

المجلة الأردنية للعلوم الحياتية  
**Jordan Journal of Biological Sciences (JJBS)**

<http://jjbs.hu.edu.jo>

**Jordan Journal of Biological Sciences (JJBS)** (ISSN: 1995–6673): An International Peer- Reviewed Research Journal funded by the Scientific Research Support, Ministry of Higher Education and Scientific Research, Jordan and published quarterly by the Deanship of Research and Graduate Studies, The Hashemite University, Jordan.

**Editor-in-Chief**

**Professor Abu-Elteen, Khaled H.**  
The Hashemite University

**Editorial Board (Arranged alphabetically):**

- **Professor Abdalla, Shtaywy S.**  
University of Jordan
- **Professor Abdel-Hafez, Sami K.**  
Yarmouk University
- **Professor Al-Hadidi, Hakam F.**  
Jordan University of Science and Technology
- **Professor Bashir, Nabil A.**  
Jordan University of Science and Technology
- **Professor Elkarmi, Ali Z.**  
The Hashemite University
- **Professor Oran, Sawsan A.**  
University of Jordan
- **Professor Sallal, Abdul-Karim J.**  
Jordan University of Science and Technology
- **Professor Tarawneh, Khaled A.**  
Mutah University

**International Advisory Board:**

- Prof. Abdul-Haque, Allah Hafiz**  
National Institute for Biotechnology and Genetic Engineering, Pakistan
- Prof. El Makawy, Aida, I**  
National Research Center, Giza, Egypt
- Prof. Ghannoum, Mahmoud A**  
University Hospital of Cleveland and Case Western Reserve University, U.S.A.
- Prof. Hanawalt, Philip C**  
Stanford University, California, U.S.A
- Prof. Kaviraj, Anilava**  
India University of Kalyani, Kenya
- Prof. Martens, Jochen**  
Institute Fur Zoologie, Germany
- Prof. Stanway, Glyn**  
University of Essex, England
- Prof. Wan Yusoff, Wan Mohtar**  
University Kebangsaa Malaysia, Malaysia
- Prof. Bamburg, James**  
Colorado State University, U.S.A,
- Prof. Garrick, Michael D**  
State University of New York at Buffalo, U.S.A.
- Prof. Gurib-Fakim, Ameenah F**  
Center for Phytotherapy and Research, Ebene, Mauritius.
- Prof. Hassanali, Ahmed**  
Kenyatta University, Nairobi, Kenya
- Prof. Matar, Ghassan M**  
American University of Beirut, Lebanon
- Prof. Nasher, Abdul Karim**  
Sanna' University, Yemen
- Prof. Waitzbauer, Wolfgang**  
University of Vienna, Austria

**Submission Address**

**Professor Abu-Elteen, Khaled H.**  
Deanship of Scientific Research and Graduate Studies  
The Hashemite University  
P.O. Box 330127, Zarqa, 13115, Jordan  
Phone: +962-5-3903333 ext. 4399  
E-Mail: [jjbs@hu.edu.jo](mailto:jjbs@hu.edu.jo)

**Editorial Board Support Team**

**Language Editor**      **Publishing Layout**  
**Dr. Qusai Al-Debyan**      **Mohannad Oqdeh**



Hashemite Kingdom of Jordan



Hashemite University

# Jordan Journal of Biological Sciences

*An International Peer-Reviewed Scientific Journal  
Funded by HYSscientific Research Support*

## Instructions to Authors

### Scopes

Study areas include cell biology, genomics, microbiology, immunology, molecular biology, biochemistry, embryology, immunogenetics, cell and tissue culture, molecular ecology, genetic engineering and biological engineering, bioremediation and biodegradation, bioinformatics, biotechnology regulations, gene therapy, organismal biology, microbial and environmental biotechnology, marine sciences. The JJBS welcomes the submission of manuscript that meets the general criteria of significance and academic excellence. All articles published in JJBS are peer-reviewed. Papers will be published approximately one to two months after acceptance.

### Type of Papers

The journal publishes high-quality original scientific papers, short communications, correspondence and case studies. Review articles are usually by invitation only. However, Review articles of current interest and high standard will be considered.

### Submission of Manuscript

Manuscript, or the essence of their content, must be previously unpublished and should not be under simultaneous consideration by another journal. The authors should also declare if any similar work has been submitted to or published by another journal. They should also declare that it has not been submitted/ published elsewhere in the same form, in English or in any other language, without the written consent of the Publisher. The authors should also declare that the paper is the original work of the author(s) and not copied (in whole or in part) from any other work. All papers will be automatically checked for duplicate publication and plagiarism. If detected, appropriate action will be taken in accordance with International Ethical Guideline. By virtue of the submitted manuscript, the corresponding author acknowledges that all the co-authors have seen and approved the final version of the manuscript. The corresponding author should provide all co-authors with information regarding the manuscript, and obtain their approval before submitting any revisions. Electronic submission of manuscripts is strongly recommended, provided that the text, tables and figures are included in a single Microsoft Word file. Submit manuscript as e-mail attachment to the Editorial Office at: [JJBS@hu.edu.jo](mailto:JJBS@hu.edu.jo). After submission, a manuscript number will be communicated to the corresponding author within 48 hours.

### Peer-review Process

It is requested to submit, with the manuscript, the names, addresses and e-mail addresses of at least 4 potential reviewers. It is the sole right of the editor to decide whether or not the suggested reviewers to be used. The reviewers' comments will be sent to authors within 6-8 weeks after submission. Manuscripts and figures for review will not be returned to authors whether the editorial decision is to accept, revise, or reject. All Case Reports and short Communication must include at least one table and/or one figure.

### Preparation of Manuscript

The manuscript should be written in English with simple lay out. The text should be prepared in single column format. Bold face, italics, subscripts, superscripts etc. can be used. Pages should be numbered consecutively, beginning with the title page and continuing through the last page of typewritten material.

The text, excluding the abstract, if required, can be divided into numbered sections with brief headings. Starting from introduction with section 1. Subsections should be numbered (for example 2.1 (then 2.1.1, 2.1.2, 2.2, etc.), up to three levels.

Manuscripts in general should be organized in the following manner:

- Title
- Abstract
- Key words
- Introduction
- Materials & Methods
- Results & Discussion
- Conclusion
- Acknowledgements
- References

### **Brief guidelines**

#### **Title Page**

The title page should contain a brief title, correct first name, middle initial and family name of each author and name and address of the department(s) and institution(s) from where the research was carried out for each author. The title should be without any abbreviations and it should enlighten the contents of the paper. All affiliations should be provided with a lower-case superscript number just after the author's name and in front of the appropriate address.

The name of the corresponding author should be indicated along with telephone and fax numbers (with country and area code) along with full postal address and e-mail address.

#### **ABSTRACT**

The abstract should be concise and informative. It should not exceed **350 words** in length for full manuscript and Review article and 150 words in case of Case Report and/ or Short Communication.. It should briefly describe the purpose of the work, techniques and methods used, major findings with important data and conclusions. No references should be cited in this part. Generally non-standard abbreviations should not be used, if necessary they should be clearly defined in the abstract, at first use.

#### **Keywords**

Immediately after the abstract, **about 4-8 keywords** should be given. Use of abbreviations should be avoided, only standard abbreviations, well known in the established area may be used, if appropriate. These keywords will be used for indexing.

#### **Abbreviations**

Non-standard abbreviations should be listed and full form of each abbreviation should be given in parentheses at first use in the text.

#### **INTRODUCTION**

Provide a factual background, clearly defined problem, proposed solution, a brief literature survey and the scope and justification of the work done.

#### **MATERIALS AND METHODS**

Give adequate information to allow the experiment to be reproduced. Already published methods should be mentioned with references. Significant modifications of published methods and new methods should be described in detail. Capitalize trade names and include the manufacturer's name and address. Subheading should be used.

## **RESULTS**

Results should be clearly described in a concise manner. Results for different parameters should be described under subheadings or in separate paragraph. Results should be explained, but largely without referring to the literature. Table or figure numbers should be mentioned in parentheses for better understanding.

## **DISCUSSION**

The discussion should not repeat the results, but provide detailed interpretation of data. This should interpret the significance of the findings of the work. Citations should be given in support of the findings. The results and discussion part can also be described as separate, if appropriate. The Results and Discussion sections can include subheadings, and when appropriate, both sections can be combined

## **CONCLUSIONS**

This should briefly state the major findings of the study.

## **Acknowledgment**

A brief acknowledgment section may be given after the conclusion section just before the references. The acknowledgment of people who provided assistance in manuscript preparation, funding for research, etc. should be listed in this section.

## **Tables and Figures**

Tables and figures should be presented as per their appearance in the text. It is suggested that the discussion about the tables and figures should appear in the text before the appearance of the respective tables and figures. No tables or figures should be given without discussion or reference inside the text.

Tables should be explanatory enough to be understandable without any text reference. Double spacing should be maintained throughout the table, including table headings and footnotes. Table headings should be placed above the table. Footnotes should be placed below the table with superscript lowercase letters. Each table should be on a separate page, numbered consecutively in Arabic numerals.

Each figure should have a caption. The caption should be concise and typed separately, not on the figure area. Figures should be self-explanatory. Information presented in the figure should not be repeated in the table. All symbols and abbreviations used in the illustrations should be defined clearly. Figure legends should be given below the figures.

## **References**

References should be listed alphabetically at the end of the manuscript. Every reference referred in the text must be also present in the reference list and vice versa. In the text, a reference identified by means of an author's name should be followed by the year of publication in parentheses ( e.g.( Brown,2009)). For two authors, both authors' names followed by the year of publication (e.g.( Nelson and Brown, 2007)). When there are more than two authors, only the first author's name followed by "*gv'cn*" and the year of publication ( e.g. ( Abu-Elteen *gv'cn*, 2010)). When two or more works of an author has been published during the same year, the reference should be identified by the letters "a", "b", "c", etc., placed after the year of publication. This should be followed both in the text and reference list. e.g., Hilly, (2002a, 2002b); Hilly, and Nelson, (2004). Articles in preparation or submitted for publication, unpublished observations, personal communications, etc. should not be included in the reference list but should only be mentioned in the article text ( e.g., Shtyawy,A., University of Jordan, personal communication). Journal titles should be abbreviated according to the system adopted in Biological Abstract and Index Medicus, if not included in Biological Abstract or Index Medicus journal title should be given in full. The author is responsible for the accuracy and completeness of the references

and for their correct textual citation. Failure to do so may result in the paper being withdrawn from the evaluation process. Example of correct reference form is given as follows:-

**Reference to a journal publication:**

Bloch BK. 2002. Econazole nitrate in the treatment of *Ecophycocyclospora*. *UChit "Ogf "L"*, **58**:314-323.

Ogunseitan OA and Ndoye IL. 2006. Protein method for investigating mercuric reductase gene expression in aquatic environments. *CrrnGpaxkqp "Olet qdkqn*, **64**: 695-702.

Hilly MO, Adams MN and Nelson SC. 2009. Potential fly-ash utilization in agriculture. *Rtqi t guu'kp"* *Pcwrcn'Uek* **19**: 1173-1186.

**Reference to a book:**

Brown WY and White SR. 1985. **The Elements of Style**, third ed. MacMillan, New York.

**Reference to a chapter in an edited book:**

Mettam GR and Adams LB. 2010. How to prepare an electronic version of your article. In: Jones BS and Smith RZ (Eds.), **Introduction to the Electronic Age**. Kluwer Academic Publishers, Netherlands, pp. 281–304.

**Conferences and Meetings:**

Embabi NS. 1990. Environmental aspects of distribution of mangrove in the United Arab Emirates. Proceedings of the First ASWAS Conference. University of the United Arab Emirates. Al-Ain, United Arab Emirates.

**Theses and Dissertations:**

El-Labadi SN. 2002. Intestinal digenetic trematodes of some marine fishes from the Gulf of Aqaba. MSc dissertation, Hashemite University, Zarqa, Jordan.

**Nomenclature and Units**

Internationally accepted rules and the international system of units (SI) should be used. If other units are mentioned, please give their equivalent in SI.

For biological nomenclature, the conventions of the *KpvtpcvqpcnEqf g'qhDqvcpkcnPqo gperwtg. 'lj g" KpvtpcvqpcnEqf g'qhPqo gperwtg'qhDc evgtkc.* and the *KpvtpcvqpcnEqf g'qh\ qqrqi kecn' Pqo gperwtg'* should be followed.

Scientific names of all biological creatures (crops, plants, insects, birds, mammals, etc.) should be mentioned in parentheses at first use of their English term.

Chemical nomenclature, as laid down in the *KpvtpcvqpcnWpkqp"qhRwtg"cpf "Crrrkf "Ej go km{* and the official recommendations of the *KWRCE/KWD'Eqo dkgf "Eqo o kulkqp"qp'Dkqej go kecn'Pqo gperwtg* should be followed. All biocides and other organic compounds must be identified by their Geneva names when first used in the text. Active ingredients of all formulations should be likewise identified.

**Math formulae**

All equations referred to in the text should be numbered serially at the right-hand side in parentheses. Meaning of all symbols should be given immediately after the equation at first use. Instead of root signs fractional powers should be used.

Subscripts and superscripts should be presented clearly. Variables should be presented in italics. Greek letters and non-Roman symbols should be described in the margin at their first use.

To avoid any misunderstanding zero (0) and the letter O, and one (1) and the letter l should be clearly differentiated.

For simple fractions use of the solidus (/) instead of a horizontal line is recommended. Levels of statistical significance such as: \* $R < 0.05$ , \*\* $R < 0.01$  and \*\*\* $R < 0.001$  do not require any further explanation.

### **Copyright**

Submission of a manuscript clearly indicates that: the study has not been published before or is not under consideration for publication elsewhere (except as an abstract or as part of a published lecture or academic thesis); its publication is permitted by all authors and after accepted for publication it will not be submitted for publication anywhere else, in English or in any other language, without the written approval of the copyright-holder. The journal may consider manuscripts that are translations of articles originally published in another language. In this case, the consent of the journal in which the article was originally published must be obtained and the fact that the article has already been published must be made clear on submission and stated in the abstract. It is compulsory for the authors to ensure that no material submitted as part of a manuscript infringes existing copyrights, or the rights of a third party.

### **Ethical Consent**

All manuscripts reporting the results of experimental investigation involving human subjects should include a statement confirming that each subject or subject's guardian obtains an informed consent, after the approval of the experimental protocol by a local human ethics committee or IRB. When reporting experiments on animals, authors should indicate whether the institutional and national guide for the care and use of laboratory animals was followed.

### **Plagiarism**

The JJBS hold no responsibility for plagiarism. If a published paper is found later to be extensively plagiarized and is found to be a duplicate or redundant publication, a note of retraction will be published, and copies of the correspondence will be sent to the authors' head of institute.

### **Galley Proofs**

The Editorial Office will send proofs of the manuscript to the corresponding author as an e-mail attachment for final proof reading and it will be the responsibility of the corresponding author to return the galley proof materials appropriately corrected within the stipulated time. Authors will be asked to check any typographical or minor clerical errors in the manuscript at this stage. No other major alteration in the manuscript is allowed. After publication authors can freely access the full text of the article as well as can download and print the PDF file.

### **Reprints**

Twenty (20) reprints are provided to corresponding author free of charge within two weeks after the printed journal date. For orders of more reprints, a reprint order form and prices will be sent with article proofs, which should be returned directly to the Editor for processing.

### **Disclaimer**

Articles, communication, or editorials published by JJBS represent the sole opinions of the authors. The publisher shoulders no responsibility or liability what so ever for the use or misuse of the information published by JJBS.

### **Indexing**

JJBS is indexed and abstracted by the Chemical Abstract Service, CAB International Abstracts, Zoological Abstract, EBSCO Database, Directory of Open Access Journals, Index Copernicus Master List, Open J Gate, NDL Japanese Periodicals Index, Genomics Journal Seek Database, SCImago and SCIRUS.

**Hashemite University**  
Deanship of Scientific Research and Graduate Studies  
**TRANSFER OF COPYRIGHT AGREEMENT**

Journal publishers and authors share a common interest in the protection of copyright: authors principally because they want their creative works to be protected from plagiarism and other unlawful uses, publishers because they need to protect their work and investment in the production, marketing and distribution of the published version of the article. In order to do so effectively, publishers request a formal written transfer of copyright from the author(s) for each article published. Publishers and authors are also concerned that the integrity of the official record of publication of an article (once refereed and published) be maintained, and in order to protect that reference value and validation process, we ask that authors recognize that distribution (including through the Internet/WWW or other on-line means) of the authoritative version of the article as published is best administered by the Publisher.

To avoid any delay in the publication of your article, please read the terms of this agreement, sign in the space provided and return the complete form to us at the address below as quickly as possible.

Article entitled:-----

Corresponding author: -----

To be published in the journal: Jordan Journal of Biological Sciences (JJBS)

I hereby assign to the Hashemite University the copyright in the manuscript identified above and any supplemental tables, illustrations or other information submitted therewith (the "article") in all forms and media (whether now known or hereafter developed), throughout the world, in all languages, for the full term of copyright and all extensions and renewals thereof, effective when and if the article is accepted for publication. This transfer includes the right to adapt the presentation of the article for use in conjunction with computer systems and programs, including reproduction or publication in machine-readable form and incorporation in electronic retrieval systems.

Authors retain or are hereby granted (without the need to obtain further permission) rights to use the article for traditional scholarship communications, for teaching, and for distribution within their institution.

- I am the sole author of the manuscript
- I am signing on behalf of all co-authors of the manuscript
- The article is a 'work made for hire' and I am signing as an authorized representative of the employing company/institution

Please mark one or more of the above boxes (as appropriate) and then sign and date the document in black ink.

Signed: \_\_\_\_\_ Name printed: \_\_\_\_\_

Title and Company (if employer representative) : \_\_\_\_\_

Date: \_\_\_\_\_

Data Protection: By submitting this form you are consenting that the personal information provided herein may be used by the Hashemite University and its affiliated institutions worldwide to contact you concerning the publishing of your article.

Please return the completed and signed original of this form by mail or fax, or a scanned copy of the signed original by e-mail, retaining a copy for your files, to:

Hashemite University  
Deanship of Scientific Research and Graduate Studies  
Zarqa 13115 Jordan  
Fax: +962 5 3903338  
Email: [jjbs@hu.edu.jo](mailto:jjbs@hu.edu.jo)



## **EDITORIAL PREFACE**

Jordan Journal of Biological Sciences (JJBS) has had another great year. We have seen a significant increase in articles submission from both regional and international scholars. The editorial board members of JJBS have been very busy throughout the year to maintain excellence in the quality publication of accepted papers. As a result, JJBS has been indexed by CABI's Full-Text Repository, EBSCO and is currently under evaluation to be indexed in National Library of Medicine's MEDLINE\ PubMed system and Elsevier's SciVerse Scopus. As in the previous two years, this sixth volume of JJBS will include four issues, ten to twelve articles in each issue. In the coming year, it is my vision to have JJBS publishes more outstanding papers and review articles from distinguished scholars in various areas of biological sciences. In addition, I will be working on the inclusion of JJBS in ISI, which will lead to a wider readership and good impact factor. As you read throughout this inaugural volume of JJBS, I would like to remind you that the success of our journal depends directly on the number of quality articles submitted for review. Accordingly, I would like to request your participation by submitting quality manuscripts for review and by encouraging your colleagues to do the same. One of the great benefits we can provide to our prospective authors, regardless of acceptance of their manuscript or not, is the mentoring nature of our review process. JJBS provides authors with high quality, helpful reviews that are shaped to assist authors in improving their manuscripts.

I would like to thank the JJBS International Advisory Board members for their continuous support of JJBS. Furthermore, I would like to thank the JJBS Editorial Board members for their exceptional work and continuous support to JJBS. My thanks are also extended to the Hashemite University and Jordanian Scientific Research Support Fund for their continuous financial and administrative support to JJBS.

Moreover, and as always, I would like to highlight and proudly thank the group of authoritative reviewers, both local and international, who have done an outstanding work. We are honored to have you on our review list and many thanks for your valuable mentorship and contributions you provided to authors. Indeed, we count on your excellent reviews to include only high quality articles worthy of publication in JJBS. Together, we strive to make JJBS reach a remarkable rank among other international journals. I very much appreciate your support to make JJBS one of the most authoritative journals in biological sciences.

Prof. Khaled H. Abu-Elteen  
Editor-in-Chief  
Hashemite University  
Zarqa, Jordan  
March 2013



## CONTENTS

**Review Articles**

- 1 - 4 Use of Anabolic Androgenic Steroids in Jordan: Mini- Review.  
*Lubna H. Tahtamouni*

**Original Articles**

- 5 - 10 Development of *Dermestes maculatus* (DeGeer, 1774) (Coleoptera, Dermestidae) on Different Fish Substrates.  
*Usman Zakka, Jonathan N. Ayertey and Millicent A. Cobblah*
- 11 - 16 Composition and Larvicidal Activity of *Artemisia vulgaris* L. Stem Essential Oil Against *Aedes aegypti*  
*Sujatha Govindaraj and Bollipo D. Ranjitha Kumari*
- 17 - 20 Susceptibility of the Hymenopteran Parasitoid, *Habrobracon hebetor* (Say) (Braconidae) to the Entomopathogenic Fungi *Beauveria bassiana* Vuillemin and *Metarhizium anisopliae* Sorokin.  
*Vahid Mahdavi, Moosa Saber, Hooshang Rafiee-Dastjerdi and Ali Mehrvar*
- 21 - 24 Accumulation of Copper in Different Tissues and Changes in Oxygen Consumption Rate in Indian Flying Barb, *Esomus danricus* (Hamilton-Buchanan) Exposed to Sub-lethal Concentrations of Copper.  
*Suchismita Das and Abhik Gupta*
- 25 - 30 Modification of the Mechanical Properties of Red Blood Cell Membrane by Spent *Plasmodium falciparum* Culture Supernatant.  
*Nii A. Aryee and Yuichi Takakuwa*
- 31 - 38 The Antihyperglycaemic Effect of the Aqueous Extract of *Origanium vulgare* Leaves in Streptozotocin-Induced Diabetic Rats.  
*Nema A. Mohamed and Omimah A. Nassier*
- 39 - 44 Fecundity of Bigfin squid, *Sepioteuthis lessoniana* (Lesson, 1830) (Cephalopoda: Loliginidae).  
*Venkatesan Vellathi and Rajagopal Santhanam*
- 45 - 50 Effect of Dredging on the Macrozoobenthos of Hazratbal Basin in the Dal Lake Srinagar Kashmir, India.  
*Basharat Mushtaq, Rajni Raina, Abdul R. Yousuf, Ashwani Wanganeo and Ummer Rashid*
- 51 - 60 Evaluation of the Physicochemical Properties and Antimicrobial Activities of Bioactive Biodegradable Films.  
*Mary S. Khali, Zahra S. Ahmed and Aml S. Elnawawy*
- 61 - 66 Effects of Three Medicinal Plant Products on Survival, Oviposition and Progeny Development of Cowpea Bruchid, *Callosobruchus maculatus* (Fab.) [Coleoptera: Chrysomelidae] Infesting Cowpea Seeds in Storage.  
*Kayode D. Ileke, Daniel S. Bulus and Ayisat Y. Aladegoroye*
- 67 - 72 Reserve Mobilization, Total Sugars and Proteins in Germinating Seeds of Durum Wheat (*Triticum durum* Desf.) under Water Deficit after Short Period of Imbibition.  
*Amal M. Harb*

---

**Short Communication**

---

73 - 76

A Novel Report on the Prevalence of Enterohaemorrhagic *Escherichia coli* non-O157 Isolated from Cattle in Kaduna State, Nigeria.

*Jasini A. Musa, Mashood A. Raji, Haruna M. Kazeem and Nicodemus M. Useh*

# Use of Anabolic Androgenic Steroids in Jordan: Mini- Review

Lubna H. Tahtamouni\*

*Department of Biology and Biotechnology, Faculty of Science, The Hashemite University,*

*P.O. Box 150459, Zarqa 13115, Jordan*

*Received: October 5, 2012; Accepted November 15, 2012*

## Abstract

Until recently, there has been a complete lack of prevalence data regarding anabolic androgenic steroid (AAS) abuse locally in Jordan and the Arab world at large. Encouraged by their trainers, many Jordanian athletes are abusing AAS. The sale of anabolic steroids in Jordan is still unregulated; selling AAS without a prescription is not banned. The Jordanian Anti-Doping Organization (JADO) was founded in 2007, and as a response to the increasing concern about steroid abuse among Jordanian youth JADO accepted the World Anti-Doping Code late 2009, committing Jordan to fighting all types of doping including AAS abuse. However, most Jordanians are not aware of the adverse side effects of AAS abuse. Promoting public awareness about the harmful consequences of AAS abuse, in addition to the role of the law enforcement authorities will hopefully curb the abuse of AAS in Jordan.

**Keywords:** Doping, the Code, Bodybuilder.

## 1. Introduction

Research about the use of anabolic androgenic steroids (AAS) in Jordan and in the Arab world at large is scarce. In addition to two prevalence studies (Al-Falasi *et al.*, 2008; Tahtamouni *et al.*, 2009), few reports from the Arab world came out after the year 2000 where only case reports of AAS abuse were discussed (Alaraj *et al.*, 2005; Samaha *et al.*, 2008). However, more attention has been drawn to this issue as a result to the growing concern of steroid abuse among Jordanian youth and athletes (Tahtamouni *et al.*, 2009).

As a political response to this concern, the Jordanian Anti-Doping Organization (JADO) was founded in 2007 ([www.jado.jo](http://www.jado.jo)). Late that year, government and national Olympic committee officials from Jordan and four other Arab countries met under the supervision of the West Asia Regional Anti-Doping Organization (West Asia RADO), which is a part of the World Anti-Doping Agency (WADA), and established the West Asia Anti-Doping Organization (WAADO). The members of WAADO (located in Amman, the capital of Jordan) have agreed to pull resources together to fight doping throughout the region ([www.wada-ama.org](http://www.wada-ama.org)).

JADO acts as an independent anti-doping organization for Jordan. It has the necessary authority and responsibility for implementing and advocating improvements in the doping control such as testing for AAS abuse among athletes, promoting anti-doping research and planning and monitoring information and education programs

([www.jado.jo](http://www.jado.jo)). Late 2009, JADO accepted the revised 2009 World Anti-Doping Code (Jordan Olympic Committee Anti-Doping Rules, 2009) (the "Code") (World Anti-Doping Code, 2012). Simultaneously, co-operation between various public authorities have led to new strategies which aim at reducing the supply of steroids and as such limiting their use within the general population.

### 1.1. The Anabolic Steroid Users

Despite signs of public concern and disapproval of anabolic steroid use, some athletes – competing as well as recreational – are using these drugs. In fact, competing bodybuilders, weightlifting athletes and students at Jordanian colleges have been identified as specific AAS-using groups (Tahtamouni *et al.*, 2009). Extractions from interviews with trainers/trainees in various gyms across this region - taken from newspapers - indicate that the motives behind the use of steroids are:

A salesman: "People want to build strength, lose fat and have the perfect bodies they see on TV" (Luck, 2009).

Iraqi gym owner: "So many people now want to work out. They want to look good" (Luck, 2009).

A-17-year old Jordanian weightlifting-trainee admitted using "supplements." He said: "I used to be 60 kilos and weak. Now, with the new products and a good coach, I'm going to be better than Rambo" (Luck, 2009).

A Kuwaiti male in his 30's said: "I wanted to bulk up, everyone at my gym had a lot of muscle, and I wanted to be like them. My friends started telling me about the courses [of steroids] they take, injections they use to get

\* Corresponding author. e-mail: [lubnatahtamuni@hu.edu.jo](mailto:lubnatahtamuni@hu.edu.jo).

bigger. So I tried it, they showed me how” (Kolarov, 2009).

Competing bodybuilders and weightlifters that were surveyed by the author (Tahtamouni *et al.*, 2009) described how trainers were advocating “the wonderful properties” of anabolic steroids. As one trainer explained: “One month of hormonal injections and you will be a clone of Sylvester Stallone” and: “Injections work much quicker than pills in blowing the muscles” (Personal Communication). Despite the general disapproval of the use of performance enhancing drugs (AAS), still some seem quite willing to use them in order to obtain the desired body look they want to have.

## 2. The Public Health Concern

Some examples of the users’ behavior when training and while using steroids indicate that there is a real cause for a public health concern. In a correspondence with an MD who is an expert in substance abuse and who is employed at a major hospital in Jordan, he explained how almost every week he was called upon for a consultation with a patient who was admitted to the hospital due to complications resulting from anabolic steroid overdosing. Even autopsies where the cause of death (apparently) was found to be excessive use of steroids are not uncommon (Personal Communication). However, exact numbers of such patients, numbers of autopsies conducted on steroid users and case reports demonstrating a direct relationship between use of steroids and cause of death are unavailable. The information provided in the correspondence should be viewed as an indication that some users are using steroids to an extent where they can be fatal or can cause serious health complications.

In general, very little information about the side effects associated with steroid use has been available in Jordan. It seems that many coaches and gym owners dealing with anabolic steroids in gyms are in fact unaware of the potential harmful side effects of these drugs (Büttner and Thieme, 2010); and if they are, these effects are rarely discussed with the users (e.g. competing bodybuilders and weightlifters) (Tahtamouni *et al.*, 2009; Personal Communication). The quality of advice may therefore vary greatly. Some may even disregard the potential risks of using AAS, as expressed by one weightlifting trainer: “If I can bench press hundreds of pounds and have low fat, then in my opinion I’m healthy” (Luck, 2008). These available examples do point to the conclusion that something needs to be done.

Some abusers do not have the nerve to administer the injectable AAS for themselves, so they ask a friend to do it for them who in turn might be inexperienced (Personal Communication). Also, Jordan currently does not have a syringe exchange program (Syringe Exchange Programs Around the World, 2012), and therefore it is possible that the same syringe is used again and again, or is circulated among users, in which case, a high risk of infection will arise. It is worth mentioning that injection equipments, syringes and needles, are readily available over-the-counter in most Jordanian pharmacies; a pack of 100-1ml syringes cost less than 5\$ (www.jomid.net).

## 3. The Drug Supply

The official sale of anabolic steroids in Jordan is still unregulated. Selling anabolic steroids at pharmacies - without a prescription - is not banned in Jordan and these drugs are so far unmentioned in any Jordanian law or any medicine regulation act (www.moh.gov.jo). Many pharmacists provide AAS as an over-the-counter drug (www.moh.gov.jo). Many AAS drugs are not registered in the Jordanian Pharmacist Association (www.jpa.org.jo). There are only three AAS that are prescribed in Jordan, namely Deca-durabolin, Andriol and Sustanon. Besides their use by athletes, these drugs are prescribed in Jordan as a treatment for reproductive dysfunction and breast cancer (Smith, 2009).

However, most of the steroids used in the gyms surveyed by the author (Tahtamouni *et al.*, 2009) are smuggled into the country, mainly from Pakistan, Egypt and India, which is against the customs law and regulations. The customs law regulates all types of smuggling into Jordan. Customs and police authorities prosecute violators and the Ministry of Health and the Food and Drug Administration handles any drugs being confiscated (www.customs.gov.jo). The drugs that are being smuggled into Jordan are supposedly of low quality (e.g. contaminated or less active compound than stated) (Evans-Brown and McVeigh, 2009). Still, to our knowledge no analysis of drugs confiscated at the borders has been published. Despite the supposedly low quality, these products are being used, presumably because they are sold at a cheaper price than steroids available from pharmacies.

### 3.1. The health policy answer

In a rather conservative society, such as that of Jordan, drug use is negatively conceived of, especially when it comes to drugs such as cocaine and cannabis. With anabolic steroids, however, there seems to be public confusion and some people are unaware of the adverse side effects of these drugs (e.g. think anabolic steroids are vitamins) (based on field work by Tahtamouni, 2009-2011). It is a priority for health authorities to inform the public since “people are using all these products (steroids) without knowing the side effects and this is very dangerous” as Jordan’s WHO representative explained (Luck, 2008). Can the formation of JADO increase the amount of available information about use of anabolic steroids?

With the establishment of JADO, competing athletes will be tested for use of performance enhancing drugs. However, in every gym which is licensed under the Higher Council of Youth – in all likelihood some non-competing athletes are found among the gym members – testing may be carried out. Does this mean that recreational athletes with no intention of competing may be subjected to testing regardless of whether or not they intend to compete as bodybuilders or weightlifters at competitions? Is this an unintended consequence of the testing system or does JADO deliberately seek to test recreational athletes?

Will doping tests have a deterrent effect on recreational athletes in these gyms?

There are gyms in Jordan which are not member of any kind of sports federation thus precluded from the testing system set up by JADO. The effects of this testing scheme remain to be seen and evaluated.

Alliances between public authorities - which could enforce and further implement new anti drug strategies – have been formed and institutions such as the Ministry of Health, Ministry of Education, Ministry of Higher Education, Higher Council of Youth, Department of Customs and Jordan Anti-Narcotics Police Department have been working together under the supervision and coordination of JADO. Between these co-operating alliances new legislation which will prohibit pharmacy sale of anabolic steroids without a prescription and introduce severe punishment for smuggling of steroids are currently in the process of being formulated. The upcoming law is still in working progress and sanctions for illegal sale or smuggling into the country remains undecided upon.

#### 4. Conclusion

Possession of anabolic steroids without prescription in Jordan is currently legal, and new legislation attempts to limit supply rather than criminalize the individual user. However, so far use of anabolic steroids in Jordan has to a great extent been unregulated.

#### Acknowledgements

The author gratefully acknowledges The Deanship of Research and Graduate Studies, The Hashemite University.

#### Conflicts of interest

None declared.

#### References

- Alaraj A, Chamoun R, Dahdaleh N, Haddad G and Comair Y. 2005. Spontaneous subdural haematoma in anabolic steroids dependent weight lifters: reports of two cases and review of literature. *Acta Neur (Wien)*, **147**: 85-88.
- Al-Falasi O, Al-Dahmani K, Al-Eisaei K, Al-Ameri S, Al-Maskari F, Nagelkerke N and Schneider J. 2008. Knowledge, attitude and practice of anabolic steroids use among gym users in Al-Ain district, United Arab Emirates. *Open Sports Med J*, **2**:75-81.
- Büttner A and Thieme D. 2010. Side effects of anabolic steroids: pathological findings and structure-activity relationships. *Handb Exp Pharmacol*, **195**:459-484.
- Custom Law, Law No. (20) of the Year 1998 [website] ([http://www.customs.gov.jo/English/customs\\_en.shtml](http://www.customs.gov.jo/English/customs_en.shtml)).
- Evans-Brown M and McVeigh J. 2009. Injecting human growth hormone as a performance enhancing drug—perspectives from the United Kingdom. *J Sub Use*, **14**: 267–288.
- Jordan Anti-Doping organization [website] (<http://www.jado.jo/index.php?lang=eng>).
- Jordan Medical Information Directory [website] (<http://www.jomid.net>).
- Jordan Ministry of Health [website] (<http://www.moh.gov.jo>).
- Jordan Olympic Committee Anti-Doping Rules, Version 1.0-July 2009 [website] ([http://www.jado.jo/pdf/eng\\_sund.pdf](http://www.jado.jo/pdf/eng_sund.pdf)).
- Kolarov H. 2009. Steroid abuse: A reality in Kuwaiti gyms. *Kuwait Times*, 6 February.
- Luck T. 2008. Looking good-but at what price. *The Jordan times*, 15 February.
- Samaha A, Nasser-Eddine W, Shatila E, Haddad J, Wazne J and Eid A. 2008. Multi-organ damage induced by anabolic steroid supplements: a case report and literature review. *J Med Case Reports*, **2**: 340.
- Smith R. 2009. Diseases, Disorders, and Common Problems, Section II. In: **Netter's Obstetrics and Gynecology**. Elsevier Inc. Canada, pp. 20-21.
- Syringe Exchange Programs Around The World: The Global Context [website] ([http://www.gmhc.org/files/editor/file/gmhc\\_intl\\_seps.pdf](http://www.gmhc.org/files/editor/file/gmhc_intl_seps.pdf)).
- Tahtamouni L, Mustafa N, Alfaouri A, Hassan I, Abdalla M and Yasin S. 2008. Prevalence and risk factors for anabolic-androgenic steroid abuse among Jordanian collegiate students and athletes. *Eur J Pub Health*, **18**: 661-665.
- The Jordanian Pharmacist Association [website] (<http://www.jpa.org.jo>).
- World Anti-Doping Code [website] ([http://www.wada-ama.org/Documents/World\\_Anti-Doping\\_Program/WADP-The\\_Code/WADA\\_Anti-Doping\\_CODE\\_2009\\_EN.pdf](http://www.wada-ama.org/Documents/World_Anti-Doping_Program/WADP-The_Code/WADA_Anti-Doping_CODE_2009_EN.pdf)).
- World Anti-Doping Organization [website] (<http://www.wada-ama.org>).



# Development of *Dermestes maculatus* (DeGeer, 1774) (Coleoptera, Dermestidae) on Different Fish Substrates

Usman Zakka<sup>1,\*</sup>, Jonathan N. Ayertey<sup>2</sup> and Millicent A. Cobblah<sup>2</sup>

<sup>1</sup>Faculty of Agriculture, Department of Crop/Soil Science, University of Port Harcourt.

East West Road Chaba Campus PMB 5323 Port Harcourt;

<sup>2</sup>African Regional Post Graduate Programme in Insects Science, University of Ghana, Legon, Ghana

Received: June 6, 2012, Accepted: August 12, 2012

## Abstract

The development of *Dermestes maculatus* (DeGeer) on four smoked fish substrates [Trigger fish (*Balistes capriscus*), Catfish (*Synodontis sp.*), African catfish (*Clarias gariepinus*) and Nile Tilapia (*Oreochromis niloticus*)] were investigated as completely randomized design under laboratory temperature of 30°C, relative humidity of 65±5% and a light: darkness regimen of 12:12 hours. Females laid eggs within 24 hours of copulation. The numbers of eggs laid and the period between larval instars were not significantly different ( $P > 0.05$ ). Mean total egg laying period varied from 18 days on *B. capriscus* to 30 days on *Synodontis sp.* with about 75% of eggs laid on days 13, 15, 15 and 17 for *O. niloticus*, *B. Capriscus*, *C. gariepinus*, and *Synodontis sp.*, respectively. Hatching started 48 hours after copulation on all fish substrates. The mixed fish substrate (comprising all species) gave the longest ( $P < 0.05$ ) developmental period of 42.75 days. Except *C. gariepinus* which recorded five larval instars, all others gave six instars. The total development period of *D. maculatus* from egg → larva → pre-pupa → pupa → adult emergence on the fish substrates decreased in the order *Synodontis* > *O. niloticus* > Mixed > *C. gariepinus* > *B. capriscus*.

**Keywords:** *Dermestes Maculatus*, Developmental Period, Emergence Pattern, Fish Substrates, Copulation.

## 1. Introduction

Fish has remained an important source of food and income to many people in the developing world including Africa where as much as 25% of the population depend on it (Essuman, 1992). It is a very rich source of good quality protein in diets of man (Amusan and Okorie, 2001; Fasakin and Aberejo, 2002; Azam *et al.*, 2004; Aderolu and Akpabio, 2009). Don-Pedro (1989) concurs that during storage, transportation and marketing, dried fish is readily attacked by several species of insects notably *D. maculatus*, *D. frischii*, *D. ater* and *Necrobia rufipes*. FAO (1990) reported that *Dermestes spp.* and *N. rufipes* were major pests of smoked fish, poultry products (Geden and Hogsette, 2001), museums (Linnie and Keatinge, 2000), Egyptian mummies (Adams, 1990) and stored cocoons of silk-worm *Bombyx mori* (Sahaf, 2007). Lale and Sastawa (1996) and Odeyemi *et al.* (2000) recorded about 50% losses during the storage of smoked fish products due to deterioration. The losses have been attributed to net reductions in the amount of nutrients available to the consumer (nutritive quality) resulting to declining consumer acceptability and market prices (economic losses) or both quantitative and qualitative

losses (Odeyemi *et al.*, 2000; Atijegbe, 2004). Thus, the experiment was designed to investigate the developmental processes of *D. maculatus* on substrates from four species of smoked fish with the aim of understanding the biology of the pest for effective and efficient management measures against losses caused by the pests in stored fish products.

## 2. Materials and Methods

The studies were carried out between October 2004 and May 2005 under controlled temperature (30°C), relative humidity (65±5 %) and light-to-darkness regimen of 12:12 hours. Smoked fish from four species of fish – the Trigger fish (*Balistes capriscus* Gmelin), Catfish (*Synodontis sp.*), African catfish (*Clarias gariepinus* Burchel) and Nile tilapia (*Oreochromis niloticus* Linnaeus) – were purchased from Madina and Makola local Markets in Accra, Ghana and used for the experiment. Treatments were arranged as completely randomized design (CRD), replicated four times and kept on open air shelves. The life cycle of the pest was determined on each food medium and appropriate records taken as outlined below.

\* Corresponding author. e-mail: uzakka@yahoo.com.

### 2.1. Rearing of *D. Maculatus*

Several unsexed adults of *D. maculatus* obtained from naturally infested smoked fish materials served as sources of the pests. The glass jars and cured fish species- *B. caprisicus* (Trigger fish), *Synodontis sp.* (Catfish), *C. gariepinus* (African catfish) and *O. niloticus* (Nile Tilapia) used for the experiment were heat-sterilized at 60°C for one hour in a hot-air Gallenkamp oven in the laboratory to kill all insect pests that may be present. The experimental bottles had their lids opened and sealed with 4 cm diameter mesh to facilitate aeration of the culture and placed on inverted Petri dishes submerged in white oil on shallow trays to keep out mites and other insect pests. Adult *D. maculatus* were then transferred into sterilized jars containing the disinfested smoked fish from *B. caprisicus*, *Synodontis sp.*, *C. gariepinus* and *O. niloticus* species to initiate new colonies of the parent stock for rearing the pests. The insect pests were fed on the four different fish substrates.

About 1000 g of the sterilized fish substrates were then conditioned for 2 days under ambient laboratory conditions with about 600 g of each substrate later poured into a series of 1-litre jars. About 20 unsexed adults of *D. maculatus* were introduced into each jar and their offspring allowed developing up to the pupa stage. The adults were sieved out after fourteen days of oviposition to ensure that offspring of relatively same age were obtained as pure F<sub>1</sub> *D. maculatus*. The pupae were then transferred from each of the substrates into separate sub-culture bottles containing each pure fish substrates. The rearing cultures were left undisturbed over a long period of time but pupae were isolated from each substrate at 7-day intervals and introduced into separate test tubes prior to adult emergence to ensure that adults were 47 days old before sexing them and kept unmated until required. The males are distinguished from the females by their possession of a deep depression and brush of hairs on the 4th abdominal sternite (Imai *et al.*, 1990). On emergence, the adults were placed in test tubes containing similar fish substrates to the ones on which they were bred and maintained under same conditions.

### 2.2. Egg Laying Bioassay

The experimental bottles were sterilized in a Gallenkamp oven, as described above, to obtain the number of egg(s) laid per female per day. Each fish species was then carefully dissected using entomological scissors and compacted with rubber band as in whole fish substrates. A male and a female adult *D. maculatus* (each 47-day old) were then introduced into separate tubes containing each fish species to serve both as food and oviposition medium and incubated for 30 days. Water was provided as soaked cotton wool and insects were allowed to drink for five minutes while egg count was done. The jars were monitored twice daily for the presence of eggs using hand lens and any egg seen was counted and removed using soft brush after which the medium was returned to its original position.

### 2.3. Larval Development

Larval instars were examined after collecting and placing eggs laid on each fish substrate from each of the experimental jars into glass tubes for incubation. On

hatching, the larvae were separated into individual tubes of 2.2 cm × 15 cm dimensions containing 10 g of each fish species and kept under observation for their development. Duration of each larval instar was determined on each substrate by the presence of exuviate after each moult. All larvae were derived from eggs laid by individuals maintained on smoked fish substrates used in determining the number of eggs laid.

### 2.4. Pupal Period

The pre-pupal stage occurs when the last instar larva becomes almost C-shaped, shortened and remains non-motile for some days, while the pupal stage is what follows immediately. Pupae were removed and placed individually in clear tubes and held under laboratory conditions until adult emergence. The sexes, length and pupal periods were recorded for each emerged adult.

Data collected on eggs laid, developmental pattern, larval instars, prepupal and pupal durations were transformed using square roots of  $\sqrt{(x+1)}$  and analysed using Genstat software version 5 Release 3.2 (Lawes Agricultural Trust, 1995) and subjected to analysis of variance at 95% level of significance and significant means were separated using LSD at 0.05 error limit.

## 3. Results

### 3.1. Pattern of oviposition by *D. maculatus* on four different fish substrates

Eggs laid on each fish species were random but gradually increased in number during the first week and subsequently declined with time. About 75% of eggs were laid on *Synodontis* by day 17 while the same level was reached on *C. gariepinus* and *B. caprisicus* by day 15 and on *O. niloticus* by day 13 (Figure 1). Maximum egg laying periods were recorded on *Balistes sp.* and *Synodontis sp.* in 30 days while the minimum egg laying period recorded on *O. niloticus* was 18 days. The maximum recorded number of eggs laid in a batch per day was 28 on *O. niloticus* and a minimum of a single egg per day was recorded on all the fish substrates.

### 3.2. Total Eggs Laid By *D. Maculatus* Within 30 Days on Different Fish Substrates

The total number of eggs laid on the different fish substrates over 30 days was not statistically significant ( $P > 0.05$ , F Prb. = 0.275). The highest mean number of eggs was, however, laid on *O. niloticus* (151±33.67) and the lowest were on *B. caprisicus* (103±43.5) with *Synodontis sp.* (132±31.11) and *C. gariepinus* (117±14.39) as intermediates (Table 1).

**Table 1.** Total number of egg laid on different fish substrates within 30 days

Fish substrates	Total eggs laid ±SE*	Range (eggs/day)
<i>O. niloticus</i>	151±33.67	1-28
<i>Synodontis sp</i>	132±31.11	1-20
<i>C. gariepinus</i>	117±14.39	1-19
<i>B. caprisicus</i>	103±43.51	1-18

### 3.3. Larval Instars of *D. Maculatus* on Different Fish Substrates

There were 6 larval instars of *D. maculatus* on all the fish substrates except on *C. gariepinus* substrates where only 5 instars were recorded (Table 2). The mean duration (in days) of the various larval instars on the different fish substrates was not statistically significant ( $P > 0.05$ , F Prb. = 0.287) without a clear trend in larval developmental periods on the substrates over 50 days. While larval development on *O. niloticus* had uniform periods except in the 6th larval instar, those on *Synodontis* and *B. capricus* had the highest number of days recorded in the 2nd larval instar and least in the 6th instar. However, on *C. gariepinus* the highest period was recorded in the 5th instar. When placed on mixed substrates, there was generally a progressive increase in duration in each subsequent larval instar (Table 2). The mean duration of larval instars was highest in the 2nd instar (range: 6-12 days) followed by the 5th instar (range: 4-11 days), and the least duration was recorded on the 6th larval instar.

