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Protective effect of *Ferula hermonis* root extract against cycram-induced DNA, biochemical and testicular damage in rats

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Abstract

Cycram (CYC), or cyclophosphamide is an alkylating drug that has been widely used in the acute treatment of various neoplastic diseases and in the chronic treatment of autoimmune disorders. The major limitation of CYC chemotherapy is the injury of normal tissue, leading to multiple organ toxicity and alter male fertility in mice, rats and human, leading to testicular damage and decreasing in RNA, DNA and protein synthesis. The present study aims to evaluate the protective role of methanol extract of Ferula hermonis (FH) against CYC induced changes in testicular RNA, DNA and protein content as well sperm morphology, count and motility, blood levels of testosterone (T) and hemoglobin (Hb) in adult male albino rats. Ninety adult albino male rats were divided into 3 main treated groups (30 animals each) according to treatment periods (15, 30 and 60 days), every one divided to 3 subgroups according to CYC dose (50, 100 and 200 mg/kg bw/day, 10 animals each), and each subgroup divided to 2 sub-sub groups treated with CYC and CYC+ FH 0.025 ml/100 g bw/day by gavage, respectively, (5 animals each), besides the control (10 animals). Animals were sacrificed, sperm morphology with its count and motility, as well RNA, DNA and protein content, and testosterone and hemoglobin levels were determined. The results revealed that CYC treatment induced alterations in RNA, DNA and protein synthesis, as well as affected T and Hb levels and sperm abnormalities of male rats. However, supplementation of FH extract protects these parameters against CYC toxicities. The protective actions of FH extract seem to be closely involved with the suppressing of plasma lipid peroxidation and increasing of antioxidant enzyme activities. Therefore, FH extract might be used in combination with CYC in cancer patients' therapy, transplantation and autoimmune diseases as a protective agent against CYC-induced reproductive toxicity.

Keywords: Ferula hermonis, protect, Cycram, DNA, Damage, Testicular, Biochemical, Rats.

1. Introduction

Cycram (CYC) or cyclophosphamide is a cytotoxic alkylating drug that has been widely used in the acute treatment of various neoplastic diseases and in the chronic treatment of autoimmune disorders. The major limitation of CYC chemotherapy is the injury of normal tissue, leading to multiple organ toxicity mainly in the heart, testes and urinary bladder (Fraiser et al., 1991; Ghobadi et al., 2017). CYC was reported to alter human fertility and is cytotoxic to rapidly dividing cells, which makes the highly proliferative testes a target for the damaging effects of this drug. Use of CYC for the treatment of cancer in male patients increases the incidence of oligo- and azoospermia and results in male infertility (Howell and Shalet, 1998). Previous studies have demonstrated that chronic administration of CYC to male rats/mice leads to decreased testicular weight, transitory oligospermia, decreased DNA synthesis in spermatogonia and protein synthesis in spermatids (Anderson et al., 1995; Meistrich et al., 1995; Kaur et al., 1997).

Molecular components of sperm like RNA have the potential to be important indicators of reproductive

toxicity. This is especially important for long-term exposures, such as those experienced in chemotherapeutic regimens, in which cytotoxic drugs are often given for months rather than days. Exposures that target germ cells for extended periods of time, as chemotherapy drugs do, may produce changes in the molecular contents of the sperm that can serve as biomarkers of testicular dysfunction (Dere et al., 2013). Rats treated with CYC were displayed lowest weight of testicles and epididymis (reproductive organs) and the lowest motility, progression, viability, and sperm count (Shabanian et al., 2017), a decrease in serum hormone levels (testosterone [T], follicle stimulating hormone [FSH], and luteinizing hormone [LH]) (Johari et al., 2011), as well as 20 % reduction in the hemoglobin (Hb) level were found (Cengiz, 2018).

Numerous studies have shown that CYC exposure can disrupt the redox balance of tissues to suggest that biochemical and physiological disturbances may result from oxidative stress (Selvakumar *et al.*, 2004). It is, therefore, important to continue the search for an effective natural compound that will protect against CYC-induced reproductive toxicity associated with chemotherapy. These studies suggest that natural compounds with antioxidative

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^{**} Abbreviations: FH: Ferula hermonis; CYC: Cycram; ANOVA: Analysis of variance; T: testosterone; Hb: hemoglobin; FSH: follicle stimulating hormone; LH: luteinizing hormone.

properties may have the potential to ameliorate CYC-mediated testicular injury. In the past decade, the bioactivities of flavonoids on human health have given rise to much attention, especially the antioxidant activity (Zhang, 2005). The protective role of flavonoids involves several mechanisms of action: direct antioxidant effect, inhibition of enzymes of oxygen-reduction pathways and sequestration of transient metal cations (Rice-Evans, 2001).

