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In silico Tools and Techniques for Screening and Development of Peptide-Based Spike Protein Inhibitors against Novel Coronavirus (Sars-CoV-2)

Raghunath Satpathy*

School of Biotechnology, Gangadhar Meher University, Amruta Vihar, Sambalpur Odisha, 768004, India Received: December 17, 2021; Revised: March 13, 2022; Accepted: April 27, 2022

Abstract

The recent pandemic situation created by the novel coronavirus (SARS-CoV-2) across the globe is a great concern. So, the discovery of novel antiviral agents is desirable to address this challenge. In this context, the antiviral peptides (AVPs) possess an enormous potential and can be considered to develop novel therapeutic strategies to combat SARS-CoV-2 infection. The anti-viral peptides are mostly preferable over small inhibitor molecules for having high target specificity and lower side effects. The spike protein is an important structural protein of SARS-CoV-2 that binds with the human angiotensin-converting enzyme-2 leading to host entry of the virus. Hence, the activity of the anti-viral peptides will be based on the interference of the peptide inhibitor between the binding site of spike protein, and the ACE2 protein ultimately will prevent the virus invasion process. Several database resources are available that contain many anti-viral peptides from natural sources. However, the experimental basis of establishing the therapeutic importance of every protein from the database is a difficult and time-consuming task. Hence the available bioinformatics tools and techniques can be suitably used to screen, structure prediction, evaluation of ant-viral peptide- SARS-CoV-2 spike protein interaction, toxicity prediction, molecular dynamics simulation, and so on. In this review, the implementation of some of the major computational tools, their availability, and effectiveness in predicting the peptides against the Spike protein have been discussed.

Keywords: Antiviral peptides, novel coronavirus, Spike Protein, Bioinformatics, Screening Methods, Binding Affinity

1. Introduction

The novel coronavirus disease was first identified in China during the last month of 2019. Further, this novel virus (SARS-CoV-2) infection continued to outbreak globally at an alarming rate. The genomic sequence study of the virus has been observed to share some homology with the related virus such as (Middle East respiratory syndrome-CoV) and the coronavirus (SARS-CoV) [1-3]. High infection, as well as the death rate of the novel coronavirus in comparison to other previously known coronaviruses, created the research challenge to discover the potential drug candidates for the pharmacological treatments. From the beginning of the pandemic, several molecules have been repurposed and proposed to prevent the infection of the virus; however, till yet, no such effective drug is available for the treatment [4-6]. Some of the repurposed drugs including remdesivir, favipiravir, lopinavir, ritonavir, ribavirin for the inhibitor of viral RNA- replication have been approved but there is no evidence regarding their clinical efficacy [7-11]. In the host-virus interaction process of SARS-CoV-2, the Spike glycoproteins (molecular weight 180-200 kDa) play a major role. The Spike glycoproteins (S) contain an Nterminus region (extracellular), transmembrane (TM)

* Corresponding author. e-mail: rnsatpathy@gmail.com.

anchored region, and a short C-terminal region (Figure 1). It was studied that, the *open* conformation of the S-protein facilitates the virus interaction with ACE2 protein of host (human) and further leads to the fusion process with the host cell membrane. The S protein also contains the polysaccharide coating that prevent the virus from the host immune system recognition process [12-14].

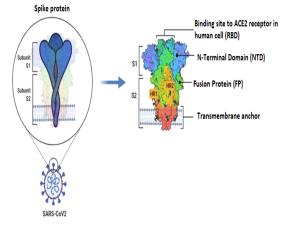


Figure 1. Structural components of the spike glycoprotein (SARS-CoV-2)

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| Subunit of Spike Protein | S. No | Specific target region of S protein | Sequences regions | Function |
|-----------------------------|-------|-------------------------------------|----------------------|---|
| | 1 | N-terminal domain (NTD) | 13-305 | Helps in attachment and recognition of virus |
| S1 | 2 | Receptor binding domain (RBD) | 319-541 | Binds to the cell receptor hACE2 |
| | 3 | Receptor binding motif (RBM) | 437-508 | Binds to the cell receptor of hACE2 |
| | 1 | Fusion peptide (FP) | 788-806 | Helps in anchoring the target membrane with the S protein, and play important role in mediating membrane fusion |
| | 2 | Heptad repeat 1 (HR1) | 912-984 | Essential for the viral fusion and cell entry mechanism |
| S2 | 3 | Heptad repeat 2 (HR2) | 1163-1213 | Essential for the viral fusion and entry function of the S2 subunit |
| | 4 | Transmembrane domain (TM) | 1213-1237 | The downstream TM domain anchors the S protein to the viral membrane |

Table 1. Target regions of the Sars -CoV-2 spike glycoprotein

The length of the spike protein of the SARS-CoV-2 virus comprises 1273 amino acids long, and it is further divided into two subunits, S1and S2, which are actively involved in the host cell entry process. The spike protein is trimeric in its native form and the S1 and S2 subunits of the virus form a "crown" structure from which the term "corona" is originated in the Latin language translation. The S1 subunit is classified into N-terminal domain (NTD), the receptor-binding domain (RBD), and the carboxyterminal domain (CTD). The CTD domain also contains two subdomains such as SD1 and SD2. Similarly, the S2 subunit consists of two regions such as heptapeptide repeat 1 (HR1) and heptapeptide repeat 2 (HR2). The HR1 act as a fusion peptide and the HR2 act as the transmembrane domain (TM) (Figure 1). In the viral entry process, the S protein directly interacts with the cell surface receptor, human angiotensin-converting enzyme-2 (hACE2). Particularly, the open conformation state of the RBD domain is accessible for binding with the hACE2. After the binding event to the human ACE2 receptor, S1 and S2 cleavage sites are accessed by the host protease due to the conformational change in the S protein [15-21]. The detailed sequence position and the function of the different components of the spike protein have been shown in Table and the mechanism is shown in Figure 1, 2.

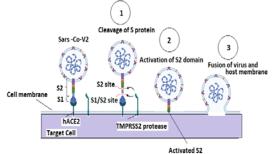


Figure 2. Basic entry mechanism of SARS-CoV-2 into the host cell

Soon after the process, the proteolytic cleavage occurs at the position of S1 and S2 subunit junction site as well as at the S2 subunit region with the help of a serine protease enzyme TMPRSS2. Following this cleavage event, HR1 and HR2 regions of the S2 subunit interact to form a structural arrangement known as *fusion core*. The *fusion core* formation further enhances the entry of the virus into the host cell by the cellular endocytic mechanism. Finally, the viral RNA is released into the host cell and the viral replication process is initiated. Interestingly, research has established that the SARS-CoV-2 shows an enhanced affinity towards binding (about 10 to 20-fold) with hACE2, as compared to another related virus such as SARS-CoV [22-23].