**Table 2.** Mean duration (days) of larval instars of *D. maculatus* on the different fish substrates

Fish species	Duration (days) of larval instars ±S.E*						Total
	1	2	3	4	5	6	
<i>O. niloticus</i>	5.75±0.48	5.75±0.48	5.75±0.48	5.75±1.03	5.25±0.48	7±0.00	35.25
<i>Synodontis</i> sp.	6.75±0.75	10±0.82	5.25±0.48	6.00±0.91	6.5±0.65	5±0.00	35.75
<i>C. gariepinus</i>	5.00±0.00	7.25±0.25	5.75±0.25	5.00±0.00	8.00±0.00	.	31.00
<i>B. capricus</i>	5.75±0.48	9.00±0.41	5.75±0.48	6.00±0.71	5.25±0.48	5.5±0.35	37.25
Mixed subs.	5.00±0.00	6.50±0.29	7.25±0.48	6.50±0.65	9.00±1.35	8.5±0.35	42.75

\*Values are means of four replicates ± SE. (Standard error)

### 3.4. Developmental Periods of Pre-Pupa and Pupal Stages of *D. Maculatus*

Statistical analysis showed significant differences ( $P < 0.05$ , F Prb. = 0.083) in the mean period between pre-pupa and the emergence of external adults. *O. niloticus* recorded higher pre-pupal period while the least pre-pupal period was recorded on mixed substrates. The results further showed significant difference ( $P < 0.05$ , F Prb. = 0.083) in pupal period with *B. capricus* recording the least from the other fish substrates (Table 3).

**Table 3.** Mean period of development from pre-pupa to adult emergence of *D. maculatus* on different fish substrates

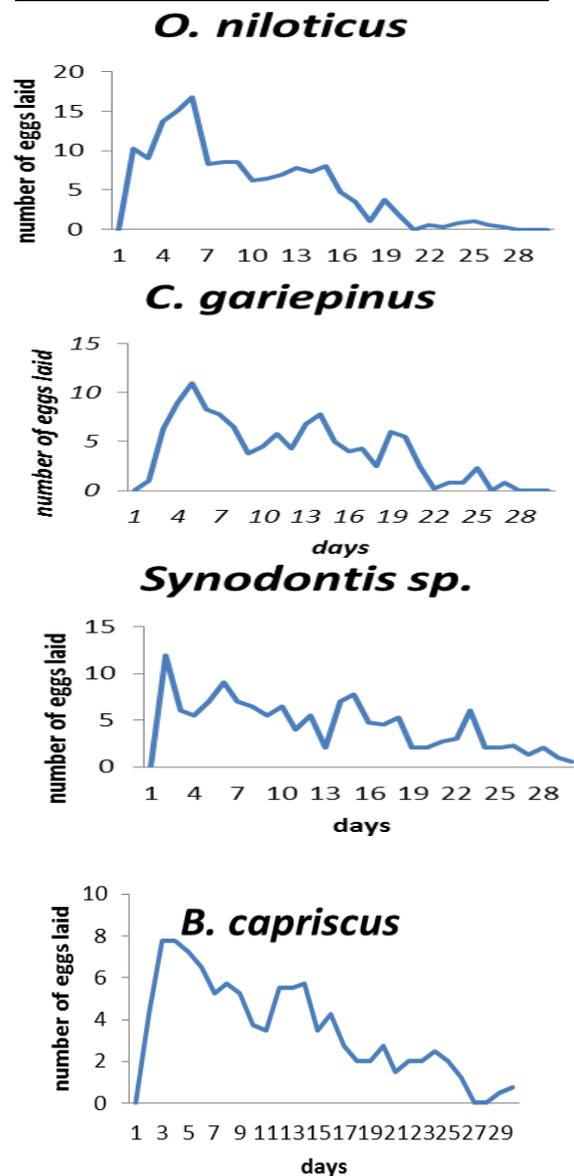
Fish species	Developmental period ± SE*			
	Pre-Pupa	Range	Pupa	Range
<i>O. niloticus</i>	12.75 <sup>a</sup> ±1.65	9-16	9.00 <sup>a</sup> ±1.29	6-10
<i>Synodontis</i> sp.	11.00 <sup>ab</sup> ±1.08	8-13	9.50 <sup>b</sup> ±0.96	8-12
<i>C. gariepinus</i>	8.75 <sup>bc</sup> ±1.11	6-9	7.50 <sup>b</sup> ±0.65	6-9
<i>B. capricus</i>	8.00 <sup>c</sup> ±0.82	9-12	6.75 <sup>b</sup> ±1.38	5-10
Mixed substrates	7.25 <sup>c</sup> ±0.48	6-8	8.00 <sup>a</sup> ±1.08	6-11
LSD 2.73				

Means with the same superscripts in the same column are not significantly ( $P > 0.05$ ) different.

However, the cumulative developmental periods of *D. maculatus* from pre-pupa to pupa on the various fish substrates did not show any significant differences ( $P > 0.05$ , F Prb. = 0.135) (Table 4). The total development period of *D. maculatus* from egg → larva → pre-pupa → pupa → adult emergence on the fish substrates decreased in the order *Synodontis* > *B. capricus* > *O. niloticus* > *C. gariepinus*.

**Table 4.** Mean development period (days) of pre-pupa and pupa of *D. maculatus* on different fish substrates

Developmental stages	Days± SE	Range
Pre-pupa	9.55±1.14	7.25-12.75
Pupa	8.15±0.56	6.75-9.50
LSD	1.74	
Values are means of four replicates ±SE		



**Figure 1.** Oviposition pattern of *D. maculatus* on different fish substrates

## 4. Discussion

### 4.1. Oviposition in *D. maculatus* on fish substrates

The study revealed that in *D. maculatus* copulation occurred immediately the adults were paired and this could be explained as reported that both male and female *D. maculatus* produce sex pheromone (Rakowski and Cymborowski, 1986; Jaskulska *et al.*, 1987) which enhances communication and within 48 hours creamy white eggs were laid. The eggs laid were oval in shape and bluntly pointed at both ends as earlier reported (Archer and Elgar, 1999; Jones and Elgar, 2004; Ezenwaji and Obayi, 2004). The results of this study confirm the works of various researchers that *D. maculatus* females copulate within 30 minutes of pairing with the males initiating copulation (Archer and Elgar, 1999; Jones and Elgar, 2004).

Egg laying in *D. maculatus* per female within 30 days on each of the different fish substrates was random and varied. The egg laying capacity of the pest tested on different fish substrates followed the descending order: *O. niloticus* > *Synodontis* > *C. gariepinus* > *B. caprisacus*. *O. niloticus* was thus the most suitable medium for egg laying on the four substrates tested and expected to carry the heaviest infestation in the field. These differences may be attributed to the fact that oviposition in insects on a specific host is determined by various factors that may determine its suitability or otherwise as a breeding medium, such as nutritional quality, host abundance (Jansen and Nylin, 1997; Barros and Zucoloto, 1999), morphology, environmental conditions, age and size of individual (Stejskal and Kucerova, 1996; Johnson and Kistler, 1987) and competition (Siemens *et al.*, 1991). The study showed that none of the fish substrates deterred egg laying, though some of the media proved to be better than others as more suitable for oviposition by *D. maculatus*.

Generally there was an initial increase in total number of eggs laid during the first week of oviposition on all the fish substrates but subsequently there was reduction in numbers as the days progressed and insects got older (Ezenwaji and Obayi, 2004). Oviposition behaviour in insects is an important contributor to the fitness of insects because of the consequent effect on the number and quality of offspring (Honek, 1993; Stejskal and Kucerova, 1996). The study further showed that 75% of eggs were laid between the 13th and 17th day on all the fish substrates. The peak laying period agrees with the results of Ezenwaji and Obayi (2004) who indicated that full oviposition in *D. maculatus* is attained during the first 6-8 days, becoming fairly uniform in about 16 days, indicating reduction in rate of oviposition with time as sperm viability also declined with age (Kidd *et al.*, 2001; Oakes *et al.*, 2003; Szczesny *et al.*, 2003).

Eggs were laid in different batches, ranging from 2-6 batches with 28 eggs per batch as the highest and this contradicts the results of (Osuji, 1975) who recorded upto 38 eggs in a batch. Maximum number of eggs recorded on *O. niloticus* was 151 within 30 days confirming the findings by previous workers (Amusan and Okorie, 2001), but differs from the findings of (Seal and Tilton,

1985 and Ezenwaji and Obayi, 2004) who recorded 407 and 598 eggs respectively.

The maximum egg laying period of 30 days recorded contradicts those of Taylor (1964) and Osuji (1975) who found maximum egg laying period in *D. maculatus* to be 14 and 189 days, but similar to the works of Coombs (1978). These may be attributed to the differences in temperature, relative humidity, age of the insects and the amount and kind of food supplied.

### 4.2. Larval instars in *D. maculatus* reared on different fish substrates

Observations made during the study showed that the hairy creamy larva on emergence darkens to light grey within a few hours. Although there were no significant differences in the duration of development of the larvae in the various fish species, larval development was shorter on *C. gariepinus* than on the other fish substrates. Assuming that a short development time on a certain fish species is an indication of good host suitability then, *C. gariepinus* may be marginally more suitable for development of *D. maculatus* which suggests higher level of infestation on *C. gariepinus*. Five larval instars were recorded on *C. gariepinus* but six on each of the other substrates. These differences could be attributed to the nutritional composition of the fish species as reported by Samish *et al.* (1992) that *D. maculatus* larva prefers substrates with high protein content. The second instar was longer on *Synodontis sp.* and *B. caprisacus*, while it was longer for the fifth instar on *C. gariepinus* and the mixed substrates. This conforms with the findings of Osuji (1975) and Rustin and Munro (1984), but different from Lale *et al.* (2000) who observed no differences in the numbers of larval instars on different fish species.

Larval development in *D. maculatus* does not involve any visible morphological change but only an increase in size from the previous instars (Osuji, 1975). The total development period of 31 days observed on *C. gariepinus* to 42.75 days on mixed substrates greatly differed from 91 days by Scoggin and Tauber (1951) and 16 days by Kreyenberg (1928). No differences in total developmental period between males and females were observed as was reported by (Kreyenberg, 1928).

### 4.3. Pre-pupal periods

In each of the fish substrates a quiescent period was observed at the end of the last larval instar where it became almost C-shaped, thickened and reduced in length from 10.99 to 6.54 mm on *O. niloticus*, 12.50 to 9.38 mm on *Synodontis sp.*, 12.95 to 8.42 mm on *C. gariepinus*, 11.38 to 8.34 mm on *B. caprisacus* and 11.97 to 9.22 mm on mixed substrates. The observation of a non-motile nature of the pre-pupa agrees with those of Osuji (1975), Anonymous (1980) and Cloud and Collison (1986). However, the pre-pupal and pupal duration differed from those of Ezenwaji and Obayi (2004) and Rustin and Munro (1984). It was also observed that pupal duration was not sex dependent as reported by (Kreyenberg, 1928).

### 4.4. Adult emergence

Comparing developmental period from eggs to adult emergence of the four fish substrates indicated that *C.*

*garipepinus* proved to be the most suitable for *D. maculatus* development because this fish substrate recorded the shortest period, even though it did not differ statistically from the others. These differences recorded may be due to evolutionary trend, physical form of the fish or its nutritional composition as reported by (Zakka *et al.*, 2009). It is not clear why *D. maculatus* females would prefer one fish substrate for oviposition and a different fish for feeding, since more eggs were laid on *O. niloticus*.

### Acknowledgement

The authors wish to thank Deutscher Akademischer Austauschdienst (DAAD) for funding the work

### References

- Adams RG. 1990. *Dermestes leechi* Kalik (Coleoptera: Dermestidae) from an Egyptian mummy. *Entomology Gazette* **41**:119–120
- Aderolu AZ and Akpabio VM. 2009. Growth and economic performance of *Clarias garipepinus* juveniles fed diets containing velvet bean, *Mucuna pruriens*, seed meal. *African J Aquatic Sci.*, **34**: 131-135.
- Amusan A A S and Okorie T G. 2001. The use of *Piper guineense* fruit oil (PFO) as protectant of dried fish against *Dermestes maculatus* (DeGeer) infestation. *Global J Pure and Applied Sci.*, **8**: 197-201.
- Anonymous, 1980. Insects in Poultry House. Ministry of Agricultural, Fisheries and Food. Leaflet 537. Agricultural Development and Advisory Services, Department of Agriculture and Fisheries for Scotland, Welsh Office UK.
- Archer M S and Elgar M A. 1999. Female preference for multiple partners: Sperm competition in the hide beetle, *Dermestes maculatus* (DeGeer). *Animal Behaviour*, **58**: 669-675.
- Atijegbe S R. 2004. Infestation of smoked fish in Ghana. M.Phil. Thesis in Entomology University of Ghana, Legon, Ghana. 103pp.
- Azam K, Ali M Y, Asaduzzaman M, Basher M Z and Hossain M M. 2004. Biochemical assessment of selected fresh fish. *J Biol Sci.*, **4**: 9-10.
- Barros H C H and Zucoloto F S. 1999. Performance and host preference of *Ascia monuste* (Lepidoptera: Pieridae). *J Insect Physiol.*, **45**: 7–14.
- Cloud J A and Collison C H. 1986. Comparison of various poultry house litter components for hide beetle (*Dermestes maculatus*, DeGeer) Larval development in the laboratory. *Poultry Sci.*, **65**: 1911-1914.
- Coombs CW. 1978. The effect of temperature and relative humidity upon the development and fecundity of *Dermestes lardarius* L. (Coleoptera, Dermestidae). *J Stored Product Res.*, **14**: 111-119.
- Don-Pedro K N. 1989. Insecticidal activity of some vegetable oils against *D. maculatus* (DeGeer) (Coleoptera; Dermestidae) on dried fish. *J Stored Product Res.*, **25**: 81-86.
- Essuman K M. 1992. Fermented fish in Africa. A study of processing, marketing and consumption. FAO Fisheries Technical Paper 329.
- Ezenwaji H M G and Obayi N S. 2004. The effect of fish moisture content on oviposition, fecundity and development of the hide beetle *Dermestes maculatus* DeGeer (Coleoptera: Dermestidae). *Animal Res Inter.*, **1(1)**: 47-51
- Fasakin E A and Aberejo BA. 2002. Effect of smoked pulverized plant material on the developmental stages of fish beetle, *Dermestes maculatus* Degeer in smoked catfish (*Clarias garipepinus*) during storage. *Bioscience Technology*, **85**:173-177.
- FAO Regional Office For Africa, 1990. **Artisanal Fish Containers in Ghana** RAFR/FI/ 90/1. Domak Press, Accra, Ghana. 33p
- Geden C J and Hogsette J A. 2001. Research and extension needs for integrated pest management for Arthropods of veterinary importance. Proceedings of a workshop in Lincoln, Nebraska, 12–14 April 1994, Center for medical, agricultural, and veterinary entomology USDA-ARS, Gainesville, Florida
- Honek A. 1993. Intraspecific variation in body size and fecundity of insects: a general relationship. *Oikos*, **67**: 483–492.
- Imai TI, Kodama H, Mori M and Kohno M. 1990. Morphological and chemical studies of male abdominal exocrine glands of the black larder beetle, *Dermestes ater* De Geer (Coleoptera: Dermestidae). *Applied Entomology and Zoology*, **25**:113–118
- Jansen D H and Nylin S. 1997. Seed-eaters versus Seed size, number, toxicity and dispersal. *Evolution*, **23**: 1–27.
- Jaskulska B, Rakowski G and Cymborowski B. 1987. The effect of juvenile hormone on aggregative behaviour of *Dermestes maculatus*. *Biochem Physiol.*, **87A**:771–773
- Johnson C and Kistler R A. 1987. Nutritional ecology of bruchid beetles. In: SlanskyF Jr. and Rodriguez JG (eds.), **Nutritional Ecology of Insects, Mites, Spiders and Related Invertebrates**, John Wiley, New York. pp. 259–276.
- Jones T M and Elgar M A. 2004. The role of male age, sperm age and mating history on fecundity and fertilization success in the hide beetle. *Pro. R. Soc. Lond.*, **271**: 1311-1318.
- Kidd S A, Eskenazi B and Wyrobek A J. 2001. Effects of male age on semen quality and fertility: A review of the literature. *Fertility and Sterility*, **75**: 237-248.
- Kreyenberg J. 1928. Biology of *Ladarius* and *Dermestes vulpinus*. *Z. Angew. Ent.*, **14**: 114-118.
- LaleN E S and Sastawa B M. 1996. The effects of sun drying on the infestation of the African catfish (*Clarias garipepinus*) by post harvest insects in the Lake Chad District of Nigeria. *Int J Pest Management*, **42**: 281-283.
- Lale N E S, Ajayi F A and Sastawa B M. 2000. Evaluation of processing methods and insectistatic essential oils for the control of skin beetles (*Dermestes maculatus*, DeGeer) infesting dried fish in the Lake Chad district of Nigeria. *Applied Tropical Agriculture*, **5**: 135-143.
- Linnie MJ and Keatinge MJ. 2000. Pest control in museums: toxicity of para-dichlorobenzene, 'vapona'™, and naphthalene against all stages in the life-cycle of museum pests. *Dermestes maculatus* Degeer, and *Anthrenus verbasci* (L.) (Coleoptera: Dermestidae). *Int Biodeter Biodegr.*, **45**:1–13

- Oakes C C, Smiragli D J, Plass C, Travler J M and Rofaire B. 2003. Aging results in hypermethylation of ribosomal DNA in sperm and liver of male rats. *Proc National Acad Sci., USA*, **100**: 278-282.
- Odeyemi O O, Owoade R A and Akinkulore O. 2000. Toxicity and population suppression effects of *Parkia clappatoniana* on dried fish pests (*Dermestes maculatus* and *Necrobia rufipes*). *Global J Pure and Applied Sci.*, **6**: 191-195.
- Osuji F N C. 1975. Some aspects of the biology of *Dermestes maculatus* DeGeer (Coleoptera, Dermestidae) in dried fish. *J Stored Product Res.*, **11**: 25-31.
- Rakowski G and Cymborowski B. 1986. Some environmental influences and physiological factors influencing the response of the hide beetle, *Dermestes maculatus*, to aggregation pheromone. *Inter J Invertebrate Reproductive Development*, **9**:35-41
- Rustin M H A and Murno D D. 1984. Popular ulcercia caused by *Dermestes maculatus* DeGeer. *Clin Meeting of the St John's Hospital Dermatological Society*: **9**: 317-321.
- Sahaf KA. 2007. Studies on *Dermestes maculatus* Degeer (Coleoptera: Dermestidae), a pest of stored silk cocoons of silkworm, *Bombyx mori* L. *J Entomological Res.*, **31**:163-164
- Samish M, Argaman Q and Perelman D. 1992. Research notes: The hide beetle, *Dermestes maculatus* Deg (Dermestidae), feeds on live turkey. *Poultry Sci.*, **71**: 388-390.
- Scoggin J K and Tauber O E. 1951. The bionomics of *Dermestes maculatus* Deg. Larval and pupal development at different moisture levels and on various media. *Annals of the Entomological Society of America*, **44**: 544-550.
- Seal D R and Tilton EW. 1985. Effects of gamma radiation on the metamorphic stages of *Dermestes maculatus* DeGeer (Coleoptera: Dermestidae). *Applied Radiation information*, **37**: 531-535.
- Siemens D H, Johnson C D and Woodman R L. 1991. Determinants of host range in bruchid beetles. *Ecology*, **72**: 1560-1566.
- Smith R H. 1986. Oviposition, competition and population dynamics in storage insects. Proceedings of the 4th International Working Conference on Stored-Product Protection, Tel Aviv, Israel, September, pp. 427-433.
- Stejskal V and Kucerova Z. 1996. The effect of grain size on the biology of *Sitophilus granaries* (L.) (Coleoptera: Curculionidae). I. Oviposition, distribution of eggs and adult emergence. *J Applied Entomology*, **120**: 143-146.
- Szczesny B, Hazra T K, PapaCastontinou J, Mitra S and Boldogh I. 2003. Age dependent deficiency in import of mitochondrial DNA glycolysis required for repair of oxidatively damaged bases. *Proc National Acad of Sci., USA*, **100**: 10670-10675.
- Taylor T A. 1964. Observations on the biology and habits of *Dermestes maculatus* DeGeer-a dried fish pest in Nigeria. *Nigerian Agriculture J.*, **1**:12-17.
- Zakka U, Ayertey JN and Cobblah MA. 2009. Suitability of four smoked fish species to *Dermestes maculatus* De Geer (Coleoptera: Dermestidae). *Nigerian J Entomology*, **26**:35-39.

# Composition and Larvicidal Activity of *Artemisia vulgaris* L. Stem Essential Oil Against *Aedes aegypti*

Sujatha Govindaraj<sup>1,2,\*</sup> and Bollipo D. RanjithaKumari<sup>2</sup>

<sup>1</sup>Department of Botany, Govt. Arts College for Women, Pudukkottai – 622 001,

<sup>2</sup>Department of Plant Science, Bharathidasan University, Tiruchirappalli – 620 024 Tamil Nadu. India.

Received: May 30, 2012, Accepted: August 17, 2012

## Abstract

This study investigates the composition and potential larvicidal activity of the essential oils isolated from the elite plant, *Artemisia vulgaris* L. which is commonly known as mugwort. Essential oils were hydrodistilled from *in vitro* raised stems and analysed by GC-MS. The major components of this oil were camphor, camphene,  $\alpha$ -thujone, 1,8-cineole,  $\gamma$ -muurolene and  $\beta$ -caryophyllene. Mosquito larvicidal assays were carried out to test the efficacy of the oil against the dengue vector, *Aedes aegypti*. Laboratory reared third instar larvae were exposed to different concentrations of the oil solution and activity was assessed at different exposure times according to standard WHO procedure. Results showed that 100% larval mortality was achieved when the larvae were treated with 500 ppm oil solution with an exposure time of 8 h. The present study has shown that mugwort oil is a potential larvicide against *Aedes aegypti* even in low doses of the test solution. The results indicate that the natural insecticides could be used in the place of synthetic insecticides and save our environment from chemical hazards.

**Keywords:** *Artemisia vulgaris*, stem essential oil, larvicidal, *Aedes aegypti*, exposure time.

## 1. Introduction

The mosquito species, *Aedes aegypti* L. is a vector of major diseases such as dengue haemorrhagic fever, chikungunya and yellow fever. *A. aegypti* is reported to infect more than hundred million people every year in more than 110 countries in the tropics (Halstead, 2000). The present resurgence of these diseases is due to the higher number of breeding places in today's throwaway society (Ravikiran *et al.*, 2006). Further, the indiscriminate use of synthetic insecticides is creating multifarious problems like environmental pollution, insecticide resistance, and toxic hazards to humans. Globally, there have been conscientious efforts to overcome these problems, and great emphasis has been placed recently on enviro-friendly and economically viable methodologies for pest control.

Phytochemicals obtained from the huge diversity of plant species are important source for safe and biodegradable chemicals, which can be screened for mosquito repellent, larvicidal, and insecticidal activities; and tested for mammalian toxicity. A large number of plant products have been reported to have mosquito larvicidal and/or repellent activity against adult mosquitoes. The discovery of insecticide activity in Asteraceae species has stimulated interest in this plant

family as part of the search for new plant derived insecticides (Rawls, 1986; Prashant *et al.*, 2006; Masotti *et al.*, 2012). In recent years, essential oils have received much attention as potent bioactive compounds against various mosquito species (Tripathi *et al.*, 2009).

*Artemisia vulgaris* L. (mugwort) is a member of the Asteraceae family. It is a tall (0.8-1.4 m), aromatic, threatened perennial herb distributed throughout the northern temperate regions of Africa, Asia, Europe, India, and North America. In traditional medicine, this plant is widely used for the treatment of diabetes and extracts of the whole plant is used for epilepsy and in combination for psychoneurosis, depression, irritability, insomnia and anxiety states (Lewis and Elwin-Lewis, 2003). Numerous medicinally active components of *A. vulgaris* have been identified, including coumarins, essential oils, flavonoids, polyacetylenes, sesquiterpene lactones, and sterols (USDA-ARS-NGRL, 2004). Essential oils make a major contribution to the plant's biological activity (Judzentiene and Buzelyte, 2006).

Mugwort essential oil is used in India for its insecticidal, antimicrobial and antiparasitical properties (Judzentiene and Buzelyte, 2006). It was reported to exhibit 90% repellence against *Aedes aegypti*, a mosquito that transmits yellow fever (Hwang *et al.*, 1985). Repellent and fumigant activity of *A. vulgaris* essential oil against *Musca domestica* L. and the stored-product

\* Corresponding author. e-mail: : sujathagovindaraj@gmail.com.

insect pest *Tribolium castaneum* (Herbst) were also reported (Judzentiene and Buzelyte, 2006; Wang *et al.*, 2006). The essential oil exhibits many other biological activities such as analgesic, anaesthetic, antidiarrheic, antineuralgic, antiseptic, antispasmodic, antiasthmatic, anti-inflammatory, sedative, CNS-stimulant, decongestant, expectorant, antiacne, larvicidal, nematocidal, pesticide, antibacterial, and it is also used in the flavour and perfumery industry (Teixiera da Silva, 2004). To date there are no published reports on larvicidal activity of *in vitro* grown stem essential oils of *A. vulgaris*. Therefore, we herein present a brief report on the composition of *A. vulgaris* stem essential oil and its larvicidal activity against *Aedes aegypti*.

## 2. Materials and Methods

### 2.1. Plant material and essential oil extraction

*A. vulgaris* seeds were collected from National Medicinal Plants Board, India and cultures were raised *in vitro* as previously reported (Sujatha and Ranjitha Kumari, 2007). Stem material was collected from *in vitro* raised plants before the onset of flowering and subjected to essential oil analyses. A 100 g sample of the air-dried stems was mixed with 1000 ml distilled water and subjected to hydrodistillation in a Clevenger-type distilling apparatus for 2 h. The resulting oil was dried over anhydrous sodium sulphate and stored in airtight fuscous glassware in a refrigerator at 4 °C until analysis.

### 2.2. Gas chromatography – mass spectrometry (GC-MS)

GC/EIMS analyses were performed with a Varian CP-3800 gas-chromatograph equipped with a DB-5 capillary column (30 m x 0.25 mm; coating thickness 0.25 µm) and a Varian Saturn 2000 ion trap mass detector. Analytical conditions: injector and transfer line temperatures at 220° and 240° C respectively; oven temperature was programmed from 60° C to 240° C at 3° C/min; carrier gas helium at 1 ml/min; injection of 0.2 µl (10% hexane solution); split ratio 1:30. Identification of the constituents was based on comparison of the retention times with those of authentic samples, comparing their Linear Retention Indices relative to the series of *n*-hydrocarbons, and by computer matching against commercial (NIST 98 and ADAMS 95) and home-made library mass spectra built up from pure substances and components of known essential oils and MS literature data (Stenhagen *et al.*, 1974; Jennings and Shibamoto, 1980; Adams, 1995). Moreover, the molecular weights of all the identified substances were confirmed by GC/CIMS, using MeOH and CI ionizing gas.

### 2.3. Procurement of eggs and rearing of *Aedes Aegypti* mosquito larvae

*Aedes aegypti* eggs (Fig. 1a) were obtained from a colony maintained at the Center for Medical Entomology, Indian Council for Medical Research, Madurai, Tamil Nadu, India. The eggs of *A. aegypti* were obtained as egg rafts on a filter paper. The egg rafts were brought to the laboratory and kept in a tray containing tap water (as culture medium) at laboratory conditions (29 ± 1°C; RH 70–75%; photoperiod 14 : 10 (light : dark)). On the next

day, the eggs were observed to hatch out into first instar larvae. Appropriate amounts of nutrients (yeast powder and dog biscuit (1:2 w/w) were added to the culture medium. On the third day after hatching, the first instar larvae moulted into second instar larvae. On the fifth day, third instar larvae were observed. The third instar larvae of *A. aegypti* were used for the larvicidal experiments in the present study (Fig. 1b).

### 2.4. Preparation of the oil solution

A sufficient amount of target (TR) oil was dissolved in tap water using 2 ml of 100 % acetone to produce a stock solution of 500 ppm. This solution was used to prepare other serial dilutions of target oil in concentrations of 500 – 1 ppm through dilution of stock with tap water. Five replicates of each concentration were made, in addition to five control replicates containing 0 ppm of oil, 2 ml of 100 % acetone and tap water (WHO, 1981).

### 2.5. Larvicidal bioassay

The tests were conducted at room temperature. The oil dilutions were tested against the third instar larvae of *A. aegypti* mosquitoes (WHO [World Health Organization], 1981; Ansari *et al.*, 2000; Rey *et al.*, 2001; Amer and Mehlhorn, 2006) to detect their toxicity on mosquito larvae. Five replicates of each oil dilution (1-500 ppm) were prepared. Each replicate containing 200 ml of the described oil solution was placed in a 500 ml glass beaker. Ten third-instar larvae of target mosquito were transferred into each beaker (Mohtar *et al.*, 1999). After that, the beakers were left on the laboratory table for 24 h. The number of dead larvae in each beaker was counted after 1, 8, 16, 20 and 24 h.

### 2.6. Statistical analysis

Experiments were set up in a complete randomized block design and each experiment was repeated five times. Data were recorded on the percentage of larval mortality. The analysis of variance (ANOVA) appropriate for the design was carried out to detect the significance of differences among the treatment means, and the treatment means were compared using Duncan's Multiple Range Test (DMRT) at a 5 % level of significance (Gomez and Gomez, 1976). The results were analyzed statistically using SPSS version 12.0 (SPSS Inc., Chicago, IL, USA) for windows.

## 3. Results and Discussion

### 3.1. Essential oil analyses

The oil was a fluid liquid, slightly greenish in colour and possessed powerful fresh-camphoraceous aroma with sweet bitter taste. The intense odour could be due to the presence of a large number of oxygenated mono- and sesquiterpene components in the stem oil. Fifty one compounds were identified, accounting for 91.0% of the essential oil. The identified constituents and corresponding percentage of the essential oil composition are listed in Table (1).

**Table 1.** Composition of *in vitro* stem essential oils of *A. vulgaris*

Compound	Rt	% MS	RI <sup>#</sup>	identification <sup>o</sup>
cis-salvene	3.39	trace	846	MS, RI
santolinatriene	4.44	0.1	906	MS, RI
tricyclene	4.86	0.2	920	MS, RI
artemisiatriene	5.17	0.8	930	MS, RI
alpha-fenchene	5.57	3.9	943	MS, RI
camphene	5.61	6.0	944	MS, RI, co-GC
sabinene	6.37	2.4	969	MS, RI, co-GC
beta-pinene	6.47	0.8	973	MS, RI, co-GC
2,3-dehydro-1,8-cineole	6.94	0.1	988	MS, RI
2-carene	7.92	0.1	1012	MS, RI
p-cymene	8.28	0.8	1020	MS, RI, co-GC
1,8-cineole	8.58	5.1	1026	MS, RI, co-GC
arbusculone	10.24	0.4	1061	MS, RI
alpha-thujone	12.52	10.7	1105	MS, RI, co-GC
beta-thujone	13.04	2.8	1114	MS, RI, co-GC
camphor	14.88	17.3	1140	MS, RI, co-GC
isoborneol	15.39	trace	1147	MS, RI, co-GC
borneol	16.16	3.7	1158	MS, RI, co-GC
3-thujanol	16.80	0.6	1167	MS, RI
myrtenal	18.00	0.6	1185	MS, RI
myrtenol	18.10	trace	1186	MS, RI
verbenone	18.85	trace	1197	MS, RI
trans-carveol	19.66	trace	1210	MS, RI
cumin aldehyde	21.05	0.2	1232	MS, RI
perilla aldehyde	23.55	0.2	1272	MS, RI
bornyl acetate	24.58	0.6	1288	MS, RI
iso-3-thujyl acetate	25.38	0.8	1301	MS, RI
delta-elemene	28.35	0.4	1334	MS, RI
alpha-copaene	31.26	0.8	1366	MS, RI
beta-cubebene	32.46	0.6	1379	MS, RI
beta-elemene	32.64	0.3	1381	MS, RI
alpha-gurjunene	33.83	1.3	1394	MS, RI
beta-caryophyllene	34.71	5.8	1405	MS, RI
beta-gurjunene	35.30	0.3	1413	MS, RI
alpha-humulene	37.17	1.9	1438	MS, RI
allo-aromadendrene	37.64	0.3	1444	MS, RI
gamma-murolene	39.58	9.0	1469	MS, RI
bicyclogermacrene	40.58	1.6	1482	MS, RI
alpha-murolene	41.00	0.6	1488	MS, RI
gamma-cadinene	41.90	0.6	1499	MS, RI
davana ether isomer *	42.42	0.4	1506	MS, RI
delta-cadinene	42.85	2.5	1512	MS, RI
davana ether isomer *	43.87	0.2	1525	MS, RI
germacrene D-4-ol	46.60	trace	1560	MS, RI
spathulenol	46.65	1.1	1561	MS, RI
caryophyllene oxide	46.88	1.2	1564	MS, RI
cis-davanone	47.94	2.5	1577	MS, RI, co-GC
cadinol-epi-alpha	51.32	0.4	1623	MS, RI
alpha-cadinol	52.29	0.5	1637	MS, RI
davanone-2-ol-beta	57.50	0.4	1712	MS, RI
cadinene-14-hydroxy-delta	62.67	0.1	1789	MS, RI

Rt = Retention time expressed in minutes; Trace: <0.05; # the retention index was calculated using a homologous series of n-

alkanes C8-C18; \* correct isomer not determined; <sup>o</sup>Co-GC: co-injection with an authentic sample.

There are no previous reports on the essential oil composition of *in vitro* raised *A. vulgaris* stems. Monoterpenes reached about 70% of the whole essential oil. The main ones were camphene (6.0%) and  $\alpha$ -fenchene (3.9%). Oxygenated monoterpenes constituted the main chemical class of the oil (51.3%) and they were represented with camphor (17.3%) and  $\alpha$ -thujone (10.7%) as principal chemicals. Sesquiterpenes constituted about 26% of the whole oil. Among them, hydrocarbons (27 compounds) reached 21.5%, whereas oxygenated derivatives (12 compounds) represented 4.4% of the oil. Among sesquiterpene hydrocarbons,  $\beta$ -caryophyllene (5.8%) was the main constituent. In the case of oxygenated sesquiterpenes, the principal one was *cis*-davanone (2.5%). Thus, the isolated essential oil was characterized by a high content of oxygenated monoterpenes [camphor (17.3 %),  $\alpha$ -thujone (10.7 %), and 1,8-cineole (5.1%)]; the monoterpene hydrocarbon camphene (6.0%); and by the sesquiterpene derivatives,  $\beta$ -caryophyllene (5.8%) and  $\gamma$ -murolene (9.0%).

It has been demonstrated that *A. vulgaris* grown in different countries possessed qualitative and quantitative differences in their essential oil composition. The oil from Italy were rich in camphor (47.7%), camphene (9.1%) and verbenone (8.6%) (Mucciarely *et al.*, 1995). The oil from the Republic of Bashkortostan was found to contain high amounts of  $\alpha$ -pinene (53.7 %), trans-chrysanthenol (13.1%),  $\beta$ -myrcene (8.8%) and  $\beta$ -pinene (7.4%) (Khalilov *et al.*, 2001). Whereas in Croatia, the chief components reported were  $\beta$ -thujone (20.8%),  $\alpha$ -pinene (15.1%) and 1,8-cineole (11.7%) (Jerkovic *et al.*, 2003). The oil isolated from North Lithuania was high in amounts of sabinene,  $\beta$ -pinene, 1,8-cineole, artemisia ketone, *cis*- and *trans*- thujone, chrysanthenyl acetate, germacrene D, and  $\beta$ -caryophyllene (Judzentiene and Buzelyte, 2006). The oil isolated from Indian grown plants was characterized with high amount of camphor (38.7%), isoborneol (8.2%) and artemisia alcohol (4.5%) (Haider *et al.*, 2003).  $\alpha$ -thujone was stated as the main constituents of *A. vulgaris* oil (Misra and Singh, 1986), while we found an average value of 10.7% in this study. Thus the oils isolated from plants native to different countries significantly differ in their composition. This clearly depicts that variations in population genetics, environmental conditions, and the stress factors the plant faces during its survival and growth influence the accumulation of essential oils.

### 3.2. Larvicidal activity of stem essential oil

Different concentrations (1-500 ppm) of stem essential oil solutions were bioassayed against the third instar larvae of *Aedes aegypti*. The results were recorded after 1, 8, 16, 20 and 24 h of treatment (Table 2). In control treatments, no larvicidal effect was observed; the larvae remained alive, and they moulted into fourth instar larvae. Whereas, when the different oil concentrations were tested, different mortality rates were recorded with respect to exposure time.

At 1 ppm oil solution, the larvae remained immobile after 24 h of treatment. When 10 ppm oil solution was

tested, 5.0 % and 12.3 % larval mortality was recorded after 20 h and 24 h of treatment respectively. When 100 ppm oil solution was tested, 78.2 % mortality was recorded after 24 h (Table 2).

**Table 2.** Effect of different concentrations of oil solution and exposure time on larvicidal bioassay of third instar larvae of *Aedes aegypti*

Oil Solution (ppm)	Larval Mortality Rate (%) After				
	1 h	8 h	16 h	20 h	24 h
Control (0)	-	-	-	-	-
1	-	-	-	-	-
10	-	-	-	5	12.3 d
100	10.4 e	45.9 e	64.8 d	70.1 c	78.2 c
200	20.1 d	52.7 d	72.9 c	85.3 b	91.5 b
300	59.4 c	71.2 c	89.5 b	100 a	100 a
400	65.1 b	83.2 b	100 a	100 a	100 a
500	89.7 a	100 a	100 a	100 a	100 a

\* Treatment means followed by different letters are significantly different from each other ( $p < 0.05$ ) according to Duncan's Multiple Range Test

At 300 ppm, complete mortality was recorded after 20 h of exposure, and at 400 ppm, 100 % mortality was recorded after 16 h (Table 2). The maximum result (100 %) was recorded with 500 ppm oil concentration after 8 h exposure (Fig. 1c). Lethal concentration (50 % larvicidal activity) was observed from 100 ppm oil concentration onwards. The exposure time is very important for 50 % larvicidal activity of the oil solution. Very low concentrations of the oil led to high mortality rates.



**Figure 1.** Larvicidal Activity of *Artemisia vulgaris* stems essential oil solution. a – Eggs of *Aedes aegypti* b – Different stages of larval development (1, 2, 3 & 4 instars) c – Treated larvae (at 500 ppm test solution) exhibiting complete mortality (after 8 h) exposure time on larvicidal activity was also previously reported (Amer and Mehlhorn, 2006).

The present study has shown that *in vitro* produced *A. vulgaris* stem essential oil is a potential larvicide against *Aedes aegypti* in low concentrations (100 ppm) of the oil solution. High doses of the oil solution will be required for large breeding habitats, to be effective. However, the oil might be used as a selective larvicide in small breeding places where water is stagnant, such as in domestic containers and desert coolers. *A. vulgaris* essential oils were previously reported as potent larvicidals against *Aedes aegypti* (Ram and Mehrotra, 1995). But this is the first larvicidal bioassay carried out using essential oil extracted from *in vitro* propagated stems. *In vitro* production of source material helps in conserving the wild resources and thus this protocol can be used for the steady production of *A. vulgaris* plants for essential oil production.

This study indicates that the essential oil of *in vitro* propagated *A. vulgaris* stems has larvicidal properties and its use as a larvicide against mosquitoes should be explored. It is worthwhile to study extensively the larvicidal properties of the plant's essential oil by isolating and identifying the active components responsible for larval mortality, and then test them in field trials in order to assess their potential as an alternative to synthetic chemical larvicides.

## References

- Adams RP. 1995. **Identification of Essential Oil Components by Gas Chromatography-Mass Spectroscopy**. Allured Publishing Corporation, Carol Stream, Illinois.
- Amer A and Mehlhorn H. 2006. Larvicidal effects of various essential oils against *Aedes*, *Anopheles*, and *Culex* larvae (Diptera, Culicidae). *Parasitol Res.*, **99**:466-472.
- Ansari MA, Vasudevan P, Tandon M, and Razdan RK. 2000. Larvicidal and mosquito repellent action of peppermint (*Mentha piperita*) oil. *Biores Technol.*, **71**:261-271.
- Dwivedi PD, Naqvi AA, and Bagchi GD. 2003. Essential oil composition of *Artemisia vulgaris* harvested at different growth periods under Indo-Gangetic plain conditions. *J Essen Oil Res.*, **15**(6):376-378.
- Gomez KA and Gomez KA. 1976. **Statistical Procedures for Agricultural Research with Emphasis on Rice**. Philippines International Rice Research Institute, Los Bans. Haider F.
- Halstead SB. 2000. Global perspective on Dengue Research. *Dengue Bull.*, **24**:77-82.
- Hwang YS, Wu KH, Kumamoto J, Axelrod H and Mulla MS. 1985. Isolation and identification of mosquito repellents in *Artemisia vulgaris*. *J Chem Ecol.*, **11**:1297-1306.
- Jennings W and Shibamoto T. 1980. **Qualitative Analysis of Flavor and Fragrance Volatiles by Glass Capillary Chromatography**. Academic Press, New York.
- Jerkovic I, Mastelic J, Milos M, Jutcau F, Masotti V and Viano J. 2003. Chemical variability of *Artemisia vulgaris* L. essential oils originated from the Mediterranean area of France and Croatia. *Flavour Fragr J.*, **18**:436-40.
- Judzentiene A and Buzelyte J. 2006. Chemical composition of essential oils of *Artemisia vulgaris* L. (mugwort) from plants grown in North Lithuania. *Chemija*, **17**(1):12-15.
- Khalilov L, Paramonov EA, Khalilova AZ, Odinkov VN, Muldashev AA, Baltaev UA and Dzehemilev UM. 2001. Identification and biological activity of volatile organic

- compounds emitted by plants and insects. IV- Composition of vapor isolated from certain species of *Artemisia* plants. *Chem Nat Compd.*, **37(4)**:339-342.
- Lewis WH and Elvin-Lewis MPF. 2003. **Medical Botany: Plants Affecting Human Health**, 2nd ed., John Wiley and Sons Inc., New Jersey, pp 345.
- Masotti V, De Dong L, Moreau X, Rabier J, Laffont-Schwob I and Thiery A. 2012. Larvicidal activity of extracts from *Artemisia* species against *Culex pipens* L. mosquito: Comparing endemic versus ubiquitous species for effectiveness. *C R Biol.*, **335(1)**:19-25.
- Misra LN and Singh SP. 1986.  $\alpha$ -Thujone, the major component of the essential oil from *Artemisia vulgaris* growing wild in Nilgiri hills. *J Nat Prod.*, **49(5)**: 941.
- Mohtar M, Yarmo MA and Kadri A. 1999. The effects of *Nerium indicum* leaf extract on *Aedes aegypti* larvae. *J Trop For Prod.*, **5(1)**:87-92.
- Mucciarely M, Caramiello R and Maffei M. 1995. Essential oils from some *Artemisia* species growing spontaneously in northwest Italy. *Flavour Fragr J.*, **10**:25-32.
- Prashant RV, Subburaju T and Balakrishnan N. 2006. Larvicidal activity of *Artemisia nilagirica* (Clarke) Pamp. and *Ocimum sanctum* L.: A preliminary study. *J Nat Med.*, **6(2)**:157-161.
- Ram PR and Mehrotra MN. 1995. **Compendium of Indian Medicinal Plants**. Publication and Information Directorate, CSIR, New Delhi. 4 (1985-1989): pp 74.
- Ravikiran S, Bhavani K, Sita Devi P, Rajeswara Rao BR, and Janardhan Reddy K. 2006. Composition and larvicidal activity of leaves and stem essential oils of *Chloroxylon swietenia* DC against *Aedes aegypti* and *Anopheles stephensi*. *Biores Technol.*, **97**:2481-2484.
- Rawls RL. 1986. Experts probe issues, chemistry of light activated pesticides. *Chem Eng News*, **22**:2124.
- Rey D, David JP, Besnard G, Jullien JL, Lagnean C and Meyran JC. 2001. Comparative sensitivity of larval mosquitoes to vegetable polyphenols versus conventional insecticides. *Entomol Exp Appl.*, **98**:361-367.
- Sujatha G and Ranjitha Kumari BD. 2007. Effect of phytohormones on micropropagation of *Artemisia vulgaris* L. *Acta Physiol Plant*, **29(3)**:189-195.
- Stenhagen E, Abrahamsson S, and McLafferty FW. 1974. **Registry of Mass Spectral Data**. John Wiley and Sons, New York.
- Teixiera da Silva JA. 2004. Mining the essential oils of the Anthemideae. *Afr J Biotechnol.*, **3(12)**:706-720.
- Tripathi AK, Upadhyay S, Bhuiyan M, and Bhattacharya PR. 2009. A review on prospects of essential oils as biopesticide in insect-pest management. *J Pharmacognosy and Phytother.*, **1(5)**:52-63.
- USDA-ARS-NGR. 2004. Dr. Duke's Phytochemical and Ethnobotanical Databases <http://www.ars-grin.gov/cgi-bin/duke/farmacy2.pl> Accessed June 21, 2012; last updated 10 March 1998.
- Wang J, Zhu F, Zhou XM, Niu CY and Lei CL. 2006. Repellent and fumigant activity of essential oil from *Artemisia vulgaris* to *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *J Stored Prod Res.*, **42**:339-347.
- WHO. 1981. Instructions for determining the susceptibility or resistance of mosquito larvae to insecticides. WHO/VBC/81, 807. World Health Organization, Geneva.



# Susceptibility of the Hymenopteran Parasitoid, *Habrobracon hebetor* (Say) (Braconidae) to the Entomopathogenic Fungi *Beauveria bassiana* Vuillemin and *Metarhizium anisopliae* Sorokin

Vahid Mahdavi<sup>1,\*</sup>, Moosa Saber<sup>1</sup>, Hooshang Rafiee-Dastjerdi<sup>2</sup> and Ali Mehrvar<sup>3</sup>

<sup>1</sup>Department of Plant Protection, College of Agriculture, University of Maragheh, Maragheh

<sup>2</sup>Department of Plant Protection, College of Agriculture, University of Mohaghegh Ardabili, Ardabil

<sup>3</sup>Department of Plant Protection, College of Agriculture, Azarbaijan Shahid Madani University, East-Azarbaijan, Tabriz, Iran.

