

Parentage Analysis of the Progenies of the Reciprocal Crosses of *Pangasianodon hypophthalmus* (Sauvage, 1878) and *Clarias gariepinus* (Burchell, 1822) using Cytochrome b Gene

Okomoda V. Tosin¹, Koh I. C. Chong², Hassan Anuar², Amornsakun Thumronk³ and Shahreza Md Sheriff^{2,4*}

¹Department of Fisheries and Aquaculture, University of Agriculture, Makurdi, Nigeria

²School of Fisheries and Aquaculture Sciences, Universiti Malaysia Terengganu, Malaysia

³Department of Technology and Industries, Prince of Songkla University, Pattani campus Thailand, ⁴Institute of Tropical Aquaculture, Universiti Malaysia Terengganu, Malaysia

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Abstract

In this study, parentage analysis of the progenies from reciprocal crosses of *Pangasianodon hypophthalmus* (PH) and *Clarias gariepinus* (CG) was accomplished by sequencing the cytochrome b gene and running phylogenetic analysis. The result obtained showed that the nucleotide composition of the cytochrome b gene of the reciprocal hybrid crosses exhibited high (99 %) similarity with those of the maternal parents notwithstanding the different morphology within the hybrid pool (i.e. Panga-like and Clarias-like). The analysis of 1076bp of the cytochrome b gene using three methods for inferring molecular relationships (Maximum Likelihood, Neighbour Joining, and Maximum Parsimony) revealed two main clusters. The upper cluster was a mix of *C. gariepinus* and the two distinct progenies of ♀CG × ♂PH (i.e. Panga-like and Clarias-like), while the lower cluster was a mix of the *P. hypophthalmus* and the progenies of ♀PH × ♂CG (i.e. Panga-like only). Hence, despite, phenotypic differences within the hybrid pool, progenies still inherit cytochrome b gene from the maternal parent alone. As a result, the direction of crosses of the reciprocal crosses could be accurately determined.

Keywords: Asian catfish, African catfish, Cytochrome b, Hybrid morphotype.

1. Introduction

The main aim of hybridization between different fish groups or species is to produce offsprings that perform better than the parental species (Bartley *et al.*, 2001; Okomoda *et al.*, 2017). However, unintended consequences have been experienced as a result of accidental backcrossing, hence, threatening the diversity of many freshwater fish species (Epifanio and Nielsen, 2000; Perry *et al.*, 2002; Senanan *et al.*, 2004; Na-Nakorn *et al.*, 2004). Aside from the fear of genetic homogenization of farmed and natural fish stocks (Hashimoto *et al.* 2010); hybrids may compete successfully with the native parental lineages in several ways (Ryman and Utter, 1987; Allendorf *et al.*, 2001; Rosenfield *et al.*, 2004), hence, the need for characterization.

The efficacy of molecular markers in the determination of the hybridization status, direction of crosses, and genetic introgression cannot be over emphasized (Forbes and Allendorf, 1991; Cianchi *et al.*, 2003; Hänfling *et al.*, 2005). This is because most hybrids inherit part of the genetic makeup of both species (Wilkins, *et al.*, 1994). As a result, the hybridization status of different crosses has

been well investigated using nuclear markers since inheritance of this kind of DNA is from both parents (Rieseberg *et al.*, 1990; Sang *et al.*, 1995; Buckler *et al.*, 1997; Odorico and Miller, 1997). Similarly, morphological and cytogenetic analysis has been widely exploited for the same purpose. It is important to note that these methods command the same level of accuracy and at a very lower cost compared to the molecular characterization method (Hashimoto *et al.* 2010). However, they cannot be used to determine the direction of the hybridization of the crosses.

Although the potential of erythrocyte characterization for directional cross discrimination have been demonstrated in the earlier study by Okomoda *et al.*, (2018a), the outlined limitations of this approach largely reduce their usability in a wide array of species or studies. The most accurate method still remains the characterization of the mitochondrial DNA (mtDNA). This is because the mtDNA is primarily matrilineally inherited. As a result, their analysis can conveniently identify the maternal origin of the hybrids, and hence the direction of crosses (Pitts, 1995). The mitochondrial cytochrome b gene has particularly gained recognition as an important index in the comparisons of closely related taxa such as between populations or species (Degani 2004; Pfrender, *et*

* Corresponding author. e-mail: shahreza@umt.edu.my.

al., 2004; Perdices, *et al.*, 2005). However, it is conserved enough for clarifying deeper phylogenetic relationships (Allegrucci, *et al.*, 1999; Cunha, *et al.*, 2002; Sulaiman, *et al.*, 2006; Feulner, *et al.*, 2007).

While mtDNA has been exploited for the characterization of hybrids, the phylogenetic relationships between progenies were seldom analysed. This is particularly important in distant crosses which lead to the production of ploidy polymorphism of the hybrid progenies (i.e. different cytogenetic characteristics within the hybrid pool). A good example of such cross is the study on the hybridization between *Pangasianodon hypophthalmus* (Sauvage, 1878) and *Clarias gariepinus* (Burchell, 1822) where three distinct morphotypes were observed (Okomoda *et al.*, 2017, 2018a, 2018b) with different cytogenetic characteristics (Okomoda *et al.*, 2018c). Understanding the various levels of genetic relationships between hybrids and pure crosses could significantly aid in the development of management guidelines for commercial use and for conservation purposes (Birstein *et al.*, 2005; Freyhof *et al.*, 2005; Ludwig, 2008). Haven established the hybridization status of the reciprocal crosses between *P. hypophthalmus* and *C. gariepinus* in the study by Okomoda *et al.*, (2018b, 2018c), this study was designed for parentage determination and to establish phylogenetic relationship between pure and reciprocal progenies of the fishes using the cytochrome b gene.

2. Materials and Methods

Progenies of pure and reciprocal crosses between of *C. gariepinus*, *P. hypophthalmus* used in this study were obtained using the method previously described by Okomoda *et al.*, (2017, 2018 b, c). In brief, mature *P. hypophthalmus* and *C. gariepinus* were spawned by induced breeding using the Ovaprim[®] hormone (0.5mL/kg). Eggs and sperm were mixed to produce pure *C. gariepinus* (♀CH × ♂CH), pure *P. hypophthalmus* (♀PH × ♂PH), and the reciprocal crosses Clariothalmus (♀CG × ♂PH) and Pangapinus (♀PH × ♂CG). The progenies obtained from six different breeding trials were then cultured for one month in fibreglass tanks. At this point, two morphotypes were conspicuous in the Clariothalmus (Clarias-like and Panga-like), while all Pangapinus were of one morphotype (Panga-like) (See figure 1). All groups were continuously fed with commercial diet (35 % CP), and water quality was maintained at optimum (temperature = 33.0 ± 1.6^oC; pH = 7.0 ± 0.41; conductivity = 251 ± 0.31mg l⁻¹; total dissolved solids = 78.0 ± 0.89 mg l⁻¹; dissolved oxygen = 4.7 ± 0.39 mg l⁻¹).