Recently, efforts have been given to discover potential antiviral therapeutic molecules. The traditional biochemical approach of small molecule inhibitors is associated with several obstacles like drug resistance, huge cost, timeconsuming as well as toxicity [24-25]. In this context, the antiviral peptides (AVPs) are considered as one of the important therapeutic molecules due to their specificity, efficacy, potency, and desirable pharmacokinetic properties [26]. For the last 50 years, Antiviral peptides (AVPs) have shown their effectiveness, hence generating a perspective for treating viral infections such as novel coronavirus. The key feature of antiviral peptides are, they are involved in the specific interaction and inhibition of target protein. Therefore, targeting the AVPs in the form of novel therapeutics against the pathogenic virus-like SARS-CoV-2 might be established as a promising tool to develop an effective treatment process. Prediction and application of effective anti-viral peptides against respiratory diseasecausing viruses like SARS-CoV, SARS-CoV2, MERS-CoV have been described and reported by many researchers [27-33].

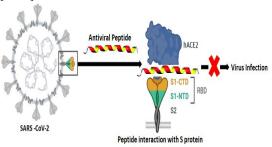


Figure 3. Possible mechanism of action of the anti-viral peptides with the spike protein

The potential AVPs are to be identified, hence they can be targeted specifically to the RBD domains in the S protein of novel coronavirus so that, the interference will restrict the host cell receptor-mediated viral entry. The possible mechanism of AVP- based inhibition of spike protein is presented in Figure 3. The binding features of the Spike protein of SARS-CoV-2 with ACE2 have been studied, and it was revealed several charged amino acids of ACE2 are important for its interaction [34]. Li et al. also studied the binding affinity of peptides that binds to the

730

specific hotspot area of the spike protein by computing the rate of inhibition [35].

The objective of the review is to provide a basic methodology that is involved in the searching and development of an effective spike inhibitor peptide molecule. Additionally, the challenges and scope of the process have been discussed.

2. In-silico development process of peptide molecules against the SARS-CoV-2 spike protein

An experimental basis for searching for novel and effective anti-viral peptide molecules that can specifically block the S protein function of the novel coronavirus is a challenging task. However, the computational approach facilitates the process for searching and predicting an effective antiviral peptide. This method will also lead to discovering and understanding the mechanism behind the spike protein-peptide interaction that corresponds to the antiviral effects [36-39]. Several steps are to be followed in the in-silico based novel effective peptide discovery against the spike protein inhibitor are schematically presented in Figure 4 and described in the sections below.

3. Retrieval of peptide sequence information

The information about the antiviral peptide sequence information from natural sources can be obtained from several available databases. Searching the literature regarding the specific peptides that can inhibit the spike protein of the virus may also help to explore the potential one. Along with the amino acid sequences, several databases also contain an analytical tool for peptide analysis. Some of the important databases and their availability that contains anti-viral peptides are represented in Table 2.

Table 2. Important databases contain information about anti-viral peptides

| Name of the database | Availability |
|---|--|
| AVPdb Database | https://webs.iiitd.edu.in/raghava/satpdb/catalogs/avpdb |
| Antimicrobial Peptide Database (APD) | https://aps.unmc.edu/database/anti |
| Database of Antimicrobial Activity and Structure of Peptides (DBAASP) | https://dbaasp.org/ |
| LAMP2 | http://biotechlab.fudan.edu.cn/database/lamp/ |
| CAMPR3 (Collection of Anti-Microbial Peptides) | http://www.camp.bicnirrh.res.in/index.php |
| DRAMP (Data repository of antimicrobial peptides) | http://dramp.cpu-bioinfor.org/ |
| | AVPdb Database Antimicrobial Peptide Database (APD) Database of Antimicrobial Activity and Structure of Peptides (DBAASP) LAMP2 CAMPR3 (Collection of Anti-Microbial Peptides) |

4. Screening criteria for the peptides

After retrieval of the anti-viral peptide sequences from the database search, it is essential to screen the spike binding activity of these peptide sequences based on a certain feature. For example, Mustafa et al. studied several features of the probable active peptides against the MERS-CoV spike protein [40]. As the MERS-CoV spike protein resembles the SARS-CoV-2 spike protein, hence similar type of criteria can be used to screen the potential peptides [41-42]. In addition to this, several strategies have been employed by many researchers for screening probable spike peptides having spike binding characteristics [43-46]. Some of the criteria for screening of potential peptide molecules are:

(i) *length of amino acids* should be 20aa to 55aa (length of the peptide sequence is proportional to the antiviral activity)

(ii) *positively charged residues* should be more abundant

(iii) *net charge* of the peptide > 0 as the spike protein of Sars- CoV-2 is neutral and less negatively charged in nature

(iv) the peptide should be *non-toxic* to human cells

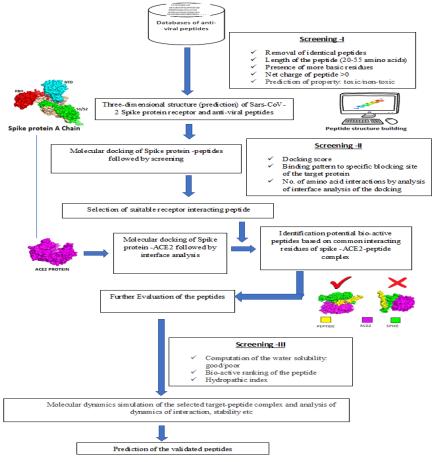


Figure 4. steps for in silico prediction of novel bioactive peptides against spike proteins

5. Prediction of the 3D structure of spike glycoproteins and anti-viral peptides

The protein data bank (PDB) available at https://www.rcsb.org/ may be searched for the availability of the three-dimensional structure of the (SARS-CoV-2) spike proteins as well as for the small peptides. Recently, Gowthaman et al. developed a database Cov 3D (available at https://cov3d.ibbr.umd.edu), which contains coronavirus

structural data from the PDB including the spike protein [47]. Also, the 3D structure prediction of the spike protein as a target can be performed from the amino acid sequence followed by validation. UniProt database (https://www.uniprot.org) is usually used to retrieve the amino acid sequence information of the spike protein. Several tools available for the 3D structure prediction of spike protein, are being used by many researchers (Table 3).

