Clostridium: Pathogenic Roles, Industrial Uses and Medicinal Prospects of Natural Products as Ameliorative Agents against Pathogenic Species

Sabina M. Num¹ and Nicodemus M. Useh^{2, 3,*}

1Department of Veterinary Pathology and Microbiology, University of Agriculture, Makurdi,

2Department of Veterinary Pathology and Microbiology, Ahmadu Bello University, Zaria, Nigeria;

3 Department of Population Medicine and Diagnostic Sciences, College of Veterinary Medicine, Cornell University,

Ithaca, New York, United States of America

Received: November 30, 2013 Revised: January 4, 2014 Accepted: January 13, 2014

Abstract

The genus *Clostridium* is made up of species that cause disease to both human beings and animals, with zoonotic species/strains playing critical roles in disease dynamics. Clostridial organisms posses pathogenic, therapeutic and industrial uses. Pathogenic clostridia cause lethal or life threatening infections that must be treated, while the industrial clostridia produce bio-fuels, to serve as alternative energy sources in the face of very high global prices of conventional fuels. Genetic engineering is currently developing novel strains by replacing or altering the gene configuration of pathogenic strains (gene knockout/down) to convert them to solvenogenic and non pathogenic strains for industrial uses. Therapeutic clostridia serve as vehicles for treatment of diseases, especially solid tumors. Because of the global problem of antibiotic resistance, which is a survival strategy by microbial pathogens, some of the well known chemotherapeutic protocols have failed and there is the need to evolve new treatment approaches, using novel agents that have superior mechanisms of action, compared to conventional antibiotics that are becoming inconsequential because of resistance. It is believed that natural products may be effective alternatives to resolving this puzzle, since they are cheaper, readily available and possibly effective against clostridial species. In this review article, the authors discussed some of the life threatening and solvenogenic clostridia and listed some natural products that may possibly be employed as drug targets for ameliorating the problem of resistance in the future, if their active principles are thoroughly researched. It is concluded that medical and veterinary research should be re-jigged to evolve active principles from natural products that are not so easily surmounted by the surge in drug resistant strains.

Keywords: Clostridium; Industrial Uses; Medicinal Prospects; Natural Products; Ameliorative Agents; Pathogenic Species...

1. Introduction

The genus *Clostridium* consists of over 100 species, ranking second in size next to Streptomyces (Dong *et al.*, 2010). Many *Clostridium* species are known to cause disease to human beings. These include neurotoxigenic clostridia (*C. botulinum* and *C. tetani*) (Weingart *et al.*, 2010), clostridia involved in gas gangrene and necrotizing infections (*C. perfringens, C. sordellii* and *C. septicum*) (Hatheway, 1990; Aldape *et al.*, 2006), and the enteropathogenic *C. difficile* (Twine *et al.*, 2009). *Clostridium chauvoei*, which has a strong phylogenetic relationship with *C. septicum*, a human pathogen, has a long history of veterinary importance. It

was believed for a very long time that this species was exclusively a veterinary pathogen, only associated with blackleg in ruminants (Useh et al., 2006a). Recent reports of the disease in human beings have placed the pathogen on the list of very important lethal disease agents of human beings (Nagano et al., 2008; Weatherhead and Tweardy, 2011). Indeed zoonotic strains of the Clostridium spp pose a great threat to health in endemic community areas. The aforementioned notwithstanding, genetic engineering of pathogenic strains is beginning to suggest that members of the genus could also serve therapeutic purposes. In tumor managements, the use of viral vehicles in gene therapy could be successful or not, depending on the delivery systems employed. There are many tumors in

^{*} Corresponding author. e-mail: nicodemus.useh@fulbrightmail.org.

which this procedure has not been successful. Thus, there is intensive on-going research in so many expert laboratories to develop alternative management approaches (Minton, 2003). On the other hand, some *Clostridium* species are of great industrial importance. For example, *C. thermocellum* can produce ethanol from lignocellulosic waste at high temperature, while *C. acetobutylicum* and *C. beijerinckii* produce solvents (acetone, butanol, and ethanol) by utilizing a variety of substrates from monosaccharides to polysaccharides (Ezeji *et al.*, 2007; Otte *et al.*, 2009; Ezeji *et al.*, 2010; Jia *et al.*, 2011).

