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Relationship of Biometric Size-Weight, Nutritive Value, and Metal Concentrations in *Clarias lazera* (Cuvier and Valenciennes) Reared in Treated Wastewater

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Abstract

The objective of the present study was to investigate the nutritive value and heavy metals accumulation in the flesh of *Clarias lazera* (Cuvier and Valenciennes) reared in the discharge canal of Soba wastewater treatment station, south Khartoum, Sudan. A total of 57 fish were collected from the canal and 52 from the White Nile which served as the control. The proximate composition of fish and concentrations of eight hazardous heavy metals i.e. chromium (Cr), iron (Fe), copper (Cu), zinc (Zn), lead (Pb), robidium (Rb), strontium (Sr), and mercury (Hg) in the flesh of *C. lazera* were measured. The results which were statistically analyzed revealed insignificant differences in moisture, ash, fat contents, energy value, and fat: protein ratio (P>0.05) between the two studied sites. A significant difference (P<0.05) was evident in the protein content of fishes from both locations. Heavy metals accumulation in *Clarias* tissues differ from one element to another depending on each element characteristics and local environmental conditions e.g. Sr was higher in the White Nile fishes than in treated wastewater fishes (P<0.05). Accumulation of Pb and Hg was comparable in wastewater and White Nile (P<0.05). Cr was also higher in treated wastewater fishes (P<0.05). Nevertheless, the concentrations of most considered elements were lower than levels recommended by various international agencies.

Key words: Clarias lazera, Heavy metals, Accumulation, White Nile, Treated wastewater.

1. Introduction

Treated wastewater was used in 19th century in Europe to irrigate crops (Ensink and van der Hoek, 2007). Reuse of treated wastewater for fish aquaculture is practiced in many countries including India (Bunting, 2006), Egypt (Misheloff, 2010), and Netherland (Oberdieck and Verreth 2009). As water demand becomes an increasingly important concern in many places, and especially essential for the increasing human population, fish farming falls within the many options which exist for productive wastewater treatment design systems (WHO, 2006). Popular fish species suitable for fish farming in treated wastewater include catfish, tilapia spp. trout, carp, and many others. The annual production depends on fish species, local and environmental conditions. According to Girard (2011), there are many constraints to reusing treated wastewater for rearing fish, such as lack of knowledge, limited available sites; rapid urbanization, rapid eutrophication, improved sanitation, rapid industrialization contamination, social and cultural acceptance and climate. The pathogen transmission risk through treated wastewater fish farming represents a controllable risk; i.e., pathogen loads can be reduced to acceptable levels if adequate measures are adopted (Straus 1996). Microbial requirements for waste-fed aqua cultural schemes should be compatible with background levels in natural waters, since the harvesting of fish and other aquatic animals is generally unrestricted and socially accepted.

Fish is one of the most important available sources of animal protein in the tropics, and has been widely accepted as a good source of protein and other elements for the maintenance of a healthy body (Tidewell and Allan, 2001). Heavy metals are persistent contaminants in the environment that come to the forefront of dangerous substances such as cadmium, lead, mercury, copper and zinc that cause serious health hazards to humans and animals (Ahmed *et al.*, 1998). The agricultural and

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industrial wastes, partially treated or untreated regularly, are being discharged into surface water (Forstner and Wittmann, 2007). Heavy metals present in such polluted waters are absorbed through gills, skin and digestive tract of fish by bio-concentration and bio-magnification. Heavy metals are natural trace components of the aquatic environment, but their levels are increased due to domestic, industrial, mining and agricultural activities (Mance, 1987; Kalay and Canli 2000). At low levels, some heavy metals such as copper, cobalt, zinc, iron and manganese are essential for enzymatic activity and many biological processes. However, the essential metals may also become toxic at high concentrations (Bryan 1976 and Authman et al., 2012). Other metals, such as Cd, Hg, Pb play unknown essential roles in living organisms, and are toxic at even low concentrations (Authman et al., 2013).

Clarias lazera (Cuvier. and Valenciennes) is a freshwater fish which attracts attention as a potential fish for aquaculture (Babiker, 1984). According to Chervinski (1984), presence of a breathing apparatus enables the fish to withstand low level of oxygen and a wide range of temperatures. It is an omnivorous feeder found mainly in shallow waters; young ones feed on ostracods and aquatic insects while adults feed on any potential food like zooplankton and molluscs, but mainly on fish e.g. *Oreochromis niloticus* (Amirthalingam and Khalifa, 1965).

The aim of this study was to highlight the implications of the reuse of treated wastewater for fish culture, and to examine and compare the levels of toxic heavy metals accumulation in the muscle tissue of *C. lazera* from the polluted discharge canal of Soba wastewater treatment station and non-polluted of Jebel Aulia Reservoir on the White Nile, south Khartoum, Sudan. The proximate chemical composition and condition factor of fish were studied as complementary aspects. The present study is expected to shed light on the importance of the reuse of treated wastewater for fish farming and its promising role in contributing to provide a safe food for the increasing Sudanese population.

