.8**	8.99 \pm 1.66	4.1 \pm 1.3***

Data are means \pm SEM of eight animals. (PHN; phenytoin, WBC; white blood cells). * $p < 0.05$ when compared to control, ** $p < 0.01$ when compared to control, *** $p < 0.001$ when compared to control.

3.3. Effect of PHN on Serum Antibody titer: Hemagglutination (HA) titer

Hemagglutination titer at the doses of (15, and 25 mg/kg) showed significant inhibition ($p < 0.001$) in the concentration of the anti-SRBCs antibodies expressed as

antibody titer when compared with the control group (Figure 1).

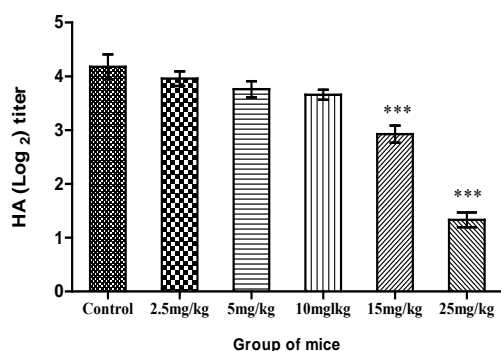


Figure 1. Effect of PHN on the anti-SRBCs antibodies production assessed by the hemagglutination log₂ titer assay. Data are means \pm SEM (n=8). *** p < 0.001 when compared to control animals

3.4. Effect of PHN on Delayed- type Hypersensitivity Response (DTH)

The subtherapeutic dose 2.5mg/kg and the therapeutic dose 5 mg/kg PHN did not show any significant change on DTH response compared to the control group. However, a significant suppression of the DTH response was determined after a subsequent injection of antigen (SRBCs) at 10 mg/kg dose or more (Figure 2).

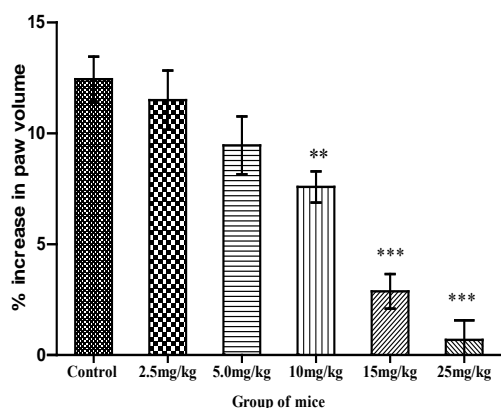


Figure 2. Effect of PHN on delayed-type hypersensitivity (DTH) measured as the difference between SRBC- injected and PBS- injected hind footpad volumes. Mice received daily i.p injection of the indicated doses of PHN for 21 days. Data are means \pm SEM of eight animals. ** p < 0.01 and *** p < 0.001 when compared to control animals.

3.5. Effect of PHN on Histology of Footpad Tissue

Microscopic examination of the footpad tissues showed an inflammatory infiltrate of leukocytes (polymorphonuclear and mononuclear leukocytes) after being challenged with SRBCs, in both hypodermis and muscle layers.

Treatment with PHN caused a significant decrease in the inflammatory infiltrate at the doses of 15 and 25mg/kg. No effects were observed in other inflammatory parameters at all doses of PHN (Table 3 and Figure 3).

Table 3. Effects of PHN treatment on right footpad tissue in mice receiving (2.5, 5, 10, 15 and 25 mg/kg), as daily i.p. injection for 21 days.

Inflamed tissue	Inflammation score (Presence of mononuclear and polymorphonuclear Leukocytes)	
Treatment mg/kg	Dermal layer	Muscular layer
Control	2.12 \pm 0.22	1.37 \pm 0.26
2.5	2.12 \pm 0.29	1.37 \pm 0.18
5	1.62 \pm 0.26	1.25 \pm 0.16
10	1.62 \pm 0.26	1.0 \pm 0.32
15	1.12 \pm 0.22*	0.87 \pm 0.22
25	0.5 \pm 0.18***	0.5 \pm 0.18*

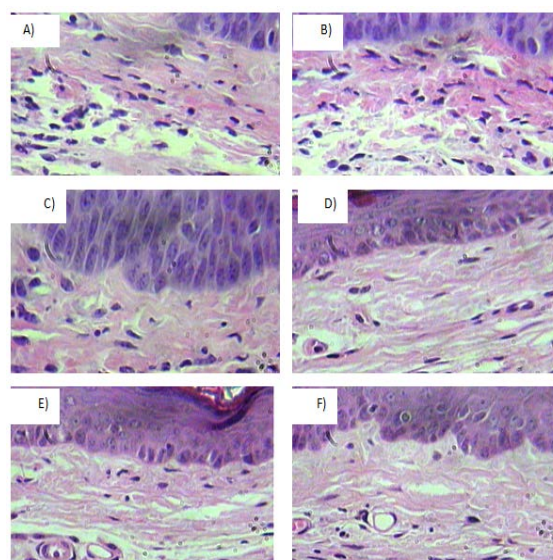


Figure 3. Inflamed footpad representative tissue sections of control and PHN-treated animals stained with hematoxylin eosin shown at high dry power field (40 x). (A); right footpad tissue of control group showing infiltration of polymorphonuclear and mononuclear leukocytes after SRBCs challenge. (B-F); PHN treated groups (2.5, 5, 10, 15 and 25 mg/kg, respectively) showing a decrease in the inflammatory infiltration of polymorphonuclear and mononuclear leukocytes.

3.6. Effect of PHN on Histology of Spleen

Microscopic examination of spleen showed a decrease in the density (atrophy) of the spleen white pulps at doses of 15 and 25 mg/kg (Figure 4). No other significant changes in other spleen histological parameters were seen at any used doses of PHN.

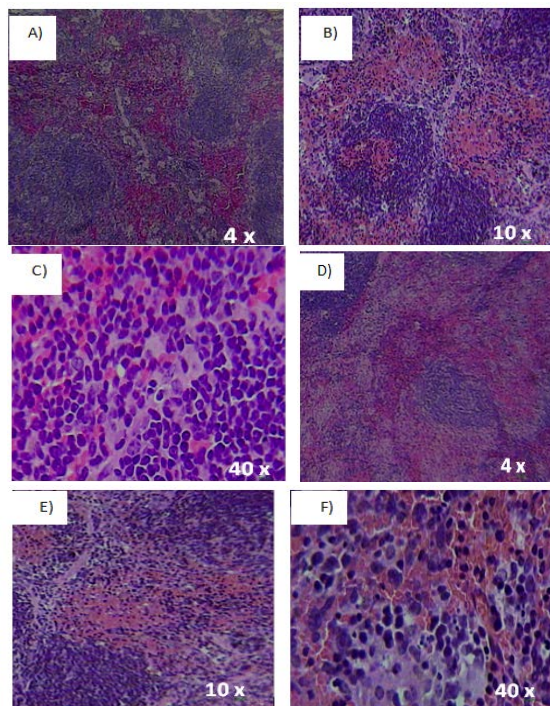


Figure 4. Spleen sections of control and PHN-treated animals stained with hematoxylin- eosin shown at different magnifications. (A-C) normal spleen (control); (D-F); 25 mg/kg PHN- treated animal showing white pulp atrophy.

4. Discussion

In the current study, the unhealthy effect of PHN has been confirmed by the significant decrease in body weights measured in PHN-treated mice at doses of 15 and 25 mg/kg. In the present study, mice treated with PHN have low food consumption. PHN alter one of the fundamental processes of the human body weight regulation, by altering appetite, metabolism, or absorption of calories. It has been noticed that mice treated with PHN exhibited mild desire for sleep which might indicate a reduction in food intake which results in body weight loss. Moreover, PHN (15, 25 mg/kg) treated mice showed a remarkable signs of behavioral abnormalities manifested by fainting and drowsiness (not calculated). These behavioral abnormalities were associated with reduction in body weights. This effect is consistent with the study of Ben-Menachem (2007) that has reported similar effect suggesting body weight loss.