Fifty fishes comprised of *P. hypophthalmus* (10), Clarias-like Clariothalmus (10), Panga-like Clariothalmus (10), Pangapinus (10), and *C. gariepinus* (10) were randomly selected and preserved in 95 % ethanol contained in appropriately labelled 1.5mL tubes. DNA was extracted from the fins of these fishes using specified protocol for Vivantis Nuclear Acid extraction kit (according to manufacturer's instruction). Amplification of the Cytochrome b, was carried out by polymerase chain reaction (PCR) using the universal primers Cyt b 3F (5'-CCACCGTTGTTATTCAACTATAGAAA-3') and Cyt b

3R (5'-AGAATRCTAGCTTTGGGAG-3') which was described by Bowen *et al.*, (2008).

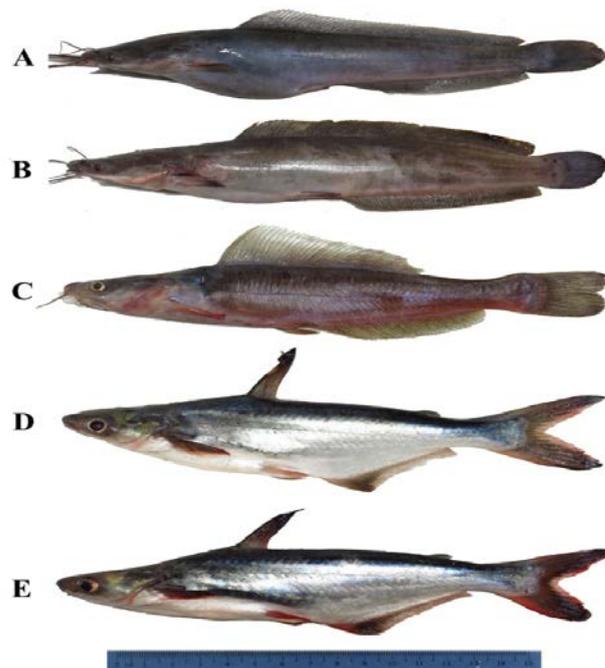


Figure 1. Morphology of (A) *Clarias gariepinus*; (B) Clarias-like Clariothalmus; (C) Panga-like Clariothalmus; (D) Pangapinus and (E) *Pangasianodon hypophthalmus*. (Source Okomoda *et al.*, 2018c)

A reaction volume of 25 µL was used containing 2.5 µL Easy Taq buffer, 2.0 µL dNTPs, 0.5 µL each of universal forward and reverse primer, 0.2µL Taq DNA polymerase (Easy Taq), 0.5µL template DNA, and 18. 8 µL sterile deionised water. Reactions were performed using an Eppendorf Mastercycler. The Cyt b PCR amplification was programmed at 95^oC for two minutes, followed by thirty-five cycles of denaturation at 95^oC (thirty seconds), annealing temperature at 47^oC (forty-five seconds) and an extension of 72^oC (one minute), this was followed by a final single extension at 72^oC (seven minutes). An aliquot of the reaction was subjected to electrophoresis on a 1.0 % agarose gel. The gels were visualized using a gel documentation system (BIO RAD, USA).

PCR products were sent for sequencing at First Base Laboratories SDN BHD, Malaysia. All sequences were aligned and edited using ClustalW as implemented in MEGA version 6. Sequence ambiguities were resolved by comparing complementary strands of ten nucleotide sequences per group of fish. The identity of the sequences was confirmed using BLAST at the NCBI website (https://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastn&GE_TYPE=BlastSearch&LINK_LOC=blasthome). The percentages of similarities of all the progenies with the available sequences on the NCBI website were gotten.

The relationship between the different groups of fish was examined using neighbour joining (Saitou and Nei 1987), maximum parsimony and maximum likelihood (Felsenstein 1983) methods (using MEGA version 6). The confidence of the branching patterns was assessed by 1000 bootstrap replicates in the NJ analysis (Hillis and Bull 1993). The out group used was *Cyprinus carpio* (NCBI

Accession number: AB158806), while *C. macrocephalus* (NCBI Accession number: KJ533248) and *Pangasius nasutus* (NCBI Accession number: HM236395) were used as the sister groups in this study.

3. Results

Amplification of the universal primers in relation to the mitochondrial cytochrome b gene showed bands of approximately 1300bp for all the groups of fish and their different morphotypes (Appendix 1). The partial sequences obtained for the cytochrome b revealed very high similarity between the nucleotide composition of the reciprocal crosses and the maternal parents irrespective of their morphotypes. Blast also showed that the target sequence was gotten as the nucleotide compositions were 99 % identical with available cytochrome b sequences in the GenBank for *Clarias gariepinus* (accession number KJ533253.1) and *Pangasianodon hypophthalmus* (accession number KM434895.1) (respectively for pure and reciprocal progenies with Clarias and Panga maternal origins) (Appendix 2, 3, 4, 5 and 6). The results obtained by sequencing allowed the analysis of 1076bp of the Cyt b gene. Similar genetic pattern were obtained from the three methods used for inferring molecular relationships in this study (Maximum likelihood, Neighbour Joining, and Maximum Parsimony Respectively for Figure 2, Appendix 7 and 8). From the result obtained, two main groups were clearly defined. The upper group was a mix of *C. gariepinus*, Clarias-like Clariothalmus and Panga-like Clariothalmus with three haplotypes. The lower group, on the other hand, was a mix of the Pangapinus and the *P. hypophthalmus* with six observed haplotypes. The variations in the difference haplotypes occurred in three bases or less.

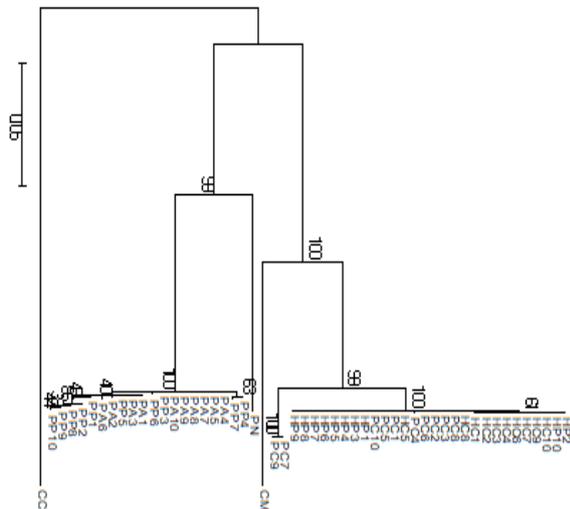


Figure 2. Phylogenetic relationships of the different progenies following Maximum likelihood analysis of 1076bp of the cytochrome b gene. PC1 - PC10 = *C. gariepinus*; HC1 - HC10 = Clarias-like Clariothalmus; HP1 - HP10 = Panga-like Clariothalmus; PA1 - PA10 = Pangapinus; PP1 - PP10 = *P. hypophthalmus*; CC = *Cyprinus carpio*; CM = *Clarias macrocephalus*; PN = *Pangasius nasutus*. (Support values are bootstrap values).