Received: August 3, 2012; Accepted: September 15, 2012

## Abstract

*Habrobracon hebetor* (Say) is an ectoparasitoid and has been studied as a biocontrol agent of various lepidopteran pests such as cotton bollworm *Helicoverpa armigera* (Hübner). With regards to the negative effects of common pesticides used in cotton fields on the parasitoids, in this study the effects of different isolates of entomopathogenic fungi (*Beauveria bassiana* and *Metarhizium anisopliae*) were evaluated on *H. hebetor*. Bioassay experiments were performed by the immersion method. For each of the treatments 15 immature individuals of the parasitoid were used. After recording the results, data were analyzed using SAS software. Bioassay results of fungi isolates on larval stages showed that the value of LC<sub>50</sub> for IRAN187C isolate of *B. bassiana* was  $1.46 \times 10^9$  conidia/ml. Because of the low mortality caused by the other isolates, the value of LC<sub>50</sub> was not set for them. Also, bioassay of fungal isolates showed that none of the isolates had any effect on the parasitoids pupal stage. According to the obtained results, it can be concluded that various fungal isolates of *B. bassiana* and *M. anisopliae* had little adverse impact on the parasitoid wasp, thus after doing field tests, the microbial control agents may be used along with these parasitoids in integrated pest management programs (IPM) in cotton.

**Keywords:** *Habrobracon hebetor*, entomopathogenic fungi, IPM, bioassay, natural enemies..

## 1. Introduction

Cotton (*Gossypium hirsutum*) is the most important economic and fiber crop worldwide (Chen *et al.*, 2002). This crop is also a major agricultural product in Iran and the area cultivated with cotton is about 91019 hectare for the years 2009-2010 (Anonymous, 2011). Insect pests are limiting factors for healthy growth of cultivated plants (Ramzan Asi *et al.*, 2009). Among insect pests, *Helicoverpa armigera* (Hübner) (Lep.: Noctuidae) is one of the most important arthropod pests of cotton crop (Matthews, 1999). This pest is a polyphagous agricultural pest which attacks a wide variety of agricultural crops including cotton, corn, tomatoes, sorghum, soybeans and groundnuts (Fitt, 1989). Early instars are foliar feeders and later instars attack seeds, fruits and bolls leading to economic loss (Fitt, 1989) and their infestations cause severe economic losses as a result of crop yield reduction (Soomro *et al.*, 1992), and the pest causes economic losses up to 30% of the total production (Yazdanpanah *et al.*,

2009). Different strategies have been employed for control of this notorious pest (Ramzan Asi *et al.*, 2009). Farmers mostly prefer chemical pesticide application for its control because it is quicker, however, indiscriminate application of broad spectrum chemical pesticides exterminates these susceptible natural enemies and leaves behind the pests that are more resistant to pesticides (Feng *et al.*, 1994) as well as these compounds can cause serious problems such as pest outbreaks (Luck *et al.*, 1977; Metcalf, 1986). In order to reduce crop losses, the use of microbial control agents which have a lower risk on the environment and humans is recommended (Hull and Beers, 1985). Among the microbial control agents are entomopathogenic fungi. Entomopathogenic fungi have a considerable potential for efficacious suppression of a variety of arthropod pests. *Beauveria bassiana* (Balsamo) Vuillemin is one of the most important entomopathogenic fungi (Leland *et al.*, 2005; Quesada-Moraga *et al.*, 2006; Al-maza *et al.*, 2006). This fungus is widely distributed in the world (St.-Leger *et al.*, 1986) and has the potential to control over 70 insect pest species (Hung and Boucias, 1992; Alizadeh *et al.*, 2007).

\* Corresponding author. e-mail: v.mahdavi@ymail.com

Another fungus effective in controlling insect pests is *Metarhizium anisopliae* Sorokin that is able to control a wide range of pests (Zimmermann, 1993).

On the other hand, one of the important methods to control pests is the use of natural enemies. Among these natural enemies is the parasitoid wasp *Habrobracon hebetor* (Say) (= *Bracon hebetor*) (Haeselbarth, 1983; Amir-Maafi and Chi, 2006) (Hymenoptera: Braconidae). *H. hebetor* is a valuable biocontrol agent of lepidopteran pests attacking crop plants and stored products, including *H. armigera* (Magro and Para, 2001). In Iran, mass rearing of *H. hebetor* is done on Mediterranean flour moth, *Ephestia (Anagasta) kuehniella* Zeller (Mudd and Corbet, 1982) and the adult wasps are released to parasitize *H. armigera* larvae in cotton fields in Ardabil and Golestan provinces in the northern parts of the country (Attaran, 1996; Navaei *et al.*, 2002).

Since the strategy of IPM includes the simultaneous use of different methods of control, different methods of control must be examined together to finally be able to utilize them for pest control. Potential effects of microbial control agents on the parasitoids must be studied (Hajek and St. Leger, 1994). In particular, *Metarhizium anisopliae* Sorokin and *Beauveria bassiana* (Balsamo) Vuillemin have been isolated from diverse species of parasitoids (Thungrabeab and Tongma, 2007).

In the present study, the effects of different isolates of the entomopathogenic fungi *B. bassiana* and *M. anisopliae* were evaluated on the immature stages of *H. hebetor* in the laboratory, in order to evaluate the possibility of simultaneous application of entomopathogenic fungi and the parasitoid *H. hebetor* in the field.

## 2. Materials and Methods

### 2.1. Insect Rearing

The *H. hebetor* colony was obtained from an insectarium maintained by the Plant Protection Bureau of Kaleibar, Iran in 2010. The colony was maintained in the laboratory at  $26 \pm 1^\circ\text{C}$ ,  $60 \pm 5\%$  RH and a photoperiod of 16:8 (L: D) on larval *E. kuehniella*, that was reared on flour in a growth chamber at the above mentioned environmental conditions. Parasitoid wasps were reared on 5<sup>th</sup> instar larvae of *E. kuehniella* for five generations and used for all experiments. Honey was provided as food for the adult parasitoids on  $5 \times 30$  mm strips of paper (Sarmadi, 2008).

### 2.2. Fungal Isolates

Fungi isolates used in this study are shown in Table 1.

**Table 1.** Isolates of fungi used in this study and origin within Iran

Fungi	Isolates	Host	Location area
<i>Beauveria bassiana</i>	IRAN 187C	<i>Leptinotarsa decemlineata</i>	Ardabil
<i>Beauveria bassiana</i>	EUT116	Lepidopteran larvae	Tehran
<i>Beauveria bassiana</i>	EUT105	Soil	Fashand-Karaj
<i>Metarhizium anisopliae</i>	M-115	<i>Parandra caspica</i>	Sari
<i>Metarhizium anisopliae</i>	M-396	<i>Parandra caspica</i>	Sari

### 2.3. Culture of Fungi

Fungi were cultured on Saboraud's Dextrose Agar Yeast extract (SDAY) in Petri dishes at  $25 \pm 1^\circ\text{C}$ ,  $80 \pm 5\%$  RH and a photoperiod 16:8 h (L:D). After preparing the medium, piece of the culture medium containing conidial fungi to be removed by a sterile scalpel were transferred to Petri dishes containing fresh medium. After 15 days the stages were full of germinated fungi. Petri dishes containing conidial of entomopathogenic fungi were used for experiments. Since strains maintained in the laboratory, after preparing them for about 2-3 weeks.

### 2.4. Production of Suspension

For producing fungi suspension, conidia were transferred into tubes with lid consist of sterile distilled water. For screening mycelium and medium, this suspension was passed through mesh fabric. A haemocytometer (Paul Marienfeld GmbH and Co. KG, Germany) was used to determine the concentration of conidia in the initial suspension. The hemocytometer is a device originally designed for the counting of blood cells. It is now also used to count other types of cells as well as other microscopic particles for example the entomopathogenic fungi conidia. After counting conidia using haemocytometer, the main concentration was determined using the formula  $Y = 5X \times 10^4$  ( $X$  = number of conidia in five squares) (Erwin, 2002). Subsequent concentrations were determined using the logarithmic distant.

### 2.5. Bioassays

The immature stages of the parasitoid were dipped in fungi solutions at the 4<sup>th</sup> or 8<sup>th</sup> day for 10 s. These days correspond to larval (without cocoon) and pupal (with cocoon) stages of the parasitoid, respectively (Rafiee-Dastjerdi *et al.*, 2008). Initial dose-setting experiments were carried out to determine the highest and lowest concentrations causing 80% and 20% mortality for both isolates (Robertson *et al.*, 2007). Concentration ranges were  $10^6$ ,  $10^7$ ,  $10^8$ ,  $10^9$ ,  $10^{10}$  and  $10^{11}$  conidia/ml. Tween 80 (Merck, Darmstadt, Germany) was used at a concentration of 200 ppm in all dilutions as a spreader (Rosenheim and Hoy, 1988). Our previous experiments showed that Tween 80 (200 ppm) has no effects on bioassays. The control plates were sprayed with distilled water plus Tween 80. After immersion, Petri dishes containing filter paper and immature stages were transferred to growth chamber with  $26 \pm 1^\circ\text{C}$  and 80% RH. Data analysis was performed by SAS program (SAS Institute, 2002).

## 3. Results and Discussion

Effects of *B. bassiana* and *M. anisopliae* isolates showed that the isolates EUT105 (*B. bassiana*) and M-396 (*M. anisopliae*) didn't cause mortality in the *H. hebetor* larval stage at all concentrations tested. The EUT116 and IRAN187C isolates of *B. bassiana* at the concentration  $10^{10}$  conidia/ml had 6.67 and 51.11% mortality on parasitoid larvae, respectively. Also, M-115 isolates of *M. anisopliae* at  $10^{10}$  conidia/ml/ml caused 2.22% mortality on larval stage of *H. hebetor* (Table 3). The above results showed that,  $LC_{50}$  values didn't apply for the listed

isolates. The only isolate for which the value of  $LC_{50}$  was determined was IRAN187C isolate of *B. bassiana* (Table 2).

**Table 2.** Probit analysis of the fungal isolates IRAN187C (*B. bassiana*) tested on parasitoid larval stage

Isolate	*N	Slope ± SE	<sup>2</sup> χ	LC <sub>50</sub> Conidia/ml	**FL (95%)
IRAN187C	270	0.6	3.4	$1.46 \times 10^9$	$(6.5 \times 10^8$ $- 3.2 \times 10^9)$
<i>B. bassiana</i>		±0.08			

\* Number of used insect

\*\* Fiducial limits

According to our results, the isolate of IRAN187C (*B. bassiana*) had an adverse influence on the larval stage of the parasitoid followed by EUT116 (*B. bassiana*) and M-115 (*M. anisopliae*). Different isolates of *Beauveria* and *Metarhizium* did not show any effect on the pupal stage of the parasitoid (Table 3). Probably, the cocoon around the parasitoid pupa was responsible for the lack of effectiveness of the fungal treatments on this developmental stage. The results showed that in the control treatment (normal conditions) in the larval and pupal stages, no losses were observed (Table 3).

**Table 3.** Mortality (± SE) of immature stages parasitoid treated with concentration of  $10^{10}$  conidia/ml of fungal isolates tested and control treatment

Isolates of fungi	Stages of parasitoid	
	Larval	Pupal
IRAN187C ( <i>B. bassiana</i> )	51.11 ± 4.01	0 ± 0
EUT116 ( <i>B. bassiana</i> )	6.67 ± 3.85	0 ± 0
EUT105 ( <i>B. bassiana</i> )	0 ± 0	0 ± 0
M-115 ( <i>M. anisopliae</i> )	2.22 ± 2.22	0 ± 0
M-396 ( <i>M. anisopliae</i> )	0 ± 0	0 ± 0
Control	0 ± 0	0 ± 0

Means in column followed by different small letters are significantly different. ANOVA with Tukey post hoc test ( $\alpha < 0.05$ )

Before this study, no investigation has been conducted on the impact of entomopathogenic fungi on *H. hebetor*, but the impact of *B. bassiana* and *M. anisopliae* on other parasitoids was studied. Rashki *et al.* (2009), in studying the effect of *B. bassiana* on *Aphidius matricariae* and its host *Myzus persicae*, showed that this pathogen had no effect on biological parameters of the parasitoid and concluded that *B. bassiana* and the parasitoid *A. matricariae* can be successfully combined for biological control of *M. persicae*. These reports are in line with the results of this study. The results of this study indicate very little effect of the entomopathogenic fungi on the parasitoid which is consistent with the results of Stolz *et al.* (2002). In evaluating the susceptibility of the parasitoids *Apoanagyrus lopezi* and *Phanerotoma sp.* to the entomopathogenic fungus *M. anisopliae*, they reported that different isolates of this fungus had very little risk on parasitoids. Also, Rosa *et al.* (2000) studying the effect of *Beauveria* and *Metarhizium* on the parasitoid *Prorops nasuta* reported that various isolates of the fungus have little negative impact on the parasitoid and can be used as a component compatible with natural enemies. Also, the

effects of fungal isolates on the host field (*Helicoverpa armigera* Hübner) showed that mentioned isolates had a good control on the *H. armigera* (Vojoudi, 2011). According to the obtained results, it can be concluded that different isolates of the fungi *B. bassiana* and *M. anisopliae* had few adverse effects on *H. hebetor*, and therefore these microbial control agents can be used of along with the parasitoid in integrated pest management (IPM) programs.

## Acknowledgements

We thank Mr. David Hill from Toronto (Canada) for proofreading the manuscript and adding valuable comments. This work received financial support from the Postgraduate Education Bureau of the University of Maragheh which is greatly appreciated.

## References

- Alizadeh A, Samih MA, Khezri M and Saberi-Riseh R. 2007. Compatibility of *Beauveria bassiana* (Bals.) Vuill. with several pesticides. *Int J Agr Biol.*, **9**: 1-4.
- Al-Maza MS, Ship LB, Roadbent B and Kevan P. 2006. Biological control of *Lygus lineolaris* (Hemiptera: Miridae) and *Frankliniella occidentalis* (Thysanoptera: Thripidae) by *Bombus impatiens* (Hymenoptera: Apidae) vectored *Beauveria bassiana* in greenhouse sweet pepper. *Biological Control*, **37**: 89-97.
- Amir-Maafi M and Chi H. 2006. Demography of *Habrobracon hebetor* (Hymenoptera: Braconidae) on two Pyralid hosts (Lepidoptera: Pyralidae). *Ann Entomol Soc Am.*, **99**: 84-90.
- Anonymous. 2011. Statistics of Agriculture, Volume 1, Agricultural and Horticultural Crops, Crop year 2009-2010, Ministry of Agriculture, Department of Economic Planning, Bureau of Statistics and Information Technology.
- Attaran MR. 1996. Effects of laboratory hosts on biological attributes of parasitoid wasp *Bracon hebetor* Say. M.Sc. Thesis. Tarbiat Modarres University, Tehran, Iran.
- Chen WX, Xiao GF and Zhu Z. 2002. Obtaining high pest-resistant transgenic upland cotton cultivars carrying cry1Ac3 gene driven by chimeric OM promoter. *Acta Bot Sinica*, **44**: 963-970.
- Erwin DC. 2002. Procedure for estimating concentration of spore suspension with Levi-Hausser hemocytometer. *Plant Pathol.*, **84**: 202-209.
- Feng MG, Poprawski TJ and Khachatourians GG. 1994. Production, formulation and application of the entomopathogenic fungus *Beauveria bassiana* for insect control: current status. *Biocontrol Sci Technol.*, **4**(1): 3-34.
- Fitt GP. 1989. The ecology of *Heliothis* in relation to agroecosystems. *Ann. Rev. Entomol.*, **34**: 17-52.
- Hajek AE and St-Leger RJ. 1994. Interaction between fungal pathogens and insect hosts. *Ann. Rev. Entomol.*, **39**: 293-322.
- Haeselbarth E. 1983. Determination list of entomophagous insects. Nr. 9. Bulletin, Section Re'gionale Ouest Palaearctique, *Organisation Internationale de Lutte Biologique*, **6**(1): 22-23.
- Hull LA and Beers EH. 1985. Ecological sensitivity modifying chemical control practices to preserve natural enemies. In: **Biological Pest Control in Agricultural Ecosystem**. Academic Press, Orlando, Florida., pp:103-121.
- Hung SY and Boucias DG. 1992. Influence of *Beauveria bassiana* on the cellular defense response of the beet armyworm, *Spodoptera exigua*. *J Invertebr Pathol.*, **60**: 152-158.

- Leland JE, McGuire MR, Grace JA, Jaronski ST, Ulloa M, Park YH and Plattner RD. 2005. Strain selection of a fungal entomopathogen *Beauveria bassiana* for control of plant bugs (Lygus spp.) (Heteroptera: Miridae). *Biological Control*, **35**: 104-114.
- Luck RF, Van den Bosch R and Garcia R. 1977. Chemical insect control a trouble pest management strategy. *Biosci.*, **27**: 606-611.
- Magro SR and Parra JRP. 2001. Biología do ectoparasitoide *Bracon hebetor* Say, 1857 (Hymenoptera: Braconidae) em sete especies de lepidopteros. *Sci Agr.*, **58**: 693-698.
- Matthews M. 1999. **Heliothine Moths of Australia**. CSIRO Publishing, 320 pp.
- Metcalf RL. 1986. The ecology of insecticides and chemical control of insects, In: **Ecological Theory and Integrated Pest Management Practice**. Kogan M. Ed., Wiley, New York. pp. 251-297.
- Mudd A and Corbet SA. 1982. Response of the ichneumonid parasite *Nemeritis canescens* to Kairomones from the flour moth, *Ephesia kuehniella*. *J. Chem. Ecol.*, **8(5)**: 843-850.
- Navaei AN, Taghizadeh M, Javanmoghaddam H, Oskoo T and Attaran MR 2002. Efficiency of parasitoid wasps, *Trichogramma pintoii* and *Habrobracon hebetor* against *Ostrinia nubilalis* and *Helicoverpa* sp. on maize in Moghan. Proceedings of 15th Iranian Plant Protection Congress, Vol. I, Pests, 193.
- Quesada-Moraga E, Maranhao EAA, Valverde-García P and Santiago-Alvarez C. 2006. Selection of *Beauveria bassiana* isolates for control of the whiteflies *Bemisia tabaci* and *Trialeurodes vaporariorum* on the basis of their virulence, thermal requirement and toxicogenic activity. *Biological Control*, **36**: 274-287.
- Rafiee-Dastjerdi H, Hejazi MJ, Nouri-Ghanbalani G and Saber M. 2008. Toxicity of some biorational and conventional insecticides to cotton bollworm, *Helicoverpa armigera* (Lepidoptera: Noctuidae) and its ectoparasitoid, *Habrobracon hebetor* (Hymenoptera: Braconidae). *J Entomol Soc Iran*, **28**: 27-37.
- Ramzan Asi M, Hamid Bashir M, Afzal M and Imran S. 2009. Effect of conidial concentration of entomopathogenic fungi on mortality of cabbage aphid, *Brevicoryne brassicae* L. *Pak J Life Soc Sci.*, **2**: 175-180.
- Rashki M, Kharazi-pakdel A, Allahyari H and Van Alphen JJM. 2009. Interactions among the entomopathogenic fungus, *Beauveria bassiana* (Ascomycota: Hypocreales), the parasitoid, *Aphidius matricariae* (Hymenoptera: Aphididae). *J Biocontrol*, **50(3)**: 324-328.
- Robertson JL, Russell RM, Preisler HK and Savin NE. 2007. **Bioassay with Arthropods**. CRC Press, London.
- Rosa W, Segura HR, Barrera JF and Williams T. 2000. Laboratory evaluation of the impact of entomopathogenic fungi on *Prorops nasuta* (Hymenoptera: Bethyridae), a parasitoid of the Coffee Berry Borer. *Environ Entomol.*, **29(1)**: 126-131.
- Rosenheim JA and Hoy MA. 1988. Sublethal effects of pesticides on the parasitoid *Aphytis melinus* (Hymenoptera: Aphelinidae). *J Econ Entomol.*, **81**: 476-483.
- Sarmadi S. 2008. Laboratory investigation on lethal and sublethal effects of imidacloprid, indoxacarb and deltamethrin on parasitoid wasp *Habrobracon hebetor* Say (Hymenoptera: Braconidae). M.Sc. Thesis, University of Mohaghegh Ardabili, Ardabil, Iran. 102 pp.
- SAS Institute. 2002. **The SAS System for Windows**. SAS Institute, Cary, NC.
- Soomro MH, Khalid S and Aslam M. 1992. Outbreak of banana bunchy top virus in Sindh, Pakistan. *FAO Tech. Plant Prot. Bull.*, **40**: 5-99.
- St-Leger RJ, Charnley AK and Cooper RM. 1986. Cuticle-degrading enzymes of entomopathogenic fungi: synthesis in culture on cuticle. *J Invertebr Pathol.*, **48**: 85-95.
- Stolz I, Nagel P, Lomer C and Peveling R. 2002. Susceptibility of the hymenopteran parasitoids *Apoanagyrus* (= Epidinocarsis) lopezi (Encyrtidae) and *Phanerotoma* sp. (Braconidae) to the entomopathogenic fungus *Metarhizium anisopliae* var. acridum (Deuteromycotina: Hyphomycetes). *Biocontrol Sci Techn.*, **12**: 349-360.
- Thungrabeab M and Tongma S. 2007. Effect of entomopathogenic fungi, *Beauveria bassiana* (Balsam) and *Metarhizium anisopliae* (Metsch) on non target insects. *KMITL Sci Tech J.*, **7(1)**: 8-12.
- Vojoudi S. 2011. Effect of abamectin, thiacloprid, chlorpyrifos and entomopathogenic fungi *Beauveria bassiana* on cotton bollworm, *Helicoverpa armigera*. M.Sc. thesis, University of Maragheh, Iran, 80 pp.
- Yazdanpanah F, Tohidfar M, Esna Ashari M, Ghareyazi B, Karimi Jashni M and Mosavi M. 2009. Enhanced insect resistance to bollworm (*Helicoverpa armigera*) in cotton containing a synthetic cry1Ab gene. *Indian J Biotechnol.*, **8**: 72-77.
- Zimmermann G. 1993. The entomopathogenic fungus *Metarhizium anisopliae* and its potential as a biocontrol agent. *Pestic Sci.*, **37**: 375-379.

# Accumulation of Copper in Different Tissues and Changes in Oxygen Consumption Rate in Indian Flying Barb, *Esomus danricus* (Hamilton-Buchanan) Exposed to Sub-lethal Concentrations of Copper

Suchismita Das<sup>1,\*</sup> and Abhik Gupta<sup>2</sup>

<sup>1</sup>Department of Life Science and Bioinformatics, <sup>2</sup>Department of Ecology and Environmental Science, Assam University, Silchar, 788011, India

Received: June 27, 2012; Accepted: September 2, 2012

## Abstract

In the present work, the accumulation pattern of sub-lethal doses (0.005, 0.0025 and 0.001  $\mu\text{g l}^{-1}$ ) of copper (Cu) in different tissues of Indian flying barb, *Esomus danricus* and its influence on the rate of oxygen consumption of this fish at the end of 28 days of exposure were studied. It was found that the pattern of accumulation changed with concentration. Cu concentration increased in all the tissues except bone and brain and the rate of uptake of Cu increased with time in the gill, liver and flesh but remained same in kidney. Treatment of 0.005 and 0.0025  $\mu\text{g l}^{-1}$  of Cu produced significant decline in the rates of oxygen consumption of the fish at all exposure duration, while treatment of 0.001  $\mu\text{g l}^{-1}$  produced similar decline only after 14 days of exposure when compared to control. Higher doses of exposure had more severe effects.

**Keywords:** Teleost, copper, accumulation, oxygen consumption.

## 1. Introduction

Copper (Cu) is an essential heavy metal that possess the ability to enter and concentrate in various tissues many times higher than the ambient levels and alter oxygen consumption rates of fish (Farkas *et al.*, 2002; De Boeck *et al.*, 1995). Free Cu may catalyze the formation of highly reactive hydroxyl radicals, which can result in oxidative damage to cells (Gaetke and Chow, 2003). Major sources of Cu in aquatic environment are sewage, industrial effluents and pesticides (Palacios and Risbourg, 2006). Fishes have shown to concentrate Cu in their tissues but the metal accumulation capacity is dependent on the assimilation and excretion capacities of species concerned (Rao and Patnaik, 1999). In a study, Gupta (1998) had shown that wetlands like floodplain lakes, marshes and swamps of Barak Valley, in Assam state of India, serve as sinks for heavy metals including Cu. Indian flying barb, *Esomus danricus* (Hamilton-Buchanan), a minnow having food and ornamental value, commonly inhabits such water bodies and are susceptible to Cu which target gill, liver, kidney and other tissues. It would, thus, be interesting to study the accumulation pattern of sublethal doses of Cu in different tissues of *E. danricus* and its influence on the rate of oxygen consumption of this fish.

## 2. Materials and Methods

### 2.1. Fish and experimental system

Fish of similar length ( $46.77 \pm 4.30$  mm) and weight ( $0.86 \pm 0.16$  g) were collected from unpolluted, freshwater ponds near Assam University campus, Barak valley, South Assam, India (Das and Gupta, 2009). They were acclimatized under laboratory conditions seven days prior to experimentation and commercially available fish food was given *ad libitum* twice daily. Temperature, pH, hardness and dissolved oxygen under laboratory condition were  $29^\circ\text{C}$ , 6.8, 30  $\text{mg l}^{-1}$  and 5.5  $\text{mg l}^{-1}$  respectively. Stock solution of Cu was prepared from  $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$  (Merck, Germany) and serial dilutions were prepared using chlorine free tap water as per dilution techniques (APHA, 2005). Three sub-lethal concentrations (0.001, 0.0025 and 0.005  $\mu\text{g l}^{-1}$ ) of Cu were selected based on 96 hours  $\text{LC}_{50}$  value of Cu for *E. danricus* (0.01  $\mu\text{g l}^{-1}$ ) determined in a prior study (Das and Gupta, 2010).

### 2.2. Metal accumulation study

Two hundred fish were randomly selected into four groups of 50 fish each. The 50 fish in each group were housed in five bowls, each containing ten animals. Each of the bowls contained 2 liters of water. Bowls of group I, II, III and IV contained 0.005, 0.0025 and 0.001  $\mu\text{g l}^{-1}$  and

\* Corresponding author. e-mail: drsuchismita9@gmail.com.

control (tap water without Cu) respectively. 5 bowls of each group were marked as 0, 7, 14, 21 and 28d corresponding to the days of exposure to that particular concentration of Cu. For example, bowl of group I marked '7 d' had 10 fish exposed to  $0.005 \mu\text{g l}^{-1}$  that were sacrificed after 7 days of exposure and so on. Cu treatment was carried out twice daily, and on each time, fresh water was used to avoid accumulation of waste and to ensure constant metal concentration. During the study period, dead fish (if any) were removed. During exposure periods, the fish were fed twice daily. Feeding was, however, stopped 24 h before each sacrifice. After the specified duration of exposure, fish were sacrificed and the gill, kidney, liver, bone, flesh and brain were excised, dried, weighed and digested in 5ml concentrated  $\text{HNO}_3$  to dryness in oven and dissolved in 10 ml distilled water. Analysis for Cu was carried out in a Perkin-Elmer 3110 atomic absorption spectrophotometer (AAS). The detection limit of the instrument was  $0.01 \mu\text{g l}^{-1}$ . The readings were checked with those of standard solutions, and contamination errors were minimised by using blanks, acid washed glass wares, analytical grade reagents and double distilled water. Digestion of samples was based on a modification of the method of Jayakumar and Paul (2006). Statistical significance of the differences in rate of Cu incorporation between 7<sup>th</sup> day and subsequent intervals (14, 21 and 28<sup>th</sup> day) of exposure at different Cu concentrations were made by one-way ANOVA and Tukey Test using SYSTAT 13 software for Windows.

### 2.3 Oxygen Consumption

Three test chambers (each of 3 litre capacity) were marked A, B and C containing 0.005, 0.0025 and 0.001  $\mu\text{g l}^{-1}$  of Cu respectively. Each test chamber contained ten fish. At the beginning (0 day), each fish of test chamber A was transferred to respiratory chamber, which was also numbered in accordance with the test chamber and the experiment was run for a period of 1h. After the experiment, the fish was weighed and replaced in its respective test chamber. The same process was repeated for other fishes of the test chamber A (ten replicates) and for 7d, 14d, 21d and 28day. Controls were also run simultaneously in chlorine-free tap water to obtain information on the oxygen consumption of the fish in normal state. Similarly, the process was repeated for fish in test chambers B and C. Respiratory measurements were made by the closed chamber method (Fitch, 1975) and the dissolved oxygen was estimated adopting Winkler's method. Rate of oxygen consumption was measured in ml/hr/100g tissue. Statistical significance of the differences in oxygen consumption between control and exposed fish at different Cu concentrations were made by one-way ANOVA and Tukey Test using SYSTAT 13 software for Windows.

### 3. Results

The initial level (on 0 day) of Cu for all tissues was below detection limit. However, the level of Cu accumulated by each tissue after 28 days of exposure was directly proportional to the exposure concentration of Cu. Cu, irrespective of exposure level, was never detected in

the flesh within 7 days of exposure and in bone and brain within 28 days of exposure (Table 1).

**Table 1.** Accumulation ( $\mu\text{g g}^{-1}$  dry weight) of Cu in selected tissues of *Esomus danricus* exposed to sub-lethal concentrations over time

Exposure concentration of Cu ( $\mu\text{g l}^{-1}$ )	Tissue	Concentration ( $\mu\text{g g}^{-1}$ ) after days of exposure (d)			
		7d	14d	21d	28d
Control	Gill	BDL	BDL	BDL	BDL
	Kidney	BDL	BDL	BDL	BDL
	Liver	BDL	BDL	BDL	BDL
	Flesh	BDL	BDL	BDL	BDL
	Bone	BDL	BDL	BDL	BDL
	Brain	BDL	BDL	BDL	BDL
0.001	Gill	BDL	BDL	0.0003± 0.0001	0.0008± 0.0001
	Kidney	BDL	BDL	BDL	BDL
	Liver	BDL	0.0007± 0.0002	0.0015± 0.0001	0.0026± 0.002
	Flesh	BDL	BDL	BDL	BDL
	Bone	BDL	BDL	BDL	BDL
	Brain	BDL	BDL	BDL	BDL
0.0025	Gill	0.0004± 0.0006	0.0019± 0.001	0.004± 0.002	0.007± 0.002
	Kidney	BDL	BDL	BDL	0.0013± 0.001
	Liver	0.0008± 0.0001	0.0027± 0.0003	0.006± 0.002	0.015± 0.002
	Flesh	BDL	BDL	BDL	0.001± 0.0004
	Bone	BDL	BDL	BDL	BDL
	Brain	BDL	BDL	BDL	BDL
0.005	Gill	0.0035± 0.0008	0.008± 0.001	0.015± 0.002	0.021± 0.004
	Kidney	0.0015± 0.0006	0.0029± 0.001	0.004± 0.002	0.008± 0.002
	Liver	0.005± 0.002	0.016± 0.002	0.03± 0.014	0.05± 0.016
	Flesh	BDL	BDL	0.0005± 0.0002	0.004± 0.001
	Bone	BDL	BDL	BDL	BDL
	Brain	BDL	BDL	BDL	BDL

BDL – Below detection limit

At the end of 28 days of exposure, the total tissue Cu concentration followed the pattern liver>gill>kidney>flesh>bone=brain for exposure to 0.005 and 0.0025  $\mu\text{g l}^{-1}$  Cu and liver>gill; kidney for 0.001  $\mu\text{g l}^{-1}$  Cu, bone, flesh and brain showing no detectable values. The study reveals that the uptake of Cu is tissue specific. Evaluating the rate of accumulation (tissue concentration / days of exposure) it was revealed that for exposure to 0.005  $\mu\text{g l}^{-1}$  Cu, the accumulation of Cu in gill and liver was similar up to 14 days but increased thereafter up to 28 days ( $p<0.05$ ). Kidney, on the other hand, showed similar rate of accumulation for all exposure durations for the same concentration ( $p<0.05$ ). For 0.0025  $\mu\text{g l}^{-1}$ , the rate of accumulation of Cu in gill up to 14 days was similar but increased thereafter, up to 28 days. Kidney and liver, on the other hand, showed similar rate of accumulation for all exposure durations for the same concentration ( $p<0.05$ ). For exposure to 0.001  $\mu\text{g l}^{-1}$  Cu, the rate of accumulation of Cu in gill, kidney and liver showed similar pattern for all exposure durations ( $p<0.05$ ). However, the rate of accumulation increased significantly with increase in concentration. Besides, bone and brain did not show Cu accumulation at any dose and flesh did not show any accumulation at the lowest dose up to 28 days of exposure.

The present study revealed that the flying barb responded to Cu by reducing the rate of oxygen consumption. It was observed that the oxygen

consumption rate of flying barb decreased with the increase in concentration of Cu, exposure duration also played a crucial role and oxygen consumption rate was found to decline with the increase in exposure period at 0.005 and 0.0025  $\mu\text{g l}^{-1}$  of Cu, whereas, 0.001  $\mu\text{g l}^{-1}$  Cu showed no significant changes in oxygen consumption upto 14 days of exposure, but declined significantly thereafter up to 28 days of exposure (Table 2).

**Table 2.** Effect of Copper on rate of oxygen consumption in *Esomus danricus*

Cu conc. ( $\mu\text{g l}^{-1}$ )	Oxygen Consumption (ml/hr/100g tissue)			
	7 d	14 d	21 d	28 d
Control	38.94± 0.46	39.19± 0.29	39.03± 0.24	39.06± 0.15
0.001	38.39± 0.61**	38.01± 0.64**	30.31± 1.25*	24.11± 1.76*
0.0025	29.24± 1.15*	24.87± 1.64*	21.70± 1.43*	19.20± 1.70*
0.005	27.27± 1.0*	21.67± 1.25*	19.70± 1.93*	14.84± 1.94*

\*Significant;

\*\* Not significant at  $p < 0.05$

#### 4. Discussion

Gill is the primary route for Cu uptake in fish due to its direct exposure to toxicants in water (Jayakumar and Paul, 2006; Kamunde *et al.*, 2002). In flying barb, Cu accumulated progressively in gills in concentration dependent manner, reaching 4.4 fold increase after 28 days of exposure to 0.005  $\mu\text{g l}^{-1}$  of Cu and only 0.8 fold magnification observed at 0.001  $\mu\text{g l}^{-1}$  of Cu. Though both liver and kidney are typically important for metal accumulation and storage in fish (Gigue're *et al.*, 2004), Cu metabolism is controlled chiefly by the liver. The liver not only tends to accumulate Cu from medium, but also plays an important role in Cu homeostasis (Grosell *et al.*, 1997). In flying barb, Cu concentration was 10 fold in liver after 28 days of exposure to 0.005  $\mu\text{g l}^{-1}$  (Table 1). In rainbow trout exposed to radioactive Cu, the liver was shown to be the major target organ while the kidney was less important for Cu accumulation (Clearwater *et al.*, 2000). Similarly, Cu concentrations were seven times higher in the liver than the kidney after 70 day of exposure in yellow perch *Perca flavescens* (Kraemer *et al.*, 2005). Thus, Cu was predominantly removed from the body and accumulated in the liver over time. Apart from the bone and brain, which had no detectable level of Cu throughout the study, the flesh accumulated the lowest level of Cu, even after 28 days of exposure (Table 1). This may be connected with the fact that the flesh and bone are not concerned with detoxification and therefore the transportation of Cu from other tissues to flesh and bone may not arise. Lack of a detectable level of Cu in the brain can be due to blood-brain barrier that prevents the entry of Cu into the brain (Crowe and Morgan, 1997).

The measurement of oxygen consumption of fish in presence of pollutant is the best index of its activity (Delhaye and Cornett, 1975). In the present study, flying barb responded to Cu by reducing the rate of oxygen consumption. Adverse effects of Cu on respiratory capabilities, as seen in the present study, were also studied

in *Tilapia sparrmanii* (Van Aardt and Hough, 2006) and in *Esomus danricus* (Vutukuru *et al.*, 2005). Cu had a depressing effect on oxygen consumption in *Cyprinus carpio* (De Boeck *et al.*, 1995) and in *Labeo capensis* and *Micropterus salmoides* (Van Aardt and Hough, 2007). Cu reduced oxygen consumption by common carp and rainbow trout larvae in a concentration-dependent way (Jeziorska and Sarnowski, 2002). In this study, all the sublethal doses of Cu induced excessive mucous secretion in gills. This phenomenon of mucous secretion can also impair gas exchange across the secondary lamellae epithelium (Handy and Eddy, 1989).

In conclusion, the present study indicates that Cu is accumulated at alarming level in gill and liver in Indian flying barb when exposed to level of Cu above 0.001  $\mu\text{g l}^{-1}$ . The fish responds to such high accumulation by lowering oxygen uptake.

#### References

- APHA. Standard methods for the examination of water and wastewater. 2005. 21<sup>st</sup> Edn., Washington, DC: American Public Health Association, AWWA, WPCP.
- Clearwater SJ, Baskin SJ, Wood CM and McDonald DG. 2000. Gastrointestinal uptake and distribution of copper in rainbow trout. *J. Experiment. Biol.*, **203**: 2455-2466.
- Crowe A and Morgan EH. 1997. Effect of dietary cadmium on iron metabolism in growing rats. *Toxicol. Applied Pharmacol.*, **145**: 136-146.
- Das S and Gupta A. 2010. Acute toxicity studies on Indian flying barb, *Esomus danricus* (Hamilton-Buchanan), in relation to exposure of heavy metals, cadmium and copper. *J Environment Res Develop.*, **4**: 705-712.
- Das S and Gupta A. 2009. Biometrics and growth features of *Esomus danricus* (Hamilton-Buchanan), from Barak Valley, South Assam. *J. Inland Fish. Soc.*, **41**: 81-83.
- De Boeck G De, Smet H and Blust R. 1995. The effect of sublethal levels of copper on oxygen consumption and ammonia excretion in the common carp, *Cyprinus carpio*. *Aquat Toxicol.*, **32**: 127-141.
- Delhaye W and Cornett D. 1975. Contribution to the study of the effect of copper on *Mytilus edulis* during reproductive period. *Comp Biochem Physiol., A*, **50**: 511-518.
- Farkas A, Salanki J and Specziar A. 2002. Relation between growth and the heavy metal concentration in organs of bream, *Abramis brama* L. populating lake Balaton. *Archiv Environment Contam Toxicol.*, **43**: 236-243.
- Fitch DD. 1975. Oxygen consumption in the Prosobranch snail, *Viviparus contectoides* (Mollusca: Gastropoda) - Effects of weight and activity. *Comp Biochem Physiol., A*, **51**: 815-820.
- Gaetke LM and Chow CK. 2003. Cu toxicity, oxidative stress, and antioxidant nutrients. *Toxicol.*, **189**: 147-163.
- Gigue're A, Campbell PGC, Hare L, McDonald DG and Rasmussen JB. 2004. Influence of lake chemistry and fish age on Cd, Cu and Zn concentrations in various organs of indigenous yellow perch (*Perca flavescens*). *Canadian J Fish Aquat Sci.*, **61**: 1702-1716.
- Grosell MH, Hogstrand C and Wood CM. 1997. Cu uptake and turnover in both Cu-acclimated and non-acclimated rainbow trout (*Oncorhynchus mykiss*). *Aquat Toxicol.*, **38**: 257-276.

- Gupta A. 1998. Metal accumulation by riverine and lacustrine populations of *Angulyagra oxytropis* (Benson) (Gastropoda: Viviparidae) from Barak Valley, Assam, India. *Environment Monit Assess.*, **50**: 249-254.
- Handy RD and Eddy FB. 1989. Surface absorption of aluminium by gill tissue and body mucous of rainbow trout (*Salmo gairdneri*) at the onset episodic exposure. *J Fish Biol.*, **34**: 865-874.
- Jayakumar P and Paul VI. 2006. Patterns of cadmium accumulation in selected tissues of the catfish *Clarias batrachus* (Linn.) exposed to sublethal concentration of cadmium chloride. *Vet Archiv.*, **76**: 167-177.
- Jeziarska B and Sarnowski P. 2002. The effect of mercury, copper and cadmium during single and combined exposure on oxygen consumption of *Oncorhynchus mykiss* Wal. and *Cyprinus carpio* L. larvae. *Archiv Polish Fish*, **10**: 15-22.
- Kamunde C, Grosell M, Higgs D and Wood CM. 2002. Copper metabolism in actively growing rainbow trout (*Oncorhynchus mykiss*): interactions between dietary and waterborne copper uptake. *J Experiment Biol.*, **205**: 279-290.
- Kraemer LD, Campbell PGC and Hare L. 2005. Dynamics of Cd, Cu and Zn accumulation in organs and sub-cellular fractions in field transplanted juvenile yellow perch (*Perca flavescens*). *Environment. Pollut.*, **138**: 324-337.
- Palacios SP and Risbourg SB. 2006. Hepatocyte nuclear structure and subcellular distribution of copper in zebrafish *Brachydanio rerio* and roach *Rutilus rutilus* (Teleostei, Cyprinidae) exposed to copper sulphate. *Aquat Toxicol.*, **77**: 306-313.
- Rao LM and Patnaik R. 1999. Heavy metal accumulation in the catfish, *Mystus vittatus* (Bloch.) from Mehadrigeeda Stream of Visakhapatnam, India. *Pollut Res.*, **19**: 325-329.
- Van Aardt WJ and Hough M. 2007. The effect of short-term Cu exposure on the oxygen consumption and Cu accumulation of mudfish (*Labeo capensis*) and the largemouth bass (*Micropterus salmoides*) in hard water. *Water SA.*, **33**: 701-708.
- Van Aardt WJ and Hough M. 2006. Acute effects of Cu on oxygen consumption and 96 hr-LC 50 values in the freshwater fish *Tilapia sparrmanii* (Teleostei: Cichlidae) in Mooi river hard water, South Africa. *African J Aquat Sci.*, **31**: 305-311.
- Vutukuru SS, Ch. Suma K, Madhavi R, Juveria J, Pauleena S, Rao JV and Anjaneyulu Y. 2005. Studies on the development of potential biomarkers for rapid assessment of copper toxicity to freshwater fish using *Esomus danricus* as model. *Int J Environ Res Public Health*, **2**: 63-73.

# Modification of the Mechanical Properties of Red Blood Cell Membrane by Spent *Plasmodium falciparum* Culture Supernatant

Nii A. Aryee<sup>1,2,\*</sup> and Yuichi Takakuwa<sup>3</sup>

<sup>1</sup> Department of Medical Biochemistry, University of Ghana Medical School, P. O. Box 4236, Accra, Ghana,

<sup>2</sup> Department of Biochemistry, Faculty of Basic Medical Sciences, College of Medicine and Allied Health Sciences ,

University of Sierra Leone, New England, Freetown, Sierra Leone,

<sup>3</sup>Department of Biochemistry, Tokyo Women's Medical University, 8-1 Kawada-cho, Shinjuku-ku, Tokyo 162-8666, Japan.

Received: August 27, 2012; Accepted: October 12, 2012

## Abstract

The effect of spent culture supernatant on RBC membrane mechanical properties, RBC membrane deformability and stability, were assayed by ektacytometry after treating RBC ghost membranes with spent *Plasmodium falciparum* (*P. falciparum*) malaria culture supernatant fluid (Pf(+)), malaria culture medium which had not been exposed to *P. falciparum*-infected RBCs Pf(-), and malaria culture medium from an *in vitro* culture system containing uninfected RBCs (M). All treated RBC membranes were less deformable, and more stable than control, untreated RBC membranes. However, compared to controls, Pf(+)-treated RBC membranes were more deformable than both Pf(-)-treated and M-treated RBC membranes. Similarly, Pf(+)-treated RBC membranes were more stable than membranes treated with both Pf(-) and M. The deformability and stability of Pf(+)-treated RBC membranes were significantly different from membranes treated with both Pf(-) and M ( $P \leq 0.05$ ). However, there was no significant difference between deformability and membrane stability for Pf(-)-treated and M-treated RBC membranes. Our results suggest that spent *P. falciparum* culture supernatant may significantly modify RBC membrane mechanical properties.

**Keywords:** RBC membrane, deformability, stability, *P. falciparum*, parasite culture supernatant.

## 1. Introduction

The red blood cell (RBC) membrane has been well described in terms of its structure and composition (Snyder and Sheafor, 1999). It is composed of a lipid bilayer, integral proteins, a sub-membranous skeletal protein network of spectrin, and peripheral proteins (Chasis and Mohandas, 1986). Deformability and stability are two essential qualities of the RBC membrane that play a crucial role in the maintenance of normal blood flow and supply to tissues (Arai *et al.*, 1990; Chien, 1987; Mohandas and Chasis, 1993). Consequently, any variation in RBC membrane deformability, as in the case of malaria (Miller, Baruch, Marsh & Doumbo, 2002) or sickle cell diseases (Platt, 1995) can potentially compromise the micro-circulatory function. Studies have demonstrated that the protein network underlying the RBC membrane, together with the membrane bilayer and the network of membrane-associated proteins, play a key role in regulating RBC membrane deformability and stability (Takakuwa, 2001). Considerable effort and time have

been invested in the *in vitro* cultivation of the erythrocytic stages of *Plasmodium*, the stages most often associated with the pathogenesis of malaria.

A major accomplishment in this area was defining ideal *in vitro* conditions for continuous cultivation of *P. falciparum* strains from different geographical areas, using HEPES-buffered RPMI 1640 culture medium that is supplemented with human serum, RBCs, and sodium bicarbonate (Trager and Jensen, 1976; Trager and Jensen, 1977; Trager and Jensen, 1980). RBCs are an obligatory requirement in this *in vitro* medium. Indeed, fresh and stored erythrocytes appear to be equally suitable for continuous *in vitro* cultivation provided that adenosine triphosphate (ATP) levels of RBCs are within the normal range (Capps and Jensen, 1983, Schuster, 2002). While this unique medium has been shown to be suitable for *in vitro* cultivation of *P. falciparum* malaria stages, the effect of malaria culture medium which has not been exposed to *P. falciparum*-infected RBCs (iRBCs) and spent malaria parasite culture supernatant on RBC membrane mechanical properties has not been fully explored. *In vitro* cultivation of *P. falciparum* is usually carried out through

\* Corresponding author. e-mail: dmariyee@yahoo.co.uk.

the continuous preservation of iRBCs deposited in static thin layers of settled infected cells. However, not all aspects regarding how the culturing methodology affects the development and growth of the parasite are fully understood.

Although some agents have been shown to alter the RBC membrane mechanical properties (Beder *et al.*, 2002), it is uncertain how *P. falciparum* spent parasite culture supernatant will affect both RBC membrane deformability and stability. The aim of this study was to assess the effect of spent parasite culture, and malaria culture medium which has not been exposed to iRBCs, on membrane deformability and stability of RBC.