2. 2. Pathogenic Clostridia

2.1. Clostridium Difficile

Clostridium difficile is the most common cause of infectious diarrhoea in hospitalized patients in the industrialized world (Johnson and Gerding, 1998). With symptoms ranging from self-limited diarrhoea to life threatening fulminant colitis, C. difficile infection (CDI) has affected hundreds of thousands of patients worldwide and substantially burdens health care resources (Kyne et al., 2002). In particular, CDI has caused increased patient morbidity and mortality in hospitals throughout the world since 2003, when outbreaks with increased disease incidence and severity first emerged (Forgetta et al., 2011), and from 2004 to 2007 contributed to almost 1,000 deaths in the province of Quebec, Canada (Gilca et al., 2008 cited by Gilca et al., 2011). Investigations of these and other outbreaks across Canada, the United States, and Western Europe led to the recognition of a severe-disease associated (SDA) strain predominantly responsible for this epidemic (Kuijper et al., 2006). This strain has been classified as North American pulse field type 1 (NAP1), ribotype 027, toxinotype III, or restriction-endonuclease type BI. Outbreaks have also been associated with other SDA strains, such as the NAP7/toxinotype V/ribotype 078 (NAP7) strain found in cases of human and animal disease (Goohuis et al., 2008; Mulvey et al., 2010) and multiple toxin A-negative B-positive (A-B+) pulsotypes and ribotypes responsible for CDI outbreaks in Ireland, the United Kingdom, the United States, and Canada (Al-Barrack et al., 1999; Stabler et al., 2006). Clostridium difficile is the leading cause of health care-associated infectious diarrhoea. After exposure to C. difficile, some patients remain asymptomatic, whereas others have illness ranging from mild diarrhoea to fulminant colitis (Johnson and Gerding, 1998). Outbreaks of C. difficile infection in North America and Europe have been attributed to the emergence of an epidemic strain (North American pulsed-field gel electrophoresis [PFGE] type 1 [NAP1]) (McDonald, 2005).

Clostridium difficile is the most commonly identified bacterial cause of HIV-related and all nosocomial diarrhoea (Bartlett, 2007). Whereas outbreaks in North American and European hospitals have led to increased *C. difficile* infection (CDI) vigilance in developed countries, data on its significance for developing countries is scarce (Warny *et al.*, 2005). This information gap is significant since antibiotic preexposure, an important CDI risk factor, is high in many developing countries due to unregulated access (Sosa, 2005). Recent data suggest CDI is under-recognized in Latin America (Legaria *et al.*, 2003). Data from sub-Saharan Africa is sparse. Despite liberal, unregulated antibiotic use, developing countries do not have data about *C. difficile* infection. Nigeria, Africa's most populous country, has the third highest absolute HIV burden worldwide. Over-the-counter antibiotic access is widespread in Nigeria and nosocomial diarrhoea incidence is high (Nwokediuko *et al.*, 2002). However, patients are not tested for CDI due to presumptions of low prevalence and technical incapacity. Result from the study by this group showed significant CDI in patients with HIV infections in a 5 star Hospital in Nigeria.

C difficile has been reported as a pathogen of animals, affecting dogs, cats, horses and pigs (Weese et al., 2001a,b; Arroyo et al., 2005; Keel et al., 2007; Keessen et al., 2010, 2011). In one study, some authors reported C. difficile colonization in dogs and cats in a longitudinal manner, indicating that C. difficile is commonly albeit sporadically found in the faeces of healthy pets (Weese et al., 2010). Another study in the Netherlands showed high C. difficile prevalence in dogs, cats, horses, pigs, poultry and calves. Twenty-two different PCR ribotypes were identified and all C. difficile isolates from pigs, cattle and poultry were toxinogenic, whereas the majority of isolates from pet animals consisted of non-toxinogenic PCR ribotypes 010 and 039. Ribotype 012 was most prevalent in cattle and ribotype 078 in pigs. No predominant ribotypes were present in horse and poultry samples. Overall, PCR ribotypes 012, 014 and 078 were the most frequently recovered toxinogenic ribotypes from animal samples. Comparison with human isolates from the Dutch Reference Laboratory for C. difficile at Leiden University Medical Centre (LUMC) showed that these types were also recovered from human hospitalized patients in 2009/2010, encompassing 0.8%, 11.4% and 9.8% of all isolates, respectively. Application of multiple locus variable-number tandem-repeat analysis indicated a genotypic relation of animal and human ribotype 078 strains, but a clear genotypic distinction for ribotypes 012 and 014. Contrary to PCR ribotype 078, significant genetic differences were observed between animal and human PCR ribotype 012 and 014 isolates. The authors concluded that toxinogenic C. difficile PCR ribotypes found in animals correspond to PCR ribotypes associated with human disease in hospitalized patients in the Netherlands (Koene et al., 2011).