Phenytoin caused a significant decrease in the relative weight of spleen at doses of (10, 15 and 25 mg/kg), this effect was confirmed by the observation of induced atrophy of the white pulp of spleen in a similar fashion. The spleen is a major lymphoid organ that plays a critical role in the primary humoral and secondary immune response. B lymphocytes partially mature in the bone marrow enter the circulation, and then populate peripheral lymphoid organs, including spleen and lymph nodes, where they complete their maturation process. Spleen white pulp plays also an important part in the initiation of immune responses by B cells to foreign antigens in the blood. This might be the reason behind the relative lymphocytopenia found in peripheral blood of treated mice.

In addition, spleen contains a large number of tissue resident leukocytes, therefore white pulp atrophy will reflect a reduction in the percentage of total leukocytes in the peripheral blood.

It has been shown that the PHN caused a significant decrease in the Red Blood Cells (RBCs) at the highest used dose (25mg/kg) which may be due to suppression of the bone marrow, that's why PHN may reduce the production of erythropoietin (EPO) hormone, thus the erythropoiesis process which is stimulated by EPO can be also affected by PHN. Likewise, PHN caused a significant decrease in the total leukocytes counts and in the percentage of neutrophils and lymphocytes. Leucopenia, neutropenia, and lymphocytopenia may reflect the toxicity or immunomodulatory effects of PHN in treated group of mice. These findings were consistent with the common occurrence of leukopenia in human model by a cohort study which has been conducted by (Blackburn, 1998).

A significant dose dependent increase in the percentage of eosinophils have been demonstrated in PHN treated mice at doses equal to or more than 5 mg/kg which is correspondent to the human therapeutic dose and this was in concordance with the study that has reported similar effect suggesting elevation of IgE with eosinophilia (Chen *et al.*, 2010). The decrease in humoral immunity response to PHN was manifested by suppression of antibodies production, tested as anti-SRBCs antibodies titer which shows a suppression of antibodies at treated doses of 15 and 25 mg/kg. White pulp atrophy, and spleen relative weight decrease observed in this study is a common finding following the administration of certain immunosuppressive drugs, and are accompanied by the decreased ability of the animal immune system to produce antibody. Suppression of DTH reaction by PHN could be not only due to the reduction in the availability of factors essential responsible for the maintenance of T cell proliferation, but also for the recruitment and activation of macrophages. Reduction in DTH may be due to any block in the antigen of SRBCs processing or presentation of these antigens by macrophages for T lymphocytes. The exact mechanism whereby PHN inhibits T cell activation is not known yet. However, it could take place via blocking of antigen specific receptors or acquisition of responsiveness to interleukins. Suppression of the delayed type hypersensitivity response has been demonstrated by PHN in this study at doses of 10, 15, and 25 mg/kg and these results were accompanied by a study by Dietert *et al.* (2010), showing that the inhibitory effects of PHN on cell mediated immune response in animal.

Moreover, DTH responses measured in the skin of the mice footpad have been used to assess cell mediated immunity in vivo. The loss of DTH reaction serves as an indicator of deteriorating cell-mediated immune function. A decreased in the infiltration of the mononuclear and polymorphonuclear leukocytes in the mouse footpad tissue, which has been demonstrated in histological examination at 15 and 25 mg/kg doses of PHN was accompanied by suppression in DTH response. These results might explain the inhibitory effect of PHN on the cell mediated immune responses.

In conclusion, the current study shows that Phenytoin had an inhibitory effect on the innate, adaptive and cell mediated as well as humoral immune response at human subtherapeutic, therapeutic and high doses. These results demonstrate an immunosuppression effect of PHN in mice. The effectiveness of PHN in the treatment of schizophrenia and other related diseases should be counterbalanced by its immunotoxicity. Further research studies should be considered to study the significance of using PHN in patients under long-term treatment. As well, studies on human models should be required to study the toxic effect of PHN on the immune system.

Acknowledgements

This work was supported by Deanship of Scientific Research, The Hashemite University. The authors wish to acknowledge Dr. Mohammad K. Mohammad from University of Jordan for providing the instruments to accomplish this work.

References

- Beier RM and Cammann RO.1978. Phenytoin intoxication and serum level. *Psychiatr Neurol Med Psychol* , **30**: 414-423.
- Ben-Menachem EO.2007. Weight issues for people with epilepsy--a review. *Epilepsia*, **9**: 42-45.
- Bernaskova KO and Mares PS.2010. Similar effects of lamotrigine and phenytoin against cortical epileptic foci in immature rats. *Physiol Res.*, **59**: 113-119.
- Bjornsson ES, Kalaitzakis EA and Olsson RF.2007. The impact of eosinophilia and hepatic necrosis on prognosis in patients with drug-induced liver injury. *Aliment Pharmacol Ther.*, **25**: 1411-1421.
- Bin-Hafeez BK, Haque RO and Parvez SA. 2003. Immunomodulatory effects of fenugreek (*Trigonella foenum graecum* L.) extract in mice. *Int Immunopharmacol.* , **3**: 257-265.
- Blackburn SC.1998. Antiepileptics and blood dyscrasias: a cohort study. *Pharmacotherapy*, **18**: 1277-1283.
- Chen YC, Chiu HC and Chu CY.2010. Drug reaction with eosinophilia and systemic symptoms: a retrospective study of 60 cases. *Arch Dermatol* , **146**: 1373-1379.
- Descotes JO. 2006. Methods of evaluating immunotoxicity. *Expert Opin Drug Metab Toxicol.*, **2**: 249-259.
- Descotes J. 2005. Immunotoxicology: role in the safety assessment of drugs. *Drug Safety*, **28**: 127-136.
- Dietert RR, Bunn TL and Lee JE. 2010. The delayed type hypersensitivity assay using protein and xenogeneic cell antigens. *Methods Mol Biol*, **598**: 185-194.
- Dietert RR and Holsapple MP.2007. Methodologies for developmental immunotoxicity (DIT) testing. *Methods* , **41**: 123-131.
- Egenolf DD.2011. Development of a murine model of lymph node metastases suitable for immunotoxicity studies. *J Pharmacol Toxicol Methods*, **63**: 236-249.
- Fararjeh MA, Mohammad MK, Bustanji YA, Alkhatib HM and Abdalla SH.2008. Evaluation of immunosuppression induced by metronidazole in Balb/c mice and human peripheral blood lymphocytes. *Int Immunopharmacol*, **8**:341-350.
- Keppel HJ.1998. 'Smart drugs' enticements on the Internet. *Ned Tijdschr Geneesk*, **142**: 977-980.
- Kugelberg EO, Norström TB, Petersen TK, Duvold TC, Andersson DI and Hughes DM.2005. Establishment of a superficial skin infection model in mice by using *Staphylococcus aureus* and *Streptococcus pyogenes*. *Antimicrob Agents Chemother*, **49**: 3435-3441.
- Lee JK.2004. Evaluation of the potential immunotoxicity of 3-monochloro-1,2-propanediol in Balb/c mice. I. Effect on antibody forming cell, mitogen-stimulated lymphocyte proliferation, splenic subset, and natural killer cell activity. *Toxicology*, **204**: 1-11.
- Ponce RA, Abad LM, Amaravadi LN, Gelzleichter TO, Gore ES, Green JF, Gupta SB, Herzyk DL, Hurst CB, Ivens IA, Kawabata TD, Maier CO, Mounho BL, Rup BV, Shankar GA, Smith HA, Thomas PF and Wierda DR. 2009. Immunogenicity of biologically-derived therapeutics: assessment and interpretation of nonclinical safety studies. *Regul Toxicol Pharmacol* , **54**: 164-182.
- Pelekanos NT.1991. Allergic rash due to antiepileptic drugs: clinical features and management. *Epilepsia*, **32**: 554-559.
- Putman EA, van der Laan JW and H. van Loveren.2003. Assessing immunotoxicity: guidelines. *Fundam Clin Pharmacol* , **17**: 615-626.
- Ratanakorn DO.1997. Single oral loading dose of phenytoin: a pharmacokinetics study. *J Neurol Sci* , **147**: 89-92.
- Riahi BC, Rafatpanah HA and Mahmoudi ML. 2010. Immunotoxicity of paraquat after subacute exposure to mice. *Food Chem Toxicol* , **48**: 1627-1631.
- Rooney AA, Yang YS and Makris SL.2008. Recent progress and diverse effects in developmental immunotoxicology: overview of a symposium at the 46th Annual SOT Meeting, Charlotte, NC. *J Immunotoxicol*, **5**: 395-400.
- Schulte AO and Ruehl-Fehlert OG .2006. Regulatory aspects of immunotoxicology. *Exp Toxicol Pathol*, **57**: 385-389.
- Spanhaak SA.2006. The ICH S8 immunotoxicity guidance. Immune function assessment and toxicological pathology: autonomous or synergistic methods to predict immunotoxicity? *Exp Toxicol Pathol*, **57**: 373-376.
- Tomson TA, Dahl ML and Kimland ED.2007. Therapeutic monitoring of antiepileptic drugs for epilepsy. *Cochrane Database Syst Rev* , **2**:123-126.
- Veraldi AC. 2006. Immunotoxic effects of chemicals: A matrix for occupational and environmental epidemiological studies. *Am J Ind Med*, **49**: 1046-1055.
- Walia KS.2004. Side effects of antiepileptics--a review. *Pain Pract*, **4**: 194-203.
- Wyllie EA. and Wyllie RS.1991. Routine laboratory monitoring for serious adverse effects of antiepileptic medications: the controversy. *Epilepsia* , **32** 5: S74-79.
- Yang CY, Dao RL, Lee TJ, Lu CW, Yang CH, Hung SI and Chung WH. 2011. Severe cutaneous adverse reactions to antiepileptic drugs in Asians. *Neurology*, **23**:2025-2033.