2. Materials and Methods

A total of 57 specimens of *C. lazera* (Arabic Garmouth) were collected from the effluent (discharge) canal carrying the treated wastewater from the maturation ponds, in addition to 52 specimens from the White Nile in the vicinity of Jebel Aulia Dam (control). Fish samples were collected from local fishermen during the period April 2012-April 2013. The canal (Figure 1) is located in the Southern part of Khartoum State ca. 15 Km, (15°29' 55"; 15°30' 25" N; 32°32' 36" – 32°36' 06" E).

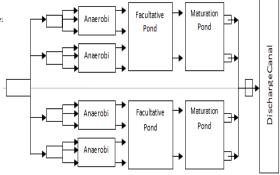


Figure 1: Ponds arrangement, flow pattern and the effluent discharge canal in Soba Wastewater Stabilization Station (Source: Ali and Hag Ibrahim, 2005).

The 2^{nd} site, Jebel Aulia dam on the White Nile, was built in 1937 ca. 50 Km south of Khartoum to store $3.5X10m^9$ of water (Rzoska *et al.* 1955). The dam length is 5 km and creates a large lake with a width ranging between 1-4 km and a maximum depth 22.5 m. As a result of the dam, a large shallow lake of about 12000 hectares was formed capable of storing 3.5 milliard m³. The maximum depth of the reservoir is ca. 15 m attained during high flood (August-mid September).

For each fish specimen, the total and standard lengths were measured, and then each fish was weighed. The condition factor (K) was calculated according to Le-Cren, 1951 equation:

$K = 100.W/L^3$

W is fish body weight (g) and L is fish total length (cm) $% \left({{{\mathbf{r}}_{\mathrm{m}}}} \right)$

Fishes were sexed and aged using vertebrae. Use of vertebrae for age determination was proved to be one of the reliable structures for fish ageing (Bishai and Abu Gideiri 1965, Mishrigi 1967, Bishai 1970, Gumaa 1974 and Tweddle 1975). Otolith was not used for aging C. lazera because its major advantage over other tissues e.g. scales and vertebrae is the presence of clearly visible daily lines (Pannella 1974). In addition, preparation of otoliths for light microscopy or scanning microscope (Liew 1974) is rather difficult and requires facilities not available in our institutes. Sexual maturity was determined according to Nikoliskii (1963). The stomach food content was investigated using the method described by Hynes (1950) where stomachs contents are examined and the individual food organisms was sorted and identified. The number of stomachs in which each item occurs is recorded and expressed as a percentage of the total number of stomachs examined. The collected fish were then skinned; the flesh was taken from different sites of the body to make sure that the examined sample is well representative to the whole body. Flesh samples were kept in air-tight plastic bags and frozen at 0.0-5.0°C till they were used for moisture determination. and ash contents

The frozen samples were then freeze-dried for 24 hrs using (Edwards High Vacuum 2507 Freeze Dryer). The dried samples were ground using a non metallic mortar. Powder samples were then kept in the air-tight plastic bags. All chemical constituents were determined according to Pearson 1976 and AOAC 1980. These include moisture; protein, fat and ash contents. Protein was evaluated by Micro-Kjeldahl Method; fat content by using soxhelt extraction method. The energy value was calculated from the fat and protein contents of samples using the values 9.02 Kcal./gm for fat content and 4.27 Kcal./gm for protein content as recommended by FAO (1989).

X-ray fluorescence spectroscopy (XRF) was used for qualitative and quantitative determination of heavy metals (Tertian and Claisse 1982). These are namely: Cr, Fe, Cu, Zn, Pb, Rb, Sr and Hg. According to Talbot (1987), XRF has the potential and capacity to give accurate linear response to a broad spectrum of elements in approximately 0.0-500 mg/Kg-1.

The data obtained was statistically analyzed using SPSS package (t-test for normal data and Mann Whitney u test for data not normally distributed).

3. Results and Discussion

The results of the present study (Table 1) showed greater sizes and weights of fishes in treated wastewater compared to those collected from the White Nile (P<0.05). The same applies to standard lengths although the difference was insignificant (P>0.05). According to Mason 1991, such results are possibly attributed to the availability of food in treated wastewater and the presence of organic matter which is rich in proteins, carbohydrates, and fats.