The genetic distances within haplotypes of the same groups in this study (i.e. pure and hybrid crosses from the same maternal origin) ranged between 0.000 - 0.056 (Table 1). However, the genetic distance between the pure *P. hypophthalmus* and *C. gariepinus* ranges from 0.472 - 0.506.

Table 1: Genetic distance range between pure and reciprocal crosses of *Pangasianodon hypophthalmus* and pure *Clarias gariepinus*.

	PC	HC	HP	PA	PP
PC	0.000 – 0.056				
HC	0.000 – 0.056	0.000 – 0.056			
HP	0.000 – 0.056	0.000 – 0.054	0.000 – 0.056		
PA	0.472 – 0.502	0.475 – 0.504	0.472 – 0.506	0.010 – 0.051	
PP	0.472 – 0.506	0.470 – 0.491	0.472 – 0.494	0.000 – 0.052	0.000 – 0.050

Keys: PC = *C. gariepinus*; HC1 = Clarias-like Clariothalmus; HP1 = Panga-like Clariothalmus; PA1 = Pangapinus; PP1 = *P. hypophthalmus*.

4. Discussion

Different researchers had justified the need to discriminate reciprocal crosses of different species because hybrid progenies could display different biological and zootechnical characteristics (Tave, 1993; Toledo-Filho *et al.* 1998; Porto-Foresti *et al.* 2008). More so, information on the directionality of mating can have significant effects on a number of factors including spatial, temporal, behavioural, physiological, and stock composition of the hybrid progenies (Pitts, 1995; Scribner *et al.*, 2001). This assumption is validated by the results of early studies which show differences in performance not only between the reciprocal crosses of *P. hypophthalmus* and *C. gariepinus*, but also between progenies of the hybrid Clariothalmus (i.e Clarias-like and Panga-like) (Okomoda *et al.*, 2018b, 2018c). Similarly, Dunham *et al.* (1982) had justified molecular discrimination of F1 progenies of the hybrids ♀*Ictalurus punctatus* × ♂*I. furcatus* and ♀*I. furcatus* × ♂*I. punctatus* on the bases of differential growth performances.

The cytochrome b gene is a highly conserved region (Moritz *et al.*, 1987; Olufeagba and Okomoda 2016); hence, the universal primer used in this study uniformly amplified 1300bp in all the progenies tested as expected. The observation of the very high similarity between the progenies of the reciprocal crosses and their maternal parent confirms the direction of crosses. Similarly, the phylogenetic analysis shows two separate groups of the test progenies in direction of the maternal origin. Observations of previous studies using different mtDNA genes are in line with the findings of this study. do-Prado *et al.*, (2011) had used 16S mitochondrial genes to discriminate reciprocal hybrids 'pintachara' and 'cachapinta' by the identification of the maternal parent of these crosses. The efficacy of differentiating reciprocal hybrids of *Leporinus macrocephalus* and *L. elongatus* using 16S mitochondrial DNA had also been reported by

Hashimoto *et al.*, (2010). In addition, Olufeagba and Okomoda (2016) confirmed the maternal origin of the hybrid ♀ *C. gariepinus* × ♂ *C. batrachus* with the cytochrome b gene. Waldbrieser and Bosworth (2008) were able to discriminate reciprocal crosses of channel catfish (*I. punctatus*) and blue catfish (*I. furcatus*) hybrid using the mitochondrial cytochrome c oxidase 1 gene.

On the whole, mitochondrial DNA have proven to be a very effective molecular tool in the identification of the parental status of hybrids since it is cytoplasmically housed and only inherited from the mother to the offspring, and has no paternal contribution (Moritz *et al.*, 1987; Wyatt *et al.*, 2006). Hence, the pattern of genetic homogenization between progenies from the same maternal parent and distinct groups observed for the different maternal origins clearly shows the direction of crosses of the progenies. In line with this, Nazia *et al.*, (2010) had earlier stated that high levels of genetic homogenization are large because of the common origin or the ongoing gene flow. Notwithstanding, different haplotype groups with close genetic distances were observed within the two main groups in this study. This observed variation within the progenies with similar maternal origins may have resulted from the different population of broodstocks used for the six different trials of breeding. Similar assumption has been advanced by several authors for the observation of haplotypes in fishes from different populations (e.g. Windsor and Hutchinson 1995; Williams, *et al.*, 1997; Doupe' and Lymbery 1999; Cross 2000; Englbrecht, *et al.*, 2002; Frost *et al.* 2006; Norfatimah *et al.*, 2009; Nazia *et al.*, 2010).

Although *C. gariepinus* and *P. hypophthalmus* belong to different families, the phylogenetic analysis of the mitochondrial cytochrome b showed a considerably close genetic distance between these two species. Pardo *et al.*, (2005) and Azevedo *et al.*, (2008), have earlier hypothesized that close genetic distances between different fish families could explain the feasibility of hybridization between them. This assumption may be the reason for the successful hybridization of *C. gariepinus* and *P. hypophthalmus*. Although the hybridization status of the reciprocal crosses produced in this study were earlier confirmed using morphological (Okomoda *et al.*, 2018b) and cytogenetic tools (Okomoda *et al.*, 2018c), the direction of hybridization or cross combination could not be determined. The use of erythrocyte characterization had also been advanced in previous studies by the authors of this research, but with some limitations (Okomoda *et al.*, 2018a). However, the characterization of the mitochondrial DNA still remains the most accurate and unambiguous tool for directional cross determination.

Acknowledgements

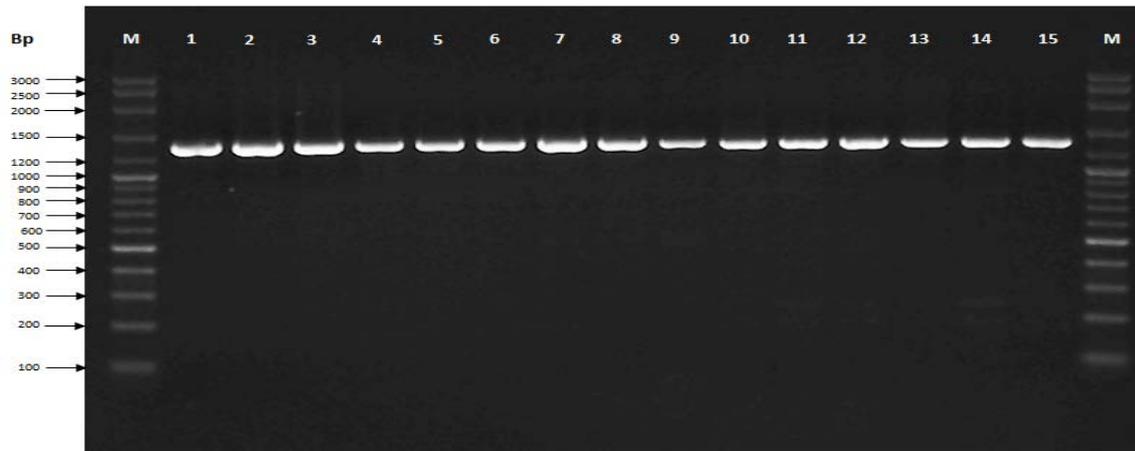
The authors are indebted to the School of Fisheries and Aquaculture Science, Universiti Malaysia Terengganu, Malaysia for providing *C. gariepinus* and *P. hypophthalmus* broodstock used in this study. The researchers also acknowledge the help of all technical staffs of the PPSPA hatchery department and laboratory officers of AKUATROP during the breeding trial and lab work of this study respectively. This study is part of the first author's Ph. D thesis.