Prevalence of Lactose Intolerance in Primary School Children in Qena Governorate, Egypt

Sawsan M A. Abuhamdah^{1,*}, Ghaleb A. Oriquat², Tahia H. Saleem³ and Mohammed H. Hassan⁴

¹Department of Biopharmaceutics and Clinical Pharmacy, Faculty of Pharmacy, University of Jordan, P.O.Box 13380, Amman, 11942.

²Faculty of Pharmacy and Medical Sciences, Al-Ahliyya Amman University, Jordan.

³Medical biochemistry department, faculty of medicine, Assiut University, Assiut,

⁴Qena Medical Biochemistry Departments, South Valley University, Qena, Egypt

Received: February 13, 2013

Revised: June 12, 2013

Accepted: June 22, 2013

Abstract

Most people are born with the ability to digest lactose. Approximately 75% of the general population loses this ability, to some extent, going into adulthood (adult hypolactasia), while others retain such ability. The aim of the present study was to determine the prevalence of lactose intolerance, and its correlation with gastrointestinal symptoms, in primary school children aged 6-12 years in Qena Governorate, Egypt. A cross-sectional study was carried out on 300 school children with clinical suspicion of lactose intolerance. Biological and clinical data were obtained from children's parents or guardians. The history of diarrheal attacks especially following ingestion of milk or dairy products, as well as the incidence of diabetes in the children or family history of such diseases was also obtained. The children were instructed to maintain a low fiber diet without lactose for 48 hours prior to the day of examination. After 12 hours of fasting lactose tolerance test was carried out. The data obtained revealed that 74% of the participants in the study were intolerant to lactose. However, only 56.8% of lactose-intolerant children had positive clinical history of abdominal pain, abdominal distension or diarrheal attacks following ingestion of milk or dairy products. The prevalence of lactose intolerance in the studied cohort increased with age. Such genetically determined intolerance was 58% at 6-7 years of age and increased to 90% by the age of 11-12.

Keywords: Lactose Intolerance, Adult Hypolactasia, Qena Governorate, Cross-Sectional Study, Egypt

1. Introduction

Lactose is a disaccharide that is abundant in mammalian milk. It is hydrolyzed into glucose and galactose by the enzyme lactase, which is located in the brush border (microvilli) of the small intestine. In most infants, intestinal lactase activity is maximal during the perinatal period. However, after 2-12 years of age two distinct groups emerge: those with low lactase activity (hypolactasia) or a "lactase non-persistence" group, and a "lactase persistence" group of individuals who retain their neonatal level of lactase activity into adulthood (Mattar *et al.*, 2012; Genauer *et al.*, 2010; NDDIC, 2009).

Lactose intolerance is generally a lifelong inherited condition, but can be a temporary result of infection or some other insult to the jejuna mucosa. Lactose malabsorption occurs in three main types: primary, secondary and congenital. The most common form is primary adult hypolactasia. Secondary or acquired hypolactasia, can result from any gastrointestinal illness that damages the brush border or significantly increases transit time in the jejunum (Swagerty *et al.*, 2002).

Lifelong complete absence of lactase [congenital hypolactasia] is rare. Recognition of these conditions is important, as its gastrointestinal symptoms are easily managed by simple dietary adjustments (Swagerty *et al.*, 2002; Matter *et al.*, 2012). Searching available literature did not reveal any previous studies regarding the prevalence of lactose intolerance in Upper Egypt. The Ethics Committee of Assiut University; approved the protocol of the present study and all experimental procedures were in accordance with the Helsinki Declaration of 1975, as revised in 1983. The present cross-sectional study was undertaken to determine the prevalence of lactose intolerance among primary school children in Qena governorate and the extent of cooperation of the community in such work. It was felt that determining the extent of this problem is important, as part of an overall study to categorize the different types of diarrheal diseases, which constitute serious morbid conditions in young school children in the less privileged communities.

* Corresponding author. e-mail: smaabuhamdah@gmail.com or s.abuhamdah@ju.edu.jo.

2. Subjects and Methods

A cohort of 300 children aged 6-12 years, including 153 males and 147 females in different primary schools in Qena governorate was recruited to participate in the study. Exclusion criteria included children with known chronic illnesses or family history of such diseases like diabetes, children with overnight fasting blood glucose level >126 mg/dl, those with any intestinal or allergic disorders or those with history of recent gastroenteritis to avoid causes of secondary hypolactasia.

The aim of the study was explained to the parents or guardians and written consents were obtained. Biological and clinical data were obtained through a written protocol including name, gender, age, weight, history of diarrheal attacks, especially following ingestion of milk or dairy products as well as chronic diseases or family history of such diseases, including diabetes mellitus.

Diagnosis of lactose intolerance depended mainly on family history and blood glucose level following ingestion of standard doses of lactose (Swagerty *et al.*, 2002; Law *et al.*, 2010). For performing the lactose tolerance test, the parents were instructed to maintain the children on a low fiber diet without lactose for 48 hours prior to examination. After 12 hours fasting, a baseline concentration of blood glucose was measured using portable glucometer (Medi Smart of Switzerland-Brilliant). Subjects then ingested an oral load of lactose of 2 g/kg body weight, with a maximum of 50 g, as 20% aqueous solution. Blood glucose level was measured again after 2 hours. The test was considered positive when intestinal symptoms occurred and the increase in blood glucose level was less than 20 mg/dl above the fasting level (Joneja, 2003; Law *et al.*, 2010).

2.1. Statistical Analysis

The data were subjected to statistical analysis and tabulation using SPSS program Version 10. Means and standard deviations were used to describe numeric variables. The variables of categorical types were given as a number and percentages. Comparisons of continuous variables were performed using the Student-t test for independent samples.

Differences were considered significant when $p < 0.05$. Comparisons for categorical variables were carried out using the χ^2 test value calculated using the Epi Info 2000 Program. Results were presented to fulfil the objectives of the study.

3. Results

There was equal sex distribution in the studied cohort, where 51% were males and 49% were females. The measured body weights of the children were used to calculate the oral lactose load. Biological and clinical history data are presented in Table 1. The overall results showed that 222 children were lactose intolerant, representing 74% of the studied group. There were no statistically significant differences between the lactose tolerant and intolerant children with regard to age and gender. However, a positive correlation could be detected between lactose intolerance and history of gastrointestinal symptoms of abdominal pain and distension or diarrhea

following ingestion of dairy products. The concentrations of fasting blood glucose and the two hours postprandial glucose level in the different age groups are presented in Table 2. It was found that the prevalence of lactose intolerance steadily increased with age from 58% in children aged 6-7 years up to 90% in the 11-12 year group (Figure 1).

The results also indicated that only 56.8% of lactose intolerant children had positive clinical history of gastrointestinal symptoms, following ingestion of milk or other dairy products, while such symptoms were negative in the remaining 43.2%. The prevalence of the positive symptoms in the children in different age groups is presented in Table 1.

Table 1. Body weight ranges and history of symptoms in the studied children.