To prevent these toxic side effects, there is a need for novel agent or natural product such as Ferula hermonis (FH) that may protect against CYC-induced reproductive toxicities in male rat. FH is a medicinal plant and a known natural antioxidant that belongs to the apiaceae family and grows abundantly in the Mediterranean region and Middle East (Naguib, 2003; Auzi et al., 2008), and used as a tonic, stimulant and aphrodisiac and as an anti-impotency medication for both sexes, as well as increasing the "stamina" of animals. Other traditional uses for FH are cauterizing wounds, curing animal infections and increasing the milk production of cows (Canogullari et al., 2009). Ferula hermonis (FH) is reported to have diverse therapeutic effects (Abutaha et al., 2019). Pharmacological reports have revealed that FH has several bioactivities which are useful to treat reproductive dysfunction, menopausal disturbances, neurological disorders (Zanoli et al., 2005) and diabetes (Raafat and El-Lakany, 2015). The plant is also reported to have antimicrobial, antiinflammatory (Saab et al., 2012; Al-Ja'fari et al., 2013), anticancer activities (Saab et al., 2012; Galal et al., 2001) and antioxidant effect due to its flavonoid composition (Abarikwu et al., 2012).

There are a little studies about effects of FH on the reproductive system and sexual appetite. The ethanolic extract of seeds and roots of FH called "masculine" was examined on male fertility and sexual functioning in rats and humans and was exhibits a high level of safety in rats, humans and cultured human fibroblasts and increases erection in rats. In addition; consumption of one tablet of masculine daily for 3 months could increase sperm number and sperm motility in men (Kassisa *et al.*, 2009). In addition, Rajeh and Al-Shehri (2019), found that FH ameliorates sperm count and T serum level due to its antioxidant activity against testicular toxicity in male rats. Another study reported that a single oral dose of FH extract can increase serum T concentration in adult male rat up to three-folds (Gunes and Fetil, 2000).

Therefore, the present study was designed to evaluate the protective effects of FH extract against CYC-induced reproductive toxicities which include sperm morphology, count and motility; RNA, DNA and protein content in rat testes as well measurement of testosterone and hemoglobin levels has been attempted.

2. Materials and Methods

2.1. Animals and treatment

Adult male albino rats (weighing 200–220 g), were obtained from animal house of National Research Centre, Dokki, Cairo, Egypt. They were housed in a standard conditions of temperature (22 \pm 2°C), relative humidity (55 \pm 10%), and 12/12 h light/dark cycle, and fed with a standard rat feed and water *ad libitum*. The experimental

designs and protocols for study conform the requirement of Ethics Committee of our Institute in accordance with the standard guide for the care and use of laboratory animals. Animals were divided into 3 main treated groups (30 animals each) according to treatment periods (15, 30 and 60 days every one divided to 3 subgroups according to CYC dose (50, 100 and 200 mg/kg bw/day, dissolved in distilled water following pretreatment with olive oil by gavage, 10 animals each), and each subgroup divided to 2 sub-sub groups treated with CYC and CYC+ FH (0.025 ml/100 g bw/day dissolved in olive oil by gavage), nd5 animals each, besides the control group (10 animals).

2.2. Drug and chemicals

CYC was purchased from Baxter Oncology GmbH, Frankfurt, Germany. All other chemicals used were of the highest purity and analytical grade.

2.3. The herbal remedy

Roots of FH were obtained from the Experimental Station of Medicinal Plants, Ministry of Agriculture, Cairo, Egypt. Sample extract was obtained by stirring 3 g of dry organ powder with 30 ml of methanol for 24 h at 120 rpm using a magnetic stirrer plate. Extract obtained was kept at 4 °C, filtered through a Whatman filter paper (No. 4) and freed of solvent under reduced pressure at 45 °C using a rotary evaporator. The dried crude concentrated extract was stored at -20 °C until further use for analyses (Rahali et al., 2018). Phenolic compounds of plant sample were extracted according to the method outlined by (Duke et al., 2003). The identification of the individual phenolic compounds was performed on high liquid chromatography (JASCO HPLC) (Table 1), using hypersil C18 reversed place column (250x4.6mm) with 5 µ particle size.

The herb was given to the treated animals orally (using oral tube), twice weekly as prescribed by herbalist (0.025 ml/ 100 g bw from plant water extract 50%) prepared in the form of water extract (by soaking in boiled water over night) for 8 successive weeks.