 Table 1: Weights and standard lengths of C. lazera in treated wastewater and White Nile

Parameter	Treated Wastewater	White Nile	Р
Weight (gm)	528.96±38.9	360±45.90	*
Standard length	35.04±1.14	30.94±1.21	NS

The condition factor of fish reared in both study sites was calculated to evaluate fish situation. The mean condition factor of fish was found to be 1.23 g/cm³ and 1.22 g/cm³ in treated wastewater and White Nile, respectively. Both values were above 1.0 indicating good conditions of the fish (Barnham and Baxter 1998), however, they were below those obtained by Nwabueze 2013 for *Clarias anguillaris*. Furthermore, King (1995) attributed differences in condition factors of fish to food abundance, adaptation to the environment and gonadal development. According to Aloo (1999), the intensity of infection with *Contracaecum* was increased with the increase in the size of bass *Micropterus salmoides*, from Lake Naivasha and the Oloidien Bay, Kenya

Only two stages "I and IV" were observed out of Nikoliskii (1963) six stages for sexual maturity (Stage 1, Immature; Stage 11, quiescent; Stage 111, maturing: Stage 1V, mature: Stage V running; Stage V1, Spent).

Table 2 illustrates the difference in body weight according to sex, age and maturity stage and the results gave an insignificant difference with sex and maturity stage (P>0.5) in both localities. However, weights of mature fish were higher than immature fish. Regarding age, a significant difference was encountered (P<0.05) in weights of fish; older fish weights were higher than younger ones in both sites.

Figure 2 shows the moisture, ash, fat and protein contents which were examined to assess the fish flesh quality. The fat/protein ratio was calculated to indicate the nutritive value of flesh. Protein content in wastewater fish was higher than in the White Nile (P<0.05). Fat, moisture content and fat/protein ratio were almost similar (P>0.05) in the two sites (0.212 for Treated wastewater and 0.217 for the White Nile).

The food value of fish is normally estimated as the percentage of the edible portion to the total weight of fish and its contents of the basic nutrients i.e. fats and proteins (Karrar, 1997). In the present study, the energy values calculated were 0.0683 and 0.0629 for wastewater and White Nile fish, respectively. These figures indicate that the nutritive values of the fish from both study sites are comparable; however, freshwater fishes are slightly more nutritious due to their low fat/protein ratio.

Table 2: Variation in body weights of C. lazera in treated wastewater and White Nile according to sex, age group and maturity stage

	Sex		Age Groups			Maturity stage			
	Treated wastewater	White Nile	Treated Whi wastewater		White Nile		ated stewater	White Nile	
Male	$459.38{\scriptstyle\pm}63.5$	392.00±86.60	1	427.20±29.8	1	308.60±29.30	А	371.04±33.5	336.8±58.7
Female	575.35+63.50	333.28±42.20	2	936.03±53.1	2	1480.00 ± 10.0	В	714.40±53.7	426.4±10.0
Р	NS	NS	*		*		*		*

*significant; NS insignificant

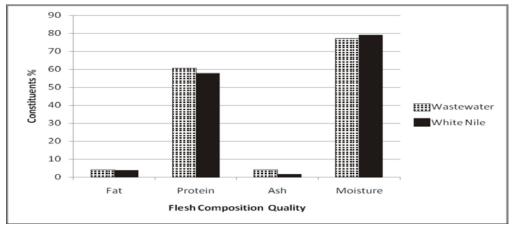


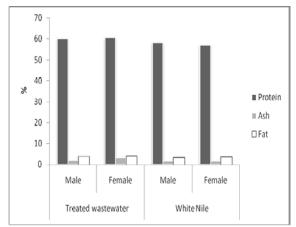
Figure2. Chemical composition of C. lazera in treated wastewater and White Nile

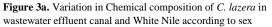
The energy value was higher in the White Nile fish (445.911+17.23 Kcal / 100 gm) than in wastewater $(414.540\pm8.394 \text{ Kcal} / 100 \text{ gm})$, but the difference was insignificant (*P*>0.05).

Figures 3a, 3b and 3c illustrate no significant difference in fat, protein and ash contents with sex, age and maturity stage, respectively (P>0.05). Exceptions were encountered in females' fat content in the White Nile that showed significantly higher values than in males (P<0.05).

Proximate composition showed no significant variation according to sex in both localities. This finding is in agreement with Mohammed *et al.* (1988) who assessed the chemical composition of *Mugil cephalus* along the Sudanese Red Sea Coast. Also proximate composition showed no significant variation with age. That result was in contrast to Dambergs (1963), but older fish reflected slight insignificant increase in fat and protein contents.

Protein and ash contents were insignificantly higher in mature fish in both study sites and that was also in contrast to Dambergs (1963), but fat content showed insignificantly higher levels in the immature fish. According to Hoar (1957), fat storage in the muscle tissue increased prior gonads maturation to provide energy for spawning activities.