Age Group (years)	Male/Female Ratio	Body Weight Range (Kg)	Positive History of G.I Symptoms
6-7	27/23	20-25	54%
7-8	23/27	20-35	30%
8-9	26/24	20-35	48%
9-10	26/24	20-50	56%
10-11	27/23	20-50	62%
11-12	24/26	31-over 50	56%
Overall	153/147		74%

Table 2. Concentrations of fasting blood glucose and two-hours postprandial in Lactose intolerant and Lactose tolerant children.

Age Group (years)	Lactose Intolerant Children		Lactose Tolerant Children	
	Fasting Blood Glucose mg/dl	Two-hours postprandial mg/dl	Fasting Blood Glucose mg/dl	Two-hours postprandial mg/dl
6-7	93.3 ± 7.77	91.3 ± 8.33	92.9 ± 9.28	139.3 ± 18.73*
7-8	93.0 ± 7.96	90.7 ± 8.17	90.6 ± 9.27	137.1 ± 13.79*
8-9	93.6 ± 8.39	94.8 ± 7.55	88.7 ± 10.54	130.0 ± 10.76*
9-10	94.1 ± 7.50	93.1 ± 7.75	93.0 ± 9.21	133.9 ± 6.25*
10-11	92.6 ± 7.28	92.0 ± 7.66	83.75 ± 6.84	126.5 ± 8.88*
11-12	92.8 ± 8.14	94.6 ± 7.60	82.40 ± 4.92	123.8 ± 10.91*

Data presented as Mean ± Standard Deviation (Mean ± SD)

*Significantly different from the fasting blood glucose level

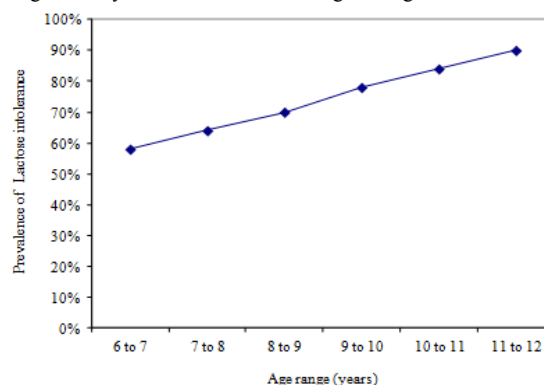


Figure 1. Prevalence of Lactose intolerance steadily increased with age

4. Discussion

Lactose intolerance, also called lactase deficiency or hypolactasia is the inability to digest lactose, a disaccharide found in milk and to a lesser extent in milk-derived dairy products. It is estimated that 75% of adults worldwide show some decrease in lactase activity during adulthood (Mattar *et al.*, 2012, Morales *et al.*, 2011, Swagerty *et al.*, 2002). Such prevalence is very close to the overall result obtained in the present work, where 74% of the studied cohort was found to be lactose intolerant. The observation in the present study that the incidence of lactose intolerance increases with age is important since many published reports treated the studied population as a single unit and paid incomplete attention to age specific considerations (Tursi, 2004).

Lactose intolerance is a slowly progressive decline in available activity of the enzyme lactase. Such activity can be influenced by intestinal transit time and/or the additional foods concomitantly consumed with lactose (Morales *et al.*, 2011; Hollox, 2005). In this respect, it should be recognized that lactose malabsorption is neither a homogeneous event nor an all-or-none phenomenon having its origins in a single etiology (Swallow, 2003). The appearance of GI symptoms depends on the amount of ingested milk and the degree of lactase deficiency. In case of partial lactase deficiency the GI symptoms appear only on ingestion of large amounts of milk or milk products (Heyman, 2006), this can be explained on the basis that the prevalence of GI symptoms depends on the amount of ingested milk and the degree of lactase deficiency, as they may have partial lactase deficiency so GI symptoms appear only on ingestion of large amounts of milk or dairy products statuses (Semenza *et al.*, 2000; Troelsen, 2005; Heyman, 2006). Children should be investigated for other causes for the appearance of the GI symptoms, like for example milk allergy (Shaw & Davies, 1999; Labayen *et al.*, 2001). The lack of correlation with gender probably indicates that if the clinical condition is genetically determined, it is not linked to the sex chromosomes. A genotyping study should be carried out to determine the variant that confers lactose tolerance in the studied population, which was beyond the scope of the present work.

5. Conclusion

The Prevalence of lactose intolerance in primary school children in Qena governorate is progressively increased with age. Not all lactose intolerant children have positive clinical history of abdominal pain and/or abdominal distension and or diarrheal attacks following ingestion of milk or milk products as this related to many factors mainly, the amount of lactose ingested and the degree of activity of lactase enzyme. Symptoms usually

disappear when you remove milk products or other lactose containing products from the diet.

References

- Enattah NS, Sahi T, Savilahti E, Terwilliger JD, Peltonen L and Järvelä I. 2002. Identification of a variant associated with adult hypolactasia. *Nat Genet.*, **30** (2):233-237.
- Genauer C and Hammer H. 2010. Maldigestion and malabsorption. In: Feldman M, Friedman LS and Sleisenger MH, (Eds), **Sleisenger & Fordtran's Gastrointestinal and Liver Disease**. 9th ed. Saunders Elsevier. Philadelphia. Chapter, 101.
- Heyman MB. 2006. Lactose intolerance in infants, children and adolescents. *Pediatr*, **118** (3):1279-1286.
- Hollox E. 2005. Evolutionary Genetics: Genetics of lactase persistence fresh lessons in the history of milk drinking. *Eur J Hum Genet.*, **13** (3): 267-269.
- Joneja JV. 2003. **Dealing with Food Allergies**. Bull Publishing, pp. 136-149.
- Labayen I, Forga L, González A, Lenoir-Wijnkoop I, Nutr R and Martínez JA. 2001. Relationship between lactose digestion, gastrointestinal transit time and symptoms in lactose malabsorbers after dairy consumption. *Aliment Pharmacol Ther.*, **15** (4):543-549.
- Law D, Conklin J and Pimentel M. 2010. Lactose intolerance and the role of the lactose breath test. *Am J Gastroenterol.*, **105** (8):1726-1728.
- Mattar R, de Campos Mazo DF and Carrilho FJ. 2012. Lactose intolerance: diagnosis, genetic, and clinical factors. *Clin Exp Gastroenterol*, **5**:113-121.
- Morales E, Azocar L, Maul X, Perez C, Chianale J and Miquel JF. 2011. The European lactase persistence genotype determines the lactase persistence state and correlates with gastrointestinal symptoms in the Hispanic and Amerindian Chilean population: a case-control and population-based study. *BMJ open*, **1** (1):1-8.
- Semenza G, Auricchio S and Mantei N. 2000. Small intestinal disaccharidases. In: **The Metabolic and Molecular Basis of Inherited Disease**. Scriver CR, Beaudel AL, Sly WS and Valle D (Eds), McGraw-Hill, NY, pp. 1623-1650.
- Shaw AD and Davies GJ. 1999. Lactose intolerance: problems in diagnosis and treatment. *J Clin Gastroenterol*, **28** (3):208-216.
- Swagerty DL, Jr., Walling AD and Klein RM. 2002. Lactose intolerance. *Am Fam Physician*, **65** (9):1845-1850.
- Swallow DM. 2003. Genetics of lactase persistence and lactose intolerance. *Annu Rev Genet*, **37**:197-219.
- The National Digestive Diseases Information Clearinghouse (NDDIC) 2009. Lactose intolerance. NIH Publication No. 09-2751.
- Troelsen JT. 2005. Adult-type hypolactasia and regulation of lactase expression. *Biochimica et Biophysica acta*, **1723** (1-3):19-32.
- Tursi A. 2004. Factors influencing lactose intolerance. *Eur J Clin Invest.*, **34** (4):314-315.