## 2. Materials and Methods

### 2.1. *In vitro* cultivation of malaria parasites

*P. falciparum* (F32 strain) used for this study was a gift from Prof. Akira Kaneko of Tokyo Women's Medical University. *P. falciparum* F32 was maintained in a continuous culture as previously described by Trager & Jensen (1976). The culture contains type O<sup>+</sup> RBCs at 2% hematocrit in malaria culture medium, Pf(-) which consists of RPMI 1640 medium buffered with 25mM HEPES and 24 mM NaHCO<sub>3</sub> and supplemented with 10 % heat-inactivated human O<sup>+</sup> serum. Culture media were kept under a standard gas mixture of 5% O<sub>2</sub>, 5%CO<sub>2</sub> and 90% N<sub>2</sub> at 37°C. From a culture system containing mixed stage *P. falciparum*-infected RBCs at 10 % parasitemia, the supernatant was harvested and centrifuged for 15 minutes at ×500 g (GS-6KR: Beckman, Fullerton, California). This supernatant (Pf(+)), typically described as "spent", was used for subsequent experiments. Similarly, supernatant from malaria culture medium incubated with uninfected RBCs (Pf(-)), under conditions similar to that of spent culture system, was harvested and treated in the same manner described for spent *P. falciparum* culture supernatant. All solvents and chemicals were of analytical grade and were purchased from Wako Chemicals (Osaka, Japan). Water was purified using the milliQ pore system (Millipore, Bedford, MA, USA).

### 2.2. RBC lysis and 5T5K ghost preparation

Fresh type O<sup>+</sup> venous blood was drawn from healthy volunteers, collected in heparin-coated tubes and processed within minutes after collection. RBCs were separated from leukocytes by filtration through polyurethane filter kit (Terumo), and washed three times with 10 mM Tris-HCl buffer (pH 7.4) that contains 120 mM KCl to remove cytosolic components. RBCs were lysed and washed once in ice-cold 1:35 diluted 5T5K buffer (5T5K buffer: 5mM Tris-HCl, pH 7.4, containing 5mM KCl) by centrifugation at ×15,000 g to obtain 5T5K RBC ghost cells. Ghost-cell suspensions were incubated at 37°C for 40 min for membrane resealing.

### 2.3. RBC lysis and MgATP ghost preparation

MgATP ghost-cells were prepared in the same manner described for 5T5K ghost-cells with slight modifications, as described previously by Mohandas & Chasis (1993). The 5T5K buffer (lysing buffer) was supplemented with 10 mM ATP and 1 mM MgCl<sub>2</sub>. To restore isotonicity, 100

µl of a mixture of 150 mM KCl, 1 mM MgCl<sub>2</sub>, and 1 mM dithiothreitol was added to RBC ghost suspensions and incubated at 37°C for 40 min for membrane resealing. MgATP-ghosts were prepared in the presence of MgATP throughout the process, whereas 5T5K ghosts were prepared in the absence of MgATP. The morphology of unfixed resealed ghosts was examined by dark-field light microscopy using a Nikon microscope.

### 2.4. Pre-treatment of ghost membranes

MgATP ghost suspensions were re-suspended in nine volumes of either spent culture supernatant (Pf(+)), malaria culture medium (Pf(-)), or a medium from *in vitro* culture containing uninfected static RBCs (M). MgATP ghost re-suspensions were incubated for 30 minutes at 37°C. Untreated MgATP ghosts served as control. After incubation, all RBC ghosts were centrifuged once at ×500 g for 10 minutes and assayed by ektacytometry for changes in membrane deformability and stability.

### 2.5. RBC membrane stability assay

RBC membrane resistance to shear-induced fragmentation was measured by subjecting MgATP ghosts to a constant high shear stress of 750 dynes cm<sup>-2</sup> in the ektacytometer, as described by Hardeman *et al.* (1994). For the present study, 500 µl of each test sample (treated and untreated ghosts) was thoroughly mixed with 3 ml of 50 % dextran (wt./vol.) in 10 Mm TBS buffer (pH 7.4), resuspended in the ektacytometer chamber, and rotated at 100 rpm to generate the desired shear stress. When the shear stress is first applied, RBC ghosts are maximally deformed into ellipsoids and produce a narrow elliptical pattern that generates a maximal value of the deformability index (DI). RBC ghosts are unable to withstand the large value of applied shear stress, and subsequently begin to fragment, with DI gradually decaying under the constant high shear stress. A decrease in the DI value reflects fragmentation of the intact membranes into small, undeformable spherical particles. T<sub>50</sub> is the time required for DI to decay to half its maximum value. A decrease in the T<sub>50</sub> value reflects a decrease in RBC membrane stability, and vice versa. Suspending RBC ghosts in a viscous medium is necessary for the ektacytometric method as cells suspended in a low viscosity fluid (e.g. Tris-buffered saline, TBS) tumble, and do not deform in response to the applied shear stress (Johnson, 1989). It has been reported that the use of dextran, a neutral polysaccharide, to increase the viscosity of the suspending medium, does not affect the integrity of RBC membrane or alter its mechanical behavior (Nash & Meiselman, 1984).

#### 2.5.1. RBC membrane stability assay

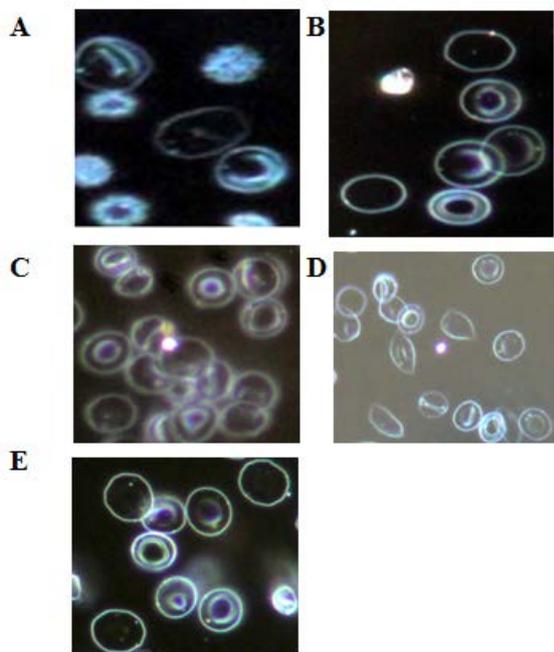
The Maximum Deformability Index (DI<sub>max</sub>) and initial rate of change of the deformability index (DI<sub>initial</sub>) which is a direct measure of membrane deformability, were determined at various shear stress values by laser diffraction analysis using an ektacytometer (LORCA: RR Mechantronics; Hoorn, The Netherlands). Details of ektacytometry have been described elsewhere (Johnson, 1989; Hardeman *et al.*, 1994). Briefly, 100 µl of each test sample was re-suspended in 3.0 ml of 50% dextran in 10 mM TBS (wt./vol.), pH 7.4, and subjected to increasing shear stress up to 200 dynes cm<sup>-2</sup>. Changes in laser

diffraction patterns were analyzed to derive  $DI_{max}$ . A decrease in  $DI_{initial}$  indicates a reduction in RBC membrane deformability. Suspending RBC ghosts in a viscous medium is necessary for the ektacytometric method as cells suspended in a low viscosity medium (e.g. TBS) tumble and do not deform in response to the applied shear stress (Johnson, 1989).

### 3. Results

#### 3.1. Morphology of resealed ghosts

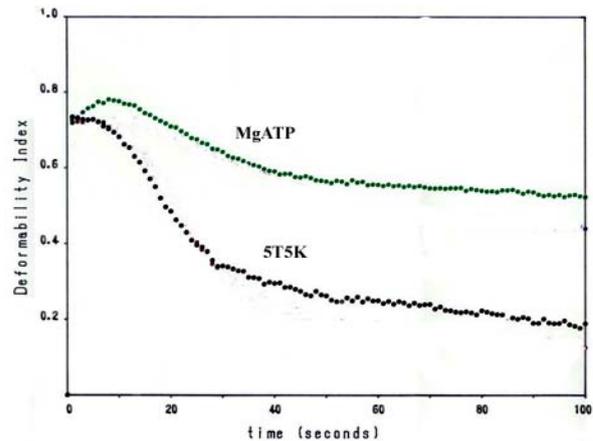
Resealed ghosts prepared without MgATP were predominantly echinocytic in shape (Fig1.A). However a few discoid-shaped ghosts were observed. Ghosts prepared in the presence of MgATP were all discoid in shape (Fig. 1B), indicating that a minimum concentration of ATP is essential for maintenance of normal discoid shape. Pre-treatment with the various media did not alter the discoid shape of MgATP ghosts (Fig.1C, D and E).



**Figure 1.** Dark-field light microscopy of ghosts. Panel A: 5T5K ghosts, Panel B: MgATP ghosts, Panel C: Pf(+)-treated MgATP ghosts, Panel D: Pf(-)-treated MgATP ghosts, Panel E: M- treated MgATP ghosts.

#### 3.2. Membrane stability profile of MgATP and 5T5K RBC ghosts

Membrane stability profile of 5T5K ghosts and MgATP ghosts is shown in Fig. 2. When ghosts were constantly subjected to a high shear stress of 750 dynes/cm<sup>2</sup>, ghost membranes were maximally deformed and fragmented soon after attaining the maximum deformability index ( $DI_{max}$ ), resulting in decreased DI value. The rate of decrease of DI is a direct measure of membrane mechanical stability. 5T5K ghosts underwent fragmentation faster than MgATP ghosts which resulted in the reduction of DI value.



**Figure 2.** Deformability profile of 5T5K and MgATP ghosts

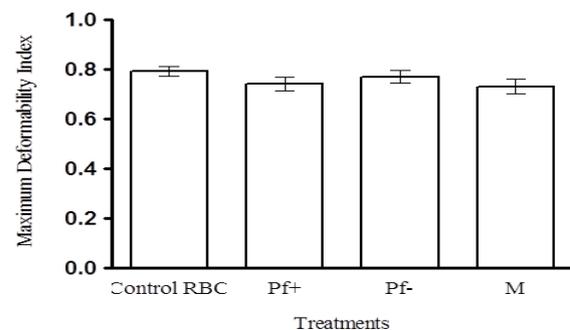
#### 3.3. Duration for DI to reach half its maximum value ( $T_{50}$ )

The Mean  $T_{50}$  values for the various treatments were 96, 40, 31 and 28 seconds for control, Pf(+), Pf(-) and M-treated MgATP ghosts, respectively (Fig. 5). Control MgATP had the longest fragmentation time, whereas M-treated MgATP ghosts had the shortest. Treated RBC ghosts exhibited varying decline in  $T_{50}$  values to about 58%, 68% and 71% of that of the control ghosts. These reductions correspond to Pf(+), Pf(-) and M-treated MgATP ghosts.

The  $T_{50}$  for all treatments were significantly different from each other, except for Pf(-) and M-treated MgATP ghosts ( $P \leq 0.05$ ).

#### 3.4. Maximum deformability index ( $DI_{max}$ )

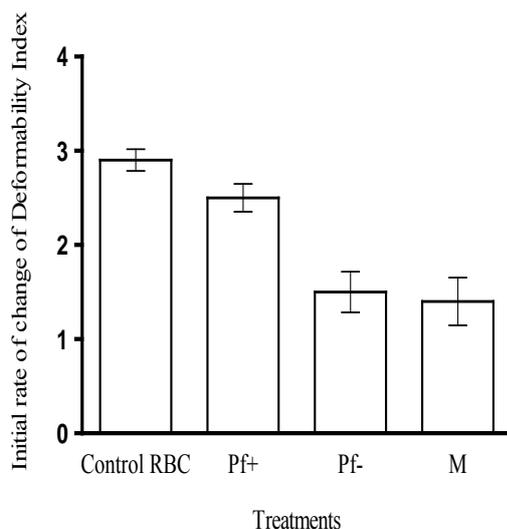
MgATP ghosts attained higher  $DI_{max}$  values than 5T5K ghosts (Fig. 3), and because of that they were used for subsequent experiments. When MgATP ghosts were subjected to increasing values of shear stress, the deformability index value increased, reaching a maximum of 0.79, 0.73, 0.77 and 0.74 for control, Pf(+), Pf(-) and M-treated ghosts, respectively (Fig.3). Although Pf(+)-treated ghosts recorded the lowest  $DI_{max}$  value, the Newman-Keuls Multiple Comparison Test showed that there was no significant difference between the mean  $DI_{max}$  values of Pf(+)- and M-treated ghosts ( $P \leq 0.05$ ). Similarly, a comparison between untreated and Pf(-)-treated ghosts showed no significant difference between their mean  $DI_{max}$  values ( $P \leq 0.05$ ).



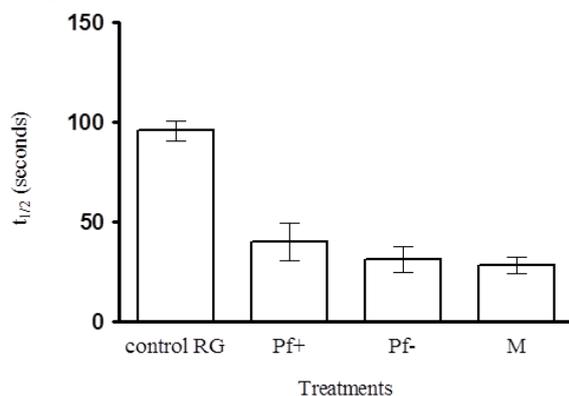
**Figure 3.**  $DI_{max}$  values for the various treatments. All values are reported as Mean  $\pm$  S.D.

### 3.5. Initial rate of change of the deformability index ( $DI_{initial}$ )

$DI_{initial}$  values for the various treatments are shown in Fig. 4. Mean  $DI_{initial}$  values obtained were 2.9, 2.5, 1.5 and 1.4 for control, Pf(+), Pf(-) and M-treated 5T5K ghosts, respectively. The mean  $DI_{initial}$  value was highest for control RBCs, and lowest for M-treated RBCs ( $P \leq 0.05$ ). Compared to control ghosts, treated RBCs exhibited reductions in  $DI_{initial}$  values to about 14%, 48% and 52% of that of the control ghosts, corresponding to Pf(+), Pf(-) and M-treated MgATP ghosts, respectively. Stastical analysis indicates that the  $DI_{initial}$  for Pf(-) and M-treated ghosts were not significantly different from each other. The mean  $DI_{initial}$  values of all other treatments were significantly different from each other ( $P \leq 0.05$ ).



**Figure 4.**  $DI_{initial}$  values for the various treatments. All values are reported as Mean  $\pm$  S.D



**Figure 5.**  $T_{50}$  values for the various treatments. All values are reported as Mean with Range

## 4. Discussion

The effects of a variety of agents on RBC membrane deformability and stability have been studied previously (Hardeman & Ince, 1999; Beder *et al.*, 2002; Tadesse *et al.*, 2004). However, the effect of *P. falciparum* culture supernatant on RBC membrane mechanical properties of deformability and stability, has not been established. The present study shows that RBC ghosts prepared in the

presence of MgATP attained higher  $DI_{max}$  values than ghosts prepared without MgATP, as indicated by the membrane stability profile (Fig. 2). This may possibly have to do with the ATP-dependent discocyte configuration, as ATP depletion of RBCs has been associated with discocyte-echinocyte transformation (Palek, Stewart, & Lionetti, 1974). The mechanism by which RBCs maintain their biconcave shape has been the subject of numerous studies. One of the critical factors for the maintenance of biconcave shape is the level of RBCs ATP (Weed *et al.* 1969; Szasz, 1970). We have used a laser diffraction method (ektacytometry) to directly measure changes in RBC membrane indices of deformability and stability. This technique was validated by subjecting resealed RBC ghosts to manipulations known to modify the membrane shear modulus (Heath *et al.*, 1982). Our results show that, compared to control untreated RBC membranes, there was a reduction in RBC membrane deformability upon pretreatment with spent parasite culture supernatant (Pf(+)), malaria culture medium (Pf(-)), and malaria culture medium containing uninfected RBCs (M). The reduction in RBC membrane deformability by spent supernatant from *P. falciparum* cultures is consistent with the observation that in *falciparum* malaria, the deformability of the entire RBC population is reduced in proportion to disease severity (Nuchsongsin *et al.*, 2007). It has been found that in patients with severe *falciparum* malaria, the entire RBC mass, containing both uninfected and infected RBCs, becomes rigid (Cooke, Mohandas, & Coppel, 2004; Parker *et al.*, 2008; Nuchsongsin *et al.*, 2007). Several mechanisms such as hemin-induced oxidative damage of the RBC membrane, alterations in the phospholipid bilayer, and attached spectrin network have been proposed as being responsible for the increased rigidity and reduced deformability of the RBCs infected with *falciparum* malaria (Parker *et al.*, 2008; Nuchsongsin *et al.*, 2007). The cause of rigidity of RBCs under *in vitro* culture conditions, however, is not well-defined, but has been attributed, according to one study, to the discharge of a multitude of proteins from infected RBCs into the culture media (Naumann *et al.*, 1991), some of which could be involved in modulating the deformability of uninfected RBCs *in vitro*. Our results indicate that both Pf(-) and M-treated RBCs caused a greater reduction in RBC membrane deformability than Pf(+)-treated RBCs. This difference in membrane deformability is most likely due to the depletion of one or more of these components during parasite cultivation. As expected, our results reveal that the control RBC ghosts had the highest membrane stability. This is due to the intact membrane skeleton that is devoid of any modulation. However, the RBC membrane stability was reduced significantly after exposure to malaria culture medium, but was partially restored as time progressed. Data from both pathologic membranes and biochemically perturbed membranes have elucidated that RBC deformability and stability are not related to one another, as decreased deformability can be associated with either increased or decreased membrane stability. (Chasis and Mohandas, 1986). These findings imply that different skeletal protein interactions may regulate RBC membrane deformability and stability.

Although multiple parasite ligand-erythrocyte receptor interactions must occur for successful *Plasmodium* invasion of the human red cell (Montero, Rodriguez, Oksov & Lobo, 2009), it is probable that a transient change in RBC membrane stability is essential for *P. falciparum* invasion of the RBC or its survival within the micro-environment of the *in vitro* culture. This change may occur independently of changes in the RBC membrane deformability.

## 5. Conclusion

In this study, we have shown that malaria culture media and spent parasite culture supernatant can modify the stability and deformability of the RBC membrane. However, these two parameters do not seem to be related. Although medium change is essential for the sustenance of *P. falciparum* *in vitro*, the premise that modulation of RBC membrane mechanical properties may be indispensable for *P. falciparum* invasion, requires further elucidation.

## Acknowledgement

The author is grateful to Prof. Yuichi Takakuwa for providing laboratory facilities for this work. The authors are also grateful to the Japan International Cooperation Agency (JICA) for funding this work.

## Authors' Contributions

NAA and YT designed the study. NAA was responsible for the study implementation and data collection. NAA and YT analyzed the data. NAA wrote the paper and all authors reviewed the manuscript.

## References

- Arai K, Iino M, Shio H and Uyesaka N. 1990. Further investigation of red blood cell deformability with nickel mesh. *Bioreheology*, **27**: 47-65.
- Beder I, Kittova M, Mataseje A, Carsky J, Orszaghova Z and Babinska K. 2002. Effect of selected substances with anti-glycative with anti-glycative and anti-oxidant properties on erythrocyte deformability in diabetic patients. *Scripta Medica (Brno)*, **75**: 239-244.
- Capps TC and Jensen JB. 1983. Storage requirements for erythrocytes used to culture *Plasmodium falciparum*. *J Parasitol.*, **69**: 158-162.
- Chasis JA and Mohandas N. 1986. Erythrocyte membrane stability and deformability: Two distinct membrane properties that are independently regulated by skeletal protein associations. *J Cell Biol.*, **103**: 343-350.
- Chien S. 1987. Red cell deformability and its relevance blood flow. *Ann Rev Physiol.*, **49**: 177-192.
- Cooke BM, Mohandas N and Coppel RL. 2004. Malaria and the red blood cell membrane. *Semin Hematol.*, **41(2)**:173-188.
- Hardeman MR and Ince C. 1999. Clinical potential of *in vitro* measured red cell deformability: A myth? *Clin Hemorheol Microcirc.*, **21**: 277-284.
- Hardeman MR., Goedhart PT, Dobbe JGG and Lettinga KP. 1994. Laser-assisted optical rotational cell analyzer (LORCA) 1. A new instrument for measurement of various structural hemorheological parameters. *Clin Hemorheol.*, **14**:605-618.
- Heath BP, Mohandas N, Wyatt JL and Shohet SB. 1982. Deformability of isolated red blood cell membranes. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, **691(2)**: 211-219.
- Johnson RM. 1989. Ektacytometry of red blood cells. *Methods Enzymol.*, **173**: 35-54.
- Miller LH, Baruch DI, Marsh K, and Doumbo OK. 2002. The pathogenic basis of malaria. *Nature*, **415**: 673-679.
- Mohandas N and Chasis JA. 1993. Red cell deformability, membrane material properties and shape: Regulation by transmembrane, skeletal and cytosolic proteins and lipids. *Semin Hematol.*, **30**: 171-192.
- Montero E, Rodriguez M, Oksov Y and Lobo CA. 2009. Babesia divergens apical membrane antigen 1 and its interaction with the human red blood cell. *Infect Immun.*, **77(11)**: 4783-4793.
- Nash GB and Meiselman H J 1984. Effects of dextran and polyvinylpyrrolidone on red cell geometry and membrane elasticity. *Ann. NY Acad Sci.*, **416**: 255-262.
- Naumann KM, Jones GL, Allan S and Smith R. 1991. A *Plasmodium falciparum* exo-antigen alters erythrocyte membrane deformability. *Federation of European Biochemical Societies*, **9**: 617-622.
- Nuchsongsin F, Chotivanich K, Charunwatthana P, Fausta O, Taramelli D, Day NP, White NJ and Dondorp AM. 2007. Effects of malaria heme products on red blood cell deformability. *Am J Trop Med Hyg.*, **77(4)**: 617-622.
- Palek J Stewart G and Lionetti FJ. 1974. The dependence of shape of human erythrocyte ghosts on calcium, magnesium, and adenosine triphosphate. *Blood*, **44(4)**: 583-597.
- Parker PD, Tilley L and Klonis N. 2004. *Plasmodium falciparum* induces reorganization of host membrane proteins during intra-erythrocytic growth. *Blood*, **103**: 2404-2406.
- Platt OS. 1995. The sickle syndrome. In: Haldin, RI, Lux SE and Stossel TP (Eds.), **Blood: Principles and Practice of Hematology**. Philadelphia, PA: Lippincott , pp. 1592-1700.
- Schuster FL. 2002. Cultivation of *Plasmodium* spp. *Clinical Microbiol Rev.*, **15**: 355-364.
- Snyder GK and Sheafor BA. 1999. Red blood cells: Centerpiece in the evolution of the vertebrate circulatory system. *Amer Zool.*, **39**: 189-198.
- Szasz I. 1970. The role of nucleotides and bivalent cations in determining the shape of normal and trypsin-treated erythrocytes. *Acta Biochim Acad Sci Hung.*, **5**: 399.
- Tadesse A, Sandoval F and Shannon E J. 2004. Stabilization of red blood cell membranes by thalidomide *in vitro*. *Immunopharmacol & Immunotoxicol.*, **26(4)**: 501 - 509.
- Takakuwa Y. 2001. Regulation of red cell membrane protein interactions: Implications for red cell function. *Current Opinion in Hematol.*, **8(2)**: 80-84.
- Trager W and Jensen JB. 1976. Human malaria parasites in continuous culture. *Science*, **193**: 673-675.
- Trager W and Jensen JB. 1977. Cultivation of erythrocytic stages. *Bull WHO*, **55**:363-365.
- Trager W and Jensen JB. 1980. Cultivation of erythrocytic and exoerythrocytic stages of plasmodia. In: Kreier JP (Ed.), **Malaria**, Vol. 2. Academic Press, New York, pp. 271-319.
- Weed RI, LaCelle P, Merrill EW, Craig G, Gregory A and Karch F. 1969. Metabolic dependence of red cell deformability. *J Clin Invest*, **48**: 795.



# The Antihyperglycaemic Effect of the Aqueous Extract of *Origanium vulgare* Leaves in Streptozotocin-Induced Diabetic Rats

Nema A. Mohamed<sup>1,\*</sup> and Omimah A. Nassier<sup>2</sup>

<sup>1</sup>Zoology Department, Faculty of Science, Alexandria University, Alexandria, Egypt;

<sup>2</sup>Biology Department, Faculty of Science for Girls, King AbdulAziz University, Saudi Arabia

Received: September 28, 2012; Accepted: October 21, 2012

## Abstract

The current study aimed to investigate the antihyperglycaemic effect of aqueous extract of *Origanum vulgare* (oregano, OV) leaves in streptozotocin (STZ) induced diabetic rat. Streptozotocin induced diabetic rats showed significant ( $P < 0.05$ ) increase in the levels of blood glucose, glycosylated haemoglobin, pancreatic amylase, liver and kidney weights/body weight ratios, urea, uric acid, creatinine and significant decrease in the levels of plasma insulin, liver and muscle glycogen and body weight. Oral administration of the aqueous extract of OV leaves (20 mg/kg) produced a significant decrease in blood-glucose levels, glycosylated haemoglobin, pancreatic amylase in STZ diabetic rats ( $P < 0.05$ ) in comparison with standard drug Glibenclamide (GB) (0.9 mg/kg body weight). Treatment with the aqueous extracts of OV leaves decreased liver weights/body weight ratios in diabetic rats, while kidney weight/body weight ratios, urea, uric acid, creatinine levels were partially improved. Oral administration of the aqueous extract of OV leaves (20 mg/kg) improved the reduction in serum insulin, liver and muscle glycogen contents and body weight in STZ diabetic rats. The data in the present study may support the use of *O. vulgare* plants as traditional remedies for the treatment of diabetes mellitus.

**Keywords:** *Origanum vulgare*, streptozotocin, glycosylated haemoglobin, glucose, insulin, rats.

## 1. Introduction

Diabetes mellitus is a chronic metabolic disease which now afflicts 10 % of the world population. It is considered a "modern-day epidemic" and is rightly recognized as a global public health issue (Gispén and Biessels, 2000; Burke *et al.*, 2003). In recent years, there has been renewed interest in the treatment of diabetes using herbal drugs, which are generally non-toxic. World Health Organization has also recommended the evaluation of the effectiveness of plants in condition where we lack safe modern drugs. Plant derivatives with hypoglycaemic properties had been used in folk medicine and traditional healing systems around the world from very ancient times (Yeh *et al.*, 2003).

*Origanum vulgare* (oregano, OV) is a member of the plant family Lamiaceae, the genus *Origanum* is an annual, perennial and shrubby herb that is native to the Mediterranean, Euro-Siberian and Irano-Siberian regions (Aligiannis *et al.*, 2001). Oregano contains oleanolic and ursolic acids, flavonoids and hydroquinones, caffeic, rosmarinic, and lithospermic acid, tannins, and phenolic

glycosides. Phenolic compounds represent 71% of the total oil. The polar phenols' thymol and carvacrol are responsible for many of the properties of the essential oil, as well as p-cymene and terpinene (Dadalioglu and Evredlik, 2004; Giordani *et al.*, 2004; Koukoulis *et al.*, 2005; Tampieri *et al.*, 2005; Bozin *et al.*, 2006). The main known pharmacological activities of OV were antibacterial (Nazia *et al.*, 2007) antifungal (Portillo-Ruiz *et al.*, 2005) antiparasitic (Force *et al.*, 2000) anti-thrombin (Goun *et al.*, 2002) anti-oxidant ( Stashenko *et al.*, 2002 ) and anti-inflammatory (Ocaña-Fuentes *et al.*, 2010). There are also some reports regarding the antimutagenic and anticarcinogenic effect of oregano; representing an alternative for the potential treatment and/or prevention of certain chronic ailments, like cancer (Arcila-Lozano *et al.*, 2004).

Oregano is a powerful antioxidant. It contains several potent antioxidants that may contribute to the findings in preliminary studies that oregano exhibits benefits towards the cardiovascular and nervous systems and relieves inflammation and modulates blood sugar and lipids (Singletary, 2010).

\* Corresponding author. e-mail: science20111@hotmail.com.

The aim of this study was directed to determine the antihyperglycaemic effect of OV extract in streptozotocin diabetic rats. Glibenclamide (GB) was used as standard antihyperglycaemic drug.

## 2. Materials and Methods

Streptozotocin was purchased from Sigma (St. Louis, MO, U.S.A.), glibenclamide (gift from Pharmacy, University hospital, Jeddah, Saudi Arabia). Dried leaves of *O.vulgare* were purchased from an authentic source at local market (Jeddah, Saudi Arabia).

### 2.1. Preparation of aqueous extracts

Plant material was prepared according to Eddouks *et al.*, 2003: 1g of powdered leaves mixed with 100 ml distilled water was boiled for 10 min. and then cooled for 15 min. Thereafter, the aqueous extract was filtered using a Millipore filter (Millipore 0.2 mm) to remove particulate matter. The filtrate was then freeze-dried and the desired dose (mg of lyophilized aqueous extract of OV leaves per kg body weight) was prepared and reconstituted in 1.5 ml of distilled water. The aqueous extracts were prepared daily, just before administration. The extracts obtained were then given orally to different groups of rats at a dose of 20 mg/kg body weight.

### 2.2. Experimental design

The experimental animals were obtained from the Animal Unit of King Fahd Medical Research Center, King Abdul Aziz University, Jeddah, Saudi Arabia. Animals were housed under environmental conditions (23 ± 1°C, 55 ± 5% humidity and 12-h light :12-h dark cycle) and maintained with free access to water and a standard chow diet. Diabetes was induced by an intraperitoneal injection of streptozotocin at a dose of 45 mg/kg body weight dissolved in a citrate buffer (0.1M, pH 4.5) (Burcelin *et al.*, 1995). After 3 days the rats with fasting blood-glucose levels more than 200 mg/dL were considered as diabetic and selected for the study. All the pharmacological experiments were carried out after obtaining approval of the Institutional animal ethics Committee of King Abdulaziz University, Saudi Arabia.

The preliminary studies of different doses (20,40,60 mg/kg) showed that the most effective dose of oregano is 20 mg/kg. The animals were randomly divided into five groups of 12 animals each. Group I (untreated controls): normal rats receiving water and fed *ad libitum* and served as a control group. Group II (untreated diabetics): diabetic rats receiving water and fed *ad libitum* and served as diabetic control rats. Group III (treated controls): normal rats receiving water and fed *ad libitum* and oregano at 20 mg/kg body weight (Lemhadri *et al.*, 2004). Group IV (treated diabetics): diabetic rats receiving water and fed *ad libitum* and oregano 20mg/kg body weight. Group V (treated diabetics): diabetic rats receiving water and fed *ad libitum* and antidiabetic drug (glibenclamide). Rat equivalent dose of glibenclamide was calculated using conversion table devised by Paget and Barnes (1964) and was 0.9 mg/kg body weight.

The drug treatment solutions were administered orally by gastric intubation using a syringe once daily at 08:00 a.m. The effect of OV aqueous extracts or

glibenclamide on blood glucose was determined in fasted rats, after 2 and 6 weeks of once daily repeated oral administration (20 mg/kg). The body, liver and kidney weights of all rats were measured at weeks 2 and 6.

### 2.3. Biochemical analysis

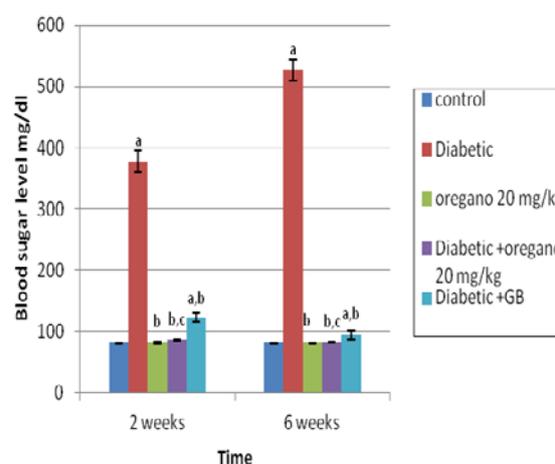
At the end of experiment, rats were anaesthetized and blood samples were collected from the tail vein. Fasting blood-glucose level was measured in the whole blood after 12 h fasting. Glycosylated haemoglobin (HbA1-test) was estimated in whole blood by fast ion – exchange resin separation method (Nuttall, 1998). Serum was separated, and insulin and pancreatic amylase were determined according to methods of Finlay and Dillard (2007) and Winn-Deen *et al.*(2008), respectively. Liver and muscle glycogen contents were determined by the method of Huijing (1970). Urea, uric acid and creatinine were estimated by using the F-200 fluorescence spectrophotometer in serum (Newman and Price, 2001).

### 2.4. Statistical analysis

Values reported are expressed as mean ± SE. Statistical significance of the difference between groups was determined by one-way analysis of variance test (ANOVA). The values were considered to be significantly different when the *P* value was less than 0.05 (Zar, 1996).

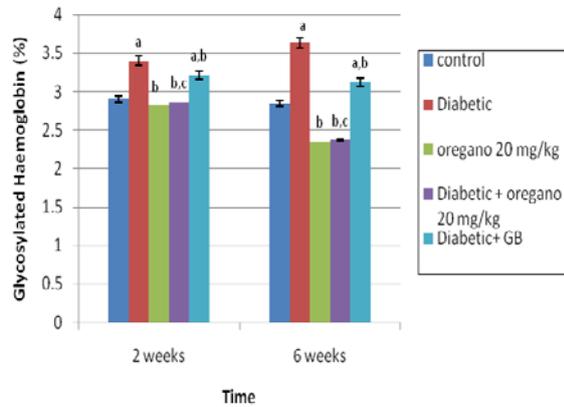
## 3. Results

In this study, the level of blood glucose, glycosylated haemoglobin (HbA1C), insulin after using a 45 mg/kg dose of streptozotocin, ensured induction of diabetes in rats (figures 1,2,3). The blood-glucose levels rose markedly after STZ administration, and the high glucose levels were maintained for 6 weeks (Fig 1). Oral administration of the aqueous OV extracts (20 mg/kg) produced a significant decrease (81.65±1.049) in blood-glucose levels in STZ diabetic rats (526.80 ±7.889) (*P* < 0.05). Treatment with glibenclamide (GB), showed reduced blood-glucose levels as compared to control group.



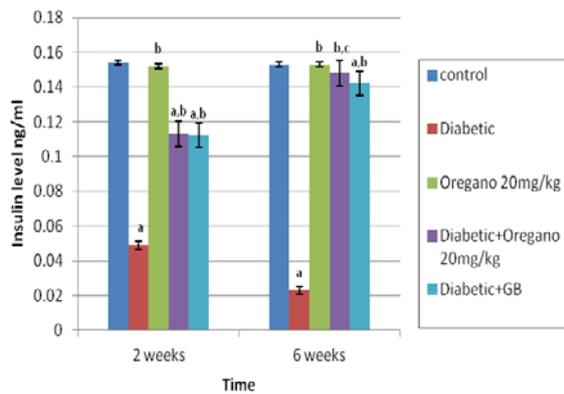
**Figure 1.** Effect of aqueous extracts of oregano leaves (20 mg/kg) on blood-sugar level (mg/dl) - Data are expressed as the mean ± SE. Each value corresponds to a mean of 6 animals. a, comparison of Group I vs Group II. b, comparison of Group III, Group IV & Group V vs Group II. c, comparison of Group IV vs Group V at *P* < 0.05.

In the untreated diabetic animals, the initial HbA1C value increased significantly ( $3.634 \pm 0.0093$ ) compared to the control. In the oregano treated diabetic groups (20 mg/kg) the HbA1C value return to the normal value as compared to control group (Fig.2).



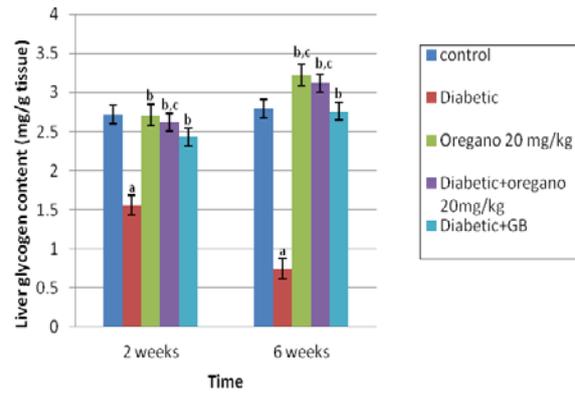
**Figure 2.** Effect of aqueous extracts of oregano leaves (20 mg/kg) on glycosylated haemoglobin (%) - Data are expressed as the mean  $\pm$  SE. Each value corresponds to a mean of 6 animals. a, comparison of Group I vs Group II .b ,comparison of Group III, Group IV & Group V vs Group II. c , comparison of Group IV vs Group V at  $P < 0.05$ .

On the other hand, the serum insulin levels decreased markedly after STZ administration. Oral administration of the aqueous OV extracts (20 mg/kg) or GB partially improved insulin levels in STZ diabetic rats ( $P < 0.05$ ) after 2 weeks. After 6 weeks, oral administration of the aqueous OV extracts (20 mg/kg) significantly improved insulin levels in STZ diabetic rats ( $P < 0.05$ ) in comparison with standard drug GB as shown in figure 3.

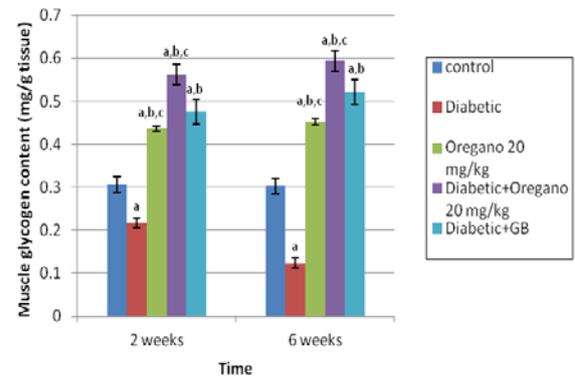


**Figure 3.** Effect of aqueous extracts of oregano leaves (20 mg/kg) on Insulin (ng/ml). Data are expressed as the mean  $\pm$  SE. Each value corresponds to a mean of 6 animals. a , comparison of Group I vs Group II .b ,comparison of Group III, Group IV & Group V vs Group II. c , comparison of Group IV vs Group V at  $P < 0.05$ .

Figures 4 and 5 revealed marked depletion in the liver and muscle glycogen contents ( $P < 0.05$ ) in STZ-induced diabetic rats compared to control. The administration of OV (20 mg/kg) or GB for six weeks significantly ( $P < 0.05$ ) increased the liver and muscle glycogen contents in diabetic rats compared to untreated diabetic and control groups. It was also noticed that OV (20 mg/kg) control groups showed a significant increase in liver and muscle glycogen contents ( $3.12 \pm 0.174$  and  $0.594 \pm 0.0209$ ) throughout the experimental period.



**Figure 4.** Effect of aqueous extracts of oregano leaves (20 mg/kg) on Liver glycogen content (mg/g tissue) - Data are expressed as the mean  $\pm$  SE. Each value corresponds to a mean of 6 animals. a, comparison of Group I vs Group II .b, comparison of Group III, Group IV & Group V vs Group II. c, comparison of Group IV vs Group V at  $P < 0.05$ .



**Figure 5.** Effect of aqueous extracts of oregano leaves (20 mg/kg) on muscle glycogen content (mg/g tissue) - Data are expressed as the mean  $\pm$  SE. Each value corresponds to a mean of 6 animals. a, comparison of Group I vs Group II .b, comparison of Group III, Group IV & Group V vs Group II. c, comparison of Group IV vs Group V at  $P < 0.05$ .

In the STZ-treated groups, the pancreatic amylase level was significantly increased. Oral administration of the aqueous OV extracts (20 mg/kg) administration produced a significant decrease in pancreatic amylase levels in STZ diabetic rats ( $P < 0.05$ ) as compared to control group (Table 1).

**Table1.** Effect of aqueous extracts of oregano leaves (20 mg/kg) on Pancreatic amylase.

Group	parameter	Pancreatic amylase(U/L)	
		2 <sup>nd</sup> week	6 <sup>th</sup> week
Control		243.80 $\pm$ 0.707	242.80 $\pm$ 0.735
Diabetic		365.42 $\pm$ 2.315 <sup>a</sup>	287.20 $\pm$ 0.583 <sup>a</sup>
Oregano(20 mg/kg)		235.20 $\pm$ 0.860 <sup>a,b</sup>	237.60 $\pm$ 0.927 <sup>b</sup>
Diabetic + Oregano(20 mg/kg)		232.20 $\pm$ 0.718 <sup>a,b</sup>	246.00 $\pm$ 1.140 <sup>b,c</sup>
Diabetic + GB		242.20 $\pm$ 1.772 <sup>b</sup>	252.60 $\pm$ 0.926 <sup>a,b</sup>

- Data are expressed as the mean  $\pm$  SE. Each value corresponds to a mean of 6 animals. a, comparison of Group I vs Group II .b, comparison of Group III, Group IV & Group V vs Group II. c, comparison of Group IV vs Group V at  $P < 0.05$ .

The mean body weight of rats was shown in table 2. Results showed that weight of diabetic controls significantly decreased during the experimental period while normal controls and OV extract rats gained significant weight ( $P < 0.05$ ). The diabetic group given oregano leaves extract (20 mg/kg) or GB administration maintained weight gain. The liver weight/body weight ratios have been shown to increase in diabetic rats significantly ( $P < 0.05$ ) in comparison to control animals (Table 2). This increase was completely reversed by OV leaves extract (20 mg/kg) administration and partially improved by GB. STZ-induced diabetes caused a significant ( $P < 0.05$ ) increase in kidney weight/body weight ratios in comparison to control. This enhancement was partially improved by OV (20 mg/kg) or GB (Table 2).

**Table 2.** Effect of aqueous extracts of oregano leaves on body weight, liver and kidney weights/body weight ratios in rats

parameter Group	body weight (g)		Liver weight / body weight		Kidney weight / body weight	
	2 <sup>nd</sup> week	6 <sup>th</sup> week	2 <sup>nd</sup> week	6 <sup>th</sup> week	2 <sup>nd</sup> week	6 <sup>th</sup> week
Control	210.00 ± 1.303	235.00 ± 1.612	0.03268 ± 0.00029	0.03278 ± 0.00025	0.00420 ± 0.00049	0.00419 ± 0.00005
diabetic	186.00 ± 4.703 <sup>a</sup>	172.00 ± 1.327 <sup>a</sup>	0.03992 ± 0.00062 <sup>a</sup>	0.05273 ± 0.00115 <sup>a</sup>	0.00602 ± 0.00022 <sup>a</sup>	0.00612 ± 0.00026 <sup>a</sup>
oregano (20 mg/kg)	205.00 ± 1.581 <sup>b</sup>	236.00 ± 1.095 <sup>b</sup>	0.03042 ± 0.00013 <sup>b</sup>	0.03321 ± 0.00070 <sup>b</sup>	0.00414 ± 0.00004 <sup>b</sup>	0.00412 ± 0.00010 <sup>b</sup>
diabetic + Oregano (20 mg/kg)	202.00 ± 1.140 <sup>b,c</sup>	225.00 ± 2.509 <sup>b,c</sup>	0.03290 ± 0.00024 <sup>b,c</sup>	0.03268 ± 0.00738 <sup>b,c</sup>	0.00519 ± 0.00010 <sup>a,b</sup>	0.00490 ± 0.00043 <sup>a,b</sup>
diabetic + GB	196.00 ± 1.673 <sup>a</sup>	211.00 ± 2.881 <sup>b</sup>	0.03814 ± 0.00153 <sup>a,b</sup>	0.04267 ± 0.02059 <sup>a,b</sup>	0.00544 ± 0.00021 <sup>a,b</sup>	0.00493 ± 0.00010 <sup>a,b</sup>

- Data are expressed as the mean ± SE. Each value corresponds to a mean of 6 animals. a, comparison of Group I vs Group II. b, comparison of Group III, Group IV & Group V vs Group II. c, comparison of Group IV vs Group V at  $P < 0.05$ .

The results in Table 3 showed significant increase ( $P < 0.05$ ) in the level of urea, uric acid and creatinine, which are markers of renal dysfunction in the diabetic group compared to control group. This enhancement was partially improved by OV (20 mg/kg) or GB (Table 3).

**Table 3.** Effect of aqueous extracts of oregano leaves on urea, uric acid and creatinine in rats

parameter Group	Urea (mg/dl)		Uric acid (mg/dl)		Creatinine (mg/dl)	
	2 <sup>nd</sup> week	6 <sup>th</sup> week	2 <sup>nd</sup> week	6 <sup>th</sup> week	2 <sup>nd</sup> week	6 <sup>th</sup> week
Control	30.40 ± 2.461	30.60 ± 2.021	1.24 ± 0.040	1.30 ± 0.032	0.30 ± 0.010	0.31 ± 0.020
Diabetic	59.21 ± 6.406 <sup>a</sup>	65.43 ± 5.137 <sup>a</sup>	1.94 ± 0.323 <sup>a</sup>	2.52 ± 0.902 <sup>a</sup>	0.72 ± 0.096 <sup>a</sup>	0.94 ± 0.137 <sup>a</sup>
Oregano (20 mg/kg)	32.00 ± 3.284	33.00 ± 3.153	1.32 ± 0.583	1.34 ± 0.058	0.32 ± 0.157	0.33 ± 0.155
Diabetic + Oregano (20 mg/kg)	53.00 ± 2.030 <sup>a,b</sup>	49.00 ± 2.144 <sup>a,b</sup>	1.84 ± 0.340 <sup>a,b</sup>	1.60 ± 0.045 <sup>a,b</sup>	0.65 ± 0.030 <sup>b</sup>	0.56 ± 0.144 <sup>a,b</sup>
Diabetic + GB	55.00 ± 3.112 <sup>a,b</sup>	52.20 ± 3.138 <sup>a,b</sup>	1.86 ± 0.082 <sup>a,b</sup>	1.65 ± 0.178 <sup>a,b</sup>	0.67 ± 0.112 <sup>a,b</sup>	0.58 ± 0.138 <sup>a,b</sup>

- Data are expressed as the mean ± SE. Each value corresponds to a mean of 6 animals. a, comparison of Group I vs Group II. b, comparison of Group III, Group IV & Group V vs Group II. c, comparison of Group IV vs Group V at  $P < 0.05$ .

#### 4. Discussion

In spite of the presence of known antidiabetic medicine in the pharmaceutical market, herbal drugs are frequently considered to be less toxic and also free from side effects, than synthetic ones. In the present study, the administration of streptozotocin (45mg/kg) induced hyperglycaemia in rats. Treatment of diabetic rats with oregano at 20 mg/kg showed a significant decrease in the blood-sugar level. This may be due to that oregano enhance the insulin sensitivity of the receptors on cells, leading to reduced levels of blood sugar and more energy production. The accumulating evidence suggests that modulation of insulin secretion and/or insulin action mechanisms could be involved in the antidiabetic effect of oregano. This evidence was confirmed with Talpur *et al.* (2005) who reported that extracts of oregano improve blood sugar levels by enhancing insulin sensitivity. Also, the hypoglycaemic effect of oregano may be due to the interference on absorption of dietary carbohydrates in small intestine or stimulation of glucose utilization by peripheral tissues. In line with this evidence of this study, Maghrani *et al.* (2003) and Ortiz-Andrade *et al.* (2007) reported that the hypoglycaemic action of medicinal plant may be due to a reduction in the intestinal absorption of glucose and/or inhibition of tubular glucose reabsorption. Oregano leaves contain phenolic glucosides that help control blood sugar as reported by Takeda *et al.* (2008).