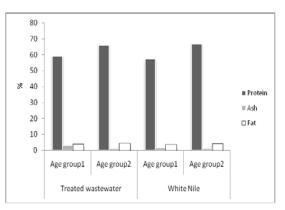


Figure 3b. Variation in Chemical composition of *C. lazera* in wastewater effluent canal and White Nile according to age group (age group 1: 1, 2 and 3 years; Age group 2: 4 and 5 years)

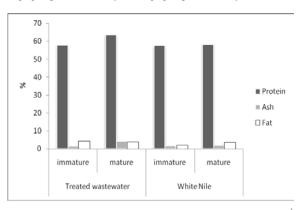


Figure 3c. Variation in Chemical composition of *C. lazera* in wastewater effluent canal and White Nile according to age

Concentrations of Fe, Cu, Zn and Rb (Table 3) were significantly higher in treated wastewater fishes than natural water fishes (P<0.05). In view of this, they were further analyzed with sex, age group, and maturity stage (Figures4a, 4b, 4c). Fe and Zn showed no variation with all the previously mentioned parameters in both study sites (P>0.05). Cr was also higher in treated wastewater fishes (P>0.05). Hg and Pb concentration was higher in treated wastewater than the White Nile. Statistically, the two heavy metal expressed insignificant difference ((P>0.05). Sr was significantly higher in the White Nile compared with treated wastewater (P<0.05).

 Table 3. Heavy metals concentrations in C. lazera in treated wastewater and White Nile

Conc. (ppm)	Wastewater	White Nile	Р	Permissible limits (µg/g wet weight)
Cu	$4.86{\pm}~0.56$	$1.26{\pm}0.16$	*	3.280 "IAEA 2003; FAO 1983a"; 20 "MAAF 2000"
Rb	$177.63{\pm}23.18$	$12.89{\pm}\ 1.54$	*	2.86 "IAEA 2003"
Cr	$0.05{\pm}\ 0.02$	0.03 ± 0.01	NS	0.730 "IAEA 2003"
Fe	$72.64{\pm}\ 12.60$	6.94±0.93	*	146 "IAEA 2003"
Zn	497.20± 44.7	37.83±7.04	*	30 FAO 1983a; 40 FAO/WHO 1989
Hg	60.03 ± 1.30	57.42±3.32	*	0.222 "IAEA 2003"; 0.50 "EC 2006"
Pb	$0.17 \pm \ 0.02$	$0.09{\pm}0.01$	*	0.2 "EC 2005"
Sr	$2.7\ 3\pm 2.46$	17.55 ± 10.35	*	130 "IAEA 2003"

These results are in agreement with the findings of many authors e.g. Clement and Lovel (1994) and Gomez (2011). However, ash content was significantly higher in treated wastewater fishes (P<0.05). The concentrations of most elements considered in this study were lower than levels recommended by the international agencies e.g.EC (2006); EC (2005); IAEA (2003); FAO/WHO (1989); FAO (1983a). Rb showed significant variation with sex and age in both study sites but not with maturity stage. Cu concentration showed no variation with sex in both study sites (P>0.05). However, it gave significant differences with age in the two studied sites. With maturity stage, Cu was much higher in treated wastewater fish and expressing a significant difference (P<0.05).

Treated wastewater fish were found to accumulate considerable amounts of heavy metals compared with White Nile fish. This is likely because metals have certain properties which make them difficult to treat or remove from wastewater if not treated by certain chemical methods (Mubarak, 2014), and living organisms usually form an intimate relationship with the chemical composition of their environment (Elgobashy et al. 2001). According to Jezierska and Witeska (2006), fish living in polluted waters tend to accumulate heavy metals in their tissues. Generally, accumulation depends on metal concentration, time of exposure, way of metal uptake, environmental conditions (water temperature, pH, hardness, salinity), and intrinsic factors (fish age, feeding habits). Concentrations of Pb and Hg in treated wastewater fish were almost similar to fish of natural nonpolluted water possibly because surface water does not always reflect the real situation of heavy metals pollution. This is expected since the physico-chemical characteristics of water normally affect the distribution of metals in water. In addition, Karrar (1997) reported that some metals have the property of being rapidly adsorbed to particulate materials like detritus and suspended sediments. Moreover, some biological processes like uptake by planktons through assimilation can exert the same effect (Abu Gideiri, 1980).

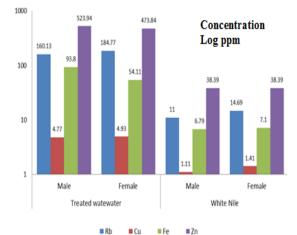
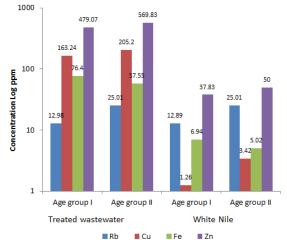
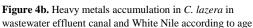


Figure 4a. Heavy metals accumulation in C. lazera in

wastewater effluent canal and White Nile according to sex





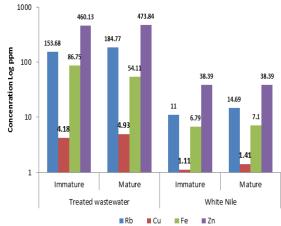


Figure 4c. Heavy metals accumulation in *C. lazera* in wastewater effluent canal and White Nile according to maturity stage