Table 1. HPLC of polyphenols in methanol extract of FH.

Ingredients	mg/100 g
Phenol	30
Resorcinol	670
Protocatchuic acid	340
Catchines	8380
Parahydroxy benzoic	3260
Caffeic acid	420
Daidzin	26
Ferulic acid	880
Coumarine	440
Paracoumaric acid unhydrate	290
Rutin	380
Myricetin	1100
Eugenol	1700
3,5 Dihydroxyisoflavon	1500
Quercedin	220
Caumpherol	3840
Pinostrobin	1810

HPLC= High pressure liquid chromatography. FH= Ferula hormones.

2.4. Sampling

At the end of the specified treatment (2, 4 and 8 weeks), the animals were euthanized by ethyl ether exposure and were killed by decapitation. Blood samples were collected in vials containing heparin. The plasma was separated for analysis of testosterone and hemoglobin using specific kits purchased from Rocky Mountain Diagnostic, Inc. 2139 Chuckwagon Road Suite 20. Colorado Springs, CO 80919, USA. Testis were obtained for RNA, DNA and protein analysis.

2.5. Sperm characteristics

Epididymal sperms were collected for sperm count and motility according to (WHO, 2010). Briefly, sperm count and motility within semen should be assessed as soon as possible after scarification, preferably at 30 minutes, but in any case within 1 hour, to limit the deleterious effects of dehydration, pH or changes in temperature on motility. Mix the semen sample well. Remove an aliquot of semen immediately after mixing, allowing no time for the spermatozoa to settle out of suspension. Remix the semen sample before removing a replicate aliquot. For each replicate, prepare a wet preparation approximately 20 m deep. Wait for the sample to stop drifting (within 60 seconds). Examine the slide with phase-contrast optics at ×200 or × 400 magnifications. Assess approximately 200 spermatozoa per replicate for the percentage of different motile categories.

For the analysis of morphological abnormalities such as head (hook less and banana shape), or abnormal tails (coiled and divided), the standard protocol for sperm morphology assay (Wyrobek *et al.*, 1983) was used. Briefly, after the treatment, the animals were sacrificed by cervical dislocation. Both of the cauda epididymises were dissected out, cut into pieces in 5 mL of saline, filtered, and smears were made. The smears were fixed in methanol and stained with 10 % Giemsa in Sörensen buffer for 10 min. A total of 1000 sperms per animal were scored under a microscope (Olympus CX21®, Japan) with 100x10 magnifications. Sperm head and tail abnormalities were determined as having either normal or abnormal morphology.

2.6. Determination of Nucleic Acids and protein

Nucleic acids (DNA and RNA) were determined using a simplified method for determination of specific DNA and RNA using quantitative PCR and an automatic DNA sequencer (Porcher *et al.*, 1992). Total protein is estimated using commercial kits (Peter, 1968).

2.7. Biochemical investigations

At the end of each experimental period and 24 h after the last dose of protective treatment, rats were sacrificed by cervical decapitation after an overnight fast. Blood was collected after 2, 4 and 8 weeks of treatment, and the separated serum was used for assaying the testosterone (T) level. Serum level of T was measured using an enzymelinked immunosorbent assay (ELISA) purchased from Rocky Mountain Diagnostic, Inc. 2139 Chuckwagon Road Suite 20. Colorado Springs, CO 80919, USA. Hemoglobin level was determined using specific kits purchased from Crystal Chem, 955 Busse Rd, Elk Grove Village, IL 60007 USA.

2.8. Statistical analysis

The values are expressed as mean \pm standard deviation (S.D). Differences between groups were assessed by one-way analysis of variance (ANOVA). Statistical analysis was performed using the SPSS for Windows 9.05 package program. Multiple comparisons were carried out by least significant difference (LSD) test. Significance at P-values <0.001, <0.01 and <0.05 have been considered in the tables

3. Results

3.1. Effect of CYC and FH extract on RNA, DNA and protein content

The results of the present study revealed that CYC treatment significantly (P <0.01) decreased RNA, DNA and protein content in male rat testes compared to control (Table 2). RNA and DNA levels decreased specially with the high dose (200 mg/kg bw of CYC) for 15 days of treatment up to the largest dose (200 mg/kg bw) for 60 days of treatment, while protein level decreased gradually in a time and dose dependent manner. However, FH administration in combination with CYC ameliorate these levels compared to CYC only treated groups.

Table 2. Effect of CYC and FH extract on RNA, DNA and protein content in rat testes.