The level of glycosylated haemoglobin (HbA1C) has been shown to be an important parameter of chronic glycaemic control in patients with DM, an elevated HbA1c almost always indicates DM (The International Expert Committee, 2009). The present data showed that, the high levels of HbA1c in diabetic rats were significantly lowered by the treatment with aqueous extract of oregano leaves. Decreased HbA1C levels in the treated diabetic rats could be due to an improvement in insulin secretion from the remnant pancreatic  $\beta$ -cells in diabetic rats, consequently, resulting in improvement in glycemic control (Vinay *et al.*, 2010).

Increased insulin level in diabetic rats after treatment with the oregano treatment (Fig. 3) this may be due to the bioactive molecules present in oregano leaf extract that may probably stimulate the  $\beta$  cells of the pancreas to produce insulin. Furthermore, this effect may be due to that oregano has been shown to have an insulin-like biological activity. This explanation agrees with that reported by Broadhurst *et al.* (2000) who showed that the positive effects of oregano extracts on insulin activity suggest a possible role of this plant in improving glucose and insulin metabolism. In contrast, Lemhadri *et al.* (2004) concluded that aqueous extract of OV (20 mg/kg) exhibits an anti-hyperglycaemic activity in STZ rats without affecting basal plasma insulin concentrations.

Glycogen is the primary intracellular storable form of glucose and its levels in various tissues, especially skeletal muscles are direct reflection of insulin activity. The observed decrease in hepatic and muscle glycogen may be due to insufficient insulin and inactivation of the glycogen synthetase system in the diabetic state (Vinay *et al.*, 2010). However, after the treatment with oregano, there was a significant increase in the liver and muscle glycogen levels

in the diabetic rats. The increased hepatic glycogen level in the treated diabetic rats may be due to increased level of insulin, which has increased glycogenesis and decreased glycogenolysis and gluconeogenesis. Thus antihyperglycemic effect of OV may be due to protection of surviving pancreatic  $\beta$  cells, increase in insulin secretion and glycogen storage (Jagtap & Patil, 2009). Inhibitory effects on glycogenolysis have been reported for glibenclamide in the presence of insulin after stimulation of glycogenolysis by glucagon (Carvalho-Martini *et al.*, 2006).

The increment in pancreatic amylase in diabetic rats was antagonized by oregano treatment. The inhibition of pancreatic amylase delay carbohydrate digestion and protract overall carbohydrate digestion time, resulting in the reduction in glucose absorption rate and consequently, dulling the postprandial plasma glucose rise. Several indigenous medicinal plants have a high potential in inhibiting pancreatic amylase enzyme activity (Valiathan, 1998). Mc Cue *et al.* (2004) reported that extracts of clonal oregano lines have strong inhibitory activity against porcine pancreatic amylase (PPA) *in vitro*. MC Cue and Shetty (2004) reported the ability of rosmarinic acid, one of the principal phenolic components of oregano, to inhibit porcine pancreatic amylase (PPA) activity. One of the potentially important components of anti-diabetic activity by oregano extracts is mild amylase inhibition by phenolic antioxidants that contribute management of hyperglycemia (Mc Cue *et al.*, 2004).

Weight loss has been known to be one of the symptoms of DM. Similar observations were detected in many experimental studies (Al-Attar & Zari, 2007; Subash-Babu *et al.*, 2008; Sellamuthu *et al.*, 2009 and Salahuddin *et al.*, 2010). In the diabetic control rats, deficiency of insulin led to decreased amino acids uptake by tissues with a consequent reduction in the level of protein synthesis. Also insulin deficiency results in lipolysis in adipose tissues and protein breakdown (Vasudenvan & Sreekumari, 2007). The increase in weight observed in the group treated with oregano, and normal control group may be a reflection of efficient insulin action. The liver and kidney weights/body weight ratios in the diabetic groups were higher than those of the normal groups (Table, 2) suggesting the occurrence of the edema and inflammation of these organs as reported by Kamath and Rajini (2006). High concentrations in serum urea, uric acid, and creatinine strongly suggested impairment of kidney function in diabetic rats. Similar effect was recorded previously by Alarcon *et al.* (2005) Jaya *et al.* (2010) and Manikandaselvi *et al.* (2012). The present data indicated that the oregano supplement has a partial renoprotective effect. These results are in agreement with other previous studies by Khan *et al.* (2011) who stated that oregano showed antiuro lithic activity both *in vitro* and *in vivo* models in addition to its antioxidant, renal epithelial cell protective, antispasmodic and diuretic activities. These different activities observed in the crude extract might be due to the presence of flavonoids that were known to possess diuretic activities (Ramamoorthy *et al.*, 2010).

Oregano has long been used in traditional medicine in the treatment of common ailments and have been potential for positive modulation of oxidation-linked diseases such

as diabetes. Flavonoids are considered as active principles in many medicinal plants and natural products with a positive effect for human health (Wollenweber, 1988). These natural compounds could act separately or synergistically to cause the hypoglycaemic effect. This could not exclude the intervention of other phytochemical constituents as bioactive hypoglycaemic agents. Most of oregano's effects on the body are due to its high content of antioxidants, which play a role in destroying the production of free radicals (Spiridon *et al.*, 2011) and modulates blood sugar and lipids (Singletary, 2010).

## 5. Conclusion

The obtained results may support the use of oregano as culturally adopted treatments for insulin resistance and hyperglycemia and support its inclusion as a natural, safe, anti-diabetic therapy for modulation of Type 2 diabetes mellitus.

## References

- Al-Attar AM and Zari TA. 2007. Modulatory effects of ginger and clove oils on physiological responses in streptozotocin - induced diabetic rats. *Int J Pharmacol.*, **3**: 34-40.
- Alarcon AFJ, Calzada BF, Hernandez GE, Ruiz AC and Roman RR. 2005. Acute and chronic hypoglycaemic effect of *Iberivillea Sonorae* root extracts. *J Ethnopharmacol.*, **97**:447.
- Aligiannis N, Kalpoutzakis E, Mitaku S and Chinou IB. 2001. Composition and antimicrobial activity of the essential oils of two *Origanum* species. *J Agric Food Chem.*, **49**:4168-4170.
- Arcila-Lozano CC, Loarca-Piña G, Lecona-Urbe S and González de Mejía E. 2004. Oregano: properties, composition and biological activity. *Arch Latinoam Nutr.*, **54**(1):100-111.
- Bozin B, Mimica-Dukic N, Simin N and Anackov G. 2006. Characterization of the volatile composition of essential oils of some lamiaceae spices and the antimicrobial and antioxidant activities of the entire oils. *J Agric Food Chem.*, **54**:1822-1828.
- Broadhurst CL, Polansky MM and Anderson RA. 2000. Insulin-like biological activity of culinary and medical plant aqueous extracts *in vitro*. *J Agric Food Chem.*, **48**:849-52.
- Burcelin R, Eddouk M, Maury J, Kande J, Assan R and Girard J. 1995. Excessive glucose production, rather than insulin resistance, accounts for hyperglycaemia in recent onset streptozotocin diabetic rats. *Dibetologia.*, **36**:283-290.
- Burke JP, Williams K, Narayan K MV, Leibson C, Haffner SM and Stern MP. 2003. A population perspective on diabetes prevention: Whom should we target for preventing weight gain? *Diabetes Care.*, **26**:1999-2004.
- Carvalho-Martini M, de Oliveira DS, Suzuki-Kemmelmeier F and Bracht A. 2006. The action of glibenclamide on glycogen catabolism and related parameters in the isolated perfused rat liver. *Res Commun Mol Pathol Pharmacol.*, **119**(1-6):115-26.
- Dadalioglu I and Evrendilek GA. 2004. Chemical compositions and antibacterial effects of essential oils of Turkish oregano (*Origanum minutiflorum*), bay laurel (*Laurus nobilis*), Spanish lavender (*Lavandula stoechas* L.), and fennel (*Foeniculum vulgare*) Polissiou MG. (2000): GC-MS analysis) on common foodborne pathogens. *J Agric Food Chem.*, **52**:8255-8260.
- Eddouks M, Jouad H, Maghrani M, Lemhadri A and Burcelin R. 2003. Inhibitor of endogenous glucose production accounts for

- hypoglycaemic effect of *Spergularia purpurea* in diabetic mice. *Phytomedicine*, **10**:594-599.
- Finlay JWA and Dillard RF. 2007. Appropriate calibration curve fitting in ligand binding assays. *AAPS J.*, **9**(2):E260-E267.
- Force M, Sparks WS and Ronzio RA. 2000. Inhibition of enteric parasites by emulsified oil of oregano in vivo. *Phytother Res.*, **14**:213-214.
- Giordani R, Regli P, Kaloustian J, Mikail C, Abou L and Portugal H. 2004. Antifungal effect of various essential oils against *Candida albicans*. Potentiation of antifungal action of amphotericin B by essential oil from *Thymus vulgaris*. *Phytother Res.*, **18**:990-995.
- Gispén WH and Biessels GJ. 2000. Cognition and produce type II or non-insulin-dependent diabetes synaptic plasticity in diabetes mellitus. Trends in mellitus and blood-glucose concentrations remained. *Neurosciences*, **23**: 542-549.
- Goun E, Cunningham G, Krasnykh O and Miles SH. 2002. Anti-thrombin activity of some constituents from *Origanum vulgare*. *Fitoterapia*, **73**: 692-694.
- Huijing F. 1970. A rapid enzymic method for glycogen estimation in very small tissue samples. *Clin Chem Acta*, **30**:567-572.
- Kamath V and Rajini PS. 2006. Altered glucose homeostasis and oxidative impairment in pancreas of rats subjected to dimethoate intoxication. *Toxicology*, **228**: 1-10.
- Khan A, Samra B, Saeed RK and Anwar HG. 2011. Antiuro lithic activity of *Origanum vulgare* is mediated through multiple pathways. *Complementary and Alternative Med.*, **11**:96.
- Jagtap AG and Patil PB. 2010. Antihyperglycemic activity and inhibition of advanced glycation end product formation by *Cuminum cyminum* in streptozotocin induced diabetic rats. *Food and Chemical Toxicol.*, **48**(8-9): 2030-2036.
- Jaya A, Shanthi P and Panchanadham S. 2010. Cytoprotective effect of *Semecarpus anacardium* against toxicity induced by Streptozotocin in rats. *J Experimental Pharmacol.*, **2**:135-143.
- Koukoulitsa C, Zika C, Geromichalos GD, Demopoulos VJ and Skaltsa H. 2006. Evaluation of aldose reductase inhibition and docking studies of some secondary metabolites, isolated from *Origanum vulgare* L. ssp. *hirtum*. *Bioorg Med Chem*, **14**:1653-1659.
- Lemhadri A, Zeggwagh NA, Maghrani M, Jouad H and Eddouks M. 2004. Anti-hyperglycaemic activity of the aqueous extract of *Origanum vulgare* growing wild in Tafilalet region. *J Ethnopharmacol.*, **92**(2-3): 251-256.
- Manikandaselvi S, Subalakshmi R, Thinagarbabu R and Ravikumar R. 2012. Evaluation of antidiabetic potential of ethanolic extract of leaves of *ficus bengalensis* linn. *Inter J Pharma and Bio Sciences*, **3**(2):B108-B113.
- Maghrani M, Lemhadri A, Jouad H, Michel JB and Eddouks M. 2003. Effect of the desert plant *Retama raetam* on glycaemia in normal and streptozotocin-induced diabetic rats. *J Ethnopharmacol.*, **87**:21-25.
- McCue P and Shetty K. 2004. Inhibitory effects of rosmarinic acid extracts on porcine pancreatic amylase *in vitro*. *Asia Pac Clin Nutr.*, **13**(1): 101-106.
- McCue P, Vattam D and Shetty K. 2004. Inhibitory effect of clonal oregano extracts against porcine pancreatic amylase *in vitro*. *Asia Pac J Clin Nutr.*, **13** (4): 401-408.
- Nazia MAC, Sabahat S and Perween T. 2007. Antibacterial effects of oregano (*Origanum vulgare*) against gram-negative bacilli. *Pak J Botany*, **39**(2): 609-613.
- Newman DJ and Price CP. 2001. **Tietz Fundamental of Clinical Chemistry**, 5<sup>th</sup> Ed., Burtis, C.A. & Ashwood, E.R. (W.B. Saunders eds. Philadelphia USA). pp.414.
- Nuttall FQ. 1998. Comparison of percent total gHb with percent HbA1c in people with and without known diabetes. *Diabetes Care*, **21**:1475-1480.
- Ocaña-Fuentes A, Arranz-Gutiérrez E, Señorans FJ and Reglero G. 2010. Supercritical fluid extraction of oregano (*Origanum vulgare*) essential oils: anti-inflammatory properties based on cytokine response on THP-1 macrophages. *Food Chem Toxicol.*, **48**(6):1568-1575.
- Ortiz-Andrade RRG, Castillo-España S, Ramirez-A PV, Villalobos-Molina RG and Estrada-Soto S. 2007.  $\alpha$ -Glucosidase inhibitory activity of the methanolic extract from *Tournefortia hartwegiana*: an anti-hyperglycemic agent. *J Ethnopharmacol.*, **109**: 48-53.
- Paget GE and Barnes JM. 1964. Toxicity tests. In: Laurence D R and Bacharach A L (Eds.), **Evaluation of Drug Activities: Pharmacometrics**. London and New York: Academic press, pp.135-166.
- Portillo-Ruiz MC, Viramontes-Ramos S, Muñoz-Castellanos LN, Gastélum-Franco MG and Nevárez-Moorillón GV. 2005. Antifungal activity of Mexican oregano (*Lippia berlandieri Shauer*). *J Food Prot*, **68**(12):2713-2717.
- Ramamoorthy J, Venkataraman S, Meera R, Christina AJM and Chidambaramathan N. 2010. Physio-Phytochemical screening and Diuretic activity of leaves of *Pavetta indica* inn. *J Pharm Sci Res.*, **2**(8):506-512.
- Salahuddin M, Jalalpure SS and Gadge NB. 2010. Antidiabetic activity of aqueous bark extract of *Cassia glauca* in streptozotocin-induced diabetic rats. *Can J Physiol Pharmacol*, **88**: 153-160.
- Sellamuthu PS, Muniappan BP, Perumal SM and Kandasamy M. 2009. Antihyperglycemic effect of mangiferin in streptozotocin induced diabetic rats. *J Health Sci.*, **55**:206-214.
- Singletary K. 2010. Oregano: Overview of the literature on health benefits. *Nutrition Today*, **45** (3): 129-138.
- Spiridon I, Colceru S, Anghel N, Teaca CA, Bodirlau R and Armatu A. 2011. Antioxidant capacity and total phenolic contents of oregano (*Origanum vulgare*), lavender (*Lavandula angustifolia*) and lemon balm (*Melissa officinalis*) from Romania. *Nat Prod Res.*, **25**(17):1657-1661.
- Stashenko EE, Puertas MA and Martinez JR. 2002. SPME determination of volatile aldehydes for evaluation of *in vitro* anti-oxidant activity. *Analytical and Bioanalytical Chem.*, **373**: 70-74.
- Subash-Babu P, Ignacimuthu S and Prince PSM. 2008. Restoration of altered carbohydrate and lipid metabolism by hyponid, a herbal formulation in streptozotocin-induced diabetic rats. *Asian J Biochem.*, **2**: 90-98.
- Takeda Y, Tomonari M, Arimoto S, Masuda T, Otsuka H, Matsunami K, Honda G, Ito M, Takaishi Y, Kiuchi F and Khodzhimatov OK. 2008. A new phenolic glucoside from an Uzbek medicinal plant, *Origanum Tyttanthum*. *J Natural Med.*, **62**(1):71-74.
- Talpur N, Echard B, Ingram C, Bagchi D and Preuss H. 2005. Effects of a novel formulation of essential oils on glucose-insulin metabolism in diabetic and hypertensive rats: a pilot study. *Diabetes Obesity and Metabolism*, **7**(2): 193-199.
- Tampieri MP, Galuppi R, Macchioni F *et al.* 2005. The inhibition of *Candida albicans* by selected essential oils and their major components. *Mycopathologia*, **159**:339-345.

- The International Expert Committee. 2009. International Expert Committee Report on the Role of the A1C Assay in the Diagnosis of Diabetes. *Diabetes Care*, **32(7)**:1327-1334.
- Zar JH.1996. **Biostatistical Analysis**. Prentice – Hall New Jersey, pp.718.
- Valiathan MS. 1998. Healing plants. *Curr Sci.*, **75**:1122.
- Vasudevan DM and Sreekumari S. 2007. **Text Book of Biochemistry for Medical Student**. New Delhi, Japee Brothers Medical Publishers Ltd., pp:151-160.
- Vinay KK, Kameswara RB, Dilip RM *et al.* 2010. Effect of *Pterocarpus santalinus* bark, on blood glucose, serum lipids, plasma insulin and hepatic carbohydrate metabolic enzymes in streptozotocin-induced diabetic rats. *Food and Chemical Toxicology*, **48(5)**:1281-1287.
- Winn-Deen ES, David H, Sigler G and Chavez R. 2008. Development of a direct assay for alpha-amylase. **Clinical Chemistry 34.Behring Diagnostics**, Somerville, NJ 08876.
- Wollenweber, E., 1988. Occurrence of flavonoid aglycones in medicinal plants. In: Cody V, Middleton E. Jr., Harborne JB and Beretz A. (Eds.) **Plant Medicine II: Biochemical Cellular and Medicinal Properties**, Progress in Clinical and Biological Research, New York, pp.45-55.
- Yeh GY, Eisenberg DM, Kaptchuk TJ and Phillips RS. 2003. Systematic review of herbs and dietary supplements for glycemic control in diabetes. *Diabetes Care*, **26**:1277-1294.



# Fecundity of Bigfin squid, *Sepioteuthis lessoniana* (Lesson, 1830) (Cephalopoda: Loliginidae)

Venkatesan Vellathi<sup>1,\*</sup> and Rajagopal Santhanam<sup>2</sup>

<sup>1</sup>Central Marine Fisheries Research Institute, Cochin, 682018; Kerala,

<sup>2</sup>Faculty of Marine Sciences, Centre of Advanced Study in Marine Biology, Amamalai University,  
Parangipettai, 608502, India.

Received: August 16, 2012; Accepted: October 22, 2012

## Abstract

A total of 25 mature female *Sepioteuthis lessoniana* (Lesson, 1830) specimens, with a dorsal mantle length between 120-196 mm, were investigated. Estimated potential fecundity varied from 180 to 1054 eggs with mean value of 497 eggs. The potential fecundity values of *S. lessoniana* in the Mandapam waters (Palk Bay) are found almost similar to those found in Tanzanian waters but higher than the ones in Sri Lankan lagoons. Larger females have higher potential fecundity. The correlation values showed that the fecundity is more closely related to the ovary weight and body weight than to the mantle length. Fecundity increased exponentially with mantle length. The Nidamental gland weight was also found more closely related to the ovary weight in logarithmic relationship. Fecundity vs. ovary weight ( $r = 0.899$ ) and Nidamental gland weight vs. ovary weight ( $r = 0.942$ ) exhibited a strong relationships while fecundity showed weak correlations with mantle length ( $r = 0.658$ ) and body weight ( $r = 0.663$ ).

**Keywords:** *Sepioteuthis lessoniana*, fecundity, Mandapam Waters (Palk Bay).

## 1. Introduction

Cephalopods (squid, cuttlefish and octopus) form increasingly essential resources for human consumption and a chief food for many top predators (Lefkaditou *et al.*, 2003). Adult cephalopods are voracious and active carnivores feeding mainly on fishes and crustaceans. Loliginid squids play an important role in near-shore ecosystems both as prey and as predators (Jackson and Pecl, 2003).

The squids and cuttlefish have fewer eggs production compared to the finfishes and prawns (Nabhitabhata, 1995). There are very few reports of fecundity estimates of Indian Cephalopods (Asokan, 2000). With the increasing trawling activity and targeting the cephalopod, the cephalopods need regular monitoring to avoid a possible collapse of the fishery. In general, fecundity is low in cephalopods because of the absence of larval stage and hatchlings are virtually in miniature adults.

There are controversies in estimating cephalopod fecundity because of the different spawning strategies of cephalopods: single synchronous terminal spawning, or multiple spawning in which several batches are spawned. Therefore, it is better to use potential fecundity where the

maximum number of eggs prior to spawning is counted (Boyle and Rodhouse, 2005).

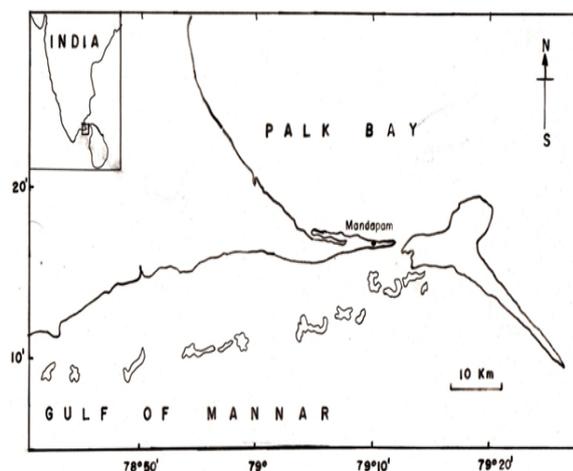
In the past, many studies were conducted on the fecundity of other squids elsewhere (Macewicz *et al.*, 2004; Salman and Onsoy, 2004; Laptikhovsky and Nigmatullin, 2005; Salman and Laptikhovsky, 2005; Nigmatullin and Markaida, 2009; Salman and Onsoy, 2010). Previous fecundity studies on *S. lessoniana* have been done in tropical regions such as Tanzania (Mhitu *et al.*, 2001), Sri Lanka (Sivashanthini *et al.*, 2010). No data on the fecundity of *S. lessoniana* in the Palk Bay waters is currently available. Hence, the objective of the work reported here is to study potential fecundity and various logarithmic relationships of fecundity in mature females of squid, *S. lessoniana* in this region.

## 2. Materials and Methods

A total of 25 mature females with dorsal mantle length (DML) between 120-196 mm and body weight (BW) between 118.3-406.1g were used for estimation of potential fecundity.

Samples were collected from commercial bottom trawlers of Mandapam landing centre (Palk Bay) between 2009-2010 (Figure 1).

\* Corresponding author. e-mail: venkatcmfri@yahoo.co.in.



**Figure 1.** Map showing the location of sampling site (Mandapam, Palk Bay).

Collected specimens were kept in ice box and transferred to laboratory for the detailed study. For each animal, the DML was measured to the nearest 1mm and BW to the nearest 0.1g. After dissecting, gonad and nidamental glands were weighed to the nearest mg. The ovary comprises the proximal and distal oviducts where the former is chiefly a store for mature oocytes and the latter a store for immature and maturing oocytes (Lipinski and Underhill, 1995). Potential fecundity of mature squid was estimated by sub-sampling the eggs from both the proximal and distal oviducts throughout the postmonsoon sampling period i.e. January - June 2010. Eggs from both the oviducts were sub-sampled to about 0.1 - 0.5 g with eggs extracted in three regions (anterior, middle and posterior position) to minimize the variation due to the different stages of development of oocytes. Two or more subsamples were taken from each ovary, and the numbers of eggs in the samples were counted in a large petriplate by adding Gilson fluids with the help of stereo zoom microscope. Gilson fluid was prepared by mixing 10 ml 60% alcohol, 15 ml 80% nitric acid, 18 ml glacial acetic acid and 20 g mercuric acid with 880 ml of distilled water.

The total number of eggs from the known weight of the samples and the full ovary is calculated by the following formula;

$$\text{Fecundity} = (N \times \text{TOW}) / \text{SOW}$$

Where, N = number of eggs in the sub-sample, TOW = Total ovary weight and SOW = sample ovary weight. The mean estimate of the number of eggs obtained in all individuals comprises the fecundity estimate of the species.

Regression analysis was used to estimate the relationship between fecundity (F) and the DML, BW, TOW and nidamental glands weight (NGW).

### 3. Results

Fecundity varied from 180 to 1054 in the individuals of DML of 120 to 196 mm (Table 1). The mean fecundity of *S. lessoniana* estimated from these samples was 497. Larger females have higher potential fecundity.

**Table 1.** Observations of potential fecundity in mature females of *S. lessoniana*.

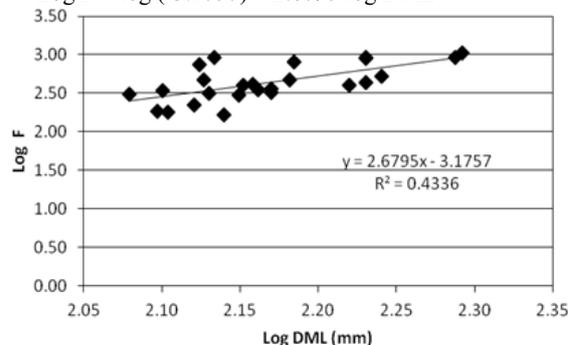
Sl. No.	DML (mm)	BW (g)	TOW (g)	F (Nos)
1	120	118.3	7.45	309
2	125	126.1	1.982	187
3	126	140.8	6.44	342
4	127	143.5	2.185	180
5	132	142.2	5.463	221
6	133	183.0	13.279	744
7	134	175.7	8.24	477
8	135	158.2	8.21	316
9	136	170.1	16.72	912
10	138	166.9	1.41	167
11	141	166.4	3.975	300
12	142	199.3	8.05	403
13	144	200.4	8.15	411
14	145	196.2	5.04	347
15	148	204.6	5.14	328
16	148	240.3	16.9	357
17	152	230.2	8.89	470
18	153	190.7	17.254	805
19	166	270.6	9.88	400
20	170	271.7	10.68	441
21	170	240.4	13.794	904
22	170	270.2	27.2	918
23	174	300.5	13.7	521
24	194	378.5	18.44	918
25	196	406.1	21.9	1054

F, DML, BW, TOW, and NGW were transformed to logarithms (base 10) and by least square method, the following relations were obtained.

#### 3.1. Relation between F and DML

A linear relationship between the F and the DML of the squid existed ( $r = 0.658$ ) (Figure 2) and regression equation is

$$\text{Log F} = \log(-3.1757) + 2.6795 \log \text{DML}$$

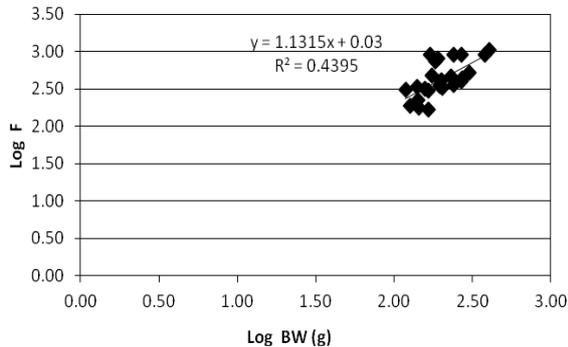


**Figure 2.** Relationship between fecundity (F) and dorsal mantle length (DML)

### 3.2. Relation between F and BW

The relation between the fecundity of squid and the weight of body was linear ( $r = 0.663$ ) and the regression equation is

$$\text{Log F} = \log(0.03) + 1.1315 \log \text{BW} \text{ (Figure 3)}$$

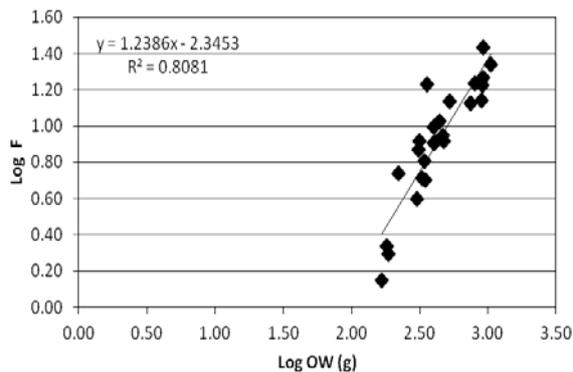


**Figure 3.** Relationship between fecundity (F) and body weight (BW)

### 3.3. Relation between F and OW

The relation between the fecundity of squid and the weight of ovary was linear (Figure 4) and the regression equation is

$$\text{Log F} = \log(-2.3453) + 1.2386 \log \text{OW} \text{ (} r = 0.899 \text{)}$$

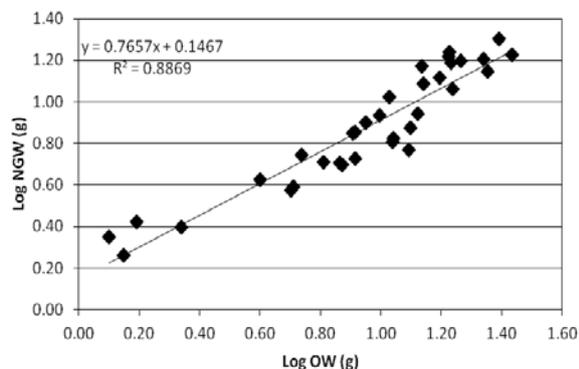


**Figure 4.** Relationship between fecundity (F) and ovary weight (OW)

### 3.4. Relation between NGW and OW

A linear relationship between the NGW and the OW of the squid existed (Figure 5) and regression equation is

$$\text{Log NGW} = \log(0.1467) + 0.7657 \log \text{OW} \text{ (} r = 0.942 \text{)}$$



**Figure 5.** Relationship between nidamental gland weight (NGW) and ovary weight (OW)

## 4. Discussion

Knowledge on the fecundity is extremely important from the viewpoint of a successful management and exploitation of the fishery. The estimated fecundity for female *S. lessoniana* from Mandapam coastal waters ranged between 180 to 1054 for individual of 120-196 mm DML with a mean of 497 eggs. The fecundity values shows wide variations within the same species caught from different area. Fecundity for *S. lessoniana* in the Zanzibar coastal waters ranged from 180 to 1180 eggs for individuals of size range 140–249 mm ML with a mean of 680 eggs (Mhithu *et al.*, 2001) which is higher than the present study. However, *S. lessoniana* of Mandapam water in this study found more fecund than those observed in Sri Lankan's lagoon waters by Sivashanthini *et al.*, (2010). Anon (1975) also estimated a low fecundity of 292 to 754 eggs on the same species in the Philippines and suggested that it may be due to multiple spawning. Tsuchiya (1981) studying in the same species in Okinawa, Japan estimated a lower fecundity of 86 to 728 eggs. However, Segawa (1987) noted the fecundity for the same species from Kominato, Japan ranged between 500 and 1000 eggs in one spawning for individuals of 200 to 250mm DML with a mean of 986 eggs. This lower and higher fecundity of *S. lessoniana* in different places may be due to the difference in short or long spawning period of the individuals in their respective places (Mhithu *et al.*, 2001). The variation in fecundity for the same species may be caused by varying environmental factors, food availability and different habitat (Sivashanthini *et al.*, 2010). In general, fecundity is low in cephalopods because of the absence of larval stage and the hatchlings are virtually miniature adults.

Fecundity increased with DML from 180 to 1054 (Figure 2) suggesting that larger individuals at maturity are likely to contribute more offspring to the next generation than smaller individuals. Nigmatullin and Markaida (2009) studying in jumbo squid *Dosidicus gigas* observed that the fecundity is closely related to female size. He also found that smaller sized female show lower fecundity compared to larger ones. A similar relationship was found in the squids *viz.* *Abraliopsis atlantica* (Laptikhovskiy, 1999), *Loligo vulgaris* (Laptikhovskiy, 2000), *L. duvaucelii* (Sang, 2007) and *S. lessoniana* (Mhithu *et al.*, 2001; Sivashanthini *et al.*, 2010). Different sizes of oocytes were observed in *S. lessoniana*, which means that the oocytes do not mature simultaneously. Lipinski and Underhill (1995) reported that there were always certain proportions of immature oocytes situated in distal portion of squid ovary. This indicates that *S. lessoniana* continue to produce eggs even after onset of spawning. This type of spawning pattern is also found in other loliginid squids, such as *L. vulgaris reynaudii* (Sauer and Lipinski, 1990) and *L. vulgaris* and *L. forbesi* (Rocha and Guerra, 1996). In Multiple spawning strategy, large eggs in the ovary causing the oviduct volume to be insufficient to accommodate all eggs. Hence, number of egg masses must be spawned in several batches (Rocha *et al.*, 2001).

There was high variation in oocyte numbers in each size (Table 1). This may be related to multiple spawning in which somatic growth occurs in between the separate batches of egg laying (Harman *et al.*, 1989) or it could be

due to different growth rates among individuals (Rocha and Guerra, 1996; Jackson *et al.*, 1997) and environmental factors (Pecl *et al.*, 2004).

In the present study the fecundity of *S. lessoniana* bore a logarithmic relationship with the total weight, mantle length and ovary weight. Fecundity exhibited a weak relationship with DML ( $r = 0.658$ ) and BW ( $r = 0.663$ ). A similar weak correlation was also obtained in squids *L. duvaucelii* (Sang, 2007) and *S. lessoniana* (Mhithu *et al.*, 2001; Sivashanthini *et al.*, 2010). These weak correlations could be related to their spawning condition in which some females of similar size have already been laid different numbers of oocytes or multiple spawning strategies. A similar observation has also been reported by Pecl (2001) in *S. lessoniana* and *S. australis* of Australian waters. The correlation values showed that the fecundity is more closely related to the ovary weight and body weight than to the mantle length. The nidamental gland weight of *S. lessoniana* shown a strong logarithmic relationship with the ovary weight ( $r = 0.942$ ) suggesting that the gland is more closely related to the ovary weight. A similar strong relationship between OW and NGW was also found by Nigmatullin and Markaida (2009). Asokan (2000) also observed the positive relationship between OW and NGW. He observed that the size of the nidamental gland increases with one set of maturation and becomes larger in the mature squids. Neethiselvan *et al.* (2001) studying in squid *Doryteuthis sibogae* found that the weight of gonad and nidamental gland increased during maturation and proved as good indices of maturation.

Maturation was size-related process in female *S. lessoniana* individuals, because OW vs. NGW and F vs. OW, BW, DML was highly correlated. Cephalopods produce lower number of eggs both in absolute numbers and per unit biomass and this reduction in numbers is compensated by reduced mortality during early stages of life cycle arising from greater parental care of the eggs and the shorter planktonic stage of the life cycle.

### Acknowledgement

Authors would like to thank the Director, CMFRI and the Dean, Centre of Advanced study in marine biology, Annamalai University for providing physical facilities to carry out this research. Gratitude is expressed to the anonymous referees for their constructive comments on the manuscript. Authors would also like to express their sincere thanks to the editor for publishing this manuscript in the journal in time.

### References

- Anon. 1975. Observation on the culture and life history of the broad finned *Sepioteuthis lessoniana* Fer. and Orb. Mindanao States Univ. *IFRO Ann Rept.*, **76** (19): 115-119.
- Asokan PK. 2000. Biology and fishery of cephalopods (mollusca: cephalopoda) along the Malabar Coast, Zoological Institute, University of Calicut, India. 121pp.
- Boyle PR and Rodhouse PG. 2005. **Cephalopods: Ecology and Fisheries**. Blackwell Science Ltd. Oxford, 452 pp.
- Harman RF, Young RE, Reid SB, Mangold KM, Suzuki T and Hixon RF. 1989. Evidence for multiple spawning in the tropical oceanic squid *Stenoteuthis oualaniensis* (Teuthoidea: Ommastrephidae). *Mar Biol.*, **101**: 513-519.
- Jackson GD, Forsythe JW, Hixon RF and Hanlon RT. 1997. Age, Growth, and maturation of *Lolliguncula brevis* (Cephalopoda: Loliginidae) in the northwestern Gulf of Mexico with a comparison of length-frequency versus statolith age analysis. *Can J Fish Aquat Sci.*, **54**: 2907-2919.
- Jackson G D and Pecl G. 2003. The dynamics of the summer-spawning population of the loliginid squid (*Sepioteuthis australis*) in Tasmania, Australia - a conveyor belt of recruits. *ICES J Mar Sci.*, **60**: 290-296.
- Laptikhovskiy VV. 1999. Fecundity and spawning in squid of families Eupoloteuthidae and Ancyrocheiridae (Cephalopoda: Oegopsida). *Sci Mar.*, **63**(1): 1 - 7.
- Laptikhovskiy VV. 2000. Fecundity of the squid *Loligo vulgaris* Lamarck, 1798 (Myopsida, Loliginidae) off northwest Africa. *Sci Mar.*, **64**(3): 275-278.
- Laptikhovskiy VV and Nigmatullin CM. 2005. Aspects of female reproductive biology of the orange-back squid, *Sthenoteuthis pteropus* (Steenstrup) eastern tropical Atlantic. *Sci Mar*, **69**(3): 383 - 390.
- Lefkaditou E, Peristeraki P, Bekas P, Tserpes G, Politou CY and Petrakis G. 2003. Cephalopods distribution in the Southern Aegean Sea Mediterranean. *Mar Sci.*, **4**(1): 79 - 86.
- Lipinski MR and Underhill LG. 1995. Sexual maturation in squid: Quantum or Continuum?" *S Afr J Mar Sci.*, **15**: 207 - 223.
- Macewicz BJ, Hunter JR, Lo NCH and LaCasella EL. 2004. Fecundity, egg deposition and mortality of market squid (*Loligo opalescens*). *Fish Bull.*, **102**: 306-327.
- Mhithu H, Mgeya Y and Ngoile M. 2001. Western Indian Ocean Marine Science Association, Zanzibar (Tanzania) and Institute of Marine Sciences, Zanzibar (Tanzania) University of Dar es Salaam. In: Richmond MD and Francis J (eds.), **Growth and Reproduction of the Big Fin Squid, *Sepioteuthis lessoniana* in the Coastal Waters of Zanzibar**. IMS/WIOMSA, Zanzibar, Tanzania.
- Nabhitabhata J. 1995. The culture of cephalopods: commercial scale attempt in Thailand. In: Nambiar KPP and Singh T. (eds.), **Aquaculture towards the 21st Century**, INFOFISH, pp. 138-146.
- Nigmatullin CM and Markaida U. 2009. Oocyte development, fecundity and spawning strategy of large sized jumbo squid *Dosidicus gigas* (Oegopsida: Ommastrephinae). *J Mar Biol Ass. U.K.*, **89**(4): 789-801.
- Neethiselvan N, Venkataramani VK and Srikrishnadhas B. 2001. Reproductive biology of the Siboga squid *Doryteuthis sibogae* (Adam) from Thuthookkudi (Tuticorin), south east coast of India. *Indian J Mar Sci.*, **30**(4): 257-260.
- Pecl GT. 2001. Flexible reproductive strategies in tropical and temperate *Sepioteuthis* squids. *Mar Biol.*, **138**: 93-101.
- Pecl GT, Moltschanivskiy NA, Tracey SR and Jordan AR. 2004. Inter-annual plasticity of squid life history and population structure: ecological and management implications. *Oecologia*, **139**: 515-524.
- Rocha F, Guerra A and Gonzalez FA. 2001. A review of reproductive strategies in Cephalopods. *Biol Rev.*, **76**: 291-304.
- Rocha F and Guerra A. 1996. Signs of an extended and intermittent terminal spawning in the squids *Loligo vulgaris* Lamarck and *Loligo forbesi* Steenstrup (Cephalopoda: Loliginidae). *J Exp Mar Biol Ecol.*, **207**: 177-189.
- Salman A and Laptikhovskiy VV. 2005. Fecundity and spawning of *Abralia veranyi* (Ruppell, 1844) (Cephalopoda: Eupoloteuthidae) in the Aegean Sea. *Sci Mar.*, **69**(2): 211 - 214.

- Salman A and Önsoy B. 2004. Analysis of fecundity of some bobtail squids of the genus *Sepiola* (Cephalopoda : Sepiolida) in the Aegean Sea (Eastern Mediterranean). *J Mar Biol Ass U.K.*, **84**: 781-782.
- Salman A and Önsoy B. 2010. Reproductive Biology of the Bobtail Squid *Rossia macrosoma* (Cephalopoda: Sepiolidea) from the Eastern Mediterranean. *Turk J Fish Aquat Sci.*, **10**: 81-86.
- Sang CK. 2007. Reproductive biology and ecology of the loliginid squid, *Loligo duvaucelii* (O rigny, 1835), in Hong Kong waters, Ph D. Thesis, University of Hong Kong. 183pp.
- Sauer WHH and Lipinski MR. 1990. Histological validation of morphological stages of sexual maturity in chokka squid *Loligo vulgaris reynaudii* D' Orb (Cephalopoda: Loliginidae). *S. Afri. J Mar Sci.*, **9**: 189-200.
- Segawa S. 1987. Life history of the oval squid, *Sepioteuthis lessoniana* in Kominato and adjacent waters, Central Honshu, Japan. *J Tokyo Univ Fish.*, **74**:67-105.
- Sivashanthini K, Thulasitha WS and Charles GA. 2010. Reproductive characteristics of squid *Sepioteuthis lessoniana* (Lesson, 1830) from the northern coast of Sri Lanka. *J Fish Aquat Sci.*, **5(1)**: 12-22.
- Tsuchiya M. 1981. On the spawning of the squid, *Sepioteuthis lessoniana* at Amitori Bay, Iriomote Island, Okinawa, Tokai Univ. Notes. *Bull Inst Oceanic Res Develop.*, **3**: 53-75.



# Effect of Dredging on the Macrozoobenthos of Hazratbal Basin in the Dal Lake Srinagar Kashmir, India

Basharat Mushtaq<sup>1,\*</sup>, Rajni Raina<sup>2</sup>, Abdul R. Yousuf<sup>3</sup>, Ashwani Wanganeo<sup>1</sup> and Ummer Rashid<sup>4</sup>

<sup>1</sup>Department of Environmental Sciences and Limnology, Barkatullah University, Bhopal 462026;

<sup>2</sup>Govt Benazeer Collage Bhopal MP ;

<sup>3</sup>Department of Environmental Science/Centre of Research for Development (CORD);

<sup>4</sup>Centre of Research for Development (CORD), University of Kashmir, Srinagar- 190006- India

Received: September 22, 2012; Accepted: October 23, 2012

## Abstract

In order to assess the effect of dredging on macrozoobenthos community, study was carried out in the Hazratbal Basin of Dal Lake from August 2010 to January 2011. The results showed strongly significant differences ( $P < 0.05$ ) between control site and the site which was sampled after dredging operation. The significant differences were also found between pre-dredging and post-dredging ( $P = 0.004$ ), while as, there were no significant differences between control site and pre-dredged site ( $P = 0.191$ ). Altogether 6 taxa were observed at post dredged site in comparison to the pre-dredged (9 taxa) and the control site (10 taxa). At control site, molluscans were found to be the dominant group (1255 ind/m<sup>2</sup>) followed by annelida (1110 ind/m<sup>2</sup>) and diptera (330 ind/m<sup>2</sup>), There were six taxa were shared both sites (control, impact sites) and one taxa *Erpobdella octoculata* of annelida was only restricted to control site. The abundance of most taxa were significantly higher at control and pre-dredged site. Three species (*Lymnea stagnalis*, *Radix auricularia* and *Radix ovata*) were completely eliminated after dredging operation. A cluster analysis showed that macrozoobenthic communities revealed 90% similarity levels between control site and pre-dredged site. After dredging similarity level was greatly declined and was recovered 76% after six months of dredging.

**Keywords:** Macrozoobenthos, dredging effects, Hazratbal Basin, Dal Lake.

## 1. Introduction

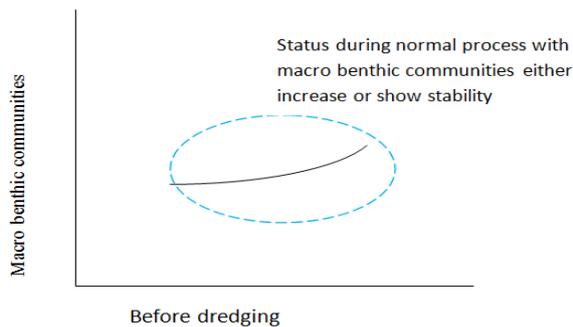
Disturbance in ecological term is an event that has profound impact on ecosystem and its resources (Pickett and White 1985). There are various disturbances which are continuously taking place at different levels in lake ecosystem. The dredging operations are long established human induced disturbances in lakes and rivers and are responsible for making change the environmental features of the water bodies (Pranovi *et al.*, 1998). Previous studies show that species spectrum of aquatic biota including fish, shellfish and benthic invertebrates depict alteration after dredging processes (Taylor and Salomon 1968, Kenny and Rees, 1996; Lewis *et al.*, 2001). In the past different researchers have investigated the impact of dredging on macrozoobenthos and have shown decreased diversity and density in macrobenthic community (Bemvenuti, *et al.*,

2005, Johnson and Nelson, 1985; Palmer, *et al.*, 2008). In addition, studies have pointed out lower diversity indices in the post dredging period in comparison to undisturbed area (Szymelfenig *et al.*, 2006). Considering the important role of the benthic fauna in the secondary production of aquatic systems, any kind of impact upon the macrobenthos may seriously harm the ecology and economy of the aquatic ecosystem, limiting its ability to function as a nursery area (Bemvenuti *et al.*, 2005).

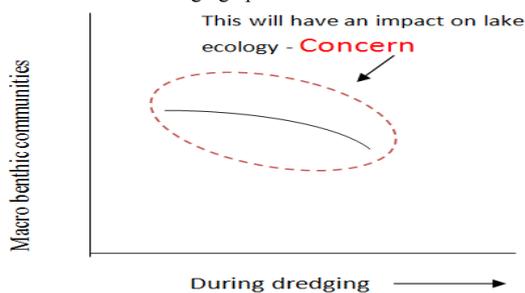
In the last 50 years, there has been drastic changes in urban lakes of Kashmir Himalaya including Dal Lake. Due to cultural eutrophication, the Dal has got silted up and consequently shrunk in size. A lake conservation plan was devised for the Dal Lake in 1980s and as per the plan dredging of the peripheral areas of the Hazratbal, Nishat and Gagribal basins was started in late 1990s' with the aim of improving the aesthetic value of the inshore area.

\* Corresponding author. e-mail: basharatmushtaq@yahoo.com.