Cu, Rb, Fe and Zn showed significantly higher levels of accumulation in fish of treated wastewater than White Nile fish (P<0.05) probably because their concentration was lower in the latter site. However, except for Zn, the concentrations of other metals did not exceed the permitted international levels (FAO 1983a, b; Huss, 1994; Al-Wher, 2008). Cr, Hg and Pb reached the wastewater

treatment plant with the wastewater transported from the Khartoum Industrial Area e.g. Khartoum Tannery, Printing facilities, paints factories and other industrial activities (Sabir et al. 2007). These metals were not subject to any biodegradation since the station depends mainly on natural micro-organisms which can only decompose organic matter. Zn, Fe and Cu concentrations were higher than Hg, Cr and Pb in both sites. Such situation could be attributed to the fact that Zn, Fe and Cu are essential micro-nutrients required in life processes, so most organisms have the ability to keep them at high concentrations in their bodies. This capacity is enhanced by certain feeding and metabolic processes which lead to high accumulation. Furthermore, many of these metals are capable of forming complexes with the available organic substances and hence have the tendency to be fixed in tissues rather than being excreted (Mara and Cairncross 1989). Deposition of heavy metals in fish tissues in treated wastewater seems to inflict no harmful effects on Clarias which survive successfully with increasing numbers and sizes. In conformity with this, Zaki (2007) stated that aquatic fauna are exposed to chronic substances i.e. pollutants that do not cause heavy mortalities but fishes survive and accumulate various amounts of microbial or chemical residues of heavy metals which might result, in extreme cases, in unpleasant tastes or are potentially dangerous. Saeed (2007) results on C. gariepinus in Lake Edku, Egypt, revealed that accumulation of heavy metals generally associated with specific tissues/organs of the fish e.g. Cu in liver, Cd in gills, liver and ovary and kidney and Pb in gills and ovary. Furthermore, Benamar and Zitouni (2013) reported that accumulation of Cu is higher than Cr in liver tissue compared with the muscle tissue of Sardinella aurita collected from Oran Coastline in Algeria. Das and Gupta (2013) concluded that Cu accumulation pattern in the Indian flying barb Esomus danricus was related to metal concentration and increased with exposure time. The findings of this study are also in agreement with Mason (1991) and Ebrahimi et al. (2007) who reported that consumption of edible tissue of mullet which is subjected to heavy metals pollution is not harmful to humans and that the heavy metals accumulation in fish tissues is below the Egyptian standards. Presence of some organic substances in treated wastewater may render heavy metals less toxic than they would be in pure uncontaminated conditions. The form of heavy metal to which aquatic organisms are exposed is important and determines its overall toxicity (Singh et al. 2011).

Cu, Rb, Fe and Zn showed slightly different pattern when analyzed according to sex. Values of Fe, Cu, and Zn showed insignificant variation between males and females in both study sites, while Rb showed significant increase in female specimens from both study areas which may be due to the high ash content of females. Patin (1982) however, reported a positive correlation between the ash content of the fish and its content of a certain metal. Authman and Abbass (2007) findings on heavy metals in *Tilapia zilli* and *Mugil cephalus* tissues indicated that accumulation was much higher than concentration of these heavy metals in their surrounding waters. The water characteristics for the two sites, obtained by Sabir *et al.* (2007) were in conformity with the results of the present study.

According to age, Cu, and Rb showed significantly higher accumulation in older fish in both localities, Zn insignificantly showed the same (Figure 4b). That is likely because fish accumulate elements throughout their life, so older fish would contain more of these substances (Connel 1975; Huss 1994). The previous finding is in agreement with Patin (1982) who found positive correlation between the actual metal content of a fish and its age, size and weight. Fe although showed no variation with age, the difference was not significant.

The pattern of variation of Cu, Rb, Fe and Zn with the stages of maturity of fish from both study sites did not follow certain models although Cu, and Rb showed insignificantly higher levels in mature fish in both study sites and Zn showed the same in treated waste water fish. According to Huss (1994), fish becomes mature when it reaches certain size not a certain age so mature fish with larger sizes are expected to contain more heavy metals as the actual metal content of a fish is positively related to its size and weight.

Insects, phytoplankton, zooplankton, weeds, molluscs and fish parts were detected as major food constituents in the stomach of Clarias lazera from both studied localities (Figure 5). Phytoplankton, fish parts and molluscs showed significant variations between the two sites whereas mean values of insects, zooplankton and weeds were almost similar (P>0.05). Fish, molluscs and zooplankton were much more detected in stomachs of freshwater fish because wastewater doesn't suit their growth. According to Jumaa (1974) the partially low oxygen, elevated CO_2 and the relatively high concentrations of nitrogen and nitrates create unfavorable conditions for zooplanktons growth. Besides, metals in both cationic and soluble complex forms can be toxic or inhibit zooplanktons growth (Biesinger and Christenser 1972). Superiority of phytoplankton and insects can be explained as due to presence of abundant organic matter in the treated wastewater which furnishes suitable conditions for their growth.