Treatment period	Dose	RNA mg/g tissue		DNA mean ± S.D		g/dl protein	
		CYC	CYC+FH	CYC	CYC+ FH	CYC	CYC+ FH
15 days	50	0.336 ± 0.030	0.403 ± 0.029	0.478 ± 0.019	0.491 ± 0.020	11.289**± 0.692	12.393 ± 0.690
	100	0.289 ± 0.015	0.387 ± 0.015	0.465 ± 0.013	$0.482^{N.S} \pm 0.018$	$9.341**\pm0.672$	$11.323*\pm0.875$
	200	0.268 ± 0.016	0.369 ± 0.027	0.430**± 0.039	0.474 ± 0.016	$8.014***\pm0.216$	$10.221**\pm0.730$
30 days	50	0.320 ± 0.030	0.392 ± 0.015	0.464*± 0.019	0.488 ± 0.030	8.745***± 0.565	10.940**± 0.644
	100	0.278 ± 0.018	0.376 ± 0.022	$0.446**\pm0.030$	0.470 ± 0.025	$8.035***\pm0.190$	9.996**± 1.005
	200	0.252 ± 0.021	0.330 ± 0.027	0.398 ± 0.021	0.398 ± 0.021	$7.47***\pm0.420$	$8.168***\pm0.488$
60 days	50	0.298 ± 0.007	0.377 ± 0.024	0.441**± 0.30	0.463 ± 0.019	8.243***± 0.547	$9.168** \pm 0.488$
	100	0.247 ± 0.014	0.340 ± 0.030	0.429 ± 0.034	0.456 ± 0.038	$7.199***\pm 0.544$	$8.273***\pm0.538$
	200	0.230 ± 0.012	0.292 ± 0.050	0.379 ± 0.018	0.436**± 0.037	6.771***± 0.3199	$7.917***\pm0.323$
Control		0.415± 0.617		0.511± 0.018		13.323 ± 0.757	

Data are presented as means \pm SD. * Significant at P \leq 0.05; ** Significant at P \leq 0.01;

^{***} Significant at P \leq 0.001. CYC= Cycram; FH= Ferula hormones.

3.2. Serum testosterone and hemoglobin levels

Table (3) showed that CYC administration caused significant reductions (P < 0.001), in serum testosterone (T) level (9.308 \pm 0.399) and hemoglobin (9.393 \pm 0.505) (P < 0.001), when compared with the control group, especially with the high dose of CYC (200 mg/kg bw) and long period treatment (60 days) in a dose and time dependent manner (9.308 \pm 0.399 ng/l and 9.393 \pm 0.505 g/dl

vs. 15.052±0.841and15.813±0.622 for T and Hb levels in treated and control groups, respectively). However, supplementation of FH improved these parameters compared to CYC only treated groups. FH was able to produce marked effects in CYC-treated rats, as demonstrated by the restoration of the levels of serum T and hemoglobin, to almost normal levels when compared with the control group.

Table 3. Effect of CYC and FH extract on serum testosterone and hemoglobin levels in male rats.

Treatments period	Dose	Testosterone hormone (r	ng/l)	Hemoglobin g/dl	Hemoglobin g/dl	
		CYC	CYC + FH	CYC	CYC + FH	
	50	11.776±0.671***	14.372±0.701	12.566± 0.783***	14.872 ± 0.584	
15 days	100	11.462±0.510***	13.668±0.559*	$11.593 \pm 0.525 ***$	13.958± 0.867*	
	200	11.03±0.424***	13.24±0.567*	$10.114***\pm0.280$	13.017± 0.719**	
	50	11.289±0.760***	13.862±0.298*	12.304± 0.651***	14.375± 0.542*	
30 days	100	10.346±0.521***	13.526±0.417*	11.359± 0.525***	13.913± 0.655**	
	200	10.037±0.292***	12.46±0.515**	9.758± 0.516***	12.755± 0.623**	
	50	10.39±0.502***	13.19±0.372**	11.958± 0.893***	13.997± 0.740**	
60 days	100	9.734±0.529***	12.728±0.505**	$10.662 \pm 0.526 ***$	13.396± 0.543**	
	200	9.308±0.399***	12.270±0.830**	9.393± 0.505***	12.708± 0.483**	
Control		15.052±0.841		15.813±0.622		

Data are presented as means \pm SD. * Significant at P \leq 0.05; ** Significant at P \leq 0.01;

3.3. Sperm abnormality in CYC and FH extract treated rats

Sperm abnormality rates in response to various treatments for 8 weeks of treatment are presented in table (4). CYC administration caused statistically significant (P < 0.01) increases in tail, head and total abnormality of sperm in comparison with the control group. A significant (P < 0.01) decrease in these abnormalities was observed in CYC+ FH groups as compared with alone CYC group. The values of head abnormality were brought near values

to control by FH administrations to CYC-treated rats, especially with the low dose of CYC; these administrations could significantly improve this parameter when compared with the alone CYC group.