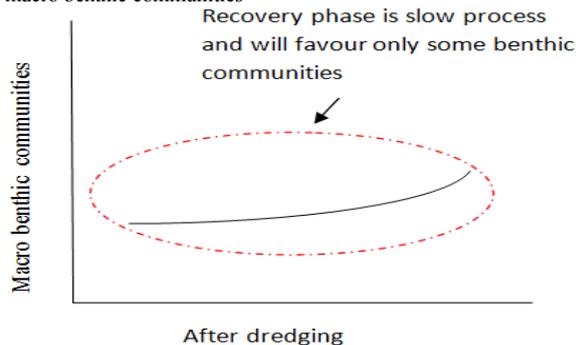
In order to have a first hand information on the impact of dredging on different aquatic communities in this water body, a lake wide study has been initiated in 2010 and shall continue till 2013. In the present communication an attempt has been made to assess the impact of the dredging programme in the lake on the macrobenthic community (important link in the food chain) living on the soft bottom in Hazratbal Basin of the Dal Lake based on the data procured from August, 2010 to January, 2011. We predicted that dredging will decrease the species diversity and density of macrozoobenthos in the areas where dredging operation was carried out. In addition, we assumed that there could be dominance of some species after dredging operation (Figure 1-3).



**Figure 1.** Hypothetical concept showing macro benthic growth before the start of dredging operation



**Figure 2.** Hypothetical concept showing the impact of dredging on macro benthic communities



**Figure 3.** Hypothetical concept showing recovery phase after dredging

## 2. Materials and Methods

### 2.1. Study area

Dal Lake is a Himalayan urban lake, located in the heart of Srinagar (34°18' N latitude and 74°9' 1E longitude) at an average altitude of 1583 m. Dal Lake comprises of four basins viz. Hazratbal Bod-Dal, Gagribal and Nageen. A perennial inflow channel known as Telbal

Nallah enters the lake from the north in its Hazratbal basin. Another inlet, Bot Kol, also joins in the Hazratbal basin, just west to Telbal stream. The water from this catchment area inclusive of water input from a high altitude lake called Marsar Lake besides this the springs present within the lake basin are also a permanent water source to the lake.

### 2.2. Sampling design

In order to assess the impact of dredging operation on macrobenthos, two sites (Figure 4) were selected one in the area which was planned to be dredged (called site 1) and the other in the area which was not to get directly impacted by the dredging (called site 2). Ekman dredge (15 × 15 cm<sup>2</sup>) was used to collect three replicate samples at each site. The unaffected site was sampled on monthly basis for the period August 2010 to January 2011. The site selected for dredging was first sampled before one week of dredging activity and then resampled after 1 day, 3 days, 6 days, 10 days, 20 days, 35 days, 50 days, 70 days, 3 months and 6 months of the dredging activity. The sampling continued till some macrobenthic recovery at the site was observed. In the field, samples were packed in polybags and immediately transferred to laboratory. In the laboratory the samples were carefully washed through 0.55mm mesh screen and preserved in 4% formalin. The sub-samples were stained with Rose Bengal, hand sorted under stereomicroscope and identified by using standard works on macrobenthos (Pennek, 1978; Milligan, 1997; Brinkhurst, 1971; Edmondson, 1959).

### 2.3. Univariate Measures of Community Structure

Univariate measures included: total abundance (N), number of taxa (S), dominance (D), Shannon-Wiener diversity index which was calculated by using base-e logarithms (H'), Simpsons diversity index (1-D), d= Margalf's species richness index and J' = equitability, these measures were calculated by using PAST programme. The significance of differences between before/after dredging and control sites was tested using One-way analysis of variance (ANOVA).

### 2.4. Multivariate analysis of community structure

Non-parametric multivariate techniques were used to measure similarity matrices. The similarity between the sites were calculated by means of Bray-Curtis index by using Bio Diversity Pro Statistical software.

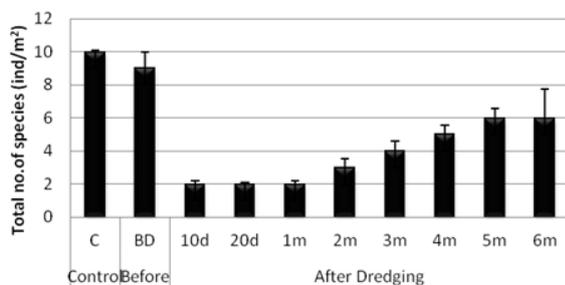


**Figure 4.** Map of Dal lake showing sampling sites in Hazratbal basin.

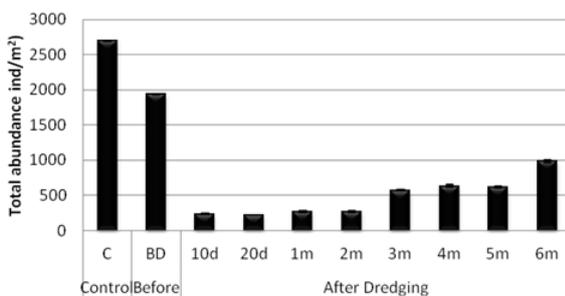
### 3. Results

A total of 10 taxa of macrozoobenthos were recorded at site 2, of which 4 were annelids, 5 molluscs and 1 dipteran. A similar species structure was recorded at site prior to dredging, there being a total of 9 taxa (3 belonged to annelida, 5 to mollusca and 1 to diptera). After the dredging of the site was performed only 6 taxa were observed in the samples from the area (3 belonged to annelida, 2 mollusca and 1 diptera). 3 species of mollusca (*Lymnea stagnalis*, *Radix auricularia* and *Radix ovate*) were completely missing from the area (site 1) after dredging (Table 1).

All the diversity indices were higher at site 2 throughout the study and at site 1 during pre-dredging period in comparison to site 1 during post-dredged period (Table 2). No benthic organism was observed at site 1 during the dredging. This position continued 6 days of dredging. The early colonizers included annelids such as, *Tubifex tubifex* and *Limnodrillus hoffmeisteri* which reappeared in the area after 10th day of dredging. The same trend was observed up to one month after dredging. The total abundance and species richness showed increased recovery rate from 2nd month of dredging and reached to maximum richness on the 5th and 6th month after dredging (Figure 5). As per our observation 9 taxa were found before dredging operation and recovered to the level of 66.6% after 6 month of dredging. It can be presumed that total (100%) recovery will be attained after 1 year. Interestingly, three species of molluscan (*Lymnea stagnalis*, *Radix ovate* and *Lymnea auricularia*) were eliminated during the process of dredging and were not recovered even after 6 months of recovery phase.



**Figure 5.** Graph depicts variation in species number at control, before dredging and decreasing trend after initial stages of dredging and then shows gradual increase in number of species (S) ind/m<sup>2</sup> along with standard errors.

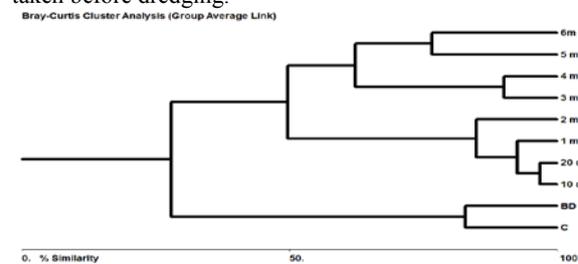


**Figure 6.** Graph depicts variation in total abundance at control site, before dredging and decreasing trend after initial stages of dredging and then shows gradual increase in abundance along with standard errors

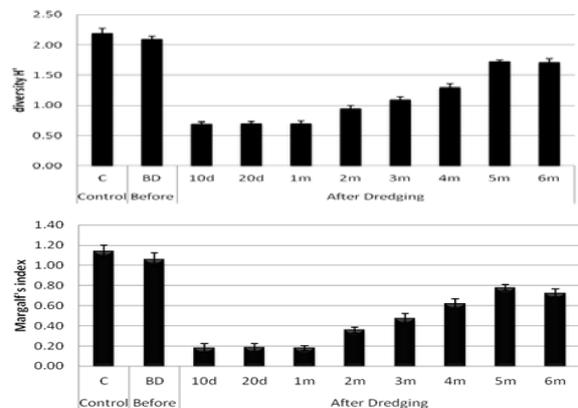
It was observed during present study that the density of some species such as, *Tubifex tubifex*, *Limnodrillus hoffmeisteri* and especially *Chironomus* sp. increased after the dredging process, while, molluscs like *Lymnea stagnalis*, *Radix ovata* and *Lymnea auricularia* were affected by the dredging operation. Interestingly, *Sphaerium* sp. (belonging to Molluscan group) was observed after 4 months of dredging.

Significance of differences between before/after dredging and control sites were tested using One-way analysis of variance (ANOVA) was used to measure the significance of differences between before/after dredging and with control site. The results showed strongly significant ( $P < 0.05$ ) differences between control site and the site which was sampled after dredging operation. The significant differences were also found between pre-dredging and post-dredging ( $P = 0.004$ ), while as, there were no significant differences between control site and pre-dredged site ( $P = 0.191$ ). No significant differences ( $P > 0.05$ ) were found between Margalf's index with diversity ( $H'$ ) and equitability ( $J'$ ).

Cluster analysis revealed that after 5 months of dredging macrozoobenthos fauna showed maximum similarity with samples taken before dredging and with control site. As it is depicted in the dendrogram (Figure 7) that samples taken after 5th and 6th month of dredging were grouped in upper cluster with samples taken at pre-dredged and at the control site showed highest similarity level (76%). However, the similarity level between control site and pre-dredged site was 83%. The middle clusters contains samples which were taken after 10th day to 3rd month of dredging showed less similarity level in terms of macrozoobenthos fauna with control site and samples taken before dredging.



**Figure 7.** Cluster analysis of samplings of control site, before dredging, after 10, 20 days, 1, 2, 3, 4, 5 and 6 months of dredging.



**Figure 8.** Graph depicting the variations in Shannon – Wiener ( $H'$ ) and Margalf's species index (d) of macrozoobenthos community with standard errors during study period in Hazratbal basin of Dal lake.



#### 4. Discussion

The present study revealed significant changes in the macrozoobenthos community of Hazratbal basin of Dal lake with dredging activities. The diversity of macrozoobenthos was found to be greatest at control and pre-dredged site in comparison to the post-dredged site. A similar development after dredging process has been reported by Jamar and Mejuto (1988). The present data reveals that dredging operation leads to significant changes in macrozoobenthos in terms of species composition, density and diversity.

The main reason for reduction of abundance, diversity and species composition in benthos of Hazratbal Basin could be because of the fact that dredging operation has disturb the microhabitats of these communities. The findings of this study fully coincide with the observation of earlier researchers who studied the impacts of dredging on benthic communities (Oliver and Slattery, 1976 and Johnson and Nelson, 1985). We have noted during present study period that after dredging the sediment structure was fine as compared to before dredging, when sediment was mixed with a lot of organic matter and other material like wood debris etc. The fine sediments may become favourable for early colonizers such as *Tubifex tubifex* and *Limnodrillus hoffmeisteri*. These two species also showed increasing trend in species richness and diversity ( $H'$ ) (Table 1.). The colonization of two species of annelid group may be due to transportation of larvae and adults from the water column, and post-settlement movement of juvenile and adult life-stages across the sediment. The results from our study correspond very well with the findings of Guerra-Garcia *et al.* (2003). It has been also reported that earlier colonists generally have similar life history patterns which include similar kind of habitat and high larval availability (Bolam and Rees, 2003).

The data shows that the number of species starts recovering 20 days after dredging, but the diversity ( $H'$ ) and Margalf's index of richness ( $d$ ) continue declining until 2 months of dredging (Figure 6) could be due to strong increase in abundance of some species such as, *Tubifex tubifex* and *Limnodrillus hoffmeisteri*. These species remain in open areas during dredging process where they grow up in large populations and recover immediately after dredging. When areas are depopulated through dredging operations, some opportunistic species usually have a good chance of building up large population in such 'open spaces' (Guerra-Garcia, *et al.*, 2003). This short-term change is reflected by the abundant increase of opportunistic species (Grassie and Sanders, 1973; Lopez-Jamer and Mejuto, 1998; Van Dalssen *et al.*, 2000).

It was observed in the present study that *Tubifex tubifex* is the only species which shows 99% recovery after 6 months of dredging. Similar studies were also reported by Lu and Wu (1998). It was also observed from the present study that *Tubifex tubifex*, *Limnodrillus hoffmeisteri* and *Chironomus* sp. were more abundant after dredging than other taxa.

In connection with the molluscan community, the present study has shown that these communities are more impacted by dredging activity in Hazratbal basin. In addition most taxa of mollusca got eliminated after

dredging. Further it was observed during present investigation that only two taxa such as, *Sphaerium* sp. and *Lymnea collumela* recovered after dredging. The recovery time of macrobenthic communities after this type of disturbance depends on the spatial scale, the hydrodynamic conditions, the bottom grain size and the structure of the community affected by the disturbance (Kaiser and Spencer, 1996; Pranovi *et al.*, 1998).

#### 5. Conclusion

Present study reveals that dredging is responsible for short-term changes in macrozoobenthos community structure which include effects on species richness, diversity and reduction of benthic habitat. The Annelida group was showed rapid recovery followed by diptera and molluscs. In fact, dredging operation initially disturbs the macrozoobenthos of lake, but it should be carried at small-scale in selected areas. The dredging of whole lake bed is dangerous for lake biodiversity. So, the lake authorities should operate dredging operation in lake as per environmental goals and should, however, avoid operating it from engineering point view.

#### References

- Bolam S G and Rees H L. 2003. Minimizing the impacts of maintenance of dredged material disposal in coastal environment: A habitat approach. *Environ Mangement*, **32** (2): 171-188.
- Bemvenuti C E, Angonesi L G and Gandra M S. 2005. Effects of dredging operation on soft bottom Macrofauna in a Harbour in the Patos Lagoon Estuarine region of Southern Brazil. *Braz J Biol.*, **65** (4): 573-581.
- Brinkhurst R O. 1974. **The Benthos of Lakes**. The Macmillan Press Ltd, London.
- Dalssen J A V, Essink K, Toxvig H, Briklund J, Romero, J and Manzanera M. 2000. Differential response of macrozoobenthos of marine sand in the North Sea and Western Mediterranean. *ICES J Mar Sci.*, **57**: 1439-1445.
- Edmonson W T. 1959. **Freshwater Biology**. John Wiley and Sons. New York, London.
- Grassle J S and Sanders H L. 1973. Life histories and role of life of disturbance. *Deep-Sea Res.*, **20**: 643-659.
- Guerra-Garcia J M, Corzo J and Garcia-Gomez C J. 2003. Short-term benthic recolonization after dredging in harbour of Ceuta, North Africa. *Marine Ecol.*, **24** (3): 217-229.
- Jhonson R O and Nelson W G. 1985. Biological effect of dredging in an offshore borrows area. *Biol Sci.*, **48** (3): 166-187.
- Kaiser M J and Spencer B E. 1996. The effects of beam trawl disturbance on infaunal communities in different habitats. *J Anim Ecol.*, **65**: 348-359.
- Kenny A J and Rees H L. 1996. The effect of marine gravel extraction on the macrozoobenthos: results 2 year post dredging. *Mar Poll Bull.*, **32** (8-9): 615-622.
- Kundankar MRD, Sarwar SG and Shah M A. 1995. Limnological characteristics of Hazratbal basin of Dal Lake 1992-93. Technical Report— submitted to Government of Jammu and Kashmir.
- Lewis M A, Weber D E, Stanley R S and Moore J C. 2001. Dredging impact on an urbanized Florida bayou: effects on benthos and algal-periphyton. *Environ Pollut.*, **115**: 161-171.

- Lopez-Jamer E and Mejuto J. 1988. Infaunal benthic recolonization after dredging operation in La Courn Bay, N W Spain. *Cah Biol Mar.*, **29**: 37-49.
- Lu L and Wu R S S. 1998. Recolonization and succession of marine macrobenthos in organic- enriched sediment deposited from fish farms. *Environ. Pollut.*, **101**: 241-251.
- Milligan M R. 1997. Identification Manual for Aquatic Oligochaeta of Florida. Volume I, Florida Department of Environmental Protection.
- Oliver J S and Slattery P N. 1976. Effect of dredging and disposal on some benthos at Monterey Bay, California, U. S. Army, Corp of Engineers, Coastal Engineering Research Center, Tech. Paper No, 76-15.
- Palmer T A, Montagna P A and Nairn R B. 2008. The Effects of a Dredge Excavation Pit on Benthic Macrofauna in Offshore Louisiana. *Environ Management.* **41**: 573-583.
- Pennak W R. 1978. **Freshwater Invertebrates of United States.** Willy Interscience Publishing New York
- Pickett S T A and White P S. 1985. **The Ecology of Natural Disturbance and Patch Dynamics.** Academic Press, London.
- Pranovi F, Giovanardi O and Franceschini G. 1998. Recolonization dynamics in areas distributed by bottom fishing gears. *Hydrobiologia*, **375/376**: 125-135.
- Qadri M Y and Yousuf A R. 1980. Limnological studies on Lake Malpur Sar. 1. The Biotope. *Geobios*, **7**: 117-119.
- Sparks – McCronky PJ and Watling L. 2001. Effects of ecological integrity of a soft bottom habitat from trawling disturbance. *Hydrobiologia*, **456**: 1-3.
- Szymelfenig M, Kotwicki L and Graca B. 2006. Benthic recolonization in post-dredging pits in the Puck Bay (Southern Baltic Sea). *Estuar Coast Shelf Sci.* **68**: 489-498.
- Taylor J L and Saloman CH. 1968. Some of hydraulic dredging in coastal development in San Boca Ciago Bay, Florida. *Bull U S Fish Wild Serv Fish*, **7(2)**: 205-241.

# Evaluation of the Physicochemical Properties and Antimicrobial Activities of Bioactive Biodegradable Films

Mary S. Khalil<sup>1</sup>, Zahra S. Ahmed<sup>2</sup> and Aml S. Elnawawy<sup>3,\*</sup>

<sup>1</sup> Department of Botany, Faculty of Science, Cairo University,

<sup>2</sup> Department of Food Sciences & Nutrition, National Research Centre (NRC), Dokki,

<sup>3</sup> Department of Food Engineering and Packaging, Food Technology Research Institute, Agricultural Research Center (ARC), Giza, Egypt.

Received: July 28, 2012; Accepted: November 10, 2012

## Abstract

The use of edible films to release antimicrobial constituents in food packaging is a form of active packaging. Different extractions of *Myrtus communis*, *Urtica urens*, *Ziziphus spina-christi*, and *Zygophyllum coccineum* were tested for their antimicrobial activity against the food pathogenic microorganisms *Escherichia coli*, *Salmonella typhimurium* and *Aspergillus niger* by using agar diffusion assay method. Soy-starch and gelatin edible films were prepared and incorporated with *Myrtus communis* and *Ziziphus spina-christi* essential oils separately and as a mixture in different concentrations. The films were characterized for their antimicrobial activity by using agar diffusion assay method and their physico-chemical properties. The films were studied on different food applications (orange, apple, lemon, tomato, pizza dough, chicken salami, meat salami, artificial cheese, mayonnaise, yoghurt and skimmed cheese). The results showed that, the films extended the shelf-life of the food products depending on the effective chemical compounds of the essential oils  $\alpha$ -pinene and limonene.

**Key Words:** *M. communis*, *Z. spina-christi*, essential oils,  $\alpha$ -pinene, limonene, edible films.

## 1. Introduction

Researches on the microbial spoilage of food has become important for food safety and keeping qualities. In food industry, all the steps of food production usually occur under sterilized condition. But at the final step where the food packaged, it usually exposes to post process surface contamination, which leading to the reduction of shelf life.

At the same way, the using of extracts from aromatic plants particularly the essential oils as antimicrobial agents are in an increasing interest (Shahidi Bonjar *et al.*, 2003), because there were considered as a rich source of biologically active compounds. They have been shown to possess antibacterial, antifungal, antiviral, insecticidal and antioxidant properties (Burt 2004, Kordali *et al.*, 2005) and have a wide range of possible applications ranging from the food industry to pharmaceuticals (Burt 2004, Holley and Patel, 2005).

There are many techniques that have been used for maintaining the quality of food products and in the recent years the edible films and coatings have received considerable amount of attention as antimicrobial

packaging materials. The main advantage is that they can be consumed with the packaged products and even if the films are not consumed they could still help in the reduction of environmental pollution (Bourtoom, 2008).

Various antimicrobial edible films can serve as a carrier for antimicrobial compounds to reduce, inhibit or retard the growth of the food pathogenic microorganisms in packed foods and packaging material (Coma *et al.*, 2001, Rodrigues and Han, 2000). The film can helps preventing brown coloration, moisture loss during storage, reducing the rate of rancidity causing lipid oxidation and also restrict of the volatile flavor loss (Pérez-Pérez *et al.*, 2006).

In general; edible films have been made from several polysaccharides, lipids, and proteins (Cagri *et al.*, 2004). Several attempts have been made in developing active packaging systems in which antimicrobial agents are incorporated into the polymeric material and are slowly released on the food surface (Devlieghere *et al.*, 2004) by diffusion through partitions (Han, 2000). Finally, there is a need to explain the advantages of using edible films as an antimicrobial food packaging materials to the consumers in order to help the industry to replace the synthetic packaging materials with the environmental friendliness biomaterials (Sonti, 2003). This study aims at

\* Corresponding author. e-mail: amalalnawawy@yahoo.com.

incorporating the naturally-derived essential oils with the edible films to use them as antimicrobial food packaging materials to extend the food shelf-life.

## 2. Materials and Method

### 2.1. Plant Materials.

Both leaves and fruits of *Myrtus communis* and *Ziziphus spina-christi* were collected from Cairo arboretum; *Zygophyllum coccineum* was collected from Sinai (ras-sydr); and *Urtica urens* was collected from El – menofeya, while its seeds were purchased from Local herbal shop in Cairo. All the specimens were identified in the Herbaria of Botany Cairo University (CAI) and Ain-Shams University (CAIA).

A food grade soy protein (prolia) with a protein content of 55% was donated by "AWA" for food additives. Gelatin powder (animal gelatin) was purchased from a local herbal shop, bloom = (180-200) g. Corn starch was purchased from a local shop. Glycerol was obtained from Technogene Company, Dokki, Giza, Egypt. Ethanol was purchased from Sigma-Aldrich, Egypt. 1N sodium hydroxide purchased from El-gomhorea Company, Cairo, Egypt.

### 2.2. Organisms and Preparation of Cultures

The food deterioration microorganisms *Aspergillus niger*, *Escherichia coli* and *Salmonella typhimurium* were used as test microorganisms.

Both *Aspergillus niger* and *Escherichia coli* were obtained from Department of Microbiology, Faculty of Science, Ain-shams University. The fungal and bacterial species were received on PDA medium and nutrient agar medium respectively. They were maintained on agar slants of both Czapek's Dox and nutrient media, respectively.

*Salmonella typhimurium* kindly provided from the Veterinary Serum & Vaccine Research Institute. The bacterial species was received on nutrient agar medium, and then cultured on *Salmonella – Shigella* agar slants.

Czapek's Dox agar and Nutrient agar media prepared according to the "Hand book of microbiological media" (Atlas, 1979). SS agar was obtained from Oxoid LTD Company, Basinocoke, Hampshire, England.

### 2.3. Preparation of Plant Extracts

Thirty six extracts of *Myrtus communis*, *Urtica urens*, *Ziziphus spina-christi* and *Zygophyllum coccineum* were used in this study. All the plants extractions (whether of air-dried or fresh parts or expressed Juice) prepared according to (Abo-Zaid, 2000), then stored at 4°C until use them in the experiments (not more than a week).

### 2.4. Extraction of Essential Oils

*Myrtus communis* and *Ziziphus spina-christi* were used for extraction of essential oils. According to (Abo-Zaid, 2000), 100 g of the air-dried leaves of both plants were crushed, then hydrodistilled for 3 hours. The yields were on average 0.5 % (v/w dried weight) and 0.01 % respectively. The essential oils were stored in dark vials at 4 °C. (The trials of the essential oils extraction from both *Urtica urens* and *Zygophyllum coccineum* through hydrodistillation method were failed).

For preparation of different concentrations of the essential oils mixture; equal volumes of the essential oils of both *Myrtus communis* and *Ziziphus spina-christi* were mixed in a dark vial and kept at room temperature for 24 hours, then stored at 4 °C (shake well before it using). 0.1, 0.2, 0.3, 0.4 and 0.5 ml of the mixture then used.

### 2.5. Edible Films Preparation

Two different edible films were prepared, one of them from soy-starch and the other from gelatin.

#### 2.5.1. Control Films

Film-forming solution of soy-starch prepared by using a modified method to that described by (Ghorpade *et al.*, 1995). While, film-forming solution of gelatin prepared according to the method that described by (de Carvalho and Grosso, 2006).

Each film-forming solution were spread in a rectangular area of (11.5 ×32.5 cm).

#### 2.5.2. Films Enriched with the Essential Oils

Films-forming solutions incorporated with the essential oils prepared by using a modified method to that described by (Maizura *et al.*, 2008) and by the same steps of preparation of control films.

Seven films sheets prepared for both of soy-starch and gelatin. A sheet containing 0.5 ml *Myrtus communis* essential oil, a sheet containing 0.5 ml *Ziziphus spina-christi* essential oil. And the other five sheets each one contained different concentration of the essential oils mixture (0.1, 0.2, 0.3, 0.4 and 0.5 ml).

### 2.6. Antimicrobial Activity

Screening of Antibacterial and antifungal activity for plants extractions, the essential oils and the edible films carried out by agar disc diffusion method.

#### 2.6.1. For plant extractions

0.2 ml of suspension of tested microorganisms spread on the solid media surfaces. Sterilized filter paper discs (5 mm in diameter) saturated with plant extractions, left to dry at room temperature for an hour. Then placed on the cultures surfaces which previously inoculated. Control cultures contained only on sterile filter paper discs.

#### 2.6.2. For Essential Oils

Sterilized filter paper discs (5 mm in diameter) were saturated with the essential oils, left to dry at room temperature for 30 minutes. Then placed on cultures surfaces which previously seeded with 0.2 ml of the inoculum. Control cultures contained only on sterile filter paper discs.

#### 2.6.3. For Edible Films

Each film sheet cut into discs (13 mm in diameter), then placed on cultures surfaces which previously seeded with 0.2 ml of the inoculum.

All the plates incubated at 30° C for 48 h for bacteria and 6 days for fungus. Diameters of inhibition zones measured in millimeters. All the tests performed in triplicates, and the results analyzed statistically using ANOVA test (Schott, 1997).

## 2.7. Physicochemical Properties of Films

**Color.** Films Color of soy-starch and gelatin examined by using Kodak camera (dimensions of photo 862 × 962) to show the color of each film.

**Thickness.** The thickness value represented by the mean of five measurements taken along the strips made on each film which used for testing tensile strength and percentage elongation at break. The films thickness measured automatically by a micrometer connected to the Universal Testing Instrument (Zwick \ Z010). The test was carried out at the National Institute for Standards.

**Solubility.** Different pH solutions (1-14 pH) were prepared by using the distilled water and solutions of HCl-KCl Buffer (1-2.2), Citrate-Phosphate Buffer (2.6-7) and Glycine-NaOH Buffer (7.4-14). Small pieces of dried films samples (3 × 4cm) were placed in Petri dishes containing 60 ml of adjusted pH solution. The Test dishes were covered and incubated at room temperature for 48 hrs.

**Water vapor permeability (WVP).** Carried out by using a modified method to that described by (Kunte *et al.*, 1997). Fan was provided for the air circulation inside the desiccator cabinets at the first for 4 hours only after that, the test completed without it. All the tests were performed in triplicates.

**Tensile strength (TS) and percent elongation (%E).** According to (Kunte *et al.*, 1997), tensile testing was performed with the Universal Testing Instrument (Zwick \ Z010) on (50 × 4 cm) dample shape film strips. Initial grip separation set at 25 mm, while cross-head speed set at 50 mm/sec, the used lot cells (100 N). The test was carried out at the National Institute for Standards.

**Scanning electron microscopy (SEM).** Carried out for both films of soy-starch and gelatin by using SEM Model Philips XL 30 attached with EDX unit with accelerating voltage (15 K.V.). Samples were coated with gold and the films surfaces pictures taken by SEM at magnification of (500x).

### Qualitative analyses by (GC-MS)

Glass jars were tightly sealed with film specimens (8 × 8 cm) of soy-starch film and gelatin film respectively. Film specimens cut from films sheets prepared in rectangular area of (11.5 × 32.5 cm) and enriched with 0.5ml of the essential oils mixture. All the samples were left at room temperature (27°C±2°C) for 7 days.

The air inside the jars examined by Gas Chromatography-Mass Spectrometer (from Agilent Technologies; 6890N, network GC system and 5975 inert XL Mass Selective Detector). It was carried out at the National Institute for Standards. The test performed in duplicate.

## 2.8. Applications on Foodstuffs

**Films Preparation.** The films-forming solutions for soy-starch and gelatin were prepared by the same method which shown before in the preparation of edible films that enriched with the essential oils. For each film-forming solution, 0.5ml from the essential oils mixture was added then spread in rectangular area of (11.5 × 32.5 cm) on foil sheet.

**Preparation of food samples.** Different selected food products were covered with the edible films, each one with

film area of (8 × 8 cm). Fruits (orange - apple - lemon), Vegetables (tomato), Bakery products (pizza dough), Meats products (chicken salami- meat salami) and Dairy products (artificial cheese) were directly attached with the films. Where, Ready meal (mayonnaise) and Dairy products (yoghurt-skimmed cheese) were packaged in glass jars and tightly sealed with films.

For control specimens, foil sheets free from films were used. The results were recorded daily during 100 days for odor changes and physical observation.

## 3. Results and Discussion

Four plants have reported in previous studies as medicinal plants were chosen; *Myrtus communis* (Montvale, 2000 and Tsybula and Kazarinova, 1996), *Ziziphus spina-christi* (Glombitza *et al.*, 1994 and Waggas, 2007), *Zygophyllum coccineum* (Batanouny *et al.*, 1999) and *Urtica urens* (Wichtl, 2002 and Randall, 2003) to be safe in using them as food additives. The antibacterial and antifungal activity for all plants extractions gave interesting results. Whereas *Myrtus communis* and *Ziziphus spina-christi* were the best of them (Table 1).

**Table 1.** Antimicrobial activities of plants extractions of *Myrtus communis*, *Urtica urens*, *Ziziphus spina-christi*, and *Zygophyllum coccineum* against the foodborne pathogens, *E. coli*, *S. typhimurium* and *Aspergillus niger*

plants species	Used part	Extraction solvent	inhibition zones diameter(mm)			
			<i>E. coli</i>	<i>S. typhimurium</i>	<i>A. niger</i>	
<i>M. communis</i>	Dry Leaves	Distilled Water	10	12	9	
		Alcohol	9	-	10	
		Water + Alcohol	-	11	14	
	Fresh Leaves	Distilled Water	9	9	-	
		Alcohol	9	10	-	
		Water + Alcohol	9	12	12	
	Dry Fruit	Distilled Water	11	15	-	
	<i>U. urens</i>	Fresh Leaves	Distilled Water	-	-	-
			Alcohol	11	9	-
fresh (Leaves+ Flowers+ Stems)		Distilled Water	6	-	-	
		Alcohol	-	7	-	
Fresh Root		Distilled Water	9	-	-	
		Alcohol	6	-	6	
Dry Leaves		Distilled Water	-	-	-	
		Alcohol	-	-	7	
Dry (Leaves+ Flowers+ Stems)		Distilled Water	-	-	7	
	Alcohol	8	-	7		
Dry Root	Distilled Water	-	-	8		
	Alcohol	11	6	10		
Dry Seeds	Distilled Water	-	-	6		
	Alcohol	7	-	6		

	Expressed Juice	Fresh Intact Plant	-	-	10
<i>Z. spina-christi</i>	Dry Leaves	Distilled Water	-	10	-
		Alcohol	-	-	6
		Water + Alcohol	-	9	7
	Fresh Leaves	Distilled Water	-	7	7
		Alcohol	-	10	7
		Water + Alcohol	-	9	7
	Dry Fruit	Distilled Water	9	-	7
	Fresh Fruit	Distilled Water	8	10	-
	Seed Embryo	Distilled Water	-	-	-
	<i>Z. coccineum</i>	Fresh Leaves	Distilled Water	-	-
Alcohol			7	-	-
Fresh Leaves and Stems		Distilled Water	9	-	-
		Alcohol	8	8	-
Expressed Juice		Fresh Intact Plant	9	8	-

Each value is the mean of three replicates.

Control Cultures containing only sterile filter paper discs & did not show any inhibition zones. Filter paper discs diameter of 5 mm.

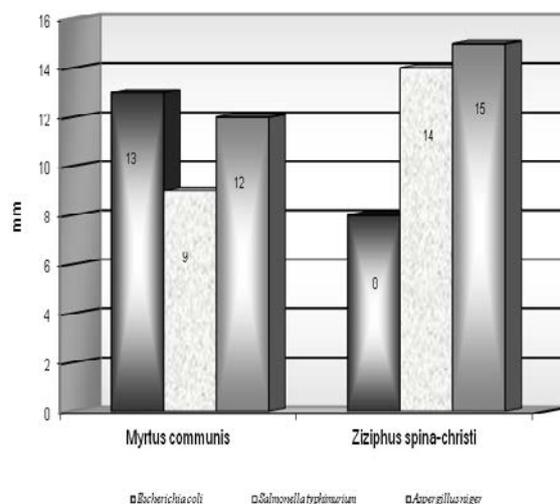
The results are not significant ( $p > 0.05$ ) according to ANOVA test.

The mode of action was attributed to the disturbance of the cytoplasmic membrane, disrupting of the proton motive force, electrolyte flow and active transport and coagulation of bacterial cell contents (Burt, 2004). Whereas, the essential oils components in plants extractions may have an inhibitory effect on the mycelial growth of fungi (Özcan *et al.*, 2005).

The extraction of the essential oils from *Urtica urens* and *Zygophyllum coccineum* through hydrodistillation method was failed. Therefore, only the essential oils of *Myrtus communis* and *Ziziphus spina-christi* were used. The method of extraction (Lemberkovics *et al.*, 2003) and the origin of the samples (Tuberoso *et al.*, 2006) have effect on the composition of essential oils in aromatic plants.

According to (Tuberoso *et al.*, 2006 and Montvale, 2000), the essential oil yields of *Myrtus communis* were on average 0.5 for the dried leaves and 0.02 for berries. Therefore, the dried leaves were used for the extraction of the essential oil. Although *Myrtus communis* fragrance clearly appeared in its extracts; but the antimicrobial activity for its essential oil against all the tested microorganisms appeared more effective than the extracts (Figure1).

The essential oil of *Ziziphus spina-christi* showed more potent antimicrobial activity against all the tested microorganisms comparing with all the plant extractions (Figure1).



**Figure 1.** The antimicrobial activity of *M. communis* and *Z. spina-christi* essential oils against the foodborne pathogens *E. coli*, *S. typhimurium* and *A. niger*.

Each value represents the mean of three replicates. Filter paper discs diameter of 5 mm.

Control cultures contained only on sterile filter paper discs & did not show any inhibition zone.

The results are not significant ( $p > 0.05$ ) according to ANOVA test.

This can be attributed to the absence of sugars that present in its extractions and include lactose, glucose, galactose, arabinose, xylose and rhamnose (Dweck, 2005). These sugars can be considered as a source of nutrient for the microorganisms.

The results of these experiments emphasize the work of Gill and Holley (2006) which concluded that, "at bactericidal concentrations of the essential oils" the bacterial-cytoplasmic membrane disrupted by increasing its non-specific permeability because the essential oils components may possess ATPase inhibiting activity. Also there were concluded that, the other secondary effects at sublethal concentrations cannot be discounted and can be expected as a consequence of membrane interactions. By the same way and according to Özcan *et al.* (2005), the essential oils has inhibitory effect on the mycelial growth of fungi. Whereas Curini *et al.* (2003) found that, the essential oil of *Myrtus communis* causing morphological alterations of fungal hyphae.

So it is obvious that, the plants extractions and essential oils have remarkable lethal effects on the tested microorganisms *Escherichia coli*, *Salmonella typhimurium* and *Aspergillus niger* by inhibiting their survival remarkably.

The results of the antimicrobial activity of edible films which incorporated with the essential oils of *Myrtus communis* and *Ziziphus spina-christi* whether individually (Table 4) or as a mixture (Table 5) demonstrated wide variation in activities against the tested microorganisms.

**Table 4.** Antimicrobial activity of the films incorporated with *M. communis* and *Z. spina-christi* Essential Oils (separately) on *E. coli*, *S. typhimurium* and *A. niger* by the disc diffusion method.

Film	Essential Oil	Diameter of inhibition Zones (mm)		
		<i>E. coli</i>	<i>S. typhimurium</i>	<i>A. niger</i>
Soy-Starch	Control	20	20	21
	<i>M. communis</i>	21	22	22
	<i>Z. spina-christi</i>	20	21	25
Gelatin	Control	18	–	20
	<i>M. communis</i>	21	25	32
	<i>Z. spina-christi</i>	18	22	33

Each value represents the mean of three replicates. Films discs diameter of 13 mm.  
Each prepared film sheet (11.5 × 32.5 cm) containing 0.5 ml of an essential oil.

Control films prepared without adding any of Essential oils. The results are significant ( $p > 0.01$ ) according to ANOVA test.

**Table 5.** Antimicrobial activity of the films incorporated with different concentrations of the essential oils mixture on *E. coli*, *S. typhimurium* and *A. niger* by the disc diffusion method.

Film	essential oils mixture Conc. (ml)	Diameter of inhibition Zones (mm)		
		<i>E. coli</i>	<i>S. typhimurium</i>	<i>A. niger</i>
Soy-Starch	0.1	28	20	30
	0.2	28	23	32
	0.3	30	20	31
	0.4	25	22	30
	0.5	26	20	25
Gelatin	0.1	20	20	44
	0.2	20	18	31
	0.3	20	20	33
	0.4	27	25	30
	0.5	27	21	30

Each value represents the mean of three replicates.

Films discs diameter of 13 mm.

Each prepared film sheet (11.5 × 32.5 cm) containing different concentration of essential oil mixture.

The results of both soy-starch and gelatin are highly significant ( $p < 0.01$ ) according to ANOVA test.

The films materials come from biological tissues; soy (protein) and starch (carbohydrate) from plants, and gelatin (protein) from animal tissue. Therefore, may be some of the chemicals residues which come from their origin attached to the powders (Pérez-Pérez *et al.*, 2006). This may be explaining the antimicrobial activity of the control films.

There is no single antimicrobial agent can cover all the requirements for food preservation. Therefore, the antimicrobial activity of the essential oils mixture of *Myrtus communis* and *Ziziphus spina-christi* were examined. And according to the presented results, the minimum inhibitory concentration not detectable and it cannot be unified to all the microorganisms (Table 5). This is because of the potential differences of the effect of the essential oils on their cell wall (Gill and Holley, 2006).

Differences between both films were appeared through the physicochemical tests.

Color is an important property because it could affect consumer acceptance of such films for both edible and nonedible packaging applications. Differences observed clearly between both films (Figure 2).

**Figure 2.** Photo of gelatin and soy -starch films.

This is attributed to the used powders color of both soy protein (yellow) and gelatin (white). The Soy protein consists of two major protein fractions referred to as the 7S (conglycinin) and 11S (glycinin). Each fraction has a quaternary (subunit) structure (Kinsella *et al.*, 1985) and make up to 37 and 31%, respectively of the total extractable proteins (Gennadios *et al.*, 1994). Both 7S and 11S contain cysteine residues (Sulfur-containing amino acid) (Kinsella *et al.*, 1985). For that soy powder has the yellow color which became faint with adding of starch. While gelatin contain on high content of the amino acids glycine, proline and hydroxyproline which are free of sulfur.

The films thicknesses were measured automatically by a micrometer connected to the Universal Testing Instrument (Zwick \ Z10). There were 0.1272 mm and 0.1704 mm for soy-starch and gelatin, respectively

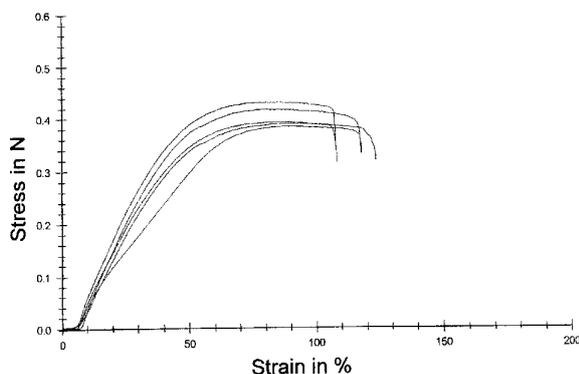
Although plasticizers are hydrophilic substances causing an increase in the films solubility (Gennadios *et al.*, 1994). It must be added to the film-forming solutions. Because it decreasing the accumulation of intermolecular forces along polymer chains. Thereby "softening" film structure, decreasing tensile strength and increasing elongation (Mellan, 1961). Whereas, films made without plasticizer were extremely brittle and shattered (Brandenburg *et al.*, 1993).

Polymers which contain groups that can associate through hydrogen or ionic bonding causing the susceptibility of films to absorb moisture (Salame, 1986).

Therefore, both gelatin and soy-starch films are susceptible to moisture absorption. All the film pieces of gelatin were completely dissolved in all the pH solutions. While, not all soy-starch film pieces dissolved. The film pieces that immersed in the pH values 1, 9 and 13 were remained stable; this result confirmed by Gennadios *et al.* (1993). So, the soy-starch film can withstand the highly acidic and highly alkaline solutions.

Also, the edible films that produced from polysaccharides and proteins were showed limited resistance to moisture transmission. This is due to the inherent hydrophilicity of the film-forming substances and to the considerable amount of hydrophilic plasticizers that incorporated into the films to ensure formation of free-standing films (Guilbert *et al.*, 1996 and Callegarin *et al.*, 1997). Therefore, both films were showing limited water vapor permeability.

Tensile strength (TS), elongation (E %) and the elastic modulus (E- Modulus), are measurements helps in the description of the mechanical properties for the films and the relation with their chemical structures (Ninnemann, 1968). Tensile strength is an important mechanical property, which expresses the maximum stress which developed on the film during the tensile testing (Briston, 1988). Whereas the elongation, is the ability of the film to stretch; it determined at the point where the film breaks under tensile testing. And It expresses as, the percentage of change of the original length of the specimen between the grips of the testing machine (Briston, 1988). While the elastic modulus, is the mathematical description of an object or substance's tendency to be deformed elastically (i.e., non-permanently) when a force is applied to it (Hartsuijker and Welleman, 2001). However, soy-starch film showed lower TS, lower E% and lower E- Modulus than gelatin film (Figure 3, Table2 and Figure 4, Table 3 respectively).



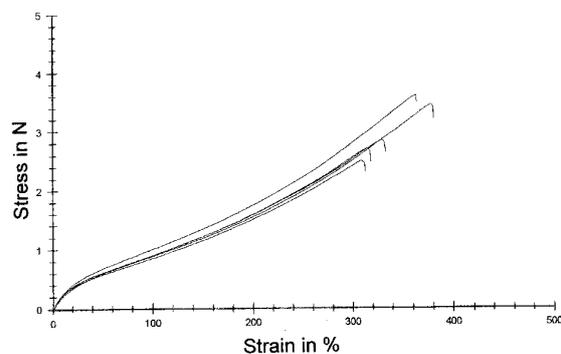
**Figure 3.** Tensile strength (TS), percent elongation (%E) and elastic modulus (E- Modulus) of Soy-starch film  
Test standard: ASTM D638-02, Load cell: 100N.  
Thickness of protein-carbohydrate film = 0.1272 mm

**Table 2.** Statistics of TS and %E of Soy-starch film.

n=5	E- Modulus MPa	RM MPa	$\epsilon$ F max %	RB MPa	$\epsilon$ Break %
<b>X</b>	1.94	0.87	81.78	0.71	110.79
<b>S</b>	0.44	0.08	6.38	0.07	6.81
<b>V</b>	22.68	9.59	7.80	9.34	6.15

**X:** the mean of five measurements. **S:** Standard deviation.

**V:** Poisson's ratio.



**Figure 4.** Tensile strength (TS), percent elongation (%E) and elastic modulus (E- Modulus) of gelatin film.

Test standard: ASTM D638-02, Load cell: 100N.

Thickness of protein film = 0.1704 mm

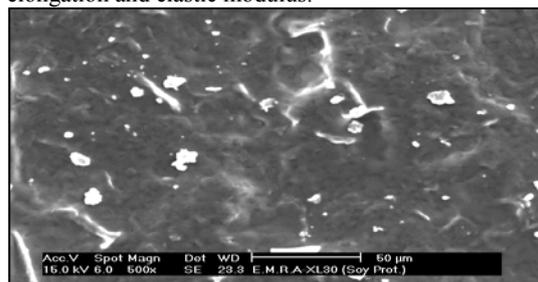
**Table 3.** Statistics of TS and %E of gelatin film.

n=5	E- Modulus MPa	RM MPa	$\epsilon$ F max %	RB MPa	$\epsilon$ Break %
<b>X</b>	4.08	5.33	338.49	4.98	341.49
<b>S</b>	0.75	0.81	30.31	0.88	29.33
<b>V</b>	18.28	15.23	8.95	17.76	8.59

**X:** the mean of five measurements. **S:** Standard deviation.

**V:** Poisson's ratio.

Those results can be explained as follow; in case of gelatin film. The amino acids chains rearranged with the help of glycerol (plasticizer) during drying the film (Mellan, 1961). This led to the formation of a uniform surface structure (Figure 6). Whereas in case of the soy-starch film, also glycerol helped in the rearrangement of amino acids chains for soy protein. But, although the addition of starch improved the mechanical properties of the film (comparing with a film was prepared free from starch). Addition of starch caused the formation of uneven surface (Figure 5) which affected the film tensile strength, elongation and elastic modulus.



**Figure 5.** SEM of soy-starch film surface.

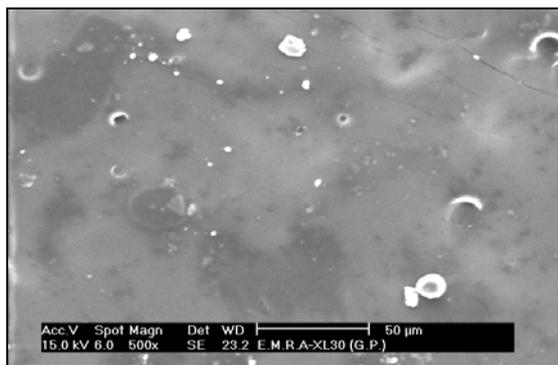


Figure 6. SEM of gelatin film surface.

The results that detected quantitatively by GC-MS showed the presence of  $\alpha$ -pinene in soy-starch film, while  $\alpha$ -pinene and limonene in gelatin (Figure 7 and Figure 8, respectively).