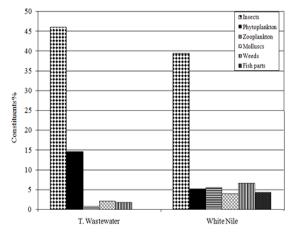


Figure 5. Food composition of *C. lazera* in treated wastewater effluent canal and White Nile

4. Conclusions

Insignificant variations were evident in proximate chemical composition of fish with age, sex, and maturity stages in both sites. However, fish collected from the treated effluent canal expressed higher sizes; weights and protein content are preferred by local consumers. Nutritive value of fishes reared in treated wastewater effluent was high and almost similar to that of natural habitat fish with significantly higher protein content. In spite of possible cultural bias against fish reared in treated wastewater effluents, the average concentrations of most hazardous heavy metals e.g. Hg and Pb are comparable to those obtained from fishes of the White Nile and are well below the values recommended by IAEA (2003), EC (2006) and other relevant agencies for human exceeded consumption. Zn concentrations the recommended level set by international agencies for human consumption and may constitute a potential health risk if ingested in large quantities.

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References

Abu Gideiri Y. B. 1980. Metal contamination in central Red Sea. Report to Red Sea Commission.

Ahmed YF, Mohamed MM, El-Nemer IZ, El-Desoky KI and Ibrahim SS. 1998. Some pathological studies on the effect of cadmium and mercuric chlorides on the gonads of catfish (*Clarias lazera*). Egypt J Comp Pathol Clin Pathol., **11**: 72-81.

Amirthalingam C and Khalifa MY.1965. A Guide to Common Commercial Freshwater Fishes in the Sudan. Government Printing Press, Khartoum, 197 pp.

Ali A K S and Hag Ibrahim S N. 2005. Soba wastewater stabilization pond (WSP) - Sudan: Treatment performance and microbiological quality. Proceedings Sardinia 2005, 10th *International Waste Management and Landfill Symposium*.

Aloo PA.1999. Ecological studies of helminth parasites of the large mouth bass, *Micropterus salmoides*, from Lake Naivasha and the Oloidien Bay, Kenya. Onderstepoort *J Vet Res.*, **66**:73-79.

Al-Wher S M. 2008. Levels of heavy metal Cd, Cu, Zn in three fish species collected from Northern Jordan Valley, Jordan. *Jordan J Biol Sci.*, **1(1)**: 41-46.

AOAC 1980. **Official Methods for Analysis**, Horwitz, N. (Ed.), 13th Ed. Association of Official Analytical Chemists, Washington D. C. 957 pp.

Authman M M N and Abbas WT. 2007. Accumulation and distribution of copper and zinc in both water and some vital tissues of two species (*Tilapia zilli* and *Mugil cephalus*) of Lake Garoun, Fayoum Province, Egypt. *Pak J Biol Sci.*, **10**(13): 2106-2122.

Authman M M N, Abbas WT and Gaafar AY. 2012. Metals concentrations in Nile tilapia *Oreochromis niloticus* (Linnaeus,

1758) from illegal fish farm in Al-Minufiya Province, Egypt, and their effects on some tissues structures. *Ecotoxicol. Environ. Saf.*, **84**: 163–172.

Authman M M N, Abbas HH and Abbas WT. 2013. Assessment of metal status in drainage canal water and their bioaccumulation in *Oreochromis niloticus* fish in relation to human health. *Env Mon Asses.*, **185(1)**: 891-907.

Babiker M M. 1984. Aspects of the biology of the catfish *Clarias lazera* (Cuv. & Val.) related to its economic cultivation. *Hydrobiol.*, **110(1)**: 295-304.

Barnham C and Baxter A. 1998. Condition Factor, K, for Salmonid Fish. Fisheries Notes. State of Victoria, Department of Primary Industries 2003:3 pp.

Benamar N and Zitouni B. 2013. Levels of chromium and copper in liver and muscle tissues of the round *Sardinella Sardinella aurita* (Valenciennes) from the Oran Coastline, Algeria. *Jordan J Biol Sci.*, **6** (4): 252 – 256.

Biesenger K E and Christenser GM. 1972. Effect of varius metals on survival, growth, reproduction and metabolism of *Daphnia mogna*. *Fish Res Bd. Canada*, **29**: 1691-1700.

Bishai H M and Abu Gideiri YB.1965. Studies on the biology of genus Synodontis at Khartoum I: Age and Growth. *Hydrobiol.*, **26**: 85 – 97.

Bishai H M. 1970. Studies on the biology of family Bagridae in the Sudan. A Ph. D Thesis submitted to Fac. Sci. Cairo University. Eygpt.

Bryan G W. 1976. Some effects of heavy metal tolerance in aquatic organisms. In: Lockwood APM (Ed.) **Effects of Pollutants on Aquatic Organisms**. Cambridge University Press. Cambridge, England. pp. 7.

Bunting S W. 2006. Confronting the realities of wastewater aquaculture in peri -urban Kolkata with bioeconomic modeling. *Water Res.*, **41**: 499-505.