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Appendix



Appendix 1. Agarose gel showing the amplified PCR products of the Cyt B of the progenies of pure and reciprocal crosses of *Pangasianodon hypophthalmus* and pure *Clarias gariepinus*. M: Marker; Lane 1-3: *C. gariepinus*; Lane 4-6: Clarias-like Clariiothalmus; Lane 7-9: Panga-like Clariiothalmus; Lane 10-12: Pangapinus; Lane 13-15: *P. hypophthalmus*.

Download [GenBank](#) [Graphics](#)

Clarias gariepinus isolate BCG2 cytochrome b gene, partial cds; mitochondrial
 Sequence ID: [KJ533253.1](#) Length: 1114 Number of Matches: 1

Range 1: 39 to 1114 [GenBank](#) [Graphics](#) ▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1982 bits(1073)	0.0	1075/1076(99%)	0/1076(0%)	Plus/Plus
Query 1	CGACGCACTCATCGACCTTCCC	CGACGCACTCATCGACCTTCCC		60
Sbjct 39	CGACGCACTCATCGACCTTCCC	CGACGCACTCATCGACCTTCCC		98
Query 61	ACTACTATTACTATGTCTTGGAGTACAAATCCTCACAGGACTATTCCTAGCCATACACTA	ACTACTATTACTATGTCTTGGAGTACAAATCCTCACAGGACTATTCCTAGCCATACACTA		120
Sbjct 99	ACTACTATTACTATGTCTTGGAGTACAAATCCTCACAGGACTATTCCTAGCCATACACTA	ACTACTATTACTATGTCTTGGAGTACAAATCCTCACAGGACTATTCCTAGCCATACACTA		158
Query 121	CACCTTCTGATATCTCAACCGCATTCTCATCAGTAGTACACATCTGCCGAGACGTCAACTA	CACCTTCTGATATCTCAACCGCATTCTCATCAGTAGTACACATCTGCCGAGACGTCAACTA		180
Sbjct 159	CACCTTCTGATATCTCAACCGCATTCTCATCAGTAGTACACATCTGCCGAGACGTCAACTA	CACCTTCTGATATCTCAACCGCATTCTCATCAGTAGTACACATCTGCCGAGACGTCAACTA		218
Query 181	CGGATGAATCATCCGAAACCTTACGCCAACGGAGCATCCTTCTTTCATCTGCATCTA	CGGATGAATCATCCGAAACCTTACGCCAACGGAGCATCCTTCTTTCATCTGCATCTA		240
Sbjct 219	CGGATGAATCATCCGAAACCTTACGCCAACGGAGCATCCTTCTTTCATCTGCATCTA	CGGATGAATCATCCGAAACCTTACGCCAACGGAGCATCCTTCTTTCATCTGCATCTA		278
Query 241	CCTTCACATTGGCCGTTGGTCTGTACTATGGCTCATACCTATACAAAGAGACCTGAAACAT	CCTTCACATTGGCCGTTGGTCTGTACTATGGCTCATACCTATACAAAGAGACCTGAAACAT		300
Sbjct 279	CCTTCACATTGGCCGTTGGTCTGTACTATGGCTCATACCTATACAAAGAGACCTGAAACAT	CCTTCACATTGGCCGTTGGTCTGTACTATGGCTCATACCTATACAAAGAGACCTGAAACAT		338
Query 301	CGGCGTCGTAATAAATAACAGCCTTCGTAAGGATACGTAACCATG	CGGCGTCGTAATAAATAACAGCCTTCGTAAGGATACGTAACCATG		360
Sbjct 339	CGGCGTCGTAATAAATAACAGCCTTCGTAAGGATACGTAACCATG	CGGCGTCGTAATAAATAACAGCCTTCGTAAGGATACGTAACCATG		398
Query 361	AGGACAAATATCCTTCTGAGGTGCCACAGTAATCACAACCTCTTATCAGCCGTACCCCTA	AGGACAAATATCCTTCTGAGGTGCCACAGTAATCACAACCTCTTATCAGCCGTACCCCTA		420
Sbjct 399	AGGACAAATATCCTTCTGAGGTGCCACAGTAATCACAACCTCTTATCAGCCGTACCCCTA	AGGACAAATATCCTTCTGAGGTGCCACAGTAATCACAACCTCTTATCAGCCGTACCCCTA		458
Query 421	CATAGGAGATGCCCTAGTCCAATGAATCTGAGGAGGCTTCTCCGTAGACAATGCAACACT	CATAGGAGATGCCCTAGTCCAATGAATCTGAGGAGGCTTCTCCGTAGACAATGCAACACT		480
Sbjct 459	CATAGGAGATGCCCTAGTCCAATGAATCTGAGGAGGCTTCTCCGTAGACAATGCAACACT	CATAGGAGATGCCCTAGTCCAATGAATCTGAGGAGGCTTCTCCGTAGACAATGCAACACT		518
Query 481	TACACGATTCTTTCGCATTCCACTTCTCCTACCATTACAATCATCGCAGCTACAATTCT	TACACGATTCTTTCGCATTCCACTTCTCCTACCATTACAATCATCGCAGCTACAATTCT		540
Sbjct 519	TACACGATTCTTTCGCATTCCACTTCTCCTACCATTACAATCATCGCAGCTACAATTCT	TACACGATTCTTTCGCATTCCACTTCTCCTACCATTACAATCATCGCAGCTACAATTCT		578
Query 541	ACACGCACTATTCTTACAGAAACAGGATCAAAACACCAATTGGACTAAAACCTCCGACGC	ACACGCACTATTCTTACAGAAACAGGATCAAAACACCAATTGGACTAAAACCTCCGACGC		600
Sbjct 579	ACACGCACTATTCTTACAGAAACAGGATCAAAACACCAATTGGACTAAAACCTCCGACGC	ACACGCACTATTCTTACAGAAACAGGATCAAAACACCAATTGGACTAAAACCTCCGACGC		638
Query 601	AGACAAAATCTCATTCCACCCTATTTCTCCTACAAAGACCTACTAGGATTTATCATTCT	AGACAAAATCTCATTCCACCCTATTTCTCCTACAAAGACCTACTAGGATTTATCATTCT		660
Sbjct 639	AGACAAAATCTCATTCCACCCTATTTCTCCTACAAAGACCTACTAGGATTTATCATTCT	AGACAAAATCTCATTCCACCCTATTTCTCCTACAAAGACCTACTAGGATTTATCATTCT		698
Query 661	ATTAACAGCCCTCGCATCTTAAGCCTATTCTCCCAAACTTCTAGGCGACCCAGAAAA	ATTAACAGCCCTCGCATCTTAAGCCTATTCTCCCAAACTTCTAGGCGACCCAGAAAA		720
Sbjct 699	ATTAACAGCCCTCGCATCTTAAGCCTATTCTCCCAAACTTCTAGGCGACCCAGAAAA	ATTAACAGCCCTCGCATCTTAAGCCTATTCTCCCAAACTTCTAGGCGACCCAGAAAA		758
Query 721	CTTACCCCCGGCCAAACCCCTAGTAACTCCACCTCACATCAAACGAGAATGATACTTCT	CTTACCCCCGGCCAAACCCCTAGTAACTCCACCTCACATCAAACGAGAATGATACTTCT		780
Sbjct 759	CTTACCCCCGGCCAAACCCCTAGTAACTCCACCTCACATCAAACGAGAATGATACTTCT	CTTACCCCCGGCCAAACCCCTAGTAACTCCACCTCACATCAAACGAGAATGATACTTCT		818
Query 781	ATTCGCATACGCCATCCTCCGATCCATCCAAACAACTAGGCGGAGTATTAGCACTATT	ATTCGCATACGCCATCCTCCGATCCATCCAAACAACTAGGCGGAGTATTAGCACTATT		840
Sbjct 819	ATTCGCATACGCCATCCTCCGATCCATCCAAACAACTAGGCGGAGTATTAGCACTATT	ATTCGCATACGCCATCCTCCGATCCATCCAAACAACTAGGCGGAGTATTAGCACTATT		878
Query 841	ATTCTCCATCCTAGTACTAATAGTAGTACCACACTACACCTCTCAAACCAACAGGGCCT	ATTCTCCATCCTAGTACTAATAGTAGTACCACACTACACCTCTCAAACCAACAGGGCCT		900
Sbjct 879	ATTCTCCATCCTAGTACTAATAGTAGTACCACACTACACCTCTCAAACCAACAGGGCCT	ATTCTCCATCCTAGTACTAATAGTAGTACCACACTACACCTCTCAAACCAACAGGGCCT		938
Query 901	AACCTTCCGACCTTTATCCCAAATCTTATTCTGAACCTTAGTAGCAGATGTAATAATCTT	AACCTTCCGACCTTTATCCCAAATCTTATTCTGAACCTTAGTAGCAGATGTAATAATCTT		960
Sbjct 939	AACCTTCCGACCTTTATCCCAAATCTTATTCTGAACCTTAGTAGCAGATGTAATAATCTT	AACCTTCCGACCTTTATCCCAAATCTTATTCTGAACCTTAGTAGCAGATGTAATAATCTT		998
Query 961	AACATGAATCGGCGGCATACCAAGTAGAACATCCGTTTCATCATTATCGGACAAATCGCCTC	AACATGAATCGGCGGCATACCAAGTAGAACATCCGTTTCATCATTATCGGACAAATCGCCTC		1020
Sbjct 999	AACATGAATCGGCGGCATACCAAGTAGAACATCCGTTTCATCATTATCGGACAAATCGCCTC	AACATGAATCGGCGGCATACCAAGTAGAACATCCGTTTCATCATTATCGGACAAATCGCCTC		1058
Query 1021	CATCCTCTACTTCTCCCTATTCTCTCATCTTAAACCCACTAGCAGCCTGACTAGAAA	CATCCTCTACTTCTCCCTATTCTCTCATCTTAAACCCACTAGCAGCCTGACTAGAAA		1076
Sbjct 1059	CATCCTCTACTTCTCCCTATTCTCTCATCTTAAACCCACTAGCAGCCTGACTAGAAA	CATCCTCTACTTCTCCCTATTCTCTCATCTTAAACCCACTAGCAGCCTGACTAGAAA		1114