| Table 3. Popular 3D structure prediction tools for peptide molecules | |
|--|--|
| | |

| S. No | Name of the tool | Availability | Recently used by researchers for spike protein structure prediction | |
|-------|------------------|---|--|--|
| 1 | SWISS MODEL | https://swissmodel.expasy.org/ | (Padilla-Sanchez 2020; Allam et al. 2020) | |
| 2 | Modeller | https://salilab.org/modeller/ | (Hall et al. 2020; Martin et al. 2020; Hassanzadeh et al. 2020) | |
| 3 | Phyre 2 | http://www.sbg.bio.ic.ac.uk/phyre2/html/p age.cgi?id=index | (Jaimes et al. 2020) | |
| 4 | RaptorX | http://raptorx.uchicago.edu/ | (King et al. 2021; Awadelkareem et al. 2020) | |
| 5 | I Tasser | https://zhanggroup.org/I-TASSER/ | (Prashantha et al. 2021; Ibrahim et al. 2020) | |

structure can be predicted by using online server like pepfold (available at https://bioserv.rpbs.univ-parisdiderot.fr/services/PEP-FOLD/). This web-based tool predicts the 3D structure of linear peptides sequences from 5 to 50 amino acid range. Also, other open-source standalone programs like open Chimera (https://www.cgl.ucsf.edu/chimera/) can be used to build peptide structure from the sequences. Many of the proteinprotein docking programs take the peptide sequence along with the target protein sequence as input and automatically predict the 3D structure thereby preparing the structural file for the docking simulation. After the prediction of the 3D structure of the peptide, the energy minimization should be performed till it attains negative free energy. Evaluation of the predicted structure can be performed by computing the Ramachandran plot, side-chain placement, and so on. For the structure verification of the predicted structures, the SAVES servers may be used (https://saves.mbi.ucla.edu/).

6. Molecular docking and molecular dynamics (MD) simulation study of anti-viral peptides with SARS-CoV-2 spike proteins

To compute the binding affinity as well as the binding pose of the selected potential anti-viral peptide with the virus spike protein, molecular docking is to be performed. Molecular docking is a computational approach in which the affinity, pose of the ligand is evaluated along with the receptors. Mainly two types of algorithms are involved in this process, searching algorithm and scoring algorithms. Based on the implementation of the combinations of algorithms, different types of docking programs are available [48-49].

The protein-peptide docking programs are broadly classified into three categories. The first one is templatebased docking (a protein-peptide input structure is searched for a known template structure from the database followed by comparative analysis). The second method is known as local docking (the peptide binding site is searched on the given input receptor by the user given peptides). The third method is global docking in which the peptide behaves as rigid in nature and binding pose and position are evaluated by the exhaustive search on the receptor. Also, in the molecular docking process of protein and peptide, the number of flexible bonds, size, a loop structure, and terminal charges in the peptide structure are the important parameters on which the accuracy of the prediction depends [50-51]. In this process, the Spike protein is behaving as the receptor and the peptide sequence as the ligand and the binding affinity, the binding pose can be computed from the receptor-ligand complex by application of suitable searching and scoring functions in several docking programs [52-60], presented in Table 4.

Table 4. Molecular docking programs that are frequently used for protein-peptide interaction study

| GalaxyPepDock Rosetta FlexPepDock PepCrawler | http://galaxy.seoklab.org/pepdock http://flexpepdock.furmanlab.cs.huji.ac.il |
|--|---|
| 1 | http://flexpepdock.furmanlab.cs.huji.ac.il |
| PepCrawler | |
| | http://bioinfo3d.cs.tau.ac.il/PepCrawler |
| HADDOCK peptide docking | http://milou.science.uu.nl/services/HADDOCK2.2/haddock.php |
| GRAMM-X | http://vakser.compbio.ku.edu/resources/gramm/grammx |
| DINC 2.0 | http://dinc.kavrakilab.org |
| pepATTRACT | http://bioserv.rpbs.univ-paris-diderot.fr/services/pepATTRACT |
| CABS-dock | http://biocomp.chem.uw.edu.pl/CABSdock |
| ClusPro PeptiDock | https://peptidock.cluspro.org |
| PIPER-FlexPepDock | http://piperfpd.furmanlab.cs.huji.ac.il |
| HawkDock server | http://cadd.zju.edu.cn/hawkdock |
| Z dock server | https://zdock.umassmed.edu |
| SwarmDock server | https://bmm.crick.ac.uk/~svc-bmm-swarmdock |
| pyDockWEB | https://life.bsc.es/pid/pydockweb |
| 3D garden | http://www.sbg.bio.ic.ac.uk/3dgarden |
| Hex server | http://hexserver.loria.fr |
| DOCKSCORE | http://caps.ncbs.res.in/dockscore |
| FRODOCK | https://chaconlab.org/modeling/frodock |
| HPEPDOCK Server | http://huanglab.phys.hust.edu.cn/hpepdock |
| MDockPeP | https://zougrouptoolkit.missouri.edu/mdockpep |
| ClusPro 2.0 | https://cluspro.org |
| | GRAMM-X DINC 2.0 pepATTRACT CABS-dock ClusPro PeptiDock PIPER-FlexPepDock HawkDock server Z dock server SwarmDock server pyDockWEB 3D garden Hex server DOCKSCORE FRODOCK HPEPDOCK Server MDockPeP |

In the computational procedure, after molecular docking simulation, the implementation of a reliable specific scoring function is the basic requirement for the prediction of the correct binding nature of the protein-peptide complex. In this context, theoretically calculation of the end-point binding free energy by the methods such as Molecular mechanics Poisson–Boltzmann surface area (MM/PBSA) is used. By using this method, the binding affinities, as well as the binding conformations, are obtained for protein-peptide complex systems [61]. In the MM-PBSA method, the free energy (ΔG) of receptor-ligand (protein-peptide) binding is estimated by deducting

the free energy between the complex and the unbound form. Previously, the MM-PBSA method has been successfully used to analyze and evaluate the proteinprotein docking complexes [62-65]. A significant number of specific viral protein-peptide interactions is used for the determination of affinity of the potential AVPs and used for the in-depth understanding of the behaviour towards the target molecule. To investigate the spike proteinpeptide interaction, it is essential to study the interface region of the docked complex. The interface analysis results from the numbers of amino acid interactions with the specific binding domains. This also leads to evaluating the different types of interaction such as cation $-\pi$, hydrogen bonding, and electrostatic interaction essential to describe the anti-viral inhibition property of the peptide [66-67].

Molecular dynamics (MD) simulation is computational method used for understanding and prediction of the structure as well as the function of biological macromolecules. This method is based on the physical principles (Newton's equations of motion). The macromolecular conformation is represented by the dynamic model from which the motion of the individual atoms and subsequent conformational changes are calculated from a trajectory file. Specifically, by the MD simulation, the macroscopic behaviour from various microscopic interactions present in molecular systems is computed. The MD simulation methods are frequently used in the drug discovery process to study the receptors (targets) and their association with ligand (drug) molecules after docking to discover the novel molecules by computing various parameters. MD simulation can also be used to study the configuration change in the protein structure that involves the monomeric or the trimeric form of the spike glycoprotein. In addition to this, a comparative structural analysis of the ligand-bound state of spike protein and native (unbound) state can be performed. Currently, the MD simulation techniques are used in the case of spike protein to study the proper binding of the ligand, total energy variation profile of the complex, dynamicity of protein domain, variation of hydrogen bonds, dynamics of actively interacting residues of the spike protein throughout the simulation period and so on [68-69].