Chervinski J. 1984. Salinity tolerance of young catfish *Clarias* lazera. J Fish Biol., 23: 147-149.

Clement S and Lovel R T. 1994. Comparison of processing yield and nutrient composition of cultured Nile Tilapia *Oreochromis niloticus* and channel cat fish *Ictaluruspunctatus*. *Aquaculture*, **119** (2 – 3): 299–310.

Connel J J. 1975. Control of Fish Quality: Fishing News Ltd. Surrey, England.

Dambergs N. 1963. Extractives of fish muscle. 3. Amount, sectional distribution, and variations of fat, water solubles, protein and moisture in cod "*Gadus morhual* L." fillets. *J Fish Res Bd Can.*, **20(4)**: 909-918.

Das S and Gupta A. 2013. Accumulation of copper in different tissues and changes in oxygen consumption rate in Indian flying barb, *Esomus danricus* (Hamilton-Buchanan) exposed to sublethal concentrations of copper. *Jordan J Biol Sci.*, **6**(1): 21 - 24.

Ebrahimi G, El Attia MA and Waffek M. 2007. Heavy metals and bacteria distribution in different organs of grey mullet (*Mugil cephalus*) cultured in different environmental conditions. The International Arab African Fish Resources Conference and Exhibition, Cairo, Egypt.

EC, 2005. European Community. Commission Regulation No 78/2005 pp.: L16/43–L16/45. *Official Journal of the European Union* (20.1.2005).

EC. 2006. European Community. Commission Regulation (EC) No. 1881/2006 of 19 December 2006. Setting maximum levels for certain contaminants in foodstuffs. OJ L 364.

Elgobashy H A, Zaghloul KH and Metwally MAA. 2001. Effect of some water pollutants on the Nile tilapia, *Oreochromis niloticus* collected from the River Nile and some Egyptian lakes. *Egypt J Aqual Biol & Fish.*, **5(4):** 251 – 219.

Ensink J H and van der Hoek W. 2007. Editorial: New international guidelines for wasetwater use in agriculture. *Tropical Medicine and International Health*. **12** (5): 575-577.

FAO, Food and Agricultural organization 1983a Compilation of legal limits for hazardous substances in fish and fishery products. *Fisheries Circular* No. 764. FAO, Rome.

FAO, Food and Agricultural Organization 1983b. Manual of Analyses of metals and organochlorines in fish. FAO Fisheries Technical Paper: 212.

FAO Food and Agricultural Organization 1989. Yield and Nutritional Value of the Commercially More Important Fish Species. *FAO Fisheries Technical Paper* No. 309, Rome, Italy, 187 pp.

FAO/WHO 1989. Evaluation of certain food additives and the contaminants mercury, lead and cadmium, *WHO Technical Report, Series* No. 505.

Forstner N and Wittmann GTW 2007. **Metal Pollution in the** Aquatic Environment. Springer- Verlag, Belin.

Girard J J. 2011. Feasibility of Wastewater Reuse for Fish Production in Small Communities in a Developing World Setting. M. SC. Thesis, Univ. South Florida.

Gomez R G. 2011. Integrated fish farming strategies. World Water Day, 2011. FAO Fisheries and Aquaculture Department: http://www.fao.org/fishery/en

Hoar W S. 1957. The gonads and reproduction. In: Brown ME (Ed), **The Physiology of Fishes.** Vol. 1. Academic Press Inc. Publishers, New York.

Huss H H. 1994. Assurance of sea food quality. FAO Fisheries Tech. Paper 334, Rome, Italy.

Hynes H B N. 1950 The food of freshwater of stickleback (*Gasterosteus aculentus* and *Pygosteus pungitus*) with a review of methods used in studies of food of fish. *J Anim Ecol.*, **19**: 36-57.

IAEA 2003. International Atomic Energy Agency, Trace elements and methylmercury in Fish Tissue.; -407: 4.

Jezierska B and Witeska M. 2006. The metal uptake and accumulation in fish living in polluted waters. In: Tawardoska I, Allen HE, Haggblom MM and Stefaniak S. (Eds), Soil and Water Pollution Monitoring, Protection and Remediation. Springer, Netherlands, pp 107-114.

Jumaa S A. 1974. Fish environment relationships. M. Sc. Thesis. University of Khartoum.

Kalay M and Canli M. 2000. Elimination of essential (Cu, Zn) and nonessential (Cd, Pb) metals from tissue of a freshwater fish *Tilapia zillii* following an uptake protocol. *Tuk. J Zool.*, **24**: 429-436.

Karrar A M H. 1997. Studies on the biochemical composition of fish and current grading. M. Sc. Thesis, University of Khartoum.

King M. 1995. Fisheries Biology, Assessment and Management. Fishing News Books, Oxford, UK.