Appendix 2. Aligned *Clarias gariepinus* cytochrome b sequence showing 99 % similarity with *Clarias gariepinus* isolate BCG2 cytochrome b gene, partial cds; mitochondrion (GenBank Accession Number KJ533253.1).

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Clarias gariepinus isolate BCG2 cytochrome b gene, partial cds; mitochondrial
 Sequence ID: [KJ533253.1](#) Length: 1114 Number of Matches: 1

Range 1: 39 to 1114 GenBank Graphics ▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1988 bits(1076)	0.0	1076/1076(100%)	0/1076(0%)	Plus/Plus
Query 1		CGACGCACCTCATCGACCTTCCCGCCCCCTCTAATATCTCCGCATGATGAAACTTTGGCTC		60
Sbjct 39		CGACGCACCTCATCGACCTTCCCGCCCCCTCTAATATCTCCGCATGATGAAACTTTGGCTC		98
Query 61		ACTACTATTACTATGTCTTGGAGTACAAATCCTCACAGGACTATTCTAGCCATACACTA		120
Sbjct 99		ACTACTATTACTATGTCTTGGAGTACAAATCCTCACAGGACTATTCTAGCCATACACTA		158
Query 121		CACCTTCTGATATCTCAACCGCATTCTCATCAGTAGTACACATCTGCCGAGACGTCAACTA		180
Sbjct 159		CACCTTCTGATATCTCAACCGCATTCTCATCAGTAGTACACATCTGCCGAGACGTCAACTA		218
Query 181		CGGATGAATCATTCCGAAACCTTCCGCAACCGGAGCATCTTCTTCTTCTCATCTGCATCTA		240
Sbjct 219		CGGATGAATCATTCCGAAACCTTCCGCAACCGGAGCATCTTCTTCTTCTCATCTGCATCTA		278
Query 241		CCTTACATTTGGCCGTGGTCTGTACTATGGCTCATACCTATACAAAGAGACCTGAAACAT		300
Sbjct 279		CCTTACATTTGGCCGTGGTCTGTACTATGGCTCATACCTATACAAAGAGACCTGAAACAT		338
Query 301		CGGCGTCGTACTACTCCTTTTAGTAATAATAACAGCCTTCGTAGGATACGTAACCATG		360
Sbjct 339		CGGCGTCGTACTACTCCTTTTAGTAATAATAACAGCCTTCGTAGGATACGTAACCATG		398
Query 361		AGGACAAATATCCTTCTGAGGTGCCACAGTAATCACAAACCTCTTATCAGCCGTACCCCTA		420
Sbjct 399		AGGACAAATATCCTTCTGAGGTGCCACAGTAATCACAAACCTCTTATCAGCCGTACCCCTA		458
Query 421		CATAGGAGATGCCCTAGTCCAATGAATCTGAGGAGGCTTCTCCGTAGACAATGCAACACT		480
Sbjct 459		CATAGGAGATGCCCTAGTCCAATGAATCTGAGGAGGCTTCTCCGTAGACAATGCAACACT		518
Query 481		TACACGATTTCTCGCATTCCACTTCTCCTACCAATCACAAATCATCGCAGCTACAATTCT		540
Sbjct 519		TACACGATTTCTCGCATTCCACTTCTCCTACCAATCACAAATCATCGCAGCTACAATTCT		578
Query 541		ACACGCACATTCTTACACGAAACAGGATCAAAACAACCAATGGACTAAGCTCCGACGC		600
Sbjct 579		ACACGCACATTCTTACACGAAACAGGATCAAAACAACCAATGGACTAAGCTCCGACGC		638
Query 601		AGACAAAATCTCATTCCACCCATATTTCTCCTACAAGACCTACTAGGATTTATCATTCT		660
Sbjct 639		AGACAAAATCTCATTCCACCCATATTTCTCCTACAAGACCTACTAGGATTTATCATTCT		698
Query 661		ATTAACAGCCCTCGCATCTCTAAGCCTATTCTCCCAACCTTCTAGGCGACCCAGAAAA		720
Sbjct 699		ATTAACAGCCCTCGCATCTCTAAGCCTATTCTCCCAACCTTCTAGGCGACCCAGAAAA		758
Query 721		CTTACCCCCCGCAACCCCTTAGTAACCTCACATCAAACAGAAATGATACTTCCT		780
Sbjct 759		CTTACCCCCCGCAACCCCTTAGTAACCTCACATCAAACAGAAATGATACTTCCT		818
Query 781		ATTCGCATACGCCATCCTCCGATCCATCCCAACAACTAGGCGGAGTATTAGCACTATT		840
Sbjct 819		ATTCGCATACGCCATCCTCCGATCCATCCCAACAACTAGGCGGAGTATTAGCACTATT		878
Query 841		ATTCTCCATCTAGTACTAATAGTAGTACCCTACTACCTCTCAAAAACAACAGGGCCT		900
Sbjct 879		ATTCTCCATCTAGTACTAATAGTAGTACCCTACTACCTCTCAAAAACAACAGGGCCT		938
Query 901		AACCTTCCGACCTTTATCCCAAATCTTATTCTGAACCTAGTAGCAGATGTAATAATCTT		960
Sbjct 939		AACCTTCCGACCTTTATCCCAAATCTTATTCTGAACCTAGTAGCAGATGTAATAATCTT		998
Query 961		AACATGAATCGGCGGCATACCAAGTAGAACATCCGTTTCATATTATCGGACAAATCGCCTC		1020
Sbjct 999		AACATGAATCGGCGGCATACCAAGTAGAACATCCGTTTCATATTATCGGACAAATCGCCTC		1058
Query 1021		CATCCTCTACTTCTCCCTATTCCTCATCTTAAACCCACTAGCAGCCTGACTAGAAA	1076	
Sbjct 1059		CATCCTCTACTTCTCCCTATTCCTCATCTTAAACCCACTAGCAGCCTGACTAGAAA	1114	