 Table 7. Molecular dynamics simulation software and their availability

7. Identification of bioactive peptide molecules

Along with the docking score and the *pose* of the binding peptide and several physicochemical screenings are also essential to select the potential peptide inhibitor molecules. The major problem is in the therapeutic use of peptides is their toxicity. Hence the peptide features like toxicity, bioavailability, in *vivo* instability, half-life, and ability to cross the membrane are to be evaluated carefully. These factors obtained from the peptide evaluation output can be utilized further in the specific rational design process of novel peptides with enhanced stability and other required physicochemical properties [70-71]. Some of the important bioinformatics tools used to compute such peptide properties are given in Table 6.

Table 6: Peptide designing and screening tools and their availability

Several scientific studies on the peptide-based inhibitors against the pathogenic virus have been developed and proposed in the last few years. The finding of small effective peptides was considered to be one of the best molecules that can be targeted to the spike protein [72]. Wong et al., in 2020 conducted *in silico* study to identify potential SARS-CoV-2 cell entry inhibitors from peptides derived from different edible insects. In-silicobased gastrointestinal (GI) digestion of the insect protein was performed to generate peptide molecules. Subsequent molecular docking study with the spike protein resulted

that, a tri-peptide generated from the source mealworm can act as the effective inhibitor [73]. Allam et al. analyzed the peptide and polyphenol-based inhibitor that can block the SARS-CoV-2 spike protein by inhibiting the glucoseregulating protein 78 receptors. The in-silico screening process was conducted by taking the available databases of bioactive peptides. Protein-peptide docking analysis resulted in five potential peptides that can inhibit the GRP78 binding site [74]. Salman et al., conducted the computational protein-protein interaction study to obtain the effective peptides from the inhalers that can bind the spike protein. Molecular docking simulation of S protein (receptor) with different compounds such as alpha-1antitrypsin, dornase-alfa, angiotensin-converting enzyme-2 (ACE-2), human palate were analyzed. The peptide molecules obtained were further predicted as the potential anti- SARS-CoV-2 agents [75]. Protein peptide docking followed by the interface analysis study also showed that the amphibian derived peptides such as Dermaceptin-9 from the amphibian genus Phyllomedusa is an effective spike protein inhibitor of Sars- CoV-2 [76-77]. In 2020, Barh et al. implemented three different in silico strategies to design the potential novel peptide inhibitors that can interact and inhibit the Sars CoV-RBD - hACE2 interaction. The key binding residues were identified from the interaction study followed by identifying the peptide binders from the bacterial peptide database. Then a chimeric peptide was designed that is capable of binding the key residues of the SARS-CoV-2 -RBD - hACE2 complex. For the screening of the best potential peptides, the parameters like physiochemical properties, numbers, and positions of key residues binding, binding energy, and antiviral properties were considered [78]. Alibakhshi et al. conducted a computational study to design the effective peptides by targeting the RBD of SARS-CoV-2 spike protein and human ACE2 interaction. The analysis was based on taking stretches of peptides from the ACE2 protein. Further, the different mutants were designed by computational method, and interaction of the peptides with spike protein was studied by using the molecular docking simulation method. As a subsequent study, molecular dynamics simulation was carried out to evaluate the best mutant peptide that interacts with the S protein in comparison to the wild peptide [79]. A similar study was conducted by Panda et al., to design a peptide having a resemblance to human ACE-2 protein. The non-interacting residues of ACE2 were mutated to generate a mutant peptide library. The molecular docking by HADDOCK server followed by molecular dynamics simulation (150 nanoseconds) by Gromacs tool was used to identify the novel mutant peptide developed the enhanced binding affinity about three times [80].

8. Challenges and Future research scope

There is a continuous increasing trend in the knowledge level in the experimental data related to the structure of SARS-CoV-2 and its entry process to the host cell. Much of the information is available now in the form of published literature and databases. This data provides a solid background to apply the computational techniques such as modeling, identification, and characterization of novel antiviral peptides that would interfere with spike-ACE2 binding and the membrane fusion process. Although the peptide molecules are advantageous over other small molecules, properties of the peptides like toxicity, immunogenicity, and stability of peptides remain as the major issues and need to be addressed in the development process of peptide-based inhibitors [81-85].

Moreover, the following specific challenges are associated with the discovery process of potential peptide inhibitors against the spike protein of SARS-CoV-2 and need to be addressed.

- S1 RBD domain of Spike glycoprotein is part of a highly mutable region, hence targeting to inhibit the domain is challenging, however, specifically targeting the peptide molecules to the HR region of the S2 subunit, with enhanced affinity might be effective to prevent the SARS-CoV-2 infections.
- Other than binding affinity property of AVPs to Sprotein, prediction of several important parameters such as penetration capacity to the tissue, stability in the plasma, protease enzyme degradation potential, immunological interference, and toxicity is necessary.
- The full-length wild-type S protein structure of SARS-CoV-2 is still not available in the database. Hence important missing amino acid residues and loops might affect the in-silico results regarding peptide-protein binding. Hence, the complete form of the crystal structure of the spike protein (both wild-type as well as variants) is to be determined.

9. Conclusions

SARS-CoV-2 is evolved as a novel pathogenic virus with a significantly enhanced infection rate. Due to the deadliest nature of this coronavirus, it is essential to search for an effective medication to respond to this infectious disease to avoid pandemic situations. Specifically, the interaction of the spike glycoprotein of the virus with the human ACE2 receptor causes the viral entry into the host cell. So, the use of peptide-based inhibitor molecules against the spike protein will be a promising approach to inhibit the process. Currently, the therapeutic application of peptide molecules is implemented against many diseases. Anti-viral peptides are most often preferred over small molecules due to their specific, few side effects, and no drug resistance activity. So, the study about targeting the spike glycoprotein of the virus with the potential novel anti-viral peptides will create a milestone in this emerging area. However, the mutation acquired by the virus, in vivo bioavailability, toxicity, and stability of anti-viral peptides are the major limitations. The available sophisticated bioinformatics tools and databases can be implemented for structure prediction of peptides, binding affinity study, validation, and dynamics analysis, in silico toxicity, bioactivity, and stability prediction, and so on. Furthermore, these results obtained from the in-silico analysis can be further combined along with the experimental results, for the development of proper validated peptide-based inhibitors by targeting the SARS-CoV-2 glycoprotein.

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Conflict of interest

Nothing

References

Santos-López G, Cortés-Hernández P, Vallejo-Ruiz V, Reyes-Leyva J. SARS-CoV-2: basic concepts, origin and treatment advances. Gac Med Mex. 2021;157(1):84-9. doi: 10.24875/GMM.M21000524, PMID 34125824.

Hu B, Guo H, Zhou P, Shi ZL. Characteristics of SARS-CoV-2 and COVID-19. Nat Rev Microbiol. 2021;19(3):141-54. doi: 10.1038/s41579-020-00459-7, PMID 33024307.