Le-Cren E D. 1951. The length-weight relationship and seasonal cycle in gonad weight and condition in perch (*Perca fluviatilis*). J Anim Ecol. ,20: 201-219.

Liew, P. K. L. 1974. Age determination of American eels based on structure of their otoliths. The Proceedings of an International Symposium on the Aging of Fish. (Edit). Bagenal, T. B. Univ. of Reading, England, 19-20 July, 1973. MAFF, 2000 Ministry of Agriculture, Fisheries and Food. Monitoring and surveillance of non-radioactive contaminants in the aquatic environment and activities regulating the disposal of wastesat sea, 1997. In Aquatic Environment Monitoring Report No. 52. Center for Environment, Fisheries and Aquaculture Science, Lowestoft, UK.

Mance G. 1987. Pollution Threat of Heavy Metals in Aquatic Environment. Elsevier, London.

Mara D and Cairncross S. 1989. Guidelines for the safe use of wastewater and excreta in agriculture and aquaculture. Measures for public health protection. World Health Organization, Geneva.

Mason C F. 1991. **Biology of Fresh Water Pollution**. Longman Singapore publishers, Singapore.

Misheloff, R. 2010. Integrated water resource management II feasibility of wastewater reuse. United States Agency for International Development (USAID). Report No. 14. 809 pp.

Mishrigi S Y. 1967. Study of Age and growth in Lates niloticus at Khartoum. *Hydrobiol.*, **30**: 45 – 56.

Mohammed G H, Mahmoud ZN and Elhag EA 1988. Chemical composition of *Mugil cephalus* of the Sudanese Red Sea Coast. *Sudan J Sci.*, **3**: 10-17.

Mubarak N M, Sahu JN, Abdulah EC and Jayakumar M. 2014. Removal of heavy metals from wastewater using carbon nanotubes. *Separation and Purification Rev.*, **43(4):** 311-338.

Nikolskii G V. 1963. **The Ecology of Fishes.** Academic Press, London, New York. 352 pp.

Nwabueze A A. 2013. Growth performance of the mudfish, *Clarias anguillaris* (Pellegrin,

1923) in Treated and Untreated Domestic Sewage. *Sustainable Agriculture Res.*, **2**(1): 62-69.

Oberdieck A and Verreth J. 2009. A handbook for sustainable aquaculture. integrated approach for a sustainable and healthy freshwater aquaculture. Sixth Framework Programme. 111 pp

Pannella G. 1974. Otolith growth patterns: an aid inage determination in temperate and tropical fishes. The Proceedings of an International Symposium on The Aging of Fish. (Edit). Bagenal, T. B. Univ. of Reading, England, 19-20 July, 1973.

Patin S A. 1982. **Pollution and the Biological Resources of the Oceans**. (English Translation). Butterworth and Comp. Ltd. England.

Pearson D. 1976. The Chemical Analysis of Foods. 7th edition, Churchill, Livingstone, Edinburgh, London and New York.

Rzoska J, Brook J and Prowse G A. 1955. Seasonal plankton development in the White Nile and the Blue Nile near Khartoum. *Verh. Int. Ver. Limnol.*,**12**: 327-337.

Sabir A A, Ahmed A A and Ali A K S. 2007. Environmental impact on biodiversity of micropopulations in three water bodies in Khartoum State, Sudan. *Egypt J Aquat Biol & Fish.*, **11(3)**: 527-543.

Saeed S M. 2007. Accumulation of heavy metals in different tissue/organs of the Nile catfish, *Clarias gariepenus* at Lake Edku, as biomarker of environmental pollution. The International Arab African Fish Resources Conf. and Exhib. 28-30 June, Cairo, Abstract 36.

Singh R, Gautam N, Mishra A, and Gupta R.2011. Heavy metals and living systems: An overview.*Indian J Pharmacol.*, **43(3):** 246–253

Strauss M. 1996. Health (Pathogen) Considerations Regarding the Use of Human Waste in Aquaculture. Water and Sanitation in Developing Countries. Talbot V. 1987. Rapid multielement analysis of oyster and cockle using XRF fluorescence and spectroscopy, with application to reconnaissance marine pollution investigations. *Sci Tot Env.*, **66**: 213-223.

Tertian R and Claisse F. 1982. Principles of Quantitative Xray Fluorescence Analysis. Hyden and Son Ltd., London, Philadelphia and Kheine.

Tidewell JH and Allan, G. L. 2001. Fish as food: aquaculture's contribution. *EMBO Rep.* **2(11):** 958–963.

Tweddle D. 1975. Age and growth of the catfish, *Bagrus meridionalis* in Southern Lake Malawi. *J Fish Biol.*, **7**: 677 – 685.

WHO 2006. Guidelines for the safe use of wastewater, excreta and greywater. Volume 3 Wastewater and excreta use in aquaculture. World Health Organization, 158 pp.

Zaki M S. 2007. Impact of heavy metals pollution on fishes. Proceedings of Egyfish (2007) Conference, Cairo, Egypt.