A significant decrease (P < 0.01) in total count and motility was found in CYC treated groups. However, the treatment with FH in combination to CYC ameliorates these parameters compared to alone CYC treated groups and brought these values near to control especially with the low dose of CYC.

Table 4. Sperm abnormality in CYC and FH extract treated rats.

Treatment	Dose	Tail abnormality		Head abnormality		Total	Total	Motility
		Divided	Coiled	Without hock	Banana shape	Abnormality	Count	%
	50	3.0± 0.581**	2.2± 0.924**	2.0± 0.581**	1.8± 0.924**	9.0± 0.581**	26.522x10 ⁶ ± 1.476	41.35% ±2.53
CYC	100	3.4± 0.302**	3.0± 0.225**	3.6± 0.789**	2.4± 0.342**	12.4± 0.302**	$24.478x10^6 \pm 3.288$	39.44% ±3.06
	200	$5.0 \pm 0.581 **$	$4.6 \pm 0.140 **$	$4.0 \pm 0.581 **$	3.2± 0.304**	16.4± 0.140**	$22.980 \text{x} 10^6 \pm 1.110$	33.59% ±2.75
CYC+ FH	50	1.4 ± 0.401	1.2± 0.837	1.0± 0.710	0.6± 0.894	4.2± 0.837*	$31.124 \times 10^6 \pm 1.755$	68.75% ±4.57
	100	$2.2 \pm 0.837 *$	$1.6 \pm 0.581 *$	$1.4{\pm}~0.140$	1.0± 0.707*	6.2± 0.280**	$28.100x10^6 \pm 0.945$	$59.35\% \pm 5.78$
	200	3.0± 0.581**	$1.2{\pm}~0.836$	$1.6 \pm 0.40 *$	2.0± 0.837**	$7.8 \pm 0.28 **$	$27.85 \text{x} 10^6 \pm 1.069$	54.69% ±4.41
Control		0.4 ± 0.548	0.6 ± 0.548	0.8 ± 0.095	0.4 ± 0.394	2.2 ± 0.924	$32.5 \times 10^6 \pm 1.714$	82.42% ±4.36

Data are presented as means \pm SD. * Significant at P \leq 0.05; ** Significant at P \leq 0.01;

4. Discussion

The present work was performed to evaluate the protective role of FH against CYC induced changes in testicular RNA, DNA and protein content, sperm morphology, count and motility, as well as blood levels of T and Hb in adult male albino rats. The main findings of this study indicated that CYC induced deleterious effect on

these parameters. That is in accordance with those of (Anderson *et al.*, 1995; O'Flaherty *et al.*, 2010; Bujan *et al.*, 2013) who found that administration of CYC leads to decreased RNA, DNA and protein synthesis in male rats/mice.

Oxidative DNA damage is caused by hydroperoxide derivative of CYC through generation of H₂O₂ (Murata *et al.*, 2004). Also, acrolein has been found to interfere with

^{***} Significant at P ≤ 0.001. CYC= Cycram; FH= Ferula hormones.

^{***} Significant at P \leq 0.001. CYC= Cycram; FH= Ferula hormones.

the tissue antioxidant defense system (Arumugam *et al.*, 1997) and produces highly reactive oxygen free-radicals (Mythili *et al.*, 2004) that are mutagenic to mammalian cells (Kawanishi *et al.*, 1998). From these studies, it is clear that the administration of a potent and safe antioxidant such as FH extract might reduce CYC-induced reproductive toxicity.

The reduction in T and Hb levels in rats exposed to CYC, especially with the high dose coincides with those of (Motawi *et al.*, 2010; Mohammadi *et al.*, 2014). The decline in serum T level after CYC treatment in consistent with previous reports, and this decline could be due to CYC-induced membrane lipid peroxidation (Das *et al.*, 2002). The decrease in serum T level in CYC-treated rats demonstrates defect in the testis. The decreased level of Hb observed in the CYC-treated group in our study may be due to the interaction of CYC with sulfhydryl-containing proteins and/or testicular Ca⁺² overload (Selvakumar *et al.*, 2006).