Yang J, Petitjean SJL, Koehler M, Zhang Q, Dumitru AC, Chen W, Derclaye S, Vincent SP, Soumillion P, Alsteens D. Molecular interaction and inhibition of SARS-CoV-2 binding to the ACE2 receptor. Nat Commun. 2020;11(1):4541. doi: 10.1038/s41467-020-18319-6, PMID 32917884.

V'kovski P, Kratzel A, Steiner S, Stalder H, Thiel V. Coronavirus biology and replication: implications for SARS-CoV-2. Nat Rev Microbiol. 2021;19(3):155-70. doi: 10.1038/s41579-020-00468-6, PMID 33116300.

Murgolo N, Therien AG, Howell B, Klein D, Koeplinger K, Lieberman LA, Adam GC, Flynn J, McKenna P, Swaminathan G, Hazuda DJ, Olsen DB. SARS-CoV-2 tropism, entry, replication, and propagation: considerations for drug discovery and development. PLOS Pathog. 2021;17(2):e1009225. doi: 10.1371/journal.ppat.1009225, PMID 33596266.

Trougakos IP, Stamatelopoulos K, Terpos E, Tsitsilonis OE, Aivalioti E, Paraskevis D, Kastritis E, Pavlakis GN, Dimopoulos MA. Insights to SARS-CoV-2 life cycle, pathophysiology, and rationalized treatments that target COVID-19 clinical complications. J Biomed Sci. 2021;28(1):9. doi: 10.1186/s12929-020-00703-5, PMID 33435929.

Martinez MA. Compounds with therapeutic potential against novel respiratory 2019 coronavirus. Antimicrob Agents Chemother. 2020;64(5):e00399-20. doi: 10.1128/AAC.00399-20, PMID 32152082.

Martinez MA. Clinical trials of repurposed antivirals for SARS-CoV-2. Antimicrob Agents Chemother. 2020;64(9):e01101-20. doi: 10.1128/AAC.01101-20, PMID 32631826.

Doi Y, Hibino M, Hase R, Yamamoto M, Kasamatsu Y, Hirose M, Mutoh Y, Homma Y, Terada M, Ogawa T, Kashizaki F, Yokoyama T, Koba H, Kasahara H, Yokota K, Kato H, Yoshida J, Kita T, Kato Y, Kamio T, Kodama N, Uchida Y, Ikeda N, Shinoda M, Nakagawa A, Nakatsumi H, Horiguchi T, Iwata M, Matsuyama A, Banno S, Koseki T, Teramachi M, Miyata M, Tajima S, Maeki T, Nakayama E, Taniguchi S, Lim CK, Saijo M, Imai T, Yoshida H, Kabata D, Shintani A, Yuzawa Y, Kondo M. A prospective, randomized, open-label trial of early versus late favipiravir therapy in hospitalized patients with COVID-19. Antimicrob Agents Chemother. 2020;64(12):e01897-20. doi: 10.1128/AAC.01897-20, PMID 32958718.

Cao B, Wang Y, Wen D, Liu W, Wang J, Fan G, Ruan L, Song B, Cai Y, Wei M, Li X, Xia J, Chen N, Xiang J, Yu T, Bai T, Xie X, Zhang L, Li C, Yuan Y, Chen H, Li H, Huang H, Tu S, Gong F, Liu Y, Wei Y, Dong C, Zhou F, Gu X, Xu J, Liu Z, Zhang Y, Li H, Shang L, Wang K, Li K, Zhou X, Dong X, Qu Z, Lu S, Hu X, Ruan S, Luo S, Wu J, Peng L, Cheng F, Pan L, Zou J, Jia C, Wang J, Liu X, Wang S, Wu X, Ge Q, He J, Zhan H, Qiu F, Guo L, Huang C, Jaki T, Hayden FG, Horby PW, Zhang D, Wang C. A trial of lopinavir–ritonavir in adults hospitalized with severe Covid-19. N Engl J Med. 2020;382(19):1787-99. doi: 10.1056/NEJMoa2001282, PMID 32187464.

Martinez MA. Lack of effectiveness of repurposed drugs for COVID-19 treatment. Front Immunol. 2021;12:635371. doi: 10.3389/fimmu.2021.635371, PMID 33777024.

Du L, He Y, Zhou Y, Liu S, Zheng BJ, Jiang S. The spike protein of SARS-CoV—a target for vaccine and therapeutic

development. Nat Rev Microbiol. 2009;7(3):226-36. doi: 10.1038/nrmicro2090, PMID 19198616.

Koley T, Madaan S, Chowdhury SR, Kumar M, Kaur P, Singh TP, Ethayathulla AS. Structural analysis of COVID-19 spike protein in recognizing the ACE2 receptor of different mammalian species and its susceptibility to viral infection. 3 Biotech. 2021;11(2):109. doi: 10.1007/s13205-020-02599-2, PMID 33552834.

Li F. Structure, function, and evolution of coronavirus spike proteins. Annu Rev Virol. 2016;3(1):237-61. doi: 10.1146/annurev-virology-110615-042301, PMID 27578435.

Henderson R, Edwards RJ, Mansouri K, Janowska K, Stalls V, Gobeil SMC, Kopp M, Li D, Parks R, Hsu AL, Borgnia MJ, Haynes BF, Acharya P. Controlling the SARS-CoV-2 spike glycoprotein conformation. Nat Struct Mol Biol. 2020;27(10):925-33. doi: 10.1038/s41594-020-0479-4, PMID 32699321.

Tang T, Bidon M, Jaimes JA, Whittaker GR, Daniel S. Coronavirus membrane fusion mechanism offers a potential target for antiviral development. Antiviral Res. 2020;178:104792. doi: 10.1016/j.antiviral.2020.104792.

Ke Z, Oton J, Qu K, Cortese M, Zila V, McKeane L, Nakane T, Zivanov J, Neufeldt CJ, Cerikan B, Lu JM, Peukes J, Xiong X, Kräusslich HG, Scheres SHW, Bartenschlager R, Briggs JAG. Structures and distributions of SARS-CoV-2 spike proteins on intact virions. Nature. 2020;588(7838):498-502. doi: 10.1038/s41586-020-2665-2, PMID 32805734.

Wrapp D, Wang N, Corbett KS, Goldsmith JA, Hsieh CL, Abiona O, Graham BS, McLellan JS. Cryo-EM structure of the 2019nCoV spike in the prefusion conformation. Science. 2020;367(6483):1260-3. doi: 10.1126/science.abb2507, PMID 32075877.

Xia S, Liu M, Wang C, Xu W, Lan Q, Feng S, Qi F, Bao L, Du L, Liu S, Qin C, Sun F, Shi Z, Zhu Y, Jiang S, Lu L. Inhibition of SARS-CoV-2 (previously 2019-nCoV) infection by a highly potent pan-coronavirus fusion inhibitor targeting its spike protein that harbors a high capacity to mediate membrane fusion. Cell Res. 2020;30(4):343-55. doi: 10.1038/s41422-020-0305-x, PMID 32231345.