Appendix A Reviewers 2013

We would like to acknowledge especially the contributions of the following reviewers during the year 2013

- | | |
|--|--|
| Abdalla, Maher Y.
<i>The Hashemite University, Jordan</i> | Alawi, Mahmoud A-S.
<i>University of Jordan</i> |
| Abdel Khalik, Kadry
<i>Sohag University, Egypt</i> | Al-Bustanji, Yasser K.
<i>University of Jordan, Jordan</i> |
| Abdel-Aziz, Nahla M.
<i>Cairo University, Egypt</i> | Al-Eisawi, Dawud
<i>University of Jordan, Jordan</i> |
| Abdelkader, Houda
<i>Monastir University, Tunisia</i> | Alemary, Gehan A.E.
<i>Al-Majmaa University, KSA</i> |
| Abdelwahab, Sayed F.
<i>Minia University, Egypt</i> | Al-Ghzawi, Abd Al-Majeed
<i>Jordan University of Science and Technology, Jordan</i> |
| Abdull-Fattah, Gamal M.
<i>King Saud University, KSA</i> | Al-Hindi, Adnan
<i>Islamic University of Gaza, Palestine</i> |
| Abini, Jwan H.
<i>The Hashemite University, Jordan</i> | Alhomida, Abdullah S.
<i>King Saud University, KSA</i> |
| Abraha, Berhanu
<i>Bahir Dar University, Ethiopia</i> | Ali, Ali Abdelaziz
<i>Ain Shams University, Egypt</i> |
| Abu-Dieyeh, Mohammed H.
<i>Hashemite University, Jordan</i> | Al-Jahdali, Mohammad O.
<i>King Abdul-Aziz University, KSA</i> |
| Abuhamdah, Sawsa M.-A.
<i>University of Jordan, Jordan</i> | Aljamal, Abdulrahim
<i>Zarqa University, Jordan</i> |
| Abuharfeil, Nizar M.
<i>Jordan University of Science and Technology, Jordan</i> | Al-Karaki, Ghazi N.
<i>Jordan University of Science and Technology, Jordan</i> |
| Abuirmeileh, Naji M.
<i>Jordan University of Science and Technology, Jordan</i> | Al-Khateeb, Asem M.A.
<i>Jordan University of Science and Technology, Jordan</i> |
| Abu-Odeh, Ra'ed
<i>Sharjah University, UAE</i> | Al-Khateeb, Hakam
<i>The Hashemite University, Jordan</i> |
| Abu-Qatouseh, Luay F.
<i>MONOJO, Jordan</i> | Al-Khatib, Faisal M.
<i>University of Jordan, Jordan</i> |
| Abu-rjai, Talal
<i>University of Jordan, Jordan</i> | Al-kofahi, Ahmad
<i>Jordan University of Science and Technology, Jordan</i> |
| Abu-Safieh, Kayed A.
<i>The Hashemite University, Jordan</i> | Al-Kurd, Refat F.
<i>Petra University, Jordan</i> |
| Abu-Shaqra, Qasem M.
<i>Al-Balqa Applied University, Jordan</i> | Almeida, Juliana
<i>Institute of Biology, Brazil</i> |
| Abu-Sharar, Taleb M.
<i>University of Jordan, Jordan</i> | Al-Nabulsi, Anas, A.-R.
<i>Jordan University of Science and Technology, Jordan</i> |
| Adam, Ishag
<i>University of Khartoum</i> | Al-Najjar, Tareq
<i>University of Jordan, Jordan</i> |
| Ahmad, Faiz
<i>Brandon University, Canada</i> | Al-Oqlah, Ahmad A.
<i>Yarmouk University, Jordan</i> |
| Ahmed, Kazi S.
<i>Bangladesh Agricultural University, Bangladesh</i> | Al-Qaoud, Khaled M
<i>MONOJO, Jordan</i> |
| Ajanwachukwu, Nnadi P.
<i>University of Nigeria, Nigeria</i> | Al-Rawashdeh, Ibrahim M.
<i>Al-Hussein Bin Talal University, Jordan</i> |
| Akhtar, Naureen
<i>University of Punjab, Pakistan</i> | Al-Shboul, Othman
<i>Jordan University of Science and Technology, Jordan</i> |
| Al-Antary, Tawfiq
<i>University of Jordan, Jordan</i> | |
| Al-Aqtum, Mousa
<i>Zarqa University, Jordan</i> | |

Al-Subeihi, Ala'

Aqaba Special Economic Authority, Jordan

Al-Wedyan, Mohammad

The Hashemite University, Jordan

Al-Zube, Loay A.W.

The Hashemite University, Jordan

Al-Zyoud, Firas A.

Mutah University, Jordan

Amarowicz, Ryszard

Polish Academy of Sciences, Poland

Amr, Zuhair S.

Jordan University of Science and Technology, Jordan

Anokwuru, Chinedu

Babcock University, Nigeria

Aqel, Amin A.

Mutah University, Jordan

Arruda, Rafael

University Federal of Mato Grosso, Brazil

Ashraf, Muhammad Aqeel

University of Malaya, Malaysia

Atteyate, Mazen A.

Al-Balqa Applied University, Jordan

Bader, Khaldoun

Al-Quds University, Palestine

Bakheit, Mohammed Ahmed

Leibniz Center for Medical and Biosciences, Germany

Bambrug, James

Colorado State University, USA

Banat, Ibrahim M.

University of Ulster, UK

Bani-Hani, Saleem A.

Jordan University of Science and Technology, Jordan

Benzohra, Ibrahim E.

University of Mostaganem, Algeria

Bidasee, Keshore R.

University of Nebraska Medical Center, USA

Bilto, Yousif Y.

University of Jordan, Jordan

Boudot, Jean-Pierre

University of Lorraine, France

Chen, Chieh-Fu

National Yang-Ming University, Taiwan

Chukeatirote, Ekachai

Mae Fah Luang University, Thailand

Coyne, Mark S.

University of Kentucky, USA

Dahal, Keshav

University of Toronto, Canada

Dakshnamurthy, Selvakumar

Wayne State University, USA

Darwish, Hisham

Jerusalem University, Palestine

Diallo, Diawo

Institute Pasteur de Dakar, Senegal

Disi, Ahmad M.

University of Jordan, Jordan

Egwurugwu, Jude N.

Imo State University, Nigeria

El Naggat, ElMoataz B.

Damanhour University, Egypt

El-Batanony, Nadia H.

University of Sadat City, Egypt

Elbilbesy, Mohamed A.

Alexandria University, Egypt

El-Demellawy, Maha A.

Genetic Engineering and Biotechnology Research Institute, Egypt

El-Ensahsy, Hesham A.

University Technology Malaysia, Malaysia

ElFiky, Zaki Ahmad

Fayoum University, Egypt

El-Leboudy, Ahlam A.

Alexandria University, Egypt

El-Migdadi, Fayig

Jordan University of Science and Technology, Jordan

Elowni, Osman A.

University of Khartoum, Sudan

Elsawy, Essam M.

Mansoura University, Egypt

Elsayed Ahmad, Elsayed Ahmad

Federal University of Rio de Janeiro, Brazil

El-Sukhon, Saeb N.

Jordan University of Science and Technology, Jordan

El-Taweel, H. A.

Alexandria University, Egypt

El-Zubeir, Ibtisam E.M.

University of Khartoum, Sudan

Emam, Azza Kamal

Cairo University, Egypt

Engidawork, Ephrem

Addis Ababa University, Ethiopia

Estrela, Carlos

Federal University of Goids, Brazil

Fahim, Saber F.

Plant Protection Research Institute, Egypt

Farshadfar, Mohsen

Paymee Noor University, Iran

Fayeun, Lawrence S.

Federal University of Technology, Nigeria

Flaczyk, Ewa

Poznan University of Life Sciences, Poland

Franceschini-Vicentini, Irene B.

Sao Paulo State University, Brazil

Gakhar, Surendra K.

Maharshi Dayanand University, India

Galal, Salma B.

Al-Azhar University, Egypt

Galani, Saddia

University of Karachi, Pakistan

Gharibeh, Mohammad

Jordan University of Science and Technology, Jordan

Ghasemi, Mohammad F.

Islamic Azad University, Iran

Glatzel, Gerhard

Austrian Academy of Sciences, Austria

Gokturk, Didem

Istanbul University, Turkey

Griffiths, Charles

University of Cape Town, South Africa

Gurib Kakim, Ameenah

University of Mauritius, Mauritius

Hamad, Mawieh A.

University of Sharjah, UAE

Hanif, Atif

University of Veterinary and Animal Sciences, Pakistan

Harb, Amal

Yarmouk University, Jordan

Hassanien, Mohamed F. Ramadan

Zagazig University, Egypt

Hassanien, Mohamed Fawzy

Zagazig University, Egypt

Hatha, Abdulla M.

Cochin University of Science and Technology, India

Herceg, Zoran

University of Zagreb, Croatia

Hijjawi, Nawal

The Hashemite University, Jordan

Howarth, Frank C.