Appendix 3. Aligned Clarias-like Clariothalmus cytochrome b sequence showing 99 % similarity with *Clarias gariepinus* isolate BCG2 cytochrome b gene, partial cds; mitochondrion (GenBank Accession Number KJ533253.1).

[Download](#) [GenBank](#) [Graphics](#)

Pangasianodon hypophthalmus isolate PSH02 cytochrome b (Cytb) gene, partial cds; mitochondrial
 Sequence ID: [KM434895.1](#) Length: 1139 Number of Matches: 1

Range 1: 42 to 1117 [GenBank](#) [Graphics](#) [Next Match](#) [Previous Match](#)

Score	Expect	Identities	Gaps	Strand
1982 bits(1073)	0.0	1075/1076(99%)	0/1076(0%)	Plus/Plus
Query 1	CGACGCACTAATTGACCTTCTGCCCCATCCAATATTTCCGCATGATGAAACTTTGGTTC	60		
Sbjct 42	CGACGCACTAATTGACCTTCTGCCCCATCCAATATTTCCGCATGATGAAACTTTGGTTC	101		
Query 61	CCTACTATTATTATGCTTATAGTACAGATCCTAACAGGACTTTTCTAGCCATACATTA	120		
Sbjct 102	CCTACTATTATTATGCTTATAGTACAGATCCTAACAGGACTTTTCTAGCCATACATTA	161		
Query 121	TACCTCAGACATCTACTGGCTTCTCATCCGTAGCCACATCTGTCGAGATGTAATA	180		
Sbjct 162	TACCTCAGACATCTACTGGCTTCTCATCCGTAGCCACATCTGTCGAGATGTAATA	221		
Query 181	CGGATGAGTCATCCGCAACTTACATGCCAACGGAGCTTCATTCTTTTTCATCTGTATTTA	240		
Sbjct 222	CGGATGAGTCATCCGCAACTTACATGCCAACGGAGCTTCATTCTTTTTCATCTGTATTTA	281		
Query 241	CCTACACATCGGACGAGGATTATATTATGGCTCTTACTTATATAAAGAAACCTGAAATAT	300		
Sbjct 282	CCTACACATCGGACGAGGATTATATTATGGCTCTTACTTATATAAAGAAACCTGAAATAT	341		
Query 301	TGGAGTAGTACTTCTCCTATTAGTTATAATAACCGCTTCGTCGGATATGTTTACCATG	360		
Sbjct 342	TGGAGTAGTACTTCTCCTATTAGTTATAATAACCGCTTCGTCGGATATGTTTACCATG	401		
Query 361	AGGTCAAATATCATTGAGGCGCCACAGTAATCACAAATCTCCTATCAGCTGTCCCTTA	420		
Sbjct 402	AGGTCAAATATCATTGAGGCGCCACAGTAATCACAAATCTCCTATCAGCTGTCCCTTA	461		
Query 421	CATAGGAGATATACTAGTACAATGAATTTGAGGTGGCTTCTCCGTAGACAATGCAACACT	480		
Sbjct 462	CATAGGAGATATACTAGTACAATGAATTTGAGGTGGCTTCTCCGTAGACAATGCAACACT	521		
Query 481	AACACGATTCCTCGCATTTCACTTCTACTTCCATTTCGTAATTGTCGAGCCACAGTATT	540		
Sbjct 522	AACACGATTCCTCGCATTTCACTTCTACTTCCATTTCGTAATTGTCGAGCCACAGTATT	581		
Query 541	ACATGCCTTATTCCTACACGAAACAGGCTCCAATAACCCAATTGGCCTAAACTCCGACGC	600		
Sbjct 582	ACATGCCTTATTCCTACACGAAACAGGCTCCAATAACCCAATTGGCCTAAACTCCGACGC	641		
Query 601	AGACAAAACTCCTCCACCATACTTCTCCTATAAAGATGATTAGGATTCATAATCCT	660		
Sbjct 642	AGACAAAACTCCTCCACCATACTTCTCCTATAAAGATGATTAGGATTCATAATCCT	701		
Query 661	CCTCAGACCCCTCGCATCTTAGCCCTCTTCTCACCAAACCTTTTAGGAGATCCAGAAAA	720		
Sbjct 702	CCTCAGACCCCTCGCATCTTAGCCCTCTTCTCACCAAACCTTTTAGGAGATCCAGAAAA	761		
Query 721	CTTACCCAGCCAAACCCATTAGTAACACCGCCACATCAAACAGAAATGATACTTCT	780		
Sbjct 762	CTTACCCAGCCAAACCCATTAGTAACACCGCCACATCAAACAGAAATGATACTTCT	821		
Query 781	ATTTGCATATGCCATCCTACGATCAATCCCAAATAAGCTAGGAGGGGTCTGGCCCTACT	840		
Sbjct 822	ATTTGCATATGCCATCCTACGATCAATCCCAAATAAGCTAGGAGGGGTCTGGCCCTACT	881		
Query 841	ATTCTCCATCCTAGTATAATAGTTGTTCCCTATTACACACCTCTAAACAACAAGGCCT	900		
Sbjct 882	ATTCTCCATCCTAGTATAATAGTTGTTCCCTATTACACACCTCTAAACAACAAGGCCT	941		
Query 901	CACCTTCGCCCCCTCTCCAATTCCTATTCTGAGCCCTAGTAGCAGACGTAGCCATTCT	960		
Sbjct 942	CACCTTCGCCCCCTCTCCAATTCCTATTCTGAGCCCTAGTAGCAGACGTAGCCATTCT	1001		
Query 961	CACCTTGAATTGGCGGTATACAGTCGAACACCCATTTCATCATTATCGGACAAATCGCCTC	1020		
Sbjct 1002	CACCTTGAATTGGCGGTATACAGTCGAACACCCATTTCATCATTATCGGACAAATCGCCTC	1061		
Query 1021	CATTTTATATTTTCTTCTTCTAGTCCTAAACCCCTAGCAGGATGACTAGAAA	1076		
Sbjct 1062	CATTTTATATTTTCTTCTTCTAGTCCTAAACCCCTAGCAGGATGACTAGAAA	1117		