It is believed that *C. difficile* is the most important cause of outbreaks of neonatal diarrhoea in pig husbandry (Songer and Uzal, 2005). Infection of neonatal piglets occurs by environmental transmission (Hopman *et al.*, 2010). The bacteria have also been detected in air samples, but the role of aerial dissemination in the transmission of *C. difficile* to piglets has yet to be conclusively elucidated (Hopman *et al.*, 2010). *C. difficile* has also been reported in retail meat in Sweden (von Abacron *et al.*, 2009), broiler chickens sold at market places in Zimbabwe (Simango and Mwakurudwa, 2008), chicken in retail poultry meat

in the United States of America (Songer *et al.*, 2009; Harvey *et al.*, 2011a,b), retail vegetables in Canada (Al-Salif and Brazier, 1996; Bakri, 2009; Metcalf *et al.*, 2010) with a zero prevalence in passerine birds in Europe (Bandelj *et al.*, 2011). The meaning of all these reports is that *Clostridium* as an organism is set to colonize the entire environment to create havoc to public health.

In the light of the ongoing uncertainty about the zoonotic potential of *C. difficile* and the emergence of hypervirulent strains, research on the possible routes of transmission from animals to human beings and between animals is becoming increasingly important. There is still little knowledge on possible transmission routes from animals to human beings (Jhung *et al.*, 2008). Possible routes other than meat consumption (direct and indirect contact) with animals, have been speculated (Weese, 2010). It is increasingly recognized that aerial dissemination of bacterial elements can be the main route of zoonotic transmission (Kuske, 2006; Keessen *et al.*, 2011).

2.2. Clostridium Chauvoei

Blackleg in ruminants has been reviewed (Useh et al., 2006a). It is a fatal disease of cattle and sheep mainly (but has also been reported in deer and other animal species), caused by C. chauvoei, and was first reported in 1870 (Armstrong and McNamee, 1950). Other synonyms of the disease are: black quarter, emphysematous gangrene, symptomatic anthrax, quarter ill (Merchant and Barner, 1964), gangrenous myositis (Williams and Andrews, 1992), clostridial myositis of ruminants (Williams, 1977) and clostridial myocarditis (blackleg of the heart) (Uzal et al., 2003). C. chauvoei was named after Professor J. A. B. Chauveau, a French bacteriologist (Cato et al., 1986), and 23 reference strains of the bacterium have been described (Holderman et al., 1977). In Nigeria, Jakari, Vom and K76 strains have been incriminated as the aetiologic agents of the disease (Bagadi, 1977), while NCTC 8076 and 8361 (Heurmann et al., 1991), strain 49 (Singh et al., 1993), strain NC 08596 (Heurmann et al., 1991), strains ch.16 and ch.22, strain Tukyu and strain Awasa (Mhoma et al., 1995) have been reported to cause blackleg in UK, India and other parts of Asia, Germany, other parts of Europe and the Americas, Mozambique, Tanzania and other parts of east Africa and Ethiopia, respectively. These strains are also responsible for the disease in both the natural and accidental hosts (Radostits et al., 2000).

2.3. Clostridium Botulinum

Botulinum neurotoxins are 150-kDa endopeptidase toxins that are produced by C. *botulinum, C. butyricum* and *C. baratii* (CDC, 1998). These neurotoxins are considered the most toxic substances known (Lamanna, 1959) and have been designated as category A biological threat agents (Rotz *et al.*, 2002). Botulism, a serious neuroparalytic and sometimes fatal illness, caused by potent neurotoxins produced by the grampositive, anaerobic, spore-forming bacterium *C. botulinum*, affects both animals and human beings. Outbreaks have been reported following consumption of unsafe canned vegetables (Date *et al.*, 2011). Rodloff and Krüger (2012) reviewed the occurrence of visceral botulism in farmers. Seven types of botulinum toxins are known (A through G), of which types A, B, E, and F cause virtually all cases of human botulism (CDC, 1998). Botulism is characterized by rapidly progressive cranial neuropathy and symmetric descending flaccid paralysis, which may progress to respiratory arrest requiring mechanical ventilation and intensive supportive care in 60% of patients and clinical recovery takes several weeks to months (Shapiro *et al.*, 1998). Botulism neurotoxin derivative has been engineered to allow self activation (Masuyer *et al.*, 2011).

2.4. Clostridium Septicum

The organism is the causative agent of muscular gangrene syndrome with clinical features that overlap with those of other clostridial diseases and acute/hyperacute syndromes seen with other diseases such as anthrax (Fasanella et al., 2010). C. septicum and C. chauvoei have a close phylogenetic relationship that made science unable to distinguish the two organisms convincingly for a very long time (Nagano et al., 2008). Traditional microbiological methods are unable to distinguish the two microbial species. Molecular diagnostics, based on real time PCR with primers designed according to sporulation, 16s rRNA and triose pentose isomerase gene sequences have been reported (Lange et al., 2010; Ham et al., 2010; Garofolo et al., 2011). Soft tissues are usually affected in the pathology of the disease with an accompanying necrotizing infection (Gnerlich et al., 2011; Kiel et al