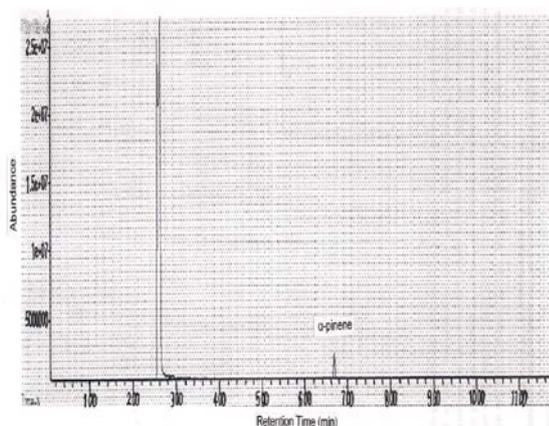


Figure 7. Chromatogram of GC-MS for soy-starch film.

Temp. :  $22 \pm 2^\circ\text{C}$  RH:  $43 \pm 5\%$

Area for  $\alpha$ -pinene = -2.76 (Representing the amount in the area of film specimen of  $(8 \times 8 \text{ cm})$ ).

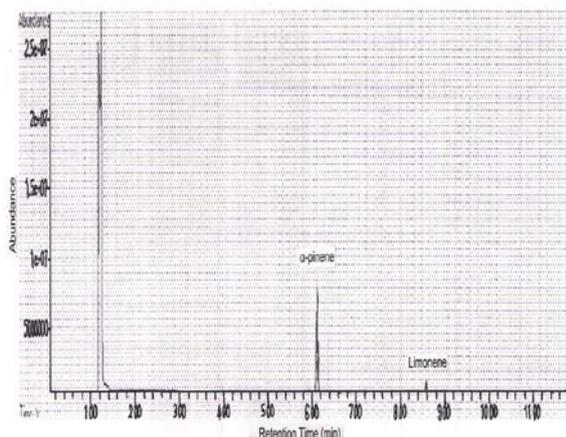


Figure 8. Chromatogram of GC-MS for gelatin film.

Temp. :  $22 \pm 2^\circ\text{C}$  RH:  $43 \pm 5\%$

Area for  $\alpha$ -pinene = -11.4 & for limonene = -0.83 (Representing the amount for both of them in the area of film specimen of  $(8 \times 8 \text{ cm})$ )

This test proved that, the antimicrobial activity of the films which directly attached to the food products was due to the existence of the essential oils components in the films. Also, the antimicrobial activity of the films which covered the jars was due to the liberalized essential oils components from the films to the space above the surface of the packaged food (not packaged under vacuum condition).

$\alpha$ -pinene is an organic compound of the terpene class, one of two isomers of pinene (Figure 9).

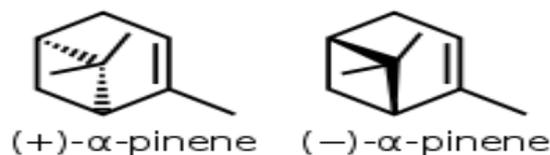


Figure 9.  $\alpha$ -pinene structure

(1S, 5S)-2, 6, 6-Trimethyl bicycle [3.1.1] hept-2-ene ((-)- $\alpha$ -Pinene).

It is an alkane, contains a reactive four-membered ring and of melting point  $64^\circ\text{C}$ . It is found in the oils of many plant species (Simonsen, 1957). The four-membered ring makes it a reactive hydrocarbon (Richter, 1945); therefore,  $\alpha$ -pinene easily attached to both soy-starch and gelatin amino acids.

Limonene is a hydrocarbon, classified as a cyclic terpene and is a chiral molecule in which biological sources produce one enantiomer (Simonsen, 1947) (Figure 10).

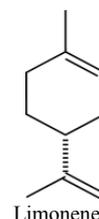


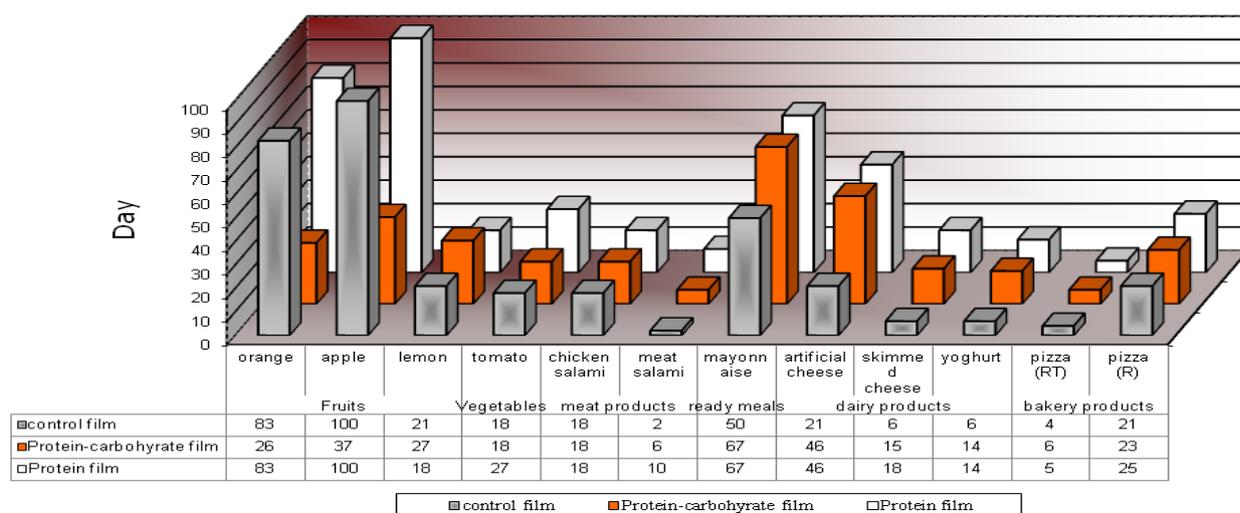
Figure 10. Limonene structure

1-methyl-4-prop-1-en-2-yl-cyclohexene (Racemic: DL-limonene).

It is a relatively stable terpene, which can be distilled without decomposition, although at elevated temperatures it cracks to form isoprene (Pakdela *et al.*, 2001). It is considered by some researchers to be a significant chemopreventive agent (Crowell, 1999).

Gelatin structure contains on high content of amino acids glycine, proline and hydroxyproline (Bourtoom, 2008). While, it contains on many of glycine (almost 1 in 3 residues arranged every third residue), proline and 4-hydroxyproline residues (Chaplin, 2009). So, the availability of free hydrogen bond in glycine amino acid in gelatin skeleton permit the binding of limonene through the second C=C double bond with gelatin. This explains the presence of limonene only in gelatin film.

The used food products showed different susceptibility towards both films of control and those enriched with the essential oils mixture (Figure 11).



**Figure 11.** The effect of films enriched with essential oils mixture on the shelf-life of different food products.

Pizza (RT): Pizza dough kept at room temperature.

Pizza (R): Pizza dough kept in the refrigerator.

Although, the direct attachment of the films which enriched with the essential oils mixture have the limited benefits because the active substances either neutralized on the food surface or diffused rapidly from the surface into the food mass (Quintavalla and Vicini, 2002). Good results obtained by this way with Meat Salami, Artificial Cheese, and the refrigerated Pizza dough. Whereas, the essential oils components can be "in some cases" insufficient to inhibit the microbial growth or adsorbed rapidly on the food stuff surface. This supposition can be explaining the results obtained with Chicken Salami, and Pizza dough which kept at room temperature.

Theoretically, food products packaged in containers and sealed with the films enriched with the essential oils mixture can be the best process. That is because; the air which filled the space over the food stuff be saturated with essential oils components (evidenced by GC-MS test), adsorbed and diffused slower than that directly attached with the food surface.

However, the use of such packaging materials is not meant to be a substitute for good preservation practices, but it should enhance the safety of food as an additional hurdle for the growth of pathogenic microorganisms.

If the types of food products can be divided in general into solid, semi-solid and soft food stuffs; this work succeeded in designing a protection process to the solid and semi-solid one by using the simplest techniques and materials. According to the results "and after the purification and chlorophyll removal process", extractions of *Myrtus communis*, *Urtica urens*, *Ziziphus spina-christi* and *Zygophyllum coccineum* can be used as antibacterial food additives.

#### 4. Conclusion

The microbicidal activities for the films of soy-starch and gelatin that enriched with 0.5ml of the essential oils mixture were attributed to the presence of  $\alpha$ -pinene in soy-starch film, and  $\alpha$ -pinene and limonene in gelatin film. From the physicochemical properties of films, those edible films were found to be suitable for packaging solid and semi-solid food products.

#### Acknowledgment

Special thanks to Dr. Eetemad Othman El-khawas, Chief Researcher in Flora and Phytotaxonomy Researches Department, Horticulture Research Institute, Agricultural Research Center for her kind assistance. Also, many thanks to the members of Herbaria of Botany of both Cairo University (CAI) & Ain-Shams University (CAIA) for identifying the experimental plants, the Department of microbiology in Faculty of Science of Ain-Shams University and the Veterinary Serum & Vaccine Research Institute for providing us with the different used microorganisms.

#### References

- Abo-Zaid EN. 2000. **Volatile Oils**, 1st ed. Dar-el-Arabeya. [In Arabic]
- Atlas RM. 1979. **Hand Book of Microbiological Media**, 2nd ed. CRC Press.
- Batanouny KH, Abou Tabl S, Shabana M and Soliman F. 1999. Wild medical plant in Egypt. An inventory to support conservation and sustainable Use. Academy of Scientific Research and Technology, Egypt. International Union for Conservation (IUCN), Switzerland. pp: 207.
- Bourtoom T. 2008. Edible films and coatings: characteristics and properties. *Inter Food Res J.*, **15** (3): 1-12.
- Brandenburg AH, Weller CL and Testin RF. 1993. Edible films and coatings from soy protein. *J Food Sci.*, **58**(5):1086-1089.
- Briston JH. 1988. **Plastics Films**, 3rd ed. Wiley: New York.
- Burt SA. 2004. Essential oils: their antibacterial properties and potential applications in foods: a review. *Inter J. Food Microbiol.* **94**:223–253.
- Cagri A, Ustunol Z and Ryser ET. 2004. Antimicrobial edible films and coatings. *J Food Prot.*, **67**(4): 833-848.
- Callegarin F, Gallo JAQ, Debeaufort F and Voilley A. 1997. Lipids and biopackaging. *J Am Oil Chem Soc.*, **74**: 1183 – 1192.
- Chaplin M. 2009. Gelatin. Licensed under a Creative Commons Attribution-Noncommercial – No Derivative Works 2.0 UK: England & Wales License.

- Coma V, Sebti I, Pardon P, Deschamps A and Pichavant FH. 2001. Antimicrobial edible packaging based on cellulosic ethers, fatty acids and nisin incorporation to inhibit *Listeria innocua* and *Staphylococcus aureus*. *J Food Prot.*, **64(4)**: 470 - 475.
- Crowell PL. 1999. Prevention and therapy of cancer by dietary monoterpenes. *J Nutr.*, **129 (3)**: 775S–778S.
- Curini M, Bianchi A, Epifano F, Bruni R, Torta L and Zambonelli A. 2003. Composition and *in vitro* antifungal activity of essential oils of *Erigeron canadensis* and *Myrtus communis* from France. *Chem Natural Compounds*, **39(2)**: 191-194.
- De Carvalho RA and Grosso CRF. 2006. Properties of chemically modified gelatin films. *Braz J Chem Eng.*, **23 (1)**: 45 - 53.
- Devlieghere F, Vermeiren L and Debevere J. 2004. New preservation technologies: Possibilities and limitations. *Inter Dairy J.*, **14**: 273-285.
- Dweck AC. 2005. A review of *Ziziphus spina- christi*. *Personal Care Magazine*, **6**: 53-55.
- Gennadios A, Mchugh TH, Weller CL and Krochta JM. 1994. Edible coatings and films based on proteins. In: Krochta JM., Baldwin EA and Nisperos-Carriedo MO, (eds). **Edible Coatings and Films to Improve Food Quality**. Technomic: Lancaster, PA. pp 201- 277.
- Gennadios A, Brandenburg AH, Weller CL and Testin RF. 1993. Effect of pH on properties of wheat gluten and soy protein isolate films. *J Agric Food Chem.*, **41**:1835-1839.
- Ghorpade VM, Gennadios A, Hanna MA and Weller CL. 1995. Soy protein isolate / poly (ethylene oxide) films. *Cereal Chem.*, **72(6)**: 559-563.
- Gill AO and Holley RA. 2006. Disruption of *Escherichia coli*, *Listeria monocytogenes* and *Lactobacillus sakei* cellular membranes by plant oil aromatics. *Inter J Food Microbiol.*, **8**: 1 – 9.
- Glombitza KW, Mahran GH, Mirhom YW, Michel KG and Motawi TK. 1994. Hypoglycemic and antihyperglycemic effects of *Ziziphus spina- christi* in rats. *Planta Med.*, **60**: 244-247.
- Guilbert S, Gontard N and Gorris LGM. 1996. Prolongation of the shelf- life of perishable food products using biodegradable films and coatings. *Lebensmittel-Wissenschaft und-Technologie*. **29(1)**:10-17.
- Han JH. 2000. Antimicrobial food packaging. *Food Technol.*, **54 (3)**: 56-65.
- Hartsuijker C and Welleman JW. 2001. **Engineering Mechanics**, Vol. 2. Springer.
- Holley RA and Patel D. 2005. Improvement of shelf-life and safety of perishable foods by plant essential oils and smoke antimicrobials. *Food Microbiol.*, **22**: 273–292.
- Kinsella JE, Damodaran S and German B. 1985. Physicochemical and functional properties of isolated proteins with emphasis on soy proteins. In: Altschul AM and Wilcke HL, (Eds), **New Protein Foods**. Orlando, FL: Academic Press. pp.107-179.
- Kordali S, Kotan R, Mavi A, Cakir A, Ala A and Yildirim A. 2005. Determination of the chemical composition and antioxidant activity of the essential oil of *Artemisia dracunculoides* and of the antifungal and antibacterial activities of Turkish *Artemisia absinthium*, *A. dracunculoides*, *Artemisia santonicum*, and *Artemisia spicigera* essential oils. *J Agric Food Chem.*, **53**: 9452–58.
- Kunte LA, Gennadios A, Cuppett SL, Hanna MA and Weller CL. 1997. Cast films from soy protein isolates and fractions. *Cereal Chem.*, **74(2)**:115–118.
- Lemberkovic E, Kéry A, Kakasy E and Simái B. 2003. Effect of extraction method on the composition of essential oils. *ISHS Acta Horticulture* 597. International Conference on Medicinal and Aromatic Plants (Part II).
- Maizura M, Fazilah A, Norziah MH and Karim AA. 2008. Antibacterial activity of modified *Sago starch-alginate* based edible film incorporated with lemongrass (*Cymbopogon citratus*) oil. *Inter Food Res J.*, **15(2)**: 233-236.
- Mellan I. 1961. **The Behavior of Plasticizers**. Pergamon press, New York.
- Montvale NJ. 2000. **PDR for Herbal Medicine**, 2nd Edition. Medical Economics Company.
- Ninnemann KW. 1968. Measurements of physical properties of flexible films. In: Sweeting OJ, (Ed), **Science and Technology of Polymer Films**. Interscience, London, England. pp. 546-649.
- Özcan M, Kasik G and Öztürk C. 2005. Inhibitory effect of essential oils of myrtle, laurel, pickling herb and thyme and savory on the mycelial growth of *Agaricus campestris* (Lange) Sing. *J Essential Oil Bearing Plants*, **8(2)**:120-125.
- Pakdela H, Panteaa D and Roy C. 2001. Production of dl-limonene by vacuum pyrolysis of used tires. *J Analytical and Applied Pyrolysi.*, **57(1)**: 91–107.
- Pérez-Pérez C, Regalado-González C, Rodríguez-Rodríguez CA, Barbosa-Rodríguez JR and Villaseñor-Ortega F. 2006. Incorporation of antimicrobial agents in food packaging films and coatings. In: Guevara-González RG and Torres-Pacheco L, (Eds.), **Advances in Agricultural and Food Biotechnology**. pp193-216.
- Quintavalla S and Vicini L. 2002. Antimicrobial food packaging in meat industry. *Meat Sci.*, **62 (3)**: 373 – 380.
- Randall C. 2003. Historical and modern uses of *Urtica*. In: Kavalali GM, (Ed.). **Urtica. Therapeutic and Nutritional Aspects of Stinging Nettles**. Taylor & Francis, London, New York. pp. 12-24.
- Richter GH. 1945. **Textbook of Organic Chemistry**, 2nd Edition. John Wiley & Sons, New York. pp. 663-666.
- Rodrigues ET and Han JH. 2000. Antimicrobial whey protein films against spoilage and pathogenic bacteria. Proceedings of the IFT Annual Meeting. Dallas, June 10-14. Chicago, Ill.: Institute of Food Technologists. pp191.
- Salame M. 1986. Barrier polymers. In: Bakker M, (Ed.), **The Wiley Encyclopedia of Packaging Technology**. New York: John Wiley and Sons. pp.48-54.
- Schott JR. 1997. **Matrix Analysis for Statistics**. New York: John Wiley and Sons.
- Shahidi Bonjar GH, Nik AK, Heydari MR, Ghasemzadeh MH, Farrokhi PR, Moein MR, Mansouri S and Foroumadi A. 2003. Anti-Pseudomonas and Anti-Bacilli activity of some medicinal plants of Iran. *DARU*. **11(4)**: 157-163.
- Simonsen JL. 1957. **The Terpenes**, 2nd Edition. (Vol. 2). Cambridge University Press.
- Simonsen JL. 1947. **The Terpenes**, 2nd Edition. (Vol. I). Cambridge University Press.
- Sonti, S. 2003. Consumer perception and application of edible coatings on fresh-cut fruits and vegetables (MSc thesis). Faculty of Agricultural and Mechanical College: Louisiana State University.
- Tsybula NV and Kazarinova NV. 1996. Sanative effect of volatile compounds produced by intact common Myrtle *Myrtus communis* L. in interiors. *Bulletin Exper Biol Med.*, **121 (5)**: 597-600.
- Tuberoso CIG, Barra A, Angioni A, Sarritzu E, and Pirisi FM. 2006. Chemical composition of volatiles in Sardinian Myrtle (*Myrtus communis* L.) alcoholic extracts and essential oils. *J Agric Food Chem.*, **54**: 1420-1426.

Waggas AM. 2007. Acute effect of Sidr leaves extract on some neurotransmitter contents in different brain areas of male albino rats. *Saudi J Biol Sci.*, **14 (1)**: 93-101.

Wichtl M (Ed.). 2002. **Teedrogen und Phytopharmaka. Ein Handbuch für die Praxis auf wissenschaftlicher Grundlage**, 4. Auflage. Wissenschaftliche Verlagsges, M.B.H., Stuttgart. pp. 617-619.

# Effects of Three Medicinal Plant Products on Survival, Oviposition and Progeny Development of Cowpea Bruchid, *Callosobruchus maculatus* (Fab.) [Coleoptera: Chrysomelidae] Infesting Cowpea Seeds in Storage

Kayode D. Ileke<sup>1,\*</sup>, Daniel S. Bulus<sup>2</sup> and Ayisat Y. Aladegoroye<sup>3</sup>

<sup>1,3</sup>Department of Environmental Biology and Fisheries, Faculty of Science, Adekunle Ajasin University, PMB 001, Akungba Akoko, Ondo State,

<sup>2</sup>Department of Food Science and Technology, Federal Polytechnic, Kaura Namoda, Zamfara State, Nigeria.

Received: August 20, 2012; Accepted: November 29, 2012

## Abstract

Powders from three medicinal plants were evaluated for their efficacy as contact and fumigant insecticides on cowpea bruchid, *Callosobruchus maculatus* (Fab.) in the laboratory at ambient tropical conditions of temperature and relative humidity. The plant powders tested included *Capsicum frutescens* L. (fruit and seed), *C. annum* Miller (fruit and seed) and *Citrus sinensis* Osbeck (peel). The powders were applied at rates 0.0 (control), 2g and 3.0g/20g of cowpea seeds either directly for contact with the insect pest or in plastic containers to assess fumigant toxicity of their volatiles. Results of contact toxicity assay showed that powders of *C. frutescens* and *C. annum* seeds were more effective against the adult *C. maculatus* evoking 100% mortality within 2 days of application at 3g/20g of cowpea seeds. There was no progeny development of the bruchid in samples treated with *Capsicum* species. The survival of the bruchid from eggs to adults when treated with the plant powders showed that there was significantly ( $P < 0.05$ ) more % progeny development in the control (69.32%) compared to others. However, the results of fumigant assays showed that *C. sinensis* had the highest insecticidal activity causing 281.25% mortality of *C. maculatus* within 4 days of application at rate 3g/20g of cowpea seeds. This study showed that all the tested plant products were toxic to cowpea bruchid and the powders can be mixed with cowpea seeds to prevent hatching of the eggs thereby helping in their management.

**Key Words:** Insecticidal activity, progeny development, *Callosobruchus maculatus*, *Capsicum frutescens*, *Capsicum annum*, *Citrus sinensis*, cowpea seed.

## 1. Introduction

Cowpea, *Vigna unguiculata* (L.) Walp, belongs to the family Leguminosae, subfamily Papilionaceae and Tribe Phaseolae (Gbaye and Holloway, 2011). It is a legume widely cultivated in tropical and subtropical countries and largely produced in West Africa, Brazil and India. Cowpea is a staple component of the diet in several developing nations and a major source of protein to combat malnutrition in young children in lieu of expensive animal protein in such countries.

One of the major problems encountered in agriculture in developing countries is post harvest losses which usually occur during storage (Adedire *et al.*, 2011). The cowpea bruchid, *Callosobruchus maculatus* has been

recognized for decades as the major post harvest insect pest of cowpea seeds. It is a cosmopolitan species (Ofuya, 2001; Ileke and Bulus, 2012a). Initial infestation of cowpea starts in the field just before harvest and the insects are carried into the store where the population builds up rapidly (Appert, 1987; Ofuya, 2001; Ileke *et al.*, 2012). The huge post-harvest losses and quality deterioration caused by this insect is a major obstacle to achieving food security in developing countries such as Nigeria. Annual production loss of 5% to *C. maculatus* in Nigeria alone would amount to 40,000 tonnes of cowpea seeds cost about \$100 million (Singh and Ntare, 1985). The larvae are the major destructive stage because adult cowpea bruchid do not feed (Ofuya, 2001; Gbaye and Holloway, 2011).

\* Corresponding author. e-mail: kayodeileke@yahoo.com.

Efficient control of stored products insect pests has long been the aim of entomologists throughout the world. Synthetic insecticides have been used for many years to control stored product insect pests (Salem *et al.*, 2007; Ashouri and Shayesteh, 2010). Development of resistance of insect pests to these conventional storage insecticides, increased concern by consumers over insecticide residues, ecological consequences and increasing cost of application call for new approaches to control stored products insect pests that are readily available, affordable and less detrimental to the environment (Adedire and Lajide, 2003; Udo, 2005; Ileke and Oni, 2011; Ashouri and Shayesteh, 2010; Ileke and Bulus, 2012b). A number of plants used locally for medicinal purposes, have also demonstrated potential as insect control agents (Arannilewa *et al.*, 2006; Oni, 2011). For example, cowpea seeds mixed with *Alstonia boonei* and *Eugenia aromatic* have been found to reduce infestation by cowpea beetle (Ofuya *et al.*, 2007; Ileke *et al.*, 2012). Plants such as *Capsicum frutescens*, *C. annum* fruit and *Citrus sinensis* peel have previously been shown to cause mortality of adult *Dasytes rugosella* in yam tuber (Ashamo, 2010). Oni (2011) examined contact toxicity of *Capsicum* species to adult *Sitophilus zeamais* and *C. maculatus*. In the present investigation, survival, oviposition and progeny development of cowpea bruchid, *C. maculatus* exposed to *Citrus sinensis* peel, *Capsicum frutescens* and *C. annum* fruit and seeds powders were evaluated.

## 2. Materials and Methods

This study was conducted in the Environmental Biology and Fisheries Research Laboratory, Faculty of Science, Adekunle Ajasin University, Akungba Akoko, Ondo State, Nigeria.

### 2.1. Insect culture

The insects used to establish a laboratory colony of *C. maculatus* came from a batch of infested cowpea seeds, *Vigna unguiculata* (L.) Walp variety Ife brown collected from Agricultural Development Project, Akure, Ondo State, Nigeria. Beetles were reared subsequently by replacement of devoured and infested cowpea seeds with fresh un-infested cowpea seeds in 2-L kilner jars covered with muslin cloth to allow air circulation. Insect rearing and the experiments were carried out at ambient temperature of 28±2°C and 75±5% relative humidity.

### 2.2. Plant collection

The plants evaluated in this work were *Capsicum frutescens* (fruit and seed), *Capsicum annum* (fruit and seed) and *Citrus sinensis* (peel). They were obtained in fresh form, free of insecticides from Oja-Oba market, Akure, Ondo State, Nigeria and authenticated by the Plant Science and Technology Department of Adekunle Ajasin University, Akungba Akoko, Ondo State. These plant materials were rinsed in clean water to remove sand and other impurities, cut into smaller pieces before air dried in a well ventilated laboratory and ground into very fine powder using an electric blender. The powders were further sieved to pass through 1mm<sup>2</sup> perforations. The powders were packed in plastic containers with tight lids and stored in a refrigerator at 4°C prior to use.

### 2.3. Collection of Cowpea Seeds

Cowpea seeds used for this study were obtained from a newly stocked seeds free of insecticides at Agricultural Development Program (ADP), Akure, Ondo State, Nigeria. Firstly, the seeds were cleaned and disinfested by keeping at -5°C for 7 days to kill all hidden infestations. This is because all the life stages, particularly the eggs are very sensitive to cold (Koehler, 2003). The disinfested cowpea seeds were then placed inside a Gallenkamp oven (model 250) at 40°C for 4 hours (Jambere *et al.*, 1995) and later air dried in the laboratory to prevent mouldiness (Adedire *et al.*, 2011) before they were stored in plastic containers with tight lids.

### 2.4. Effect of contact toxicity of plants powders on adult mortality, oviposition and progeny development of *Callosobruchus maculatus*

Fine powders of *Capsicum frutescens*, *Capsicum annum*, *Citrus sinensis* were admixed with cowpea seeds at the rates of 2 and 3g /20g of cowpea seeds in 250ml plastic containers. Ten pairs of adult *C. maculatus* (2 to 3 days old) sexed according to the methods described by Halstead (1963); Appert (1987); Odeyemi and Daramola (2000) were introduced into the treated. Male *C. maculatus* have comparative shorter abdomen and the dorsal side of the terminal segment is sharply curved downward and inward. In contrast the females have comparatively longer abdomen and the dorsal side of the terminal segment is only slightly bent downward. The female also has two dark visible spots on their elytra (Odeyemi and Daramola, 2000). Untreated cowpea seeds were similarly infested. Four replicates of the treated and untreated controls were laid out in Complete Randomized Block Design in insect cage. Insect mortality was assessed every 24 hours for four days. Adults were assumed dead when probed with sharp objects and made no responses. At the end of day 4, all insects, both dead and alive were removed from each container. The experiment was kept inside the insect cage for another 30 days to allow for the emergence of the first filial (F<sub>1</sub>) generation. The number of adults that emerged from each replicate was counted with an aspirator and recorded. The percentage adult emergence was then calculated using the method described by Odeyemi and Daramola (2000).

$$\% \text{ Progeny development} = \frac{\text{No of adult emerged} \times 100}{\text{No of eggs laid}} \times \frac{1}{1}$$

### 2.5. Fumigant effect of plants powders on adult mortality, oviposition and progeny development of *C. maculatus*

Ten grams of the cowpea seeds were weighed into 50ml transparent plastic tubes that had been cut opened at the bottom and sealed with muslin cloth. Fine powders of *Capsicum frutescens*, *Capsicum annum*, *Citrus sinensis* weighing 2g and 3g concentrations were put into another half-cut 25ml plastic tubes. The 50ml tube and 25ml tube were then joined together with the aid of gum (Ileke and Bulus, 2012a). Ten pairs of adult *C. maculatus* (2 to 3 days old) sexed according to the methods described above were introduced to the tube containing 10g of cowpea seeds and tightly sealed (Ileke and Bulus, 2012a). Untreated cowpea seeds were similarly infested. Four replicates of the treated and untreated controls were laid out in Complete Randomized Block Design in insect cage.

Bruchid mortality was assessed every 24 hours for four days. Adults were assumed dead when probed with sharp objects and made no responses. At the end of day 4, all insects, both dead and alive were removed from each container. The experiment was kept inside the insect cage for another 30 days to allow for the emergence of the first filial ( $F_1$ ) generation. The number of adults that emerged from each replicate was counted with an aspirator and recorded. Percentage adult emergence was calculated as described above.

### 2.6. Statistical analysis

Data were subjected to analysis of variance and where significant differences existed, treatment means were separated using the Tukey's test.

## 3. Results

### 3.1. Effectiveness of plants powders as contact insecticides

The effectiveness of the various plant powders on the survival of cowpea bruchid, *C. maculatus* at different periods after treatment is presented in Tables 1 and 2. The results revealed that in each treatment, the mortality of *C. maculatus* increased gradually with time of exposure. Chilly pepper, *C. frutescens* seed powder caused 100% mortality of *C. maculatus* at rate 2g/20g of cowpea seeds within 2 days of exposure (Table 1). The corresponding value for *C. sinensis*, *C. frutescens* fruit and *C. annuum* fruit and seed powders were 34.5%, 87.5%, 51.25% and 71.25% mortality of adult cowpea bruchid respectively. The contact toxicities of these plant powders increased with increase in dosage as well as increase in the period of exposure to plant powders (Table 2). At rate 3g/20g of cowpea seeds, 100% mortality was obtained in sample treated with *C. frutescens* seed at 1 day after application of powder. The results indicated that various plant powders tested as contact insecticides significantly ( $P < 0.05$ ) reduced number of tested insect. In general, *Capsicum* species seeds powders were more toxic than other tested plant powders.

**Table 1.** Percentage mortality of adult *Callosobruchus maculatus* treated with various powders at rate 2g/20g of cowpea seeds for contact toxicity.

Powder	Mortality % $\pm$ SE mean after			
	1 day	2 day	3 day	4 day
<i>Citrus sinensis</i> (p)	17.25 $\pm$ 1.44b	34.50 $\pm$ 1.44b	68.25 $\pm$ 2.39b	80.00 $\pm$ 1.25b
<i>Capsicum frutescens</i> (f)	45.00 $\pm$ 2.04c	87.50 $\pm$ 5.20e	100.00 $\pm$ 0.00c	100.00 $\pm$ 0.00c
<i>Capsicum frutescens</i> (s)	70.00 $\pm$ 1.25d	100.00 $\pm$ 0.00f	100.00 $\pm$ 0.00c	100.00 $\pm$ 0.00c
<i>Capsicum annuum</i> (f)	20.00 $\pm$ 4.05b	51.25 $\pm$ 2.39c	100.00 $\pm$ 0.00c	100.00 $\pm$ 0.00c
<i>Capsicum annuum</i> (s)	41.25 $\pm$ 2.39c	71.25 $\pm$ 2.39d	100.00 $\pm$ 0.00c	100.00 $\pm$ 0.00c
Control	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a

Each value is a mean  $\pm$  standard error of four replicates. Means within the same column followed by the same letter(s) are not significantly different at  $P > 0.05$  using Tukey's test.

Keys: f- fruit, s - seed, p – peel.

**Table 2.** Percentage mortality of adult *Callosobruchus maculatus* treated with various powders at rate 3g/20g of cowpea seeds for contact toxicity.

Powder	Mortality % $\pm$ SE mean after			
	1 day	2 day	3 day	4 day
<i>Citrus sinensis</i> (p)	28.75 $\pm$ 1.25b	47.50 $\pm$ 1.44b	78.75 $\pm$ 1.25b	88.75 $\pm$ 1.25b
<i>Capsicum frutescens</i> (f)	50.00 $\pm$ 1.25c	71.25 $\pm$ 2.39c	100.00 $\pm$ 0.00c	100.00 $\pm$ 0.00c
<i>Capsicum frutescens</i> (s)	100.00 $\pm$ 0.00e	100.00 $\pm$ 0.00d	100.00 $\pm$ 0.00c	100.00 $\pm$ 0.00c
<i>Capsicum annuum</i> (f)	32.75 $\pm$ 1.25b	60.00 $\pm$ 1.25c	100.00 $\pm$ 0.00c	100.00 $\pm$ 0.00c
<i>Capsicum annuum</i> (s)	71.75 $\pm$ 1.25d	100.00 $\pm$ 0.00d	100.00 $\pm$ 0.00c	100.00 $\pm$ 0.00c
Control	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a

Each value is a mean  $\pm$  standard error of four replicates. Means within the same column followed by the same letter(s) are not significantly different at  $P > 0.05$  using Tukey's test.

Keys: f- fruit, s- seed, p - peel

### 3.2. Fumigant effect of various plant powders

Table 3 and 4 showed the fumigant effect of various plant powder against *C. maculatus*. At day 4 after post treatment, *C. sinensis* was able to cause 66.75% mortality of adult *C. maculatus* while the corresponding values for *C. frutescens* and *C. annuum* seeds powders were 55% and 40% mortality of bruchid respectively at 2g/20g of cowpea seeds. (Table 3). The toxicities of these plant powders to cowpea bruchid increased with an increase in concentration and period of exposure to plant powders. *C. sinensis* powder caused 81.25% mortality of *C. maculatus* while the corresponding value for *C. frutescens* seed powder was 72.5% mortality of *C. maculatus* at rate 3g/20g of cowpea seeds (Table 4).

**Table 3.** Percentage mortality of adult *Callosobruchus maculatus* treated with various powders at rate 2g/20g of cowpea seeds for fumigant toxicity.

Powder	Mortality % $\pm$ SE mean after			
	1 day	2 day	3 day	4 day
<i>Citrus sinensis</i> (p)	20.00 $\pm$ 1.25b	238.75 $\pm$ 1.25c	45.00 $\pm$ 2.04d	66.75 $\pm$ 1.25d
<i>Capsicum frutescens</i> (f)	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a	10.00 $\pm$ 1.25b	20.00 $\pm$ 1.25b
<i>Capsicum frutescens</i> (s)	10.00 $\pm$ 1.25b	22.50 $\pm$ 1.44b	31.25 $\pm$ 1.25c	55.00 $\pm$ 2.04d
<i>Capsicum annuum</i> (f)	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a	12.50 $\pm$ 1.44b
<i>Capsicum annuum</i> (s)	0.00 $\pm$ 0.00a	10.00 $\pm$ 1.25b	20.00 $\pm$ 1.25bc	40.00 $\pm$ 1.25c
Control	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a	0.00 $\pm$ 0.00a

Each value is a mean  $\pm$  standard error of four replicates. Means within the same column followed by the same letter(s) are not significantly different at  $P > 0.05$  using Tukey's test.

Keys: f- fruit, s- seed, p – peel.

**Table 4.** Percentage mortality of adult *Callosobruchus maculatus* treated with various powders at rate 3g/20g of cowpea seeds for fumigant toxicity.

Powder	Mortality % ± SE mean after			
	1 day	2 day	3 day	4 day
<i>Citrus sinensis</i> (p)	45.00± 2.04c	67.50± 2.04d	78.75± 1.25e	81.25± 2.39d
<i>Capsicum frutescens</i> (f)	0.00± 0.00a	12.50± 1.44b	25.00± 1.25c	31.25± 2.39b
<i>Capsicum frutescens</i> (s)	20.00± 1.25b	38.75± 1.25c	51.25± 2.39d	72.50± 3.15cd
<i>Capsicum annum</i> (f)	0.00± 0.00a	0.00± 0.00a	8.75± 1.25b	20.00± 1.25b
<i>Capsicum annum</i> (s)	8.75± 1.25b	20.00± 1.25b	38.75± 1.2d	62.00± 1.25c
Control	0.00± 0.00a	0.00± 0.00a	0.00± 0.00a	0.00± 0.00a

Each value is a mean ± standard error of four replicates. Means within the same column followed by the same letter(s) are not significantly different at P>0.05 using Tukey's test.

Keys: f- fruit, s-seed, p - peel

### 3.3. Effect of various plant powders applied as contact and fumigant insecticides on oviposition and progeny development of *C. maculatus*

Table 5 shows the oviposition and percentage progeny development of *C. maculatus* after being exposed to various plant powders as contact insecticide at two concentrations after 4 days. Progeny development was significantly suppressed by various plant powders with *Capsicum* species and completely inhibited the emergence of *C. maculatus* (100% efficiency).

**Table 5.** Fecundity of *Callosobruchus maculatus* treated with various plant powders as contact insecticides

Plant Powder	2g/20g of cowpea seeds No of egg laid	% no of Progeny development	3g/20g of cowpea seeds No of egg laid	% no of Progeny development
<i>Citrus sinensis</i> (p)	36.25± 1.70c	15.84	31.25± 2.39c	9.60
<i>Capsicum frutescens</i> (f)	10.50± 1.32b	0.00	2.75± 1.25ab	0.00
<i>Capsicum frutescens</i> (s)	0.00± 0.00a	0.00	0.00± 0.00a	0.00
<i>Capsicum annum</i> (f)	12.75± 2.02b	0.00	7.25± 2.39b	0.00
<i>Capsicum annum</i> (s)	0.00± 0.00a	0.00	0.00± 0.00a	0.00
Control	88.75± 1.25d	69.32	88.75± 1.25d	69.32

Each value is a mean ± standard error of four replicates. Means within the same column followed by the same letter(s) are not significantly different at P>0.05 using Tukey's test.

Keys: f- fruit, s-seed, p - peel

In Table 6, fecundity of *C. maculatus* showed that more eggs were laid on samples treated with plants powders applied as fumigant. Oviposition and % progeny development were high in cowpea seeds treated with *Capsicum* species fruit powders.

**Table 6.** Fecundity of *Callosobruchus maculatus* treated with various plant powders as fumigant insecticides

Plant Powder	2g/20g of cowpea seeds No of egg laid	% no of Progeny development	3g/20g of cowpea seeds No of egg laid	% no of Progeny development
<i>Citrus sinensis</i> (p)	11.25± 2.39a	11.33	9.50± 3.15a	10.32
<i>Capsicum frutescens</i> (f)	38.75± 1.25bc	32.47	30.00± 2.04bc	28.76
<i>Capsicum frutescens</i> (s)	25.00± 2.04ab	21.67	12.00± 0.00a	19.49
<i>Capsicum annum</i>	41.25±	35.86	37.50±	31.11

<i>annum</i> (f)	2.39c		3.15c	
<i>Capsicum annum</i> (s)	32.50± 3.15bc	27.17	21.25± 2.39ab	24.58
Control	77.50± 1.44d	80.00	77.50± 1.44d	80.00

Each value is a mean ± standard error of four replicates. Means within the same column followed by the same letter(s) are not significantly different at P>0.05 using Tukey's test.

Keys: f- fruit, s-seed, p - peel

## 4. Discussion

Results reported in this study show that *Capsicum* species and *C. sinensis* powders have insecticidal effects on cowpea bruchid, *C. maculatus* at all levels of treatment but varied with the method of application, exposure period and plant powder concentrations. The two *Capsicum* species seed and fruit powders applied as contact insecticides were very effective against *C. maculatus* causing 100% mortality of adult *C. maculatus* at rate 3g/20g of cowpea seeds within 4 days of application. They also reduced oviposition and completely inhibited progeny development. This shows that *Capsicum* species probably have oviposition deterrent, ovicidal and larvicidal properties. The observed activity may be due to the "peppery" nature and pungency of the *Capsicum* species (Ashamo, 2010). The pungency of *Capsicum* species was attributed to capsaicin (Miyakado *et al.*, 1979; Ashamo, 2010). This result is in agreement with the results of Ivbijaro and Agbaje (1986), and Asawalam *et al.* (2007). They both found that *C. frutescens* considerably reduced all stages of *C. maculatus*. The result of this investigation are also similar to the observation of Ashamo (2010) who obtained 100% mortality of adult *Dasytes rugosella* in yam tuber treated with powders and oils of *Capsicum* species. Oni (2011) reported that *Capsicum* species seeds and fruits powders significantly toxic to *Sitophilus zeamais* and *C. maculatus* in stored maize and cowpea seeds, respectively.

The high mortality and low progeny development caused by the powder of *C. sinensis* can be attributed to strong choky odour disrupting respiratory activity of the beetles. Sweet orange peel powders may probably have the same insecticidal properties when applied as contact and fumigant. The results obtained from this study agreed with those reported by Don Pedro (1996a; b) in studies with six *Citrus* species peel oils against *C. maculatus*, *S. zeamais* and *Dermestes maculatus*. He reported on fumigant action of toxic vapour of *Citrus* species peel oils against *C. maculatus*, *S. zeamais* and *Dermestes maculatus*.

In this study, the lethal effect of tested plant powders on cowpea bruchid could be as a result of contact toxicity. Insects breathe by means of trachea which usually opens at the surface of the body through spiracles (Adedire *et al.*, 2011). These spiracles might have been blocked by the powders thereby leading to suffocation. The powders also prevented oviposition and progeny development when applied as contact insecticides. The choky effect of these powders also disrupt mating activities, sexual communication and inhibit locomotion an effect that have been reported by many researchers (Ofuya, 1992; Adedire 2002; Maina and Lale, 2004; Akinkulere *et al.*, 2006; 2009; Adedire *et al.*, 2011; Ileke *et al.*, 2012).

Adult bruchids do not feed on stored cowpea seeds but only deposit their eggs. Thus, the use of oviposition inhibitors would be advantageous for the management of cowpea bruchids. The powders of these plants could be mixed with stored cowpea seeds before storage.

### Acknowledgment

The authors are grateful to Dr. O. A. Obembe of the Department of Plant Science and Biotechnology, Faculty of Science, Adekunle Ajasin University, Akungba Akoko, Ondo State, Nigeria for his assistance in identification of plants used in this study. The comments by two anonymous reviewers towards improving the quality of this research work are acknowledged.

### References

- Adedire CO. 2002. Use of nutmeg, *Myristica fragrans* powder and oil for the control of cowpea storage bruchid, *Callosobruchus maculatus*. *J Plant Dis and Protect.*, **109**: 193-199
- Adedire CO, Lajide L. 2003. Ability of extract of ten tropical plant species to protect maize grains against infestation by the maize weevil *Sitophilus zeamais* during storage. *Nigerian J. Experiment. Biol.*, **4(2)**: 175-179
- Adedire CO, Obembe, OO, Akinkurolere RO, Oduleye O. 2011. Response of *Callosobruchus maculatus* (Coleoptera: Chrysomelidae: Bruchidae) to extracts of cashew kernels. *J Plant Dis and Protect.*, **118(2)**: 75-79
- Akinkurolere RO, Adedire CO, Odeyemi OO. 2006. Laboratory evaluation of the toxic properties of forest anchomanes, *Anhomanus difformis*, against pulse beetle, *Callosobruchus maculatus* (Coleoptera: Bruchidae). *Insect Sci.*, **13**: 25-29
- Akinkurolere RO, Sebastien B, Haoliang C, Hongyu Z. 2009. Parasitism and host location preference in *Habrobracon hebetor* (Hymenoptera: Braconidae): Role of refuge, choice and host instar. *J Economic Entomol.*, **102(2)**: 610-615.
- Appert, J. 1987. **The Storage of Food Grains and Seeds**. CTA Macmillan, pp 146.
- Arannilewa ST, Ekrakene T, Akinneye JO. 2006. Laboratory evaluation of four medicinal plants as protectants against the maize weevil *Sitophilus zeamais* (Mot). *Afri J Biotechnol.*, **5(21)**: 2032-2036.
- Asawalam EF, Emosairue SO, Ekeleme F, Wokocha RC. 2007. Insecticidal effects of powdered parts of eight Nigerian plant species against maize weevil, *Sitophilus zeamais* Mot [Coleoptera: Curculionidae]. *Electronic J. Environment. Agricul. Chem.*, **6(11)**: 2526-2533.
- Ashamo MO. 2010. Management of the yam moth, *Dasyses rugosella*, a pest of stored yam tubers using plant products. 10th International Working Conference on Store Product Protection 793 – 798.
- Ashouri S, Shayesteh N. 2010. Insecticidal activities of two powdered spices, black pepper and red pepper on adult *Rhyzopertha dominica* (F.) and *Sitophilus granarius* (L.). *Munis Entomol Zool.*, **5(2)**: 600 – 607.
- Don-Pedro KN. 1996a. Fumigant toxicity is the major route of insecticidal activity of citrus peel essential oils. *Pesticide Sci.*, **46**: 71 – 78.
- Don-Pedro KN. 1996b. Fumigant toxicity of citrus peel oils against adult and immature stages of storage insects. *Pesticide Sci.*, **46**: 213 – 223.
- Gbaye OJ, Holloway GJ. 2011. Varietal effects of cowpea, *Vigna unguiculata*, on tolerance to malathion in *Callosobruchus maculatus*. *J Stored Prod Res.*, **47**: 365 – 371
- Halstead DGH. 1963. External sex difference in stored products *Coleoptera*. *Bulletin Entomol Res.*, **54**: 119-134.
- Ileke KD, Oni MO. 2011. Toxicity of some plant powders to maize weevil, *Sitophilus zeamais* on stored wheat grains. *Afri J Agricul Res.*, **6(13)**: 3043 – 3048.
- Ileke KD, Bulus DS. 2012a. Evaluation of contact toxicity and fumigant effect of some medicinal plant and pirimiphous methyl powders against cowpea bruchid, *C. maculatus* (Fab.) (Coleoptera, Chrysomelidae) in stored cowpea seeds. *J Agricul Sci.*, **4(4)**: 279 - 284.
- Ileke KD, Bulus DS. 2012b. Responses of Lesser grain borer, *Rhyzopertha dominica* (F) [Coleoptera: Bostrichidae] to powders and extracts of *Azadirachta indica* and *Piper guineense* seeds in stored wheat grains. *Jordan J Biol Sci.*, **5(4)**, 315-320.
- Ileke KD, Odeyemi OO, Ashamo MO. 2012. Insecticidal activity of *Alstonia boonei* De Wild powder against cowpea bruchid, *Callosobruchus maculatus* (Fab.) [Coleoptera : Chrysomelidae] in stored cowpea seeds. *Inter J Biol.*, **4(2)**: 125- 131
- Ivbijaro MF, Agbaje M. 1986. Insecticidal activities of Piper guineense and Capsicum species in cowpea bruchid, *Callosobruchus maculatus*. *Insect Sci. Appl.*, **7**: 521 - 524.
- Jambere B, Obeng-Ofori D, Hassanali A. 1995. Products derived from the leaves of *Ocimum kilmandsharicum* as post harvest grain protectant against the infection of three major stored insect product pests. *Bulletin Entomol Res.*, **85**: 351-367.
- Koehler PG. 2003. Biopesticides Data sheet volume 2. Entomology and Nematology Dept, Cooperative extension service, Institute of Food and Agricultural Science, University of Florida, Gainesville. 326pp
- Maina YT, Lale NES. 2004. Efficacy of integrating varietal resistance and neem (*Azadirachta indica*) seed oil for the management of *Callosobruchus maculatus* infesting Bambara Groundnut in storage in storage. *Nigerian J Entomol.*, **2**: 94 – 103.
- Miyakado M, Nakayama I, Yoshoka H, Nakatani NN. 1979. The piperase amides: Structure of piperacide, a new insecticide amide from *Piper nigrum*. *Agricul Biol Chem.*, **43**: 1609 – 1611
- Odeyemi OO, Daramola AM. 2000. **Storage Practices in the Tropics: Food Storage and Pest Problems**. First Edition, Dave Collins Publication, Nigeria , pp 235.
- Ofuya TI. 1992. Oviposition deterrence and ovicidal properties of some plant powders against *C. maculatus* in stored cowpea seeds. *J Agricul Sci.*, **115**: 343 – 345.
- Ofuya TI. 2001. Pest of stored cereals and pulses in Nigeria. In: Ofuya TI and Lale NES ( Eds.), **Biology, Ecology and Control of Insect Pests of Stored Food Legumes**. Dave Collins publications, Nigeria. pp 25-58.
- Ofuya TI, Olotuah OF, Aladesanwa RD. 2007. Potential of dusts of *Eugenia aromatic* dry flower buds, and black pepper dry fruit formulated with three organic flours for controlling *Callosobruchus maculatus*. *Nigerian J Entomol.*, **24**: 98 - 106
- Oni MO. 2011. Evaluation of seed and fruit powder of *Capsicum annum* and *C. frutescens* for control of *Callosobruchus maculatus* (Fab.) in stored cowpea and *Sitophilus zeamais* in stored maize. *Inter J Biol.*, **3(2)**: 185-188
- Oni MO, Ileke KD. 2008. Fumigant toxicity of four botanical plant oils on survival, egg laying and progeny development of the dried yam beetle, *Dinoderus porcellus* (Coleoptera: Bostrichidae). *Ibadan J Agricul Res.*, **4 (2)**: 31-36.