Appendix 5. Aligned Pangapinus cytochrome b sequence showing 99 % similarity with *Pangasianodon hypophthalmus* isolate PSH02 cytochrome b (Cytb) gene, partial cds; mitochondrial sequence (GenBank Accession Number KM434895.1).

[Download](#) ▾ [GenBank](#) [Graphics](#)

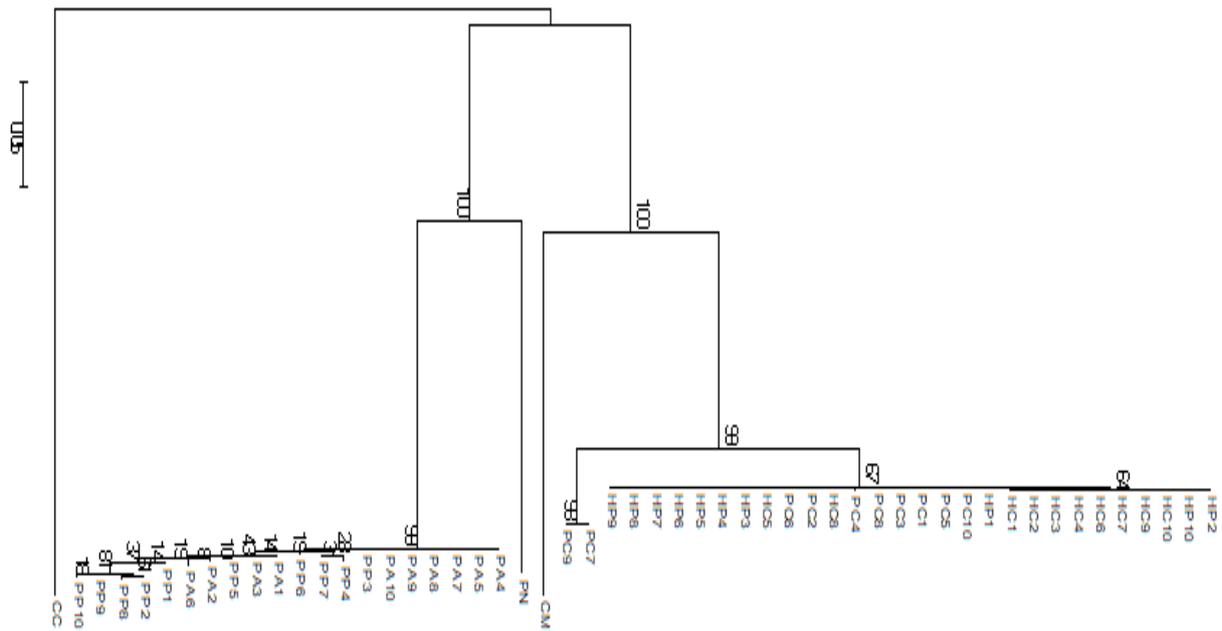
Pangasianodon hypophthalmus isolate PSH02 cytochrome b (Cytb) gene, partial cds; mitochondrial
Sequence ID: [KM434895.1](#) Length: 1139 Number of Matches: 1

Range 1: 42 to 1117 [GenBank](#) [Graphics](#)