Sakkiah S, Guo W, Pan B, Ji Z, Yavas G, Azevedo M, Hawes J, Patterson TA, Hong H. Elucidating interactions between SARS-CoV-2 trimeric spike protein and ACE2 using homology modeling and molecular dynamics simulations. Front Chem. 2020;8:622632. doi: 10.3389/fchem.2020.622632, PMID 33469527.

Walls AC, Park YJ, Tortorici MA, Wall A, McGuire AT, Veesler D. Structure, function, and antigenicity of the SARS-CoV-2 spike glycoprotein. Cell. 2020;181(2):281-292.e6. doi: 10.1016/j.cell.2020.02.058, PMID 32155444.

Kang S, Peng W, Zhu Y, Lu S, Zhou M, Lin W, Wu W, Huang S, Jiang L, Luo X, Deng M. Recent progress in understanding 2019 novel coronavirus (SARS-CoV-2) associated with human respiratory disease: detection, mechanisms and treatment. Int J Antimicrob Agents. 2020;55(5):105950. doi: 10.1016/j.ijantimicag.2020.105950, PMID 32234465.

Letko M, Marzi A, Munster V. Functional assessment of cell entry and receptor usage for SARS-CoV-2 and other lineage B betacoronaviruses. Nat Microbiol. 2020;5(4):562-9. doi: 10.1038/s41564-020-0688-y, PMID 32094589.

Chang KY, Yang JR. Analysis and prediction of highly effective antiviral peptides based on random forests. PLOS ONE. 2013;8(8):e70166. doi: 10.1371/journal.pone.0070166, PMID 23940542.

Castel G, Chtéoui M, Heyd B, Tordo N. Phage display of combinatorial peptide libraries: application to antiviral research.

 Molecules.
 2011;16(5):3499-518.

 10.3390/molecules16053499, PMID 21522083.

doi:

Charoenkwan P, Anuwongcharoen N, Nantasenamat C, Hasan MM, Shoombuatong W. In silico approaches for the prediction and analysis of antiviral peptides: a review. Curr Pharm Des. 2021;27(18):2180-8. doi: 10.2174/1381612826666201102105827, PMID 33138759.

Kaspar AA, Reichert JM. Future directions for peptide therapeutics development. Drug Discov Today. 2013;18(17-18):807-17. doi: 10.1016/j.drudis.2013.05.011, PMID 23726889.

Bruno BJ, Miller GD, Lim CS. Basics and recent advances in peptide and protein drug delivery. Ther Deliv. 2013;4(11):1443-67. doi: 10.4155/tde.13.104, PMID 24228993.

Schütz D, Ruiz-Blanco YB, Münch J, Kirchhoff F, Sanchez-Garcia E, Müller JA. Peptide and peptide-based inhibitors of SARS-CoV-2 entry. Adv Drug Deliv Rev. 2020;167:47-65. doi: 10.1016/j.addr.2020.11.007, PMID 33189768.

Mahendran ASK, Lim YS, Fang CM, Loh HS, Le CF. The potential of antiviral peptides as COVID-19 therapeutics. Front Pharmacol. 2020;11:575444. doi: 10.3389/fphar.2020.575444, PMID 33041819.

Sharma A, Pant K, Pande A, Sinha S, Pant B. Modeling novel anti-viral peptides (AVPs) with in-silico docking simulations against corona virus. Mater Today Proc. 2021;46:11169-76. doi: 10.1016/j.matpr.2021.02.377, PMID 33680868.

Agarwal G, Gabrani R. Antiviral peptides: identification and validation. Int J Pept Res Ther. 2021;27:149-68.

Mahmud S, Biswas S, Kumar Paul G, Mita MA, Afrose S, Robiul Hasan M, Sharmin Sultana Shimu M, Uddin MAR, Salah Uddin M, Zaman S, Kaderi Kibria KM, Arif Khan M, Bin Emran T, Abu Saleh M. Antiviral peptides against the main protease of SARS-CoV-2: A molecular docking and dynamics study. Arab J Chem. 2021;14(9). doi: 10.1016/j.arabjc.2021.103315, PMID 103315.

Dai C, Ma Y, Zhao Z, Zhao R, Wang Q, Wu Y, Cao Z, Li W. Mucroporin, the first cationic host defense peptide from the venom of Lychas mucronatus. Antimicrob Agents Chemother. 2008;52(11):3967-72. doi: 10.1128/AAC.00542-08, PMID 18779362.

Li Q, Zhao Z, Zhou D, Chen Y, Hong W, Cao L, Yang J, Zhang Y, Shi W, Cao Z, Wu Y, Yan H, Li W. Virucidal activity of a scorpion venom peptide variant mucroporin-M1 against measles, SARS-CoV and influenza H5N1 viruses. Peptides. 2011;32(7):1518-25. doi: 10.1016/j.peptides.2011.05.015, PMID 21620914.

Mitra D, Pandey J, Jain A, Swaroop S. In silico design of multiepitope-based peptide vaccine against SARS-CoV-2 using its spike protein. J Biomol Struct Dyn. 2021:1-14. doi: 10.1080/07391102.2020.1869092, PMID 33403946.

Stoddard SV, Wallace FE, Stoddard SD, Cheng Q, Acosta D, Barzani S, Bobay M, Briant J, Cisneros C, Feinstein S, Glasper K, Hussain M, Lidoski A, Lingareddy P, Lovett G, Matherne L, McIntosh J, Moosani N, Nagge L, Nyamkondiwa K, Pratt I, Root E, Rutledge MR, Sawyer M, Singh Y, Smith K, Tanveer U, Vaghela S. In silico design of peptide-based SARS-CoV-2 fusion inhibitors that target WT and mutant versions of SARS-CoV-2 HR1 domains. Biophysica. 2021;1(3):311-27. doi: 10.3390/biophysica1030023.

Ling R, Dai Y, Huang B, Huang W, Yu J, Lu X, Jiang Y. In silico design of antiviral peptides targeting the spike protein of SARS-CoV-2. Peptides. 2020;130:170328. doi: 10.1016/j.peptides.2020.170328.

Alibakhshi A, Ahangarzadeh S, Beikmohammadi L, Soltanmohammadi B, Bahrami AA, Ranjbar MM, Mohammadi E. Computational design of a potential therapeutic peptide against spike protein of SARS-CoV-2. J Comput Biophys Chem. 2021;20(4):337-46. doi: 10.1142/S2737416521500162.

Mustafa S, Balkhy H, Gabere M. Peptide-protein interaction studies of antimicrobial peptides targeting middle east respiratory syndrome coronavirus spike protein: an in-silico approach. Adv Bioinformatics. 2019;2019:6815105. doi: 10.1155/2019/6815105, PMID 31354813.