Salem SA, Abou-Ela RG, Matter MM, El-kholy MY. 2007. Entomocidal effect *Brassica napus* extracts on two store pests, *Sitophilus oryzae* and *Rhyzopertha dominica*. *J Applied Sci Res.*, **3(4)**: 317 – 322.

Singh BB, Ntare BR. 1985. Development of improved cowpea varieties in Africa In: Singh, S. R., Rachie, K. O. (Eds), **Cowpea**

**Research, Production and Utilization**. John Wiley and Sons, Chichester, pp 267 – 279.

Udo IO. 2005. Evaluation of the potential of some local spices as stored grain protectants against maize weevil *Sitophilus zeamais*. *J Applied Sci. Environ Manag.*, **9(1)**: 165 – 16.

# Reserve Mobilization, Total Sugars and Proteins in Germinating Seeds of Durum Wheat (*Triticum durum* Desf.) under Water Deficit after Short Period of Imbibition

Amal M. Harb\*

Department of Biological Sciences, Faculty of Science, Yarmouk Univeristy,

P.O. Box: 566 – 21163, Irbid, Jordan.

Received: November 4, 2012; Accepted: December 6, 2012

## Abstract

Imbibition during the first few hours is crucial for the success of seed germination process. The effect of water deficit after 2 and 8 hours (h) of imbibition of seeds of durum wheat was studied in terms of reserve mobilization, total sugars and proteins at 24, 48 and 72 h post treatment. Imbibition for 2 and 8 h was not enough to support normal mobilization of seed reserves and solubilization of sugars. But, a significant increase in total proteins was shown after 24 and 48 h of water deficit for the two imbibition periods (2 and 8 h). Our results indicate that most of the biochemical and molecular changes are intensified during the first 2-8 h of imbibition. Therefore, biochemical and molecular dissection of this phase of seed germination is of great value in the improvement of our understanding of plant's response to water deficit.

**Keywords:** Durum wheat, imbibition, reserve mobilization, water deficit.

## 1. Introduction

One of the most detrimental factors that plants face is water deficit. Water deficit is one of the major abiotic stresses that threaten food security worldwide (Chaves *et al.*, 2003; Passioura, 2006); hence, there is an urgent need for multilevel investigations and dissection for better and comprehensive understanding of how plants respond to water deficit. In addition, this will help find effective ways to increase crop yield without consuming too much water.

Plants are exposed to drought at all stages of their growth and development. Plant's response to water deficit is dependent on the developmental stage (Blum *et al.*, 1980). No correlation was found between the response of seeds and photosynthesizing seedlings to osmotic stress induced by polyethylene glycol (PEG) in ten cultivars of bread wheat, *Triticum aestivum* L. and one cultivar of durum wheat, *Triticum durum* Desf. (Blum *et al.*, 1980). Therefore, one cannot extrapolate results about stress response from one developmental stage to another stage, which necessitates independent studies of the response for each developmental stage.

The first stage in plant development is seed germination, which starts by water uptake (imbibition), followed by reserve mobilization and protein synthesis and

ends with the emergence of the radicle from seed tissues (Bewley, 1997). Water uptake by seeds was shown to occur in three phases: initial rapid water uptake, then a plateau phase, which is followed by rapid water uptake during which the radicle elongates and emerges (Bewley, 1997). In germinating wheat seeds, it was shown that protein synthesis starts to increase 30 min after imbibition, and it keeps increasing in the next 6 hours of imbibition (Marcus *et al.*, 1966). This coincides with the first phase of imbibition, which is characterized by the rapid water uptake. In addition, this suggests a crucial role for this phase in the success of the germination process.

Seeds of bread wheat (*Triticum aestivum* L.) were imbibed for 24 hours, and then they were dehydrated for the subsequent days. The results showed a decrease in reserve mobilization started at day 5 of germination (Miazek *et al.*, 2001). Indeed, germinating seeds of bread wheat showed tolerance to dehydration up to the 4<sup>th</sup> day post-imbibition (Miazek *et al.*, 2001). Whereas, in other studies, the effect of osmotic stress on the germination process of seeds revealed a highly significant decrease in the germination percentage, germination rate and reserve mobilization (Almansouri *et al.*, 2001; Sayar *et al.*, 2010; Soltani *et al.*, 2006). This indicates the differential response of germinating seeds to the different water deficit treatments. Indeed, exposing *Arabidopsis thaliana* plants

\* Corresponding author. e-mail: harbhope78@gmail.com.

to two different water deficit regimes resulted in significant differences in the transcriptome profiles of the two treatments (Harb *et al.*, 2010). Therefore, it is of high importance to study the response of germinating seeds to water deficit that simulates to some extent the real stress in the field. This will lead to a better understanding of how water deficit affect the germination process of seeds, and help in the development of strategies to protect plants from the negative effect of water deficit as early as possible.

In this study, the effect of water deficit after the first hours of imbibition was tested in terms of reserve mobilization, total sugars and proteins during the first 3 days of germination after the onset of water deficit.

## 2. Materials and Method

### 2.1. Plant Material

Seeds of durum wheat (*Triticum durum* Desf. cv. Hourani 27) were provided by the National Center for Agriculture Research and Extension (NCARE), Jordan.

### 2.2. Water Uptake

Six groups of seeds of durum wheat (*Triticum durum* Desf. cv. Hourani 27) each group with 20 seeds was prepared. Then, their initial weight (W1) was taken before imbibition. After that, the seeds were put on wet filter paper in 9 cm plates and the plates were kept in an incubator set at 25°C under dark conditions for different time intervals: 1, 2, 3, 4, 6, 7, and 8 h. Final weight (W2) was taken every one hour after imbibition, and water uptake percentage was calculated:

$$\text{Water uptake\%} = (W2 - W1) / W1 * 100$$

### 2.3. Reserve Mobilization

Dry seeds without imbibition were oven dried at 104°C overnight. After that, their weight was taken and considered the original seed dry weight. The effect of water deficit after few hours of imbibition was tested; wheat seeds were kept on wet filter paper in 9 cm plates at 25°C under dark conditions for 2 and 8 h. Then, wheat seeds were transferred to dry filter paper in 9 cm plates after blotting the excess water. Samples were taken from treated and control (well-watered) plates at: 24, 48 and 72 h. Samples were oven dried at 104°C overnight, and their dry weights were taken (dry weight of seed remnants).

Reserve mobilization was calculated as follows:

Weight of mobilized reserve (mg seed<sup>-1</sup>) = (Original seed dry weight – Dry weight of seed remnants)/Number of seeds

Reserve depletion % = (Weight of mobilized reserve / Original seed dry weight) \* 100

### 2.4. Germination Under Water Deficit

Seeds were considered germinated when the radicle is 1 mm long. The germination percentage of wheat seeds exposed to imbibition for 2 and 8 h, followed by water deficit was calculated at 48 and 72 h of treatment.

### 2.5. Quantification of Sugars

Sugars were quantified by anthrone method (Yemm and Willis, 1954). Briefly, seeds were ground in 80% ethanol.

Then, a volume of the supernatant was reacted with anthrone reagent under boiling for 5-10 min. Glucose was the standard, and absorbance was read at 630 nm.

### 2.6. Quantification of Total Proteins

Total proteins in wheat seeds were quantified by Bradford's method (1976). Briefly, seeds were ground in distilled water, and then the mixture was centrifuged. Four (4) µL of the supernatant was mixed with 200 µL of Bradford reagent. The absorbance of the resultant product was read at 575nm. Standard curve of bovine serum albumin was run with the samples.

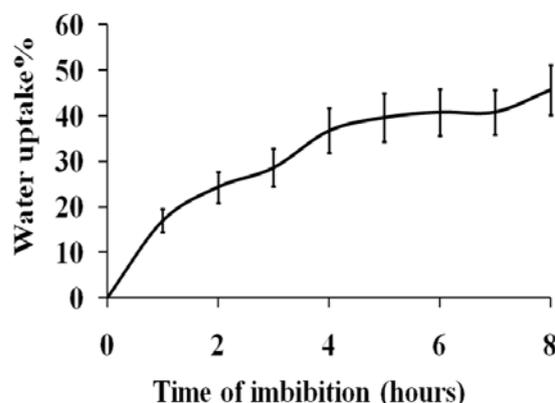
### 2.7. Statistical Analysis

All data were analyzed by Student's - T test using Excel software. Differences with p-value less than 0.05 were considered significant.

## 3. Results

### 3.1. Water Uptake

To determine at what time point water uptake by seeds will reach 25 and 50%, the water uptake of wheat seeds was monitored for the first 8 hours of imbibition. During this time period, an increase in the accumulated water content in seeds was shown (Fig. 1). After 2 h of imbibition, the accumulative water uptake was about 25%, and then it increases to reach 50% after 8 h of imbibition (Fig. 1).



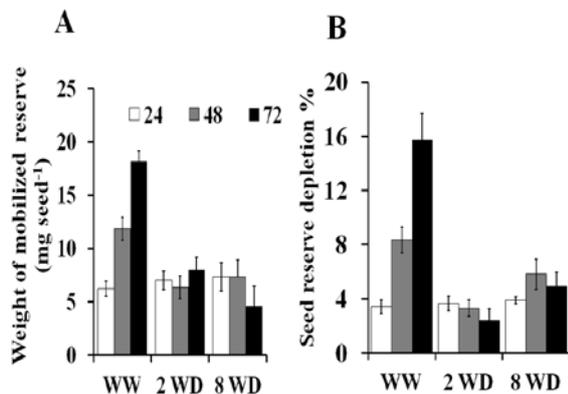
**Figure 1.** Water uptake during the first hours of imbibition in seeds of durum wheat. Error bars represent the standard errors of the means (n=8). The experiment was repeated with the same results.

### 3.2. Reserve Mobilization in Response to Water Deficit

To test if water absorbed during the first 8 h is enough to support all the phases of the germination process; wheat seeds imbibed for 2 and 8 h were dehydrated for 24, 48, and 72 h. After that, reserve mobilization was determined for both treatments at the three time points of water deficit. The results showed no change in reserve mobilization between the treated and the control seeds after 24 h of treatment. Whereas, the amount of water absorbed after 2 and 8 h was not enough for normal reserve mobilization after 48 and 72 h of water deficit. After 48 h of water deficit, reserve mobilization was 54% and 61% of the well-watered control at 2 and 8 h of imbibition, respectively

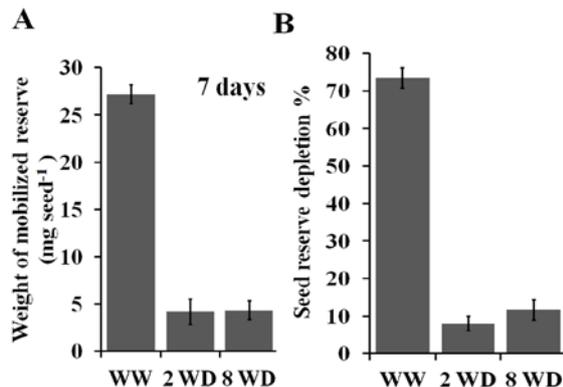
(Fig. 2A). Whereas, after 72 h of water deficit reserve mobilization decreased to 44% and 25% at 2 and 8 hours of imbibition, respectively (Fig. 2A).

The results of seed reserve depletion showed the same trend as that shown for reserve mobilization. No change in seed reserve depletion was shown after 24 h (Fig. 2B). Seed reserve depletion was reduced by about 5 and 4 % after 48 h of water deficit for the two imbibition periods (2 and 8 h), respectively (Fig. 2B). After 72 h of water deficit, the reductions were 13 and 11 % after 2 and 8 h of imbibition, respectively (Fig. 2B).



**Figure 2.** The effect of water deficit on reserve mobilization of seeds of durum wheat. A) Weight of mobilized reserve ( $\text{mg}\cdot\text{seed}^{-1}$ ) in wheat seeds exposed to water deficit at 2 and 8 hours of imbibition (2 WD and 8 WD, respectively) compared to the well-watered (WW) control. B) Seed reserve depletion % in wheat seeds exposed to water deficit at 2 and 8 hours of imbibition (2 WD and 8 WD, respectively) compared to the well-watered (WW) control. Error bars represent the standard errors of the means ( $n = 6$ ). The experiment was repeated with the same results.

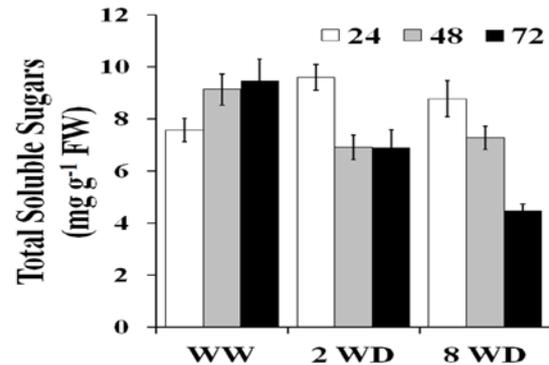
Reserve mobilization was determined after 7 days of water deficit at 2 and 8 h of imbibition. Both 2 and 8 h of imbibition showed a reduction in reserve mobilization and depletion of 84% compared to the well-watered control (Fig. 3A and B).



**Figure 3.** Reserve mobilization after 7 days of germination of seeds of durum wheat under water deficit (WD) compared to well-watered control (WW). A) Weight of mobilized reserve ( $\text{mg}\cdot\text{seed}^{-1}$ ) in seeds exposed to water deficit at 2 and 8 hours of imbibition (2 WD and 8 WD, respectively) compared to the well-watered control (WW). B) Reserve depletion percentage in seeds exposed to water deficit at 2 and 8 hours of imbibition (2 WD and 8 WD, respectively) compared to the well-watered control (WW). Error bars represent the standard errors of the means ( $n = 6$  for WD and 10 for WW). The experiment was repeated with the same results.

### 3.3. Total Sugars in Response to Water Deficit

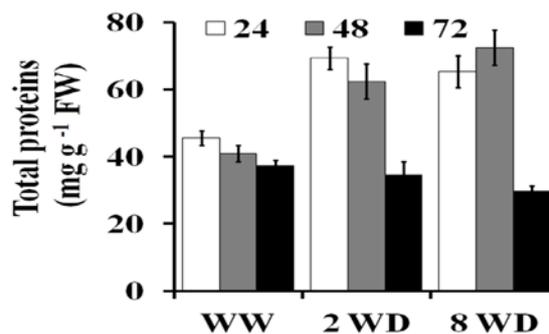
Soluble sugars were quantified in dehydrated seeds imbibed for 2 and 8 h of imbibition after 24, 48 and 72 h of water deficit. No change in soluble sugars was shown after 24 h of water deficit for the two imbibition periods (2 and 8 h) (Fig. 4). After 48 h of water deficit, soluble sugars decreased to 76 and 78 % compared to the control at 2 and 8 h of imbibition, respectively (Fig. 4). The reductions were 73 and 47 % at 2, and 8 h of imbibition after 72 h of water deficit, respectively (Fig. 4).



**Figure 4.** The effect of water deficit on the total soluble sugars in durum wheat seeds exposed to water deficit at 2 and 8 hours of imbibition (2 WD and 8 WD, respectively) compared to the well-watered (WW) control. Error bars represent the standard errors of the means ( $n = 6$ ). The experiment was repeated with the same results.

### 3.4. Total Proteins in Response to Water Deficit

After 24 and 48 h a highly significant increase in the concentration of total proteins was shown (Fig. 5). At 2 h of imbibition, the increases were 24 and 22  $\text{mg}\cdot\text{g}^{-1}$  fresh weight (FW) after 24 and 48 h of water deficit, respectively (Fig. 5). At 8 h of imbibition, the increases were 20 and 32  $\text{mg}\cdot\text{g}^{-1}$  FW after 24 and 48 h of water deficit, respectively (Fig. 5).



**Figure 5.** The effect of water deficit on the total proteins in durum wheat seeds exposed to water deficit at 2 and 8 hours of imbibition (2 WD and 8 WD, respectively) compared to the well-watered (WW) control. Error bars represent the standard errors of the means ( $n = 5$ ). The experiment was repeated with the same results.

### 3.5. Seed germination in response to water deficit

Water deficit at 2 and 8 h of imbibition resulted in a complete inhibition of germination after 48 and 72 h of water deficit.

#### 4. Discussion

The results of water uptake during the first 8 h of imbibition showed an increase in the accumulative water with time. This is consistent with the study of water uptake in barley (*Hordeum vulgare*) and bread wheat (*Triticum aestivum* L.) (Davidson *et al.*, 1976; Clarke, 1980, respectively). Factors related to the sowing medium and to the seed were found to affect water uptake by seeds (Davidson *et al.*, 1976; Clarke, 1980). Water potential and hydraulic conductivity of the soil impose limiting factors on the water uptake process (Ward and Shaykewich, 1972). Moreover, the hydraulic conductivity of the seeds is also playing a major role (Ward and Shaykewich, 1972). To exclude the effect of the initial seed size and other factors on the imbibition process, wheat seeds of the same size and from the same batch were used for the determination of water uptake in this study.

A reduction in reserve mobilization was revealed after 48, 72 h and after day 7 of water deficit at 2 and 8 h of imbibition. These results suggest that water deficit during the first hours of imbibition is detrimental to the biochemical and molecular changes needed for seed germination. In agreement, a study on durum wheat showed a highly significant decrease in reserve mobilization in response to osmotic stress after 7 days of imbibition (Soltani *et al.*, 2006).

Soluble sugars are one biochemical indicator of the efficiency of seed reserve mobilization. Water deficit at 2 and 8 h of imbibition inhibited the solubilization of sugars after 48 and 72 h of the treatment. This is in agreement with the results of the effect of osmotic stress on the degradation of sugars in durum wheat, which revealed a drastic decrease of soluble sugars after 48 h of treatment (Almansouri *et al.*, 2001). In contrast with our results, studies of germinating seeds of bread wheat showed an increase in soluble sugars started at day 2 of water deficit up to day 5 after 24 h of imbibition (Miazek *et al.*, 2001).

Proteins are major components of cereals' grain (Shewry and Halford, 2002; Šramková *et al.*, 2009; Triboi *et al.*, 2003). Seed proteins can be classified into: storage, structural, metabolic, and protective proteins (Shewry and Halford, 2002). In this study, the concentration of total proteins was increased after 24 and 48 h of water deficit following a short period of imbibition of 2 and 8 h. This increase in protein content might be explained as an acclimation strategy to water deficit (Nakashima *et al.*, 2009). It is known that structural and functional proteins are induced by environmental stresses such as water deficit (Bartels and Sunkar, 2005; Ramanjulu and Bartels, 2002). Indeed, in a study on two wheat cultivars with different tolerance to salinity, protein synthesis under salt stress was found as an adaptation strategy adopted by the salinity-tolerant cultivar (Dellaquila and Spada, 1993). In germinating wheat seeds, albumins and globulins started to increase at day 2 of water deficit under light conditions (Miazek *et al.*, 2001). In addition, in germinating wheat seeds, RNA synthesis started as early as 3 h of germination (Rejman and Buchowicz, 1973). This coincides with the first phase of imbibition, which is characterized by the rapid water uptake. This may suggest a crucial role for this phase in the success of the germination process.

In this study, germination was inhibited by water deficit after 2 and 8 h of imbibition. In bread wheat, dehydrated seeds after 24 h of imbibition were tolerant to water deficit up to the fifth day of imbibition (Miazek *et al.*, 2001). The effect of osmotic stress using polyethylene glycol (PEG) and/or mannitol on seed germination of wheat, showed a drastic inhibition of the percentage and rate of germination starting at day 2 of imbibition (Almansouri *et al.*, 2001; Blum *et al.*, 1980; Sayar *et al.*, 2010). The complete inhibition of germination as a result of water deficit can be explained by the complete absence of moisture, whereas, under osmotic stress, some moisture is available to allow low germination.

Intensive seedling growth occurs during the first 4 days of imbibition due to the intensive mobilization of seed reserves (Miazek *et al.*, 2001). Imbibition during the first hours of the germination process is crucial to provide the germinating seed with enough moisture for the biochemical and molecular changes during normal germination process. Hence, understanding the biochemical and molecular basis of water-seed relationship during the first hours of imbibition is invaluable for the improvement of plant's resistance to water deficit.

#### References

- Almansouri M, Kinet J and Lutts S. 2001. Effect of salt and osmotic stresses on germination in durum wheat (*Triticum durum* Desf.). *Plant and Soil*, **231**: 243–254.
- Bartels D and Sunkar R. 2005. Drought and salt tolerance in plants. *Crit Rev Plant Sci.*, **24**:23–58.
- Bewley J. 1997. Seed germination and dormancy. *Plant Cell*, **9**:1056–1066.
- Blum A, Sinemina B and Ziv D. 1980. An evaluation of seed and seedling drought tolerance screening tests in wheat. *Euphytica*, **29**: 727–736.
- Bradford M. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochem.*, **72**:248 – 254.
- Chaves M, Maroco J and Pereira J. 2003. Understanding plant responses to drought- from genes to the whole plant. *Functional Plant Biol.*, **30**: 239 –264.
- Clarke J. 1980. Measurement of relative water uptake rates of wheat seeds using agar media. *Canad J Plant Sci.*, **60**:1035–1038.
- Davidson D, Eastman M and Thomas J. 1976. Water uptake during germination of barley. *Plant Sci Lett.*, **6**:223–230.
- Dellaquila A and Spada P. 1993. The effect of salinity stress upon protein synthesis of germinating wheat embryos. *Annals of Botany*, **72**:97–101.
- Harb A, Krishnan A, Ambavaram MM and Pereira A. 2010. Molecular and physiological analysis of drought stress in *Arabidopsis* reveals early responses leading to acclimation in plant growth. *Plant Physiol.*, **154**:1254–1271.
- Marcus A, Feeley J and Volcani T. 1966. Protein synthesis in imbibed seeds III. Kinetics of amino acid incorporation ribosome activation, and polysome formation. *Plant Physiol.*, **41**:1167–1172.
- Miazek A Bogdan J and Zagdanska B. 2001. Effects of water deficit during germination of wheat seeds. *Biologia Plantarum*, **44**:397–403.

- Nakashima K Ito Y and Yamaguchi-Shinozaki K. 2009. Transcriptional regulatory networks in response to abiotic stresses in Arabidopsis and grasses. *Plant Physiol.*, **149**: 88–95.
- Passioura J. 2006. Increasing crop productivity when water is scarce-from breeding to field management. *Agricult Water Manag.*, **80**:176–196.
- Ramanjulu S and Bartels D. 2002. Drought- and desiccation induced modulation of expression in plants. *Plant Cell Environ.*, **25**:141–151.
- Rejman E and Buchowicz J. 1973. RNA synthesis during the germination of wheat seed. *Phytochem.*, **12**: 271–276.
- Sayar R Bchini H Mosbahi M and Ezzine M. 2010. Effects of salt and drought stresses on germination, emergence and seedling growth of durum wheat (*Triticum durum* Desf.). *J Agricult Res.*, **5**:2008–2016.
- Shewry PR and Halford NG. 2002. Cereal seed storage proteins: structures, properties and role in grain utilization. *J Experi Botany*, **53**: 947–958.
- Soltani A Gholipoor M and Zeinali E. 2006. Seed reserve utilization and seedling growth of wheat as affected by drought and salinity. *Environ Experi Botany*, **55**:195–200.
- Šramková Z , Gregová E and Šturdíka E. 2009. Chemical composition and nutritional quality of wheat grain. *Acta Chimica Slovaca* ,**2**:115–138.
- Triboř E Martre P and Triboř –Blondel AM. 2003. Environmentally-induced changes in protein composition in developing grains of wheat are related to changes in total protein content. *J Experi Botany*, **54**: 1731-1742.
- Ward J and Shaykewich C. 1972. Water absorption by wheat seeds as influenced by hydraulic properties of soil. *Canad J Soil Sci.*, **52**:99–105.
- Yemm E and Willis A. 1954. The estimation of carbohydrates in plant extracts by anthrone. *Biochem J.*, **57**:508–514.



# A Novel Report on the Prevalence of Enterohaemorrhagic *Escherichia coli* non-O157 Isolated from Cattle in Kaduna State, Nigeria

Jasini A. Musa<sup>1</sup>, Mashood A. Raji<sup>1</sup>, Haruna M. Kazeem<sup>1</sup> and Nicodemus M. Useh<sup>1, 2,\*</sup>

<sup>1</sup>Department of Veterinary Pathology and Microbiology, Ahmadu Bello University, Zaria, Nigeria

<sup>2</sup>Laboratory of Molecular Biology of Infectious Diseases, Department of Population Medicine & Diagnostic Sciences,

College of Veterinary Medicine, Cornell University, 14853 Ithaca, New York, United States of America

Received: September 22, 2012; Accepted: November 24, 2012

## Abstract

This study was carried out to investigate the presence of enterohaemorrhagic *Escherichia coli* (EHEC) of cattle that may pose a risk to human beings. Two hundred and forty (240) faecal samples were obtained from 8 randomly selected commercial cattle farms in Kaduna state, Nigeria. *E. coli* colonies from 76 (31.2 %) faecal samples were confirmed by Gram staining reactions and biochemically using indole, methyl red, Voges Proskauer and citrate (IMViC), triple sugar iron and motility tests respectively. Characterization of the isolates revealed three heterogeneous serogroups (O111, O118 and O126) from apparently healthy cattle, while no *E. coli* serogroup was isolated from diarrhoeic cattle. The prevalence of non-O157 isolates was 4.5 %. Association between the serogroups and source of samples (farms) was significant ( $P < 0.05$ ). The O126 serogroup isolated from apparently healthy cattle occurred more frequently, followed by O118 and O111 respectively. Although it is not known whether the presence of EHEC subgroups in apparently healthy cattle in the study areas may pose a health threat, it is safe to assume that the human population in these areas, including cattle rearers and veterinarians, is at risk of exposure to the EHEC subgroups reported in the study. Data from the study possibly suggest cattle as important source of enterohaemorrhagic *E. coli* in Kaduna State, Nigeria.

**Key Words:** Cattle, enterohaemorrhagic *Escherichia coli*, serogroups, Nigeria.

## 1. Introduction

The term 'enterohaemorrhagic *Escherichia coli*' (EHEC) was originally used to describe strains that cause haemorrhagic colitis (HC) and haemolytic-uraemic syndrome (HUS) (Nataro and Kaper, 1998), express shiga toxins (stx), cause attaching and effacing (A/E) lesions on epithelial cells and possess large plasmid. In accordance with the latest nomenclature, these strains are called shiga toxin-producing *E. coli* (formerly shiga-like toxin-producing *E. coli*) (WHO, 1998). Cattle appear to be the main reservoir of EHEC from which the organisms have been isolated (Clarke, 2001; Djordjevic *et al.*, 2001). *E. coli* O111 is the most frequently implicated non-O157 strain causing gastroenteritis with HUS, particularly in the United States of America and Europe (Bettelheim, 2000; Pearce *et al.*, 2006). Most studies indicated that majority of O111 serogroups were recovered from individuals with HC

and HUS than from cattle (Bettelheim, 2003). Cattle and human O118 serogroups represent the same clones and are similar in virulence attributes. Evidence for zoonotic transmission of *E. coli* O118 serogroups have been documented (Buchanan and Doyle, 1997).

*E. coli* O126 has been reportedly isolated from the faecal samples of cattle and human beings. The serogroup O126 has not been implicated in cases of haemolytic uraemic syndrome (Buchanan and Doyle, 1997; Bettelheim, 2000). Some other non-O157 serogroups of EHEC have been implicated in diarrhoea, HC and HUS in humans (Eklund *et al.*, 2001; Bettelheim, 2003). In the present study, we report the prevalence of enterohaemorrhagic *E. coli* from the faeces of cattle in Kaduna State, Nigeria for the first time.

\* Corresponding author. e-mail: nicodemus.useh@fulbrightmail.org.

## 2. Materials and Methods

### 2.1. Study Area

The study area was Kaduna State, which is located between latitude 10<sup>0</sup> and 11<sup>0</sup>N and longitude 7<sup>0</sup> and 8<sup>0</sup>E, North-Western Nigeria. Eight commercial cattle farms were randomly selected from five different local government areas of Kaduna State, Nigeria.

### 2.2. Sample Collection

A total of two hundred and forty (240) faecal samples from apparently healthy (233) and diarrhoeic (7) cattle were collected from 8 randomly selected commercial farms using stratified sampling technique (Field and Graham, 2003). Faecal material (1-2 g) was aseptically collected from the rectum of each animal using clean disposable hand gloves. The samples were placed in separate sterile bottles containing 9-10 mL of tryptone soya broth (TSB) and kept in a cold box at 4 °C, and transported to the Bacteriology Diagnostic Laboratory, Department of Veterinary Pathology and Microbiology, Ahmadu Bello University, Zaria, Nigeria and processed immediately.

### 2.3. Isolation and Identification of Suspected Colonies

Bacterial isolation, identification and biochemical tests were carried out using standard procedures described elsewhere (Barrow and Feltham, 1993; Cheesbrough, 2000). Briefly, samples were streaked on sorbitol macConkey agar and suspected positive colonies were confirmed using biochemical tests.

### 2.4. Biochemical Characterization

Colonies growing on sorbitol macConkey agar (SMAC) suspected to be *E. coli* were subjected to biochemical tests (indole, methyl red, Voges-Proskauer, citrate (IMViC), triple sugar iron, TSI and motility) (Cheesbrough, 2000).

### 2.5. Serogrouping of Somatic 'O' Isolates

All confirmed *E. coli* isolates were sub-cultured onto nutrient agar slants and stored at 4 °C for serogrouping (Blanco *et al.*, 2006). Somatic 'O' isolates of enterohaemorrhagic *Escherichia coli* O111, O118 and O126 were identified using monospecific *E. coli* antisera (SIFIN Berlin, Germany) (Blanco, 2006).

### 2.6. Statistical Analysis

Data obtained from the apparently healthy and diarrhoeic cattle were analyzed using Student's t-test and values of P<0.05 were significant.

## 3. Results

### 3.1. Spatial Distribution of Enterohaemorrhagic *E. coli*

Out of the 240 faecal samples collected from 8 randomly selected commercial cattle farms, the specific prevalence rate for each farm ranged between 0.0 % (Farm A, FA; Farm B, FB; Farm G, FG) and 17.4 % (Farm E, FE) respectively. A total of 11 (4.5 %) *E. coli* serogroups from apparently healthy cattle were found, of which 2 (8.7 %) isolated from Farm E (FE) and 1 (3.0 %) from FH were O111, 2 (8.7 %) from FE and 1 (4.4 %) from Farm F (FF) were O118, 1 (3.0 %) each from Farms C (FC), D (FD)

and H (FH), and 2 (8.7 %) from farm F (FF) were O126 serogroups respectively. *E. coli* serogroup O126 occurred more frequently, followed by O111 and O118 respectively. All the farms had one or more serogroups, except FA, FB and FG where no *E. coli* serogroup was isolated. A prevalence rate of 2.1 % was recorded for *E. coli* O126 and 1.2 % each for O111 and O118 respectively. The prevalence rate of non-O157 which was 4.5 % was statistically significant (P<0.05) (Table 1).

**Table 1.** Distribution of *E. coli* serogroups among commercial cattle farms in Kaduna State, Nigeria

Farm	Positive <i>E. coli</i> serogroups			
	Specific Prevalence (%)	O111	O118	O126
Farm A	0.0	0 (0.0)	0 (0.0)	0 (0.0)
Farm B	0.0	0 (0.0)	0 (0.0)	0 (0.0)
Farm C	3.0	0 (0.0)	0 (0.0)	1 (3.0)
Farm D	3.1	0 (0.0)	0 (0.0)	1 (3.0)
Farm E	17.4	2 (8.7)	2 (8.7)	0 (0.0)
Farm F	13.0	0 (0.0)	1 (4.4)	2 (8.7)
Farm G	0.0	0 (0.0)	0 (0.0)	0 (0.0)
Farm H	6.1	1 (3.0)	0 (0.0)	1 (3.0)
Total	4.5	3 (1.2)	3 (1.2)	5 (2.1)
$\alpha^2 = 0.04,$				
P< 0.05				

### 3.2. Distribution of Enterohaemorrhagic *E. coli* in Relation to Age

The age distribution of serogroups isolated from commercial cattle farms identified 1 (3.2 %) *E. coli* serogroup as O111, isolated from the young (0-1 year). *E. coli* O111, O118 and O126 were isolated from adults where O126 serogroup had the highest prevalence (7.4 %) rate. However, the specific prevalence varied among the adult species of cattle ranging between 4.5 % (in cattle older than 3 years) and 7.4 % (for those older than 1-2 years). The relationship observed between age and *E. coli* serogroups was not statistically significant (P>0.05) in this study.

### 3.3. Distribution of Enterohaemorrhagic *E. coli* in Relation to Breed

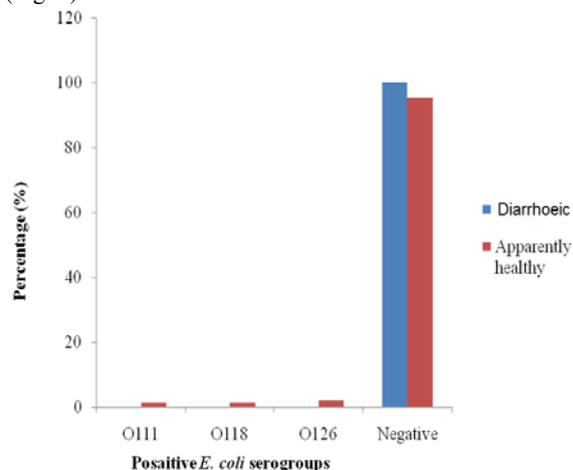
The specific prevalence rate ranged between 0.0 % (Holstein and Simmentals) and 8.1 % (Friesian) in the exotic breeds of cattle and 6.9% in locals (Rahaji). One (2.7 %) O111 and 2 (5.4 %) O126 serogroups were isolated from Friesian breed of cattle. At least one non-O157 serogroup was isolated from different types of local breeds. The relationship between breed and serogroups was not statistically significant (P>0.05).

### 3.4. Distribution of Enterohaemorrhagic *E. coli* in Relation to Sex

The relationship between sex and *E. coli* serogroups showed that *E. coli* serogroups were distributed according to the sex of cattle. A total of 1 (2.1 %) each for O111 and O118 serogroups and 2 (4.2 %) for O126 were isolated from males, while 2 (1 %) each for O111, O118 and 3 (1.5 %) for O126 were isolated in females. Overall, 11 (4.5 %) with one or more serogroups were identified. The relationship between sex and *E. coli* serogroups was not statistically significant ( $P>0.05$ ).

### 3.5. Distribution of Enterohaemorrhagic *E. coli* in Relation to Health Status

The relationship between health status and *E. coli* serogroups plummeted (0 %) in diarrhoeic and increased (4.5 %) in apparently healthy cattle respectively. Thus, the relationship between health status and *E. coli* serogroups from commercial cattle farms was not significant ( $P>0.05$ ) (Fig. 1).



**Figure 1.** Relationship between health status (%) and *E. coli* serogroups isolated from commercial cattle farms in Kaduna State, Nigeria

## 4. Discussion

The prevalence of *E. coli* non-O157 isolated from commercial cattle farms in Kaduna State, Nigeria, was 4.5 %. The authors found individual rates of 2.1 % in males and 1 % in females each for O111 and O118 serogroups respectively. This result agreed with the findings of Bettelheim (2003) and Pearce *et al.* (2006) who reported prevalence rate of 1-2 % for *E. coli* O111 and O118. Serogroup O126 is not frequently associated with disease in humans. Thus, the most common serogroups associated with disease in humans, which were also isolated from apparently healthy cattle in this study were *E. coli* O111 and O118. The prevalence of 1.7 % for O111 serogroup recorded in young animals agreed with the work of Blanco *et al.* (2000), who reported that calves are important reservoirs of *E. coli* non-O157. No *E. coli* serogroup was isolated from diarrhoeic cattle in this study, further supporting our suspicion that cattle may be reservoirs of colibacillosis in the area investigated.

The relationship between sex and the *E. coli* serogroups revealed that males (8.4 %) recorded higher number of *E.*

*coli* non-O157 serogroups as compared to females (3.5 %). Montenegro *et al.* (1990) reported 11.6 % and 3.0 % prevalence rates in cows and bulls using DNA hybridization technique, but the serogroups, except *E. coli* O126 were different. In addition, differences in the areas of study and the changing dynamics of disease may have contributed to this disparity. It is not known whether the presence of EHEC subgroups in apparently healthy cattle, reported in the current study may pose a significant health hazard to human beings residing in the areas investigated. However, it is safe to assume that the human population in these areas, including cattle rearers and veterinarians, is at risk of exposure to the EHEC subgroups reported in the study. Thus, there is the need for veterinary and human public health officials to educate the communities on the public health hazards of colibacillosis. In conclusion, this study revealed that cattle are important reservoir of EHEC in Kaduna state, Nigeria and research should be carried out to establish the extent to which human beings are at risk of being exposed, especially in tropical Nigeria, where cattle owners maintain close contact with animals in the residential areas.

## Acknowledgements

*E. coli* monospecific antiserum was generously donated by David Bukbuk, Head, Department of Veterinary Microbiology & Parasitology, University of Maiduguri, Nigeria. Manuscript was prepared using a facility provided by Professor Dr. Heinrich Neubauer, Director, Friedrich-Loeffler-Institute of Bacterial Infections and Zoonoses (*Clostridium* National Reference Laboratory), Federal Research Institute for Animal Health, Jena, Germany via a German Research Foundation (DFG) fellowship to NM Useh for which the authors are most grateful.

## References

- Barrow G I and Feltham R K A. 1993. **Cowan and Steel's Manual for the Identification of Medical Bacteria**, 3rd Edition. Cambridge University Press.
- Bettelheim K A. 2000. Role of non-O157 verocytotoxin-producing *Escherichia coli* (VTEC). *J Applied Microbiol.*, **88**: 38-50.
- Bettelheim KA. 2003. Non-O157 verotoxin-producing *Escherichia coli*: A problem, paradox, and paradigm. *Experimental Biol Med.*, **228**: 333-344.
- Blanco M, Blanco J E, Mora A, González E A, Blanco J. 2000. Serotypes and virulence genes of verocytotoxigenic *E. coli* (VTEC) isolated from cattle in Spain. In: Duffy, G., Garvey, D., Coia, P.J., Wasteson, Y. and McDowell D.A. (Eds), **Verocytotoxigenic *E. coli* in Europe, 3. Pathogenicity and virulence of verocytotoxigenic *E. coli***. Teagasc. The National Food Centre, Dublin.
- Blanco JA. 2006. A manual: Kit for *E. coli* serotyping, including O2, O26, O78, O86 and O141 sera. *Laboratorio de Referencia de E. coli (LREC)*, Espana, pp. 1-7.
- Blanco M, Blanco J E, Bahbi C A, Mora A, Alonso M P, Varela G, Gadea M P, Schelotto F, Gonzalez E A and Blanco J. 2006. Typing of intimin (eae) genes from enteropathogenic *Escherichia coli* (EPEC) isolated from children with diarrhoea in Montevideo, Uruguay: Identification of two novel intimin variants ( $\mu$ B and  $\beta$ R/ $\beta$ 2B). *J Med Microbiol.*, **55**: 1165-1174.

- Buchanan R L, Doyle M P. 1997. Food borne disease significance of *Escherichia coli* O157:H7 and other enterohaemorrhagic *E. coli*. *Food Technol.*, **51**: 69-76.
- Cheesbrough M. 2000. **District Laboratory Practice in Tropical Countries**. Lower price editions, part 2, Cambridge University Press.
- Clarke S C. 2001. Diarrhoeagenic *Escherichia coli*—an emerging problem? *Diag Microbiol Infect Dis.*, **41**: 93-98.
- Djordjevic S P, Hornitzky M A, Barley G, Gill P, Vanselow B, Walker K and K. A. Bettelheim. 2001. Virulence properties and serotypes of shiga toxin-producing *Escherichia coli* from healthy Australian slaughter-age sheep. *J Clin Microbiol.*, **39**: 2017- 2021.
- Eklund M, Scheutze F and Siitonen A. 2001. Clinical isolates of non-O157 shiga toxin-producing *Escherichia coli*: serotypes, virulence, characteristics and molecular profiles of strains of the same serotypes. *J Clin Microbiol.*, **39**: 2829-2834.
- Field A and Graham J H. 2003. **How To Design Experiments**. Sage publications Ltd.
- Montenegro M A, Bulte M, Trumpf T, Aleksic S, Reuter G, Bulling E and Helmuth R. 1990. Detection and characterization of faecal verotoxin-producing *Escherichia coli* from healthy cattle. *J Clin Microbiol.*, **25**: 1417-1421.
- Nataro J P and Kaper J B. 1998. Diarrhoeagenic *Escherichia coli*. *Clin Microbiol Rev.*, **11**: 142-201.
- Pearce M C, Evans J, McKendrick I J, Smith A W, Knight H I, Mellor D J, Woolhouse M. E J, Gunn G J and Low J C. 2006. Prevalence and virulence factors of *Escherichia coli* serogroups O26, O103, O111, and O145 shed by cattle in Scotland. *Applied Environ Microbiol.*, **7**: 653 659.
- World Health Organization (WHO). 1998. Zoonotic non-O157 shiga toxin-producing *Escherichia coli* (STEC). Report of a WHO scientific group meeting, Berlin, Germany. Department of Communicable Disease Surveillance and Response. June, 23-26, pp.1-38.



# Jordan Journal of Biological Sciences



An International Peer – Reviewed Research Journal

Published by the deanship of Research & Graduate Studies, The Hashemite University, Zarqa, Jordan

Name: ..... الاسم:

Specialty: ..... التخصص:

Address: ..... العنوان:

P.O. Box: ..... صندوق البريد:

City & Postal Code: ..... المدينة: الرمز البريدي:

Country: ..... الدولة:

Phone: ..... رقم الهاتف:

Fax No.: ..... رقم الفاكس:

E-mail: ..... البريد الإلكتروني:

Method of payment: ..... طريقة الدفع:

Amount Enclosed: ..... المبلغ المرفق:

Signature: ..... التوقيع:

Cheques should be paid to Deanship of Research and Graduate Studies – The Hashemite University.

I would like to subscribe to the Journal

**For**

- One year  
 Two years  
 Three years

### One Year Subscription Rates

	Inside Jordan	Outside Jordan
Individuals	JD10	\$70
Students	JD5	\$35
Institutions	JD 20	\$90

### Correspondence

#### Subscriptions and sales:

**Prof. Khaled H. Abu-Elteen**  
The Hashemite University  
P.O. Box 330127-Zarqa 13115 – Jordan  
Telephone: 00 962 5 3903333 ext. 4399  
Fax no. : 0096253903349  
E. mail: jjbs@hu.edu.jo



Hashemite Kingdom of Jordan



Hashemite University

# المجلة الأردنية للعلوم الحياتية

مجلة علمية عالمية محكمة  
تصدر بدعم من صندوق دعم البحث العلمي

# المجلة الأردنية للعلوم الحياتية

## مجلة علمية عالمية محكمة

المجلة الأردنية للعلوم الحياتية : مجلة علمية عالمية محكمة ومفهرسة ومصنفة، تصدر عن الجامعة الهاشمية و بدعم من صندوق دعم البحث العلمي- وزارة التعليم العالي و البحث العلمي .

### هيئة التحرير

#### رئيس التحرير:

الأستاذ الدكتور خالد حسين أبو التين

الجامعة الهاشمية، الزرقاء، الأردن .

#### الأعضاء:

الأستاذ الدكتور سوسن عطاالله العوران  
الجامعة الأردنية

الأستاذ الدكتور شتيوي صالح عبدالله  
الجامعة الأردنية

الأستاذ الدكتور عبدالكريم جبر السلال  
جامعة العلوم والتكنولوجيا الأردنية

الأستاذ الدكتور نبيل البشير  
جامعة العلوم و التكنولوجيا الأردنية

الأستاذ الدكتور حكم فائق الحديدي  
جامعة العلوم والتكنولوجيا الأردنية

الأستاذ الدكتور خالد أحمد الطراونة  
جامعة مؤتة

الأستاذ الدكتور سامي خضر عبدالحافظ  
جامعة البرموك

الأستاذ الدكتور علي زهير الكرمي"  
الجامعة الهاشمية

### فريق الدعم:

#### تنفيذ وإخراج

م. مهدي عقده

#### المحرر اللغوي

الدكتور قصي الذبيان

### ترسل البحوث إلى العنوان التالي :

رئيس تحرير المجلة الأردنية للعلوم الحياتية

عمادة البحث العلمي و الدراسات العليا

الجامعة الهاشمية

الزرقاء – الأردن

هاتف : ٣٩٠٣٣٣٣ ٥ ٠٠٩٦٢٢ فرعي ٤١٤٧

Email: jjbs@hu.edu.jo, Website: www.jjbs.hu.edu.jo