United Arab Emirates University, UAE

Hussain, Fida

Ayub Agriculture Research Institute, Pakistan

Hussein, Ayad A.

King Hussein Cancer Center, Jordan

Hussein, Emad I.

Yarmouk University, Jordan

Hussein, Hany

Biological Center of the Czech Academy of Science, Czech Republic

Hutson, A.M.

IUCN, UK

Ibrahimi, Ibrahim

University of Jordan, Jordan

Ileke, Kayode D.

Adekunle Ajasin University, Nigeria

Ismail, Naim S.

The Hashemite University, Jordan

Izquierdo, Alejandro C.

University Autonoma Metropolitana, Mexico

Jaber, Basem M.

University of Jordan, Jordan

Jacob, Jacob H.

Al-alBayte University, Jordan

Jaeschke, Hartmut

University of Kansa Medical Center, USA

Jaradat, Ziad

Jordan University of Science and Technology, Jordan

Kanani, Amit

Anand Agricultural University, India

Karsen, Hasan

Harran University, Turkey

Kasolo, Josephine N.

Makererere University, Uganda

Kasrawi, Mahmmud

University of Jordan, Jordan

Katbeh-Bader, Ahmad A.

University of Jordan, Jordan

Kaviraji, Anilava

University of Kalyani, India

Kenwright, Kathleen McLoughlin

The University of Tennessee Health Science Center, USA

Khabour, Omar

Jordan University of Science and Technology, Jordan

Khalaf, Marouf A.-K.

University of Jordan, Jordan

Khalil, Ahmad N.

Yarmouk University, Jordan

Khalil, Raida W.

Philadelphia University, Jordan

Khan, Ayaz A.

University of Malakand, Pakistan

Khan, Fauzia A.

Aga Khan University, Pakistan

Khan, Luqman Ahmad

Jamia Millia Islamia University, India

- Kharroubi, Akram**
Al-Quds University, Palestine
- Kosterin, Oleg E.**
Novosibirsk State University, Russia
- Krishnappa, Kaliyamoorthi**
Bharathidasan University, India
- Kumar, Arun**
Manipal College of Medical Sciences, Nepal
- Kumar, Suresh**
Purdue University, USA
- Kweka, Eliningaya**
Tropical Resticides Research Institute, Tanzania
- Lahham, Jameel N.**
Yarmouk University, Jordan
- Li, Huanxiu**
Sichuan Agriculture University, China
- Ligerio, Raphael**
University Federal of Minas Gerais, Brazil
- Mahasneh, Adel**
University of Jordan, Jordan
- Mahdi, Batool M.**
Baghdad University, Iraq
- Maji, Ardhendu**
Calcutta School of Tropical Medicine, India
- Maloul, Salem R.**
The Hashemite University, Jordan
- Massa, Salvatore**
Agricultural, Food and Environmental Sciences, Italy
- Massadeh, Majed M.**
Jordan University of Science and Technology, Jordan
- Massadeh, Muhannad A.**
The Hashemite University, Jordan
- Matar, Ghassan**
American University of Beirut, Lebanon
- Matar, Suzan A.-W.**
University of Jordan, Jordan
- Mattalka, Khaled Z.**
University of Petra, Jordan
- Milesi, Silvia V.**
University Federal of Rio Grande do Sul, Brazil
- Mohamed, Moemen A.**
Assiut University, Egypt
- Mohammed, Faisal I.**
University of Jordan, Jordan
- Moharam, Moustafa H.**
Sohag University, Egypt
- Molina, Bosquez E.**
University Autonoma Metropolitana- Iztapalapa, Mexico
- Momoh, A. O.**
Federal University of Technology, Nigeria
- Mosa, Ahmed A.**
Ain-Shams University, Egypt
- Mostafa, Ehab F.**
Zagazig University, Egypt
- Nahdavi, Vhid**
Research Center of Agricultural and Natural Resources, Iran
- Naji, Sameer A.**
The Hashemite University, Jordan
- Nasher, Abdel-Karim**
Sana'a University, Yemen
- Nusier, Mohamed**
Jordan University of Science and Technology, Jordan
- Nwoko, Chris O.**
Federal University of Technology, Nigeria
- O'Kane, Steve**
University of Northern Iowa, USA
- Obiad, Reyad R.**
Jordan University of Science and Technology, Jordan
- Ohenhen, Regina E.**
Ambrose Alli University, Nigeria
- Ohimain, Elijah I.**
Niger Delta University, Nigeria
- Ojo, Akinrotimi**
Center of Nigeria Institute of Oceanography and Marine Research, Nigeria
- Okokon, Esther**
Nigeria Institute for Oceanography and Marine Research, Nigeria
- Ola, Mohammad S.**
King Saud University, KSA
- Oliveria, Ademir K.-M.**
University Anhanguera-Uniderp, Brazil
- Omer, Osama H.**
Qassim University, KSA
- Owais, Saed J.**
Mutah University, Jordan
- Ozbek Hanefi**
Istanbul Medipol University, Turkey
- Ozcan, Bahri D.**
Osmaniye Korkut Ata University, Turkey
- Pifer, Linda L.**
The University of Tennessee Health Science Center, USA
- Prakasham, Reddy S.**
Indian Institute of Chemical Technology, India
- Qasem, Jamal R.S.**
University of Jordan, Jordan
- Qin, Chuanguan**
Northwestern Polytechnical University, China