Bangaru S, Ozorowski G, Turner HL, Antanasijevic A, Huang D, Wang X, Torres JL, Diedrich JK, Tian JH, Portnoff AD, Patel N, Massare MJ, Yates JR, Nemazee D, Paulson JC, Glenn G, Smith G, Ward AB. Structural analysis of full-length SARS-CoV-2 spike protein from an advanced vaccine candidate. Science. 2020;370(6520):1089-94. doi: 10.1126/science.abe1502, PMID 33082295.

Fikriani CN, Ardana IKKG, Listyorini D. The comparison of SARS-CoV-2, SARS-CoV, and MERS-CoV genome and spike protein variations. J Riset Biologi Aplikasinya. 2021;3(1):38-44. doi: 10.26740/jrba.v3n1.p38-44.

Wang G, Li X, Wang Z. APD3: the antimicrobial peptide database as a tool for research and education. Nucleic Acids Res. 2016;44(D1):D1087-93. doi: 10.1093/nar/gkv1278, PMID 26602694.

Du L, Yang Y, Zhou Y, Lu L, Li F, Jiang S. MERS-CoV spike protein: a key target for antivirals. Expert Opin Ther Targets. 2017;21(2):131-43. doi: 10.1080/14728222.2017.1271415, PMID 27936982.

Zhao H, Zhou J, Zhang K, Chu H, Liu D, Poon VKM, Chan CC, Leung HC, Fai N, Lin YP, Zhang AJ, Jin DY, Yuen KY, Zheng BJ. A novel peptide with potent and broad-spectrum antiviral activities against multiple respiratory viruses. Sci Rep. 2016;6(1):22008. doi: 10.1038/srep22008, PMID 26911565.

Wang G. Natural antimicrobial peptides as promising anti-HIV candidates. Curr Top Pept Protein Res. 2012;13:93-110. PMID 26834391.

Gowthaman R, Guest JD, Yin R, Adolf-Bryfogle J, Schief WR, Pierce BG. CoV3D: a database of high resolution coronavirus protein structures. Nucleic Acids Res. 2021;49(D1):D282-7. doi: 10.1093/nar/gkaa731, PMID 32890396.

Morris, G. M.; Lim-Wilby, M. Molecular Docking. In. Methods in Molecular Biology; Humana Press 2008, (365–382). DOI: 10.1007/978-1-59745-177-2_19

Satpathy, R. Application of Molecular Docking Methods on Endocrine Disrupting Chemicals: A Review. J. Appl. Biotechnol. Rep. 2020; 7: 74–80.

Wang, J.; Alekseenko, A.; Kozakov, D.; Miao, Y. Improved Modeling of Peptide-Protein Binding Through Global Docking and Accelerated Molecular Dynamics Simulations. Front. Mol. Biosci. 2019; 6: 112. DOI: 10.3389/fmolb.2019.00112

Ciemny, M.; Kurcinski, M.; Kamel, K.; Kolinski, A.; Alam, N.; Schueler-Furman, O.; Kmiecik, S. Protein–Peptide Docking: Opportunities and Challenges. Drug Discov. Today 2018, 23: 1530–37. DOI: 10.1016/j.drudis.2018.05.006

Padilla-Sanchez V, Padilla-Sanchez V, Padilla-Sanchez V, Padilla-Sanchez V, Padilla-Sanchez V, Padilla-Sanchez V, Padilla-Sanchez V. In silico analysis of SARS-CoV-2 spike glycoprotein and insights into antibody binding. Res Ideas Outcomes. 2020;6:e55281. doi: 10.3897/rio.6.e55281.

Allam M, Ismail A, Khumalo ZTH, Kwenda S, Van Heusden P, Cloete R, ... & Bhiman JN 2020. Whole genome sequence of the severe acute respiratory syndrome coronavirus 2 SARS-COV-2 obtained from A South African coronavirus Disease 2019 Covid19 patient. *Accessed on*, 2.

Hassanzadeh K, Perez Pena H, Dragotto J, Buccarello L, Iorio F, Pieraccini S, Sancini G, Feligioni M. Considerations around the

SARS-CoV-2 Spike Protein with particular attention to COVID-19 brain infection and neurological symptoms. ACS Chem Neurosci. 2020;11(15):2361-9. doi: 10.1021/acschemneuro.0c00373, PMID 32627524.

Martin WR, Cheng F. Repurposing of FDA-approved toremifene to treat COVID-19 by blocking the Spike glycoprotein and NSP14 of SARS-CoV-2. J Proteome Res. 2020;19(11):4670-7. doi: 10.1021/acs.jproteome.0c00397, PMID 32907334.

Jaimes JA, André NM, Chappie JS, Millet JK, Whittaker GR. Phylogenetic analysis and structural modeling of SARS-CoV-2 spike protein reveals an evolutionary distinct and proteolytically sensitive activation loop. J Mol Biol. 2020;432(10):3309-25. doi: 10.1016/j.jmb.2020.04.009, PMID 32320687.

King M, Dinulos A, DaMota J, Walton E, Dahlquist K. A bioinformatics approach to investigating the structural and functional consequences of SNPs in TMPRSS2 for COVID-19 infection. FASEB J. 2021;35(S1). doi: 10.1096/fasebj.2021.35.S1.04753.

Awadelkareem EA, Ali SA. Vaccine design of coronavirus spike (S) glycoprotein in chicken: immunoinformatics and computational approaches. Transl Med Commun. 2020;5(1):13. doi: 10.1186/s41231-020-00063-0, PMID 32869000.

Prashantha CN, Gouthami K, Lavanya L, Bhavanam S, Jakhar A, Shakthiraju RG, Suraj V, Sahana KV, Sujana HS, Guruprasad NM, Ramachandra R. Molecular screening of antimalarial, antiviral, anti-inflammatory and HIV protease inhibitors against spike glycoprotein of coronavirus. J Mol Graph Model. 2021;102:107769. doi: 10.1016/j.jmgm.2020.107769.

Ibrahim IM, Abdelmalek DH, Elshahat ME, Elfiky AA. COVID-19 spike-host cell receptor GRP78 binding site prediction. J Infect. 2020;80(5):554-62. doi: 10.1016/j.jinf.2020.02.026, PMID 32169481.

Weng G, Wang E, Chen F, Sun H, Wang Z, Hou T. Assessing the performance of MM/PBSA and MM/GBSA methods. 9. Prediction reliability of binding affinities and binding poses for protein–peptide complexes. Phys Chem Chem Phys. 2019;21(19):10135-45. doi: 10.1039/c9cp01674k, PMID 31062799.

Massova I, Kollman PA. Computational alanine scanning to probe protein–protein interactions: a novel approach to evaluate binding free energies. J Am Chem Soc. 1999;121(36):8133-43. doi: 10.1021/ja990935j.