The level of abnormalities in sperm morphology, total count and motility in CYC treated rats was high; however, FH administration improved these parameters. That is in line with the results of (Abarikwu *et al.*, 2012; Watcho et al., 2019) who found that CYC-treated rats had higher abnormal sperm morphology than the control group. However, the CYC plus Rutin (one of the FH component) treated rats had lower abnormal sperm morphology rates than the CYC-treated rats only (*P*< 0.05).

Our results support those of Ilbey et al. (2009), who reported that CYC treatment induced irregular and diminished seminiferous tubules containing a few germ cells counted. As well with Mohammadi et al. (2014), who found that spermatogonia, spermatocyte, spermatid and sperm count in seminiferous tubules were decreased significantly in CYC treated rats. These results suggest that CYC inhibits spermatogenesis. The sperm abnormalities observed in CYC-treated rats are a serious problem of reproductive function, since abnormal sperm cannot reach the oviduct after intravaginal ejaculation (Izawa et al., 2008).

The positive effects of antioxidants on human health have attracted more attention in recent times. Most of the FH component biological actions seem to be associated with its potency as an antioxidant. Administration of antioxidant FH along with CYC has been shown to prevent the testicular injury and reproductive dysfunction as revealed by restoring the normal level of sperm counts, morphology and motility, DNA, RNA and protein as well T and Hb levels. The protective role of FH involves several mechanisms of action: direct antioxidant effect, inhibition of enzymes of oxygen-reduction pathways and sequestration of transient metal cations (Rice-Evans, 2001).

5. Conclusion

In conclusion, this study suggests that CYC treatment induced a significant alteration in RNA, DNA and protein synthesis and affected T and Hb levels and sperm (morphology and motility) abnormalities of male rats. However, supplementation of FH protects morphological structure of sperms and RNA, DNA, protein content as well blood Hb and T levels against CYC toxicities. These protective actions of FH seem to be closely involved with

the suppressing of plasma lipid peroxidation and increasing of antioxidant enzyme activities. Therefore, FH might be considered as an alternative drug in combination with CYC in cancer patients, transplantation and autoimmune diseases to prevent CYC-induced reproductive toxicity in patients receiving CYC chemotherapy.

Competing interests

The authors declare that they have no competing interests.

References

Abarikwu SO, Otuechere CA, Ekor M, Monwuba K and Osobu D. 2012. Rutin Ameliorates Cyclophosphamide-induced Reproductive Toxicity in Male Rats. *Toxicol Int.*, **9(2)**: 207-214.

Abutaha N, Nasr FA, Al-zharani M, Alqahtani AS, Noman OM, Mubarak M, Abdelhabib S and Wadaan MA. 2019. Effects of Hexane Root Extract of Ferula hermonis Boiss. on Human Breast and Colon Cancer Cells: An In Vitro and In Vivo Study. *BioMed Res Int*, Volume **2019**, Article ID 3079895, 12 pages.

Al-Ja'fari A, Vila R, Freixa B, Costa J and Canigueral S. 2013. Antifungal compounds from the rhizome and roots of Ferula hermonis. *Phytother Res.*, **27**(6): 911-915.

Anderson D, Bishop JB, Garner RC, Ostrosky-Wegman P and Selby PB. 1995. Cyclophosphamide: review of its mutagenicity for an assessment of potential germ cell risks. *Mutat Res.*, **330**: 115-181.

Arumugam N, Sivakumar V, Thanislass J and Devaraj H. 1997. Effects of acrolein on rat liver antioxidant defense system. *Indian J Exp Biol.*, **35**: 1373–1374.

Auzi AA, Gray AI, Salem MM, Badwan AA and Sarker SD. 2008. Feruhermonins A-C: three daucane esters from the seeds of Ferula hermonis (Apiaceae). *J Asian Nat Prod Res.*, **10**: 711-717.

Bujan L, Walschaerts M, Moinard N, Hennebicq S, Saias J and Brugnon F. 2013. Impact of chemotherapy and radiotherapy for testicular germ cell tumors on spermatogenesis and sperm DNA: a multicenter prospective study from the CECOS netword. *Fertil Steril.*, **100(3)**: 673-680.

Canogullari S, Baylan M, Copur G and Sahin A. 2009. Effects of dietary Ferula elaeochytris root powder on the growth and reproductive performance of Japanese quail (*Coturnixcoturnix japonica*): it is not recommended in a breeder diet. *Arch Geflugelk.*, **73** (1): S. 56-60.

Cengiz M. 2018. Hematoprotective effect of boron on cyclophosphamide toxicity in rats. *Cell Mol Biol (Noisy le Grand)*, **64 (5)**: 62-65.