Rabi, Atallah Z.
Jordan University of Science and Technology, Jordan

Rassian, Ossama S E
Sinai University, Egypt

Rawashdeh, Ibrahim
Al-Hussein Bin Talal University, Jordan

Rehman, Zia ur
Massey University, New Zealand

Saeed, Ali K.
University of Jordan, Jordan

Sajid, Muhammad
Agricultural University, Pakistan

Santos, Rento de Lima
University Federal Minas Gerais, Brazil

Santoyo, Montserrat C.
Instituto Tecnológico Tepic, Mexico

Satpute, Rajendra A.
Government Institute of Science, India

Sharif, Labib A.
Jordan University of Science and Technology, Jordan

Shbailat, Saba
The Hashemite University, Jordan

Shraideh, Ziad A.
University of Jordan, Jordan

Shruthi, S.D.
The Oxford College of Science, India

Shukla, Sudhir
National Botanical Research Institute, India

Siddiqi, Nikhat J.
King Saud University, KSA

Sieiro, Garmen
University of Vigo, Spain

Silva-Aguayo, Gonzalo
University of Concepcion, Chile

Srichairatanakool, Somdet
Chiang Mai University, Thailand

Stephen, Adewole O.
Obafemi Awolowo University, Nigeria

Sultana, Nuzhat
University of Karachi, Pakistan

Suvarna, Biradar
College of Agriculture, India

Swedan, Samer F.
Jordan University of Science and Technology, Jordan

Tahhan, Raghab A.
Jordan University of Science and Technology, Jordan

Tahtamouni, Lubna H.
The Hashemite University, Jordan

Takruri, Hamed
University of Jordan, Jordan

Tamimi, Samih M.
University of Jordan, Jordan

Tarn, Der-Cherng
Taipei Veterans General Hospital, Taiwan

Teodoro, Grazielle S.
University of Campinas, Brazil

Tukan, Salma K.
University of Jordan, Jordan

Upton, Jeff D.
Canterbury Health Lab, New Zealand

Urooj, Asna
University of Mysore, India

Vaseem Baig, M.M.
Yeshwant College, Nanded, India

Vatandoost, Hassan
Tehran University of Medical Sciences, Iran

Vieira, Alexandre R.
University of Pittsburgh, USA

Wagenaar, G.M.
University of Johannesburg, South Africa

Waitzbauer, Wolfgang
University of Vienna, Austria

Wan Yusoff, Wan Mohtar
National University of Malaysia, Malaysia

Wu, Qiuye
Second Military Medical University, China

Xiong, Zhi-Qiang
Chinese Academy of Sciences, China

Xu, Xueming
Jiangnan University, China

Yadav, Taya P.
Maharshi Dayanand University, India

Yan, Weikai
Eastern Cereal and Oilseed Research Center, Canada

Yemenicioglu, Ahmet
Izmir Institute of Technology, Turkey

Yue, Ming
The University of Chicago, USA

Zapico, Florence L.
Natural Sciences and Mathematics Institute, Philippines

Appendix B

Contents of Volume 6- 2013

Number 1

Use of Anabolic Androgenic Steroids in Jordan: Mini- Review. <i>Lubna H. Tahtamouni</i>	1-4
Development of <i>Dermestes maculatus</i> (DeGeer, 1774) (Coleoptera, Dermestidae) on Different Fish Substrates. <i>Usman Zakka, Jonathan N. Ayertey and Millicent A. Cobblah</i>	5-10
Composition and Larvicidal Activity of <i>Artemisia vulgaris</i> L. Stem Essential Oil Against <i>Aedes aegypti</i> <i>Sujatha Govindaraj and Bollipo D. Ranjitha Kumari</i>	11-16
Susceptibility of the Hymenopteran Parasitoid, <i>Habrobracon hebetor</i> (Say) (Braconidae) to the Entomopathogenic Fungi <i>Beauveria bassiana</i> Vuillemin and <i>Metarhizium anisopliae</i> Sorokin. <i>Vahid Mahdavi, Moosa Saber, Hooshang Rafiee-Dastjerdi and Ali Mehrvar</i>	17-20
Accumulation of Copper in Different Tissues and Changes in Oxygen Consumption Rate in Indian Flying Barb, <i>Esomus danricus</i> (Hamilton-Buchanan) Exposed to Sub-lethal Concentrations of Copper. <i>Suchismita Das and Abhik Gupta</i>	21-24
Modification of the Mechanical Properties of Red Blood Cell Membrane by Spent <i>Plasmodium falciparum</i> Culture Supernatant. <i>Nii A. Aryee and Yuichi Takakuwa</i>	25-30
The Antihyperglycaemic Effect of the Aqueous Extract of <i>Origanum vulgare</i> Leaves in Streptozotocin-Induced Diabetic Rats. <i>Nema A. Mohamed and Omimah A. Nassier</i>	31-38
Fecundity of Bigfin squid, <i>Sepioteuthis lessoniana</i> (Lesson, 1830) (Cephalopoda: Loliginidae). <i>Venkatesan Vellathi and Rajagopal Santhanam</i>	39-44
Effect of Dredging on the Macrozoobenthos of Hazratbal Basin in the Dal Lake Srinagar Kashmir, India. <i>Basharat Mushtaq, Rajni Raina, Abdul R. Yousuf, Ashwani Wanganeo and Ummer Rashid</i>	45-50
Evaluation of the Physicochemical Properties and Antimicrobial Activities of Bioactive Biodegradable Films. <i>Mary S. Khali, Zahra S. Ahmed and Aml S. Elnawawy</i>	51-60
Effects of Three Medicinal Plant Products on Survival, Oviposition and Progeny Development of Cowpea Bruchid, <i>Callosobruchus maculatus</i> (Fab.) [Coleoptera: Chrysomelidae] Infesting Cowpea Seeds in Storage. <i>Kayode D. Ileke, Daniel S. Bulus and Ayisat Y. Aladegoroye</i>	61-66
Reserve Mobilization, Total Sugars and Proteins in Germinating Seeds of Durum Wheat (<i>Triticum durum</i> Desf.) under Water Deficit after Short Period of Imbibition. <i>Amal M. Harb</i>	67-72
A Novel Report on the Prevalence of Enterohaemorrhagic <i>Escherichia coli</i> non-O157 Isolated from Cattle in Kaduna State, Nigeria. <i>Jasini A. Musa, Mashood A. Raji, Haruna M. Kazeem and Nicodemus M. Useh</i>	73-76

Number 2

- African Flora as Potential Sources of Medicinal Plants: Towards the Chemotherapy of Major Parasitic and Other Infectious Diseases- A Review. 77-84
Ameenah Gurib-Fakim and Mohamad F. Mahomoodally
- Sodium Azide Induced Complementary Effect of Chromosomal Stickiness in *Brassica campestris* L. 85-90
Girjesh Kumar and Kshama Dwivedi
- Effects of an Ecdysteroid Analog (RH-0345) on the Ovarian and Testicular Components of *Eupolybothrus nudicornis* (Myriapoda : Chilopoda) 91-98
Ouided Daas-Maamcha, Kahina Houd-Chaker, Meriem Soucha , Tarek Daas and Patrick Scaps
- Evaluation of the Performance of Different Maize Varieties against *Sitophilus zeamais* Motsch. (Coleoptera: Curculionidae) Infestation in the Niger Delta Region of Nigeria 99- 104
Usman Zakka , Ndowa E.S. Lale and Odidika C. Umeozor
- Total Phenol, Antioxidant and Cytotoxic Properties of Wild Macrofungi Collected from Akure Southwest Nigeria. 105- 110
Olusegun V. Oyetayo , Antonio Nieto- Camacho , Teresa M. Ramirez-Apana , Rodriguez E. Balamero and Manuel Jimenez
- Isolation, Characterization and Determination of Antimicrobial Properties of Lactic Acid Bacteria from Human Milk. 111- 116
Farhana S. Diba , Khondoker M. Hossain, M. A. Azim and Md. Moinul Hoque
- Challenges Towards *Hypericum sinaicum* Conservation in South Sinai, Egypt. 117- 126
Om-Mohamed A. Khafagi , Elbialy. E. Hatab and Karim. A. Omar
- Molecular Analysis of Intracultivar Polymorphism of 'Panchadarakalasa' Mango by Microsatellite Markers 127- 136
Hameedunnisa Begum , Medagam Thirupathi Reddy, Surapaneni Malathi, Boreddy P.Reddy, Gonela Narshimulu, Javaregowda Nagaraju and Ebrahimali Abubaker Siddiq
- An Initial *In vitro* Investigation into the Potential Therapeutic Use of *Lucilia sericata* Maggot to Control Superficial Fungal Infections. 137- 142
Sulaiman M. Alnaimat , Milton Wainwright and Saleem H. Aladaileh
- Trehalose Accumulation in Wheat Plant Promotes Sucrose and Starch Biosynthesis. 143- 150
Hanaa E. Ahmed , Elhusseiny A. Youssef, Maimona A. Kord and Ebtesam A. Qaid
- Response of Three Accessions of Jordanian *Aegilops crassa* Boiss. and Durum Wheat to Controlled Drought. 151- 158
Amal M. Harb and Jamil N. Lahham
- Effects of Cigarette Smoking on Some Immunological and Hematological Parameters in Male Smokers in Erbil City. 159- 166
Farhang A. Aula and Fikry A. Qadir

Knowledge of the Use and Benefits of Applying Biotechnology and Cell Based Therapy in Orthopaedics in Jordan: Questionnaire Survey and Regulation Assessment. 167- 176

Loay A. Al-Zu'be , Thakir D. Al-Momani , Bilal M. Al-Trabsheh and Modhafar Z. Al-Zoubi

Number 3

Fermentation Studies for the Production of Dibutyl Phthalate, an Ester Bioactive Compound from *Streptomyces albidoflavus* MTCC 3662 Using Low-Priced Substrates 177- 181

Raj N. Roy and Sukanta K. Sen

In vivo Assay for Antagonistic Potential of Fungal Isolates against Faba bean (*Vicia faba* L.) Chocolate Spot (*Botrytis fabae* Sard.). 183- 189

Ermias T. Taffa , Chemed F. Gurmessa and Samuel Sahile W. Mariam

"Vinegar" as Anti-bacterial Biofilm formed by *Streptococcus pyogenes* Isolated from Recurrent Tonsillitis Patients, *in vitro*. 191- 197

Narjis F. Ismael

Effects of COX-1 and COX-2 Inhibitors in L- Nitro-L-Arginine Methyl Ester Induced Hypertensive Rats. 199- 204

Ismail M. Maulood and Almas M. R. Mahmud

Is Gaza Sandy Shoreline Region Contaminated with Human Gastrointestinal Parasites? 205-210

Ahmed H. Hilles , Adnan I. Al Hindi and Yousef A. Abu Safieh

In vitro Screening of *Lactobacillus* Species from Homemade Yoghurt for Antagonistic Effects against Common Bacterial Pathogens. 211- 216

Akhter A. Ahmed

Bioefficacy of Azadirachtin in Controlling *Culex Pipiens Pipiens* (Diptera: Culicidae). 217- 222