Hou T, Wang J, Li Y, Wang W. Assessing the performance of the MM/PBSA and MM/GBSA methods. I. The accuracy of binding free energy calculations based on molecular dynamics simulations. J Chem Inf Model. 2011;51(1):69-82. doi: 10.1021/ci100275a, PMID 21117705.

Spiliotopoulos D, Kastritis PL, Melquiond AS, Bonvin AM, Musco G, Rocchia W, Spitaleri A. dMM-PBSA: a new HADDOCK scoring function for protein-peptide docking. Front Mol Biosci. 2016;3:46. doi: 10.3389/fmolb.2016.00046, PMID 27630991.

Genheden S, Ryde U. The MM/PBSA and MM/GBSA methods to estimate ligand-binding affinities. Expert Opin Drug Discov. 2015;10(5):449-61. doi: 10.1517/17460441.2015.1032936, PMID 25835573.

Lim H, Baek A, Kim J, Kim MS, Liu J, Nam KY, Yoon J, . Hot spot profiles of SARS-CoV-2 and human ACE2 receptor protein interaction obtained by density functional tight binding fragment molecular orbital method. Sci Rep. 2020;10(1):16862. doi: 10.1038/s41598-020-73820-8, PMID 33033344.

Ortega JT, Serrano ML, Pujol FH, Rangel HR. Role of changes in SARS-CoV-2 spike protein in the interaction with the human ACE2 receptor: an in silico analysis. Excli J. 2020;19:410-7. doi: 10.17179/excli2020-1167, PMID 32210742.

Dror RO, Dirks RM, Grossman JP, Xu H, Shaw DE. Biomolecular simulation: a computational microscope for molecular biology. Annu Rev Biophys. 2012;41:429-52. doi: 10.1146/annurev-biophys-042910-155245, PMID 22577825.

Justino GC, Nascimento CP, Justino MC. Molecular dynamics simulations and analysis for bioinformatics undergraduate students. Biochem Mol Biol Educ. 2021;49(4):570-82. doi: 10.1002/bmb.21512, PMID 33844418.

Ali R, Rani R, Kumar S. New peptide based therapeutic approaches. In: Ashraf GMd, Sheikh IA, editors. Advances in protein chemistry. Jeddah: OMICS Group eBooks; 2013.

Gentilucci L, De Marco R, Cerisoli L. Chemical modifications designed to improve peptide stability: incorporation of non-natural amino acids, pseudo-peptide bonds, and cyclization. Curr Pharm Des. 2010;16(28):3185-203. doi: 10.2174/138161210793292555, PMID 20687878.

Pomplun, S. Targeting the SARS-CoV-2-spike Protein: From Antibodies to Miniproteins and Peptides. R.S.C. Med. Chem. 2020; 12:197–202. DOI: 10.1039/d0md00385a

Wong, F. C.; Ong, J. H.; Chai, T. T. Identification of Putative Cell-Entry-Inhibitory Peptides Against SARS-CoV-2 from Edible Insects: An in-Silico Study. EFOOD 2020; 1 : 357–68. DOI: 10.2991/efood.k.200918.002

Allam, L.; Ghrifi, F.; Mohammed, H.; El Hafidi, N.; El Jaoudi, R.; El Harti, J.; Lmimouni, B.; Belyamani, L.; Ibrahimi, A. Targeting the GRP78-Dependant SARS-CoV-2 Cell Entry by Peptides and Small Molecules. Bioinformatics Biol. Insights 2020; 14: 1177932220965505. DOI: 10.1177/1177932220965505

Salman, S.; Shah, F. H.; Chaudhry, M.; Tariq, M.; Akbar, M. Y.; Adnan, M. In Silico Analysis of Protein/Peptide-Based Inhalers Against SARS-CoV-2. Future Virol. 2020; 15 : 557–64. DOI: 10.2217/fvl-2020-0119.

Fakih, T. M. Dermaseptin-Based Antiviral Peptides to Prevent COVID-19 through In Silico Molecular Docking Studies Against SARS-Cov-2 Spike Protein. Pharm. Sci. Res. 2020; 7: 65–70. DOI: 10.7454/psr.v7i4.1079

Satpathy, R. In Silico Prediction of Anti-SARS-CoV-2 Effect of Dermaseptin Peptides from Amphibian Origin. Trends Pept. Protein Sci. 2020; 5: 1–9.

Barh, D.; Tiwari, S.; Silva Andrade, B.; Giovanetti, M.; Almeida Costa, E.; Kumavath, R.; Ghosh, P.; Góes-Neto, A.; Carlos Junior Alcantara, L.; Azevedo, V. Potential Chimeric Peptides to Block the SARS-CoV-2 Spike Receptor-Binding Domain. F1000Res. 2020; 9: 576. DOI: 10.12688/f1000research.24074.1

Alibakhshi, A.; Ahangarzadeh, S.; Beikmohammadi, L.; Soltanmohammadi, B.; Bahrami, A. A.; Ranjbar, M. M.; Mohammadi, E. Computational Design of a Potential Therapeutic Peptide Against Spike Protein of SARS-CoV-2. J. Comput. Biophys. Chem. 2021; 20 (4): 337–46. DOI: 10.1142/S2737416521500162

Panda, S. K.; Sen Gupta, P. S.; Biswal, S.; Ray, A. K.; Rana, M. K. ACE-2-derived Biomimetic Peptides for the Inhibition of Spike Protein of SARS-CoV-2. J. Proteome Res. 2021; 20 (2): 1296–1303. DOI: 10.1021/acs.jproteome.0c00686.

Efaz FM, Islam S, Talukder SA, Akter S, Tashrif MZ, Ali MA, Sufian MA, Parves MR, Islam MJ, Halim MA. Repurposing fusion inhibitor peptide against SARS-CoV-2. J Comp Chem. 2021;42(32):2283-93. doi: 10.1002/jcc.26758, PMID 34591335.

Chowdhury AS, Reehl SM, Kehn-Hall K, Bishop B, Webb-Robertson BM. Better understanding and prediction of antiviral peptides through primary and secondary structure feature importance. Sci Rep. 2020;10(1):19260. doi: 10.1038/s41598-020-76161-8, PMID 33159146.

Vlieghe P, Lisowski V, Martinez J, Khrestchatisky M. Synthetic therapeutic peptides: science and market. Drug Discov Today. 2010;15(1-2):40-56. doi: 10.1016/j.drudis.2009.10.009, PMID 19879957.

Thotakura N, Kaushik L, Kumar V, Preet S, Babu PV. Advanced approaches of bioactive peptide molecules and protein drug delivery systems. Curr Pharm Des. 2018;24(43):5147-63. doi: 10.2174/1381612825666190206211458, PMID 30727874.

Uusna J, Langel K, Langel Ü. Toxicity, immunogenicity, uptake, and kinetics methods for CPPs Cell-Penetrating Peptides. Methods Mol Biol. 2015;1324:(133-48). doi: 10.1007/978-1-4939-2806-4 9, PMID 26202267.

738