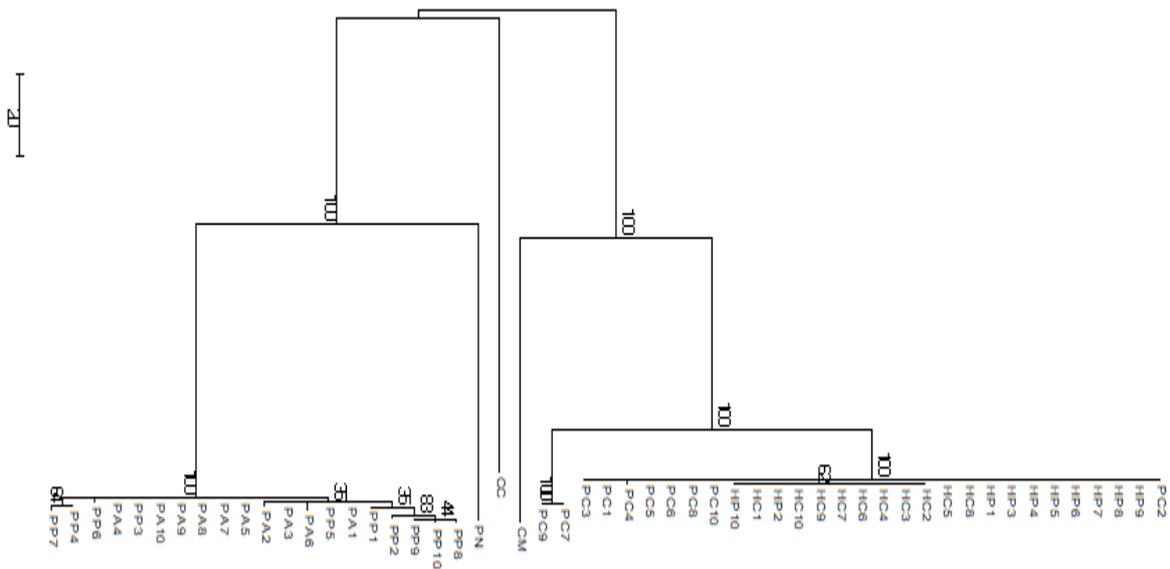
▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1977 bits(1070)	0.0	1074/1076(99%)	0/1076(0%)	Plus/Plus
Query 1	CGACGCACTAATTGACCTTCCTGGCCCATCCAATATTTCCGCATGATGAACTTTGGTTC			60
Sbjct 42	CGACGCACTAATTGACCTTCCTGGCCCATCCAATATTTCCGCATGATGAACTTTGGTTC			101
Query 61	CCTACTATTATTATGCCTTATAGTACAGATCCTAACAGGACTTTTCTAGCCATACATTA			120
Sbjct 102	CCTACTATTATTATGCCTTATAGTACAGATCCTAACAGGACTTTTCTAGCCATACATTA			161
Query 121	TACCTCAGACATCTCTACTGGCTTCTCATCCGTAGCCACATCTGTCGAGATGTAATA			180
Sbjct 162	TACCTCAGACATCTCTACTGGCTTCTCATCCGTAGCCACATCTGTCGAGATGTAATA			221
Query 181	CGGATGAGTCATCCGCAACTTACATGCCAACGGAGCTTCATTCTTTTTCATCTGTATTTA			240
Sbjct 222	CGGATGAGTCATCCGCAACTTACATGCCAACGGAGCTTCATTCTTTTTCATCTGTATTTA			281
Query 241	CCTACACATCGGACGAGGATTATATTATGGCTCTTACTTATATAAAGAAACCTGAAATAT			300
Sbjct 282	CCTACACATCGGACGAGGATTATATTATGGCTCTTACTTATATAAAGAAACCTGAAATAT			341
Query 301	TGGAGTAGTACTTCTCCTATTAGTTATAATAACCGCCTTCGTGGGATGTTTTACCATG			360
Sbjct 342	TGGAGTAGTACTTCTCCTATTAGTTATAATAACCGCCTTCGTGGGATGTTTTACCATG			401
Query 361	AGGTCAAATATCATTTTGAGGCGCCACAGTAATCACAATCTCCTATCAGCTGTCCCTTA			420
Sbjct 402	AGGTCAAATATCATTTTGAGGCGCCACAGTAATCACAATCTCCTATCAGCTGTCCCTTA			461
Query 421	CATAGGAGATATACTAGTACAATGAATTTGAGGTGGCTTCTCCGTAGACAATGCAACACT			480
Sbjct 462	CATAGGAGATATACTAGTACAATGAATTTGAGGTGGCTTCTCCGTAGACAATGCAACACT			521
Query 481	AACACGATTCCTCGCATTTCACTTCTACTTCCATTGTAATTGTCGACGCCACAGTATT			540
Sbjct 522	AACACGATTCCTCGCATTTCACTTCTACTTCCATTGTAATTGTCGACGCCACAGTATT			581
Query 541	ACATGCCTTATTCTACACGAAACAGGCTCCAATAACCCAATTGGCCTAACTCCGACGC			600
Sbjct 582	ACATGCCTTATTCTACACGAAACAGGCTCCAATAACCCAATTGGCCTAACTCCGACGC			641
Query 601	AGACAAAATCTCCTTCCACCATACTTCTCCTATAAAGATGATTAGGATTCATAATCCT			660
Sbjct 642	AGACAAAATCTCCTTCCACCATACTTCTCCTATAAAGATGATTAGGATTCATAATCCT			701
Query 661	CCTCACAGCCCTCGCATCTTAGCCCTCTTCTCACCAAACCTTTTAGGAGATCCAGAAAA			720
Sbjct 702	CCTCACAGCCCTCGCATCTTAGCCCTCTTCTCACCAAACCTTTTAGGAGATCCAGAAAA			761
Query 721	CTTCAACCCAGCCAAACCATAGTAACACCGCCACATCAAACAGAAATGATACTTCTCT			780
Sbjct 762	CTTCAACCCAGCCAAACCATAGTAACACCGCCACATCAAACAGAAATGATACTTCTCT			821
Query 781	ATTTGCATATGCCATCTACGATCAATCCCAAATAAGCTAGGAGGGGTCTGGCCCTACT			840
Sbjct 822	ATTTGCATATGCCATCTACGATCAATCCCAAATAAGCTAGGAGGGGTCTGGCCCTACT			881
Query 841	ATTCTCCATCTAGTATTAATAGTTGTTCCCTATTACACACCTCTAAACAACAAGGCCT			900
Sbjct 882	ATTCTCCATCTAGTATTAATAGTTGTTCCCTATTACACACCTCTAAACAACAAGGCCT			941
Query 901	CACCTTCCGCCCCCTCTCCAATTCCTATTCTGAGCCCTAGTAGCAGACGTAGCCATTCT			960
Sbjct 942	CACCTTCCGCCCCCTCTCCAATTCCTATTCTGAGCCCTAGTAGCAGACGTAGCCATTCT			1001
Query 961	CACCTTGAATTGGCGGTATACAGTCGAACACCCATTATCATTATCGGACAAATCGCCTC			1020
Sbjct 1002	CACCTTGAATTGGCGGTATACAGTCGAACACCCATTATCATTATCGGACAAATCGCCTC			1061
Query 1021	CATTTTATATTTTCTTCTTCTTAGTCTAAACCCCTAGCAGGATGACTAGAAA			1076
Sbjct 1062	CATTTTATATTTTCTTCTTCTTAGTCTAAACCCCTAGCAGGATGACTAGAAA			1117

Appendix 6. Aligned *Pangasianodon hypophthalmus* cytochrome b sequence showing 99 % similarity with *Pangasianodon hypophthalmus* isolate PSH02 cytochrome b (Cytb) gene, partial cds; mitochondrial sequence (GenBank Accession Number KM434895.1).



Appendix 7. Phylogenetic relationships of the different progenies following Neighbour joining analysis of 1076bp of the cytochrome b gene. PC1 - PC10 = *C. gariepinus*; HC1 - HC10 = Clarias-like Clariiothalmus; HP1 - HP10 = Panga-like Clariiothalmus; PA1 - PA10 = Pangapinus; PP1 - PP10 = *P. hypophthalmus*; CC = *Cyprinus carpio*; CM = *Clarias macrocephalus*; PN = *Pangasius nasutus*. (Support values are bootstrap values).



Appendix 8. Phylogenetic relationships of the different progenies following Maximum Parsimony analysis of 1076bp of the cytochrome b gene. PC1 - PC10 = *C. gariepinus*; HC1 - HC10 = Clarias-like Clariiothalmus; HP1 - HP10 = Panga-like Clariiothalmus; PA1 - PA10 = Pangapinus; PP1 - PP10 = *P. hypophthalmus*; CC = *Cyprinus carpio*; CM = *Clarias macrocephalus*; PN = *Pangasius nasutus*. (Support values are bootstrap values).