Das UB, Mallick M, Debnath JM and Ghosh D. 2002. Protective effect of ascorbic acid on cyclophosphamide induced testicular gametogenic and androgenic disorders in male rats. *Asian J Androl.*, **4**: 201-207.

Dere E, Anderson LM, Hwang K and Boekelheide K. 2013. Biomarkers of Chemotherapy-Induced Testicular Damage. *Fertil Steril.*, **100(5)**: 1192-1202.

Duke SO, Imando AM, Pace PF, Reddy KM and Semeda KJ. 2003. Isoflavone, glycophosate and amino methyl phosphoric acid levels in seeds of glyphosate treated, glyphosate-resistant soybean. *J Agric Food Chem.*, **51**: 344-350.

Fraiser LH, Kanekel S and Kehrer JP. 1991. Cyclophosphamide toxicity. Characterizing and avoiding the problem. *Drugs*, **42**: 781–795.

Galal AM, Abourashed EA, Ross SA, ElSohly MA, Al-Said MS and El-Feraly FS. 2001. Daucane sesquiterpenes from Ferula hermonis. *J Nat Prod.*, **64(3)**: 399-400.

Ghobadi E, Moloudizargari M, Asghari MH and Abdollahi M. 2017. The mechanisms of cyclophosphamide-induced testicular toxicity and the protective agents. *Expert Opin Drug Metab Toxicol.*, **13(5):** 525-536.

Gunes AT and Fetil E. 2000. Hormones: androgens, antiandrogns, anabolic steroids, estrogens - unapproved uses or indications. *Clin Dermatol.*, **18**(1): 55-61.

Howell S and Shalet S. 1998. Gonadal damage from chemotherapy and radiotherapy. *Endocrinol Metab Clin North Am.*, **27(4)**: 927–943.

Ilbey YO, Ozbek E, Simsek A, Otunctemur A, Cekmen M and Somay A. 2009. Potential chemoprotective effect of melatonin in cyclophosphamide-and cisplatin-induced testicular damage in rats. *Fertil Steril.*, **92**: 1124-1132.

Izawa H, Kohara M, Aizawa K, Suganuma H, Inakuma T and Watanabe G. 2008. Alleviative effects of quercetin and onion on male reproductive toxicity induced by diesel exhaust particles. *Biosci Biotechnol Biochem.*, **72**: 1235-1241.

Johari H, Mahmoudinejad F and Amjad G. 2011. An evaluation of the effect of the hydroalcoholic extract of ginger on the hypothalamus-pituitary-gonadal axis in adult female rats (Rat) treated with cyclophosphamide. *Pejoda.*, **6**: 62-70.

Kassisa E, Fuldera S, Khalilb K, Hadiehc B, Nahhasb F and Saad B. 2009. Efficacy and safety assessments of *ferula assa- foetida* L., traditionally used in greco- arab herbal medicine for enhancing male fertility, libido and erectile function. *Open Complement Med J.*, 1: 102–109.

Kaur F, Sangha GK and Bilaspuri GS. 1997. Cyclophosphamide-induced structural and biochemical changes in testis and epididymidis of rats. *Indian J Exp Biol.*, **35**: 771-775.

Kawanishi M, Matsuda T, Nakayama A, Takebe H, Matsui S and Yagi T. 1998. Molecular analysis of mutations induced by acrolein in human fibroblast cells using supF shuttle vector plasmids. *Mutat Res.*, **417**: 65-73.

Meistrich ML, Parchuri N, Wilson G, Kurdoglu B and Kangasniemi M. 1995. Hormonal protection from cyclophosphmide-induced inactivation of rat stem spermatogonia. *J Androl.*, **16**: 334-341.

Mohammadi F, Nikzad H, Taghizadeh M, Taherian A, Azami-Tameh A, Hosseini SM and Moravveji A. 2014. Protective effect of *Zingiber officinale* extract on rat testis after cyclophosphamide treatment. *Andrologia.*, **46**: 680-686.

Motawi TMK, Sadik NAH and Refaat A. 2010. Cytoprotective effects of DL-alpha-lipoic acid or squalene on cyclophosphamide-induced oxidative injury: An experimental study on rat myocardium, testicles and urinary bladder. *Food Chem Toxicol.*, **48**: 2326-2336.

Murata M, Suzuki T, Midorikawa K, Oikawa S and Kawanishi S. 2004. Oxidative DNA damage induced by a hydroperoxide derivative of cyclophosphamide. *Free Radic Biol Med.*, **37**: 793–802.

Mythili Y, Sudharsan PT, Selvakumar E and Varalakshmi P. 2004. Protective effect of DL-alpha-lipoic acid on cyclophosphamide induced oxidative cardiac injury. *Chem Biol Interact.*, **30**: 13-19.

Naguib YM. 2003. Pharmaceutically active composition extracted from Ferula hermonis and process of its extraction. United States Patent 6623768.

O'Flaherty C, Hales BF, Chan P and Robaire B. 2010. Impact of chemotherapeutics and advanced testicular cancer or Hodgkin lymphoma on sperm deoxyribonucleic acid integrity. *Fertil Steril.*, **94**: 1374–1379.

Peter T. 1968. Protein colorimetric method. *Clinical Chem.*, **14**: 1147-1159

Porcher C, Malinge MC, Picat C and Grandchamp B. 1992. A simplified method for determination of specific DNA or RNA copy number using quantitative PCR and an automatic DNA sequencer. *Biotechniques*, **13**(1): 106-114.

Raafat K and El-Lakany A. 2015. Acute and subchronic in-vivo efects of Ferula hermonis L. and Sambucus nigra L. and their potential active isolates in a diabetic mouse model of neuropathic pain. *BMC Complement Altern Med.*, **15:** 257.

Rahali FZ, Kefi S, Rebey IB, Hamdaoui G, Tabart J, Kevers C, Franck T, Mouithys-Mickalad A and Sellami IH. 2018. Phytochemical composition and antioxidant activities of different aerial parts extracts of Ferula communis L. *Plant Biosyst*, DOI: 10.1080/11263504.2018.146169.

Rajeh NA and Al-Shehri AM. 2019. Antioxidant effect of Ferula hormones Boiss on acrylamide induced testicular toxicity in male rats. *Indian J Exp Biol.*, **57**: 139-144.

Rajeh NA, Al-Shehri AM (2019) Antioxidant effect of *Ferula hermonis* Bioss on acrylamide induced testicular toxicity in male rats. *Indian J Exp Biol.*, **57**: 138-144.

Rice-Evans C. 2001. Flavonoid antioxidants: Review. *Curr Med Chem.*, **8(7)**: 797-807.

Saab AM, Lampronti I and Borgatti M, et al. 2012. Invitro evaluation of the anti-proliferative activities of the wood essential oils of three Cedrus species against K562 human chronic myelogenous leukaemia cells. *Nat Prod Res.*, **26(23)**: 2227-2231.

Selvakumar E, Prahalathan C, Mythili Y and Varalakshmi P. 2004. Protective effect of DL-alpha-lipoic acid in cyclophosphamide induced oxidative injury in rat testis. *Reprod Toxicol.*, **19**(2): 163-167.

Selvakumar E, Prahalathan C, Sudharsan PT and Varalakshmi P. 2006. Protective effect of lipoic acid on cyclophosphamide-induced testicular toxicity. *Clin Chim Acta.*, **367** (1–2): 114-119.

Shabanain S, Farahbod F, Rafieian M, Ganji F and Adib A. 2017. The effects of Vitamin C on sperm quality parameters in laboratory rats following long-term exposure to cyclophosphamide. *J Adv Pharm Technol Res.*, **8**: 73-79.

Watcho P, Mpeck I, Deeh Defo P, Wankeu-Nya M, Ngadjui E, Bonsou Fozin G, Kamtchouing P and Kamanyi A. 2019. Cyclophosphamide-induced reproductive toxicity: Beneficial effects of Helichrysum odoratissimum (Asteraceae) in male Wistar rats. *J Integ Med.*, **17**(5): 366-373.

World Health Organization. 2010. **WHO laboratory manual for the examination and processing of human semen**. 5th ed., WHO Press, Switzerland.

Wyrobek AJ, Gordon LA, Burkhart JG, Francis MW, Kapp Jr, RW and Letz G. 1983. An evaluation of the mouse sperm morphology test and other sperm tests in nonhuman mammals. A report of the U.S. Environmental Protection Agency Gene-Tox Program. *Mutat Res.*, **115**: 1-72.

Zanoli P, Rivasi M, Zavatti M, Brusiani F, Vezzalini F and Baraldi M. 2005. Activity of single components of Ferula hermonis on male rat sexual behavior. *Int J Impot Res.*, **17**(6): 513-518

Zhang YM. 2005. Protective effect of quercetin on aroclor 1254-induced oxidative damage in cultured chicken spermatogonial cells. *Toxicol Sci.*, **88**(2): 545-550.