Abdelouaheb Alouani, Nassima Rehim and Nouredine Soltani

Micro and Macronutrient Properties of *Pleurotus ostreatus* (Jacq: Fries) Cultivated on Different Wood Substrates. 223- 226

Victor O. Oyetayo and Olatomiwa O. Ariyo

Effect of Ethanol Extract of *Calotropis procera* Root Bark on Carbon Tetrachloride-Induced Hepato-nephrotoxicity in Female Rats. 227- 229

Daniel Dahiru , Augustine Amos and Sarah H. Sambo

Ameliorative Effect of the Aqueous Extract of *Zingiber officinale* on the Cadmium-Induced Liver and Kidney Injury in Females Rats. 231- 234

Sarbaz I. Mohammad, Inaam A. Mustafa and Shang Z. Abdulqader

Prevalence of Congenital Red-Green Color Vision Defects among Various Ethnic Groups of Students in Erbil City 235- 238

Karim J. Karim and Mohammed A. Saleem

Molecular Characterization and Phylogenetic Analysis of Cellulase Producing *Streptomyces griseorubens* (Strain St-1) Isolated from Indian Soil 239- 242

Pinky Prasad Tanuja Singh , Sheila Bedi and Sonika Kumari

Phenylketonuria in Sohag: A preliminary Study

Ghaleb A. Oriquat, Tahia H. Saleem, Nagwa S. Ahmed and Sawsan M.A. Abuhamdah 243- 245

Number 4

Bionanotechnology: The Novel Nanoparticles Based Approach for Disease Therapy. 246 - 251

Adel M. Mahasneh

Levels of Chromium and Copper in Liver and Muscle Tissues of the Round Sardinella *Sardinella aurita* (Valenciennes) from the Oran Coastline- Algeria. 252 - 256

Nardjess Benamar and Boutiba Zitouni

Evaluation of Antioxidant Properties of *Morus nigra* L. Fruit Extracts [II]. 258 - 265

Najlala K. Issa and Rihan S. Abd-Aljabar

Antimicrobial Activity of Xerophytic Plant (*Cotula cinerea* Delile) Extracts Against Some Pathogenic Bacteria and Fungi. 266 - 271

Djamel Bensizerara, Taha Menasria, Maimouna Melouka, Lamia Cheriet and Haroun Chenchouni

Correlation Between Numerical Profiles Generated for Soil Spore Forming Bacilli and Their Inhibitory Potential Against *Staphylococcus aureus* ATCC 6538. 272 - 276

Qasem M. Abu Shaqra

Nitrate Reductase Assay Using Sodium Nitrate for Rapid Drug Susceptibility Testing of *Mycobacterium tuberculosis* Directly on Sputum Samples. 278 - 282

Mohammed Abdul- Imam Almazini

Protective Effects of *Enantia chlorantha* Stem Bark Extracts on Acetaminophen Induced Liver Damage in Rats. 284 - 290

Olamide E. Adebisi and Mathew O. Abatan

Odonata of Wadi Al Mujib Catchment with Notes on the Impact of Wadi Al Mujib Dam, Jordan (Insecta: Odonata). 292 - 299

Zuhair S. Amr, Loay S. Al Azzam, Ahmad Katbeh-Bader and Ehab K. Eid

Comparative Studies on Anti-hyperglycemic Effects of Ethyl Acetate and Methanol Extract of *Albizzia lucida* Benth Bark in Alloxan Induced Diabetic Rats. 300 - 307

Arumugam S. Kumar, Subramanian Kavimani and Korlakunta N. Jayaveera

Bacteriological and Mycological Assessment for Water Quality of Duhok Reservoir, Iraq. 308 - 315

Yahya A. Shekha, Hero M. Ismael and Akhter A. Ahmed

Effects of *Theileria lestoquardi* Infection on Haematological and Biochemical Parameters in Experimentally Infected Desert Ewes. 316 - 319

Aisha A. Elsadig, Yousif H. Abdalla Elmansoury, Husna M. Elbasheir, Amna E. Babiker, Aza A. Adam, Tahani O. Abdelmageed and Sabri Hussein

Computational Prediction of Binding of Methyl Carbamate, Sarin, 320 - 323

Deltamethrin and Endosulfan Pesticides on Human Oxyhaemoglobin.

Padma Saxena

New Records of Arthropod Ectoparasites of Bats from North-Eastern Algeria.

Mohamed Lamine Bendjeddou, Idir Bitam, Awatef Abiadh, Zihad Bouzlama and Zuhair S. Amr

324 - 327

Evaluation of Immunomodulatory Effects of Antiepileptic Drug Phenytoin.

Mohammad A. Al-Fararjeh, Mohammad H. Jaber and Yaseen S. Abdelrahman

328 - 333

Prevalence of Lactose Intolerance in Primary School Children in Qena Governorate, Egypt.

Sawsan M A. Abuhamdah, Ghaleb A. Oriquat, Tahia H. Saleem and Mohammed H. Hassan

334 - 336

Jordan Journal of Biological Sciences



An International Peer – Reviewed Research Journal

Published by the deanship of Research & Graduate Studies, The Hashemite University, Zarqa, Jordan

Name: الاسم:
 Specialty: التخصص:
 Address: العنوان:
 P.O. Box: صندوق البريد:
 City & Postal Code: المدينة: الرمز البريدي:
 Country: الدولة:
 Phone: رقم الهاتف:
 Fax No.: رقم الفاكس:
 E-mail: البريد الإلكتروني:
 Method of payment: طريقة الدفع:
 Amount Enclosed: المبلغ المرفق:
 Signature: التوقيع:
 Cheques should be paid to Deanship of Research and Graduate Studies – The Hashemite University.

I would like to subscribe to the Journal

For

- ☐ One year
☐ Two years
☐ Three years

One Year Subscription Rates

	Inside Jordan	Outside Jordan
Individuals	JD10	\$70
Students	JD5	\$35
Institutions	JD 20	\$90

Correspondence

Subscriptions and sales:

Prof. Khaled H. Abu-Elteen
 The Hashemite University
 P.O. Box 330127-Zarqa 13115 – Jordan
 Telephone: 00 962 5 3903333 ext. 4399
 Fax no. : 0096253903349
 E. mail: jjbs@hu.edu.jo



Hashemite Kingdom of Jordan



The Hashemite University

المجلة الأردنية للعلوم الحياتية

مجلة علمية عالمية محكمة
تصدر بدعم من صندوق دعم البحث العلمي

المجلة الأردنية للعلوم الحياتية

مجلة علمية عالمية محكمة

المجلة الأردنية للعلوم الحياتية : مجلة علمية عالمية محكمة ومفهرسة ومصنفة، تصدر عن الجامعة الهاشمية و بدعم من صندوق دعم البحث العلمي- وزارة التعليم العالي و البحث العلمي .

هيئة التحرير

رئيس التحرير:

الأستاذ الدكتور خالد حسين أبو التين

الجامعة الهاشمية، الزرقاء، الأردن .

الأعضاء:

الأستاذ الدكتور سوسن عطا الله العوران
الجامعة الأردنية

الأستاذ الدكتور شتيوي صالح عبدالله
الجامعة الأردنية

الأستاذ الدكتور عبدالكريم جبر السلال
جامعة العلوم والتكنولوجيا الأردنية

الأستاذ الدكتور نبيل البشير
جامعة العلوم والتكنولوجيا الأردنية

الأستاذ الدكتور حكم فائق الحديدي
جامعة العلوم والتكنولوجيا الأردنية

الأستاذ الدكتور خالد أحمد الطراونة
جامعة مؤتة

الأستاذ الدكتور سامي خضر عبدالحافظ
جامعة البرموك

الأستاذ الدكتور علي زهير الكرمي
الجامعة الهاشمية

فريق الدعم:

تنفيذ وإخراج

م. مهند عقده

المحرر اللغوي

الدكتور قصي الذبيان

ترسل البحوث إلى العنوان التالي :

رئيس تحرير المجلة الأردنية للعلوم الحياتية

عمادة البحث العلمي و الدراسات العليا

الجامعة الهاشمية

الزرقاء – الأردن

هاتف : ٣٩٠٣٣٣٣ ٥ ٠٠٩٦٢٢ فرعي ٤١٤٧

Email: jjbs@hu.edu.jo, Website: www.jjbs.hu.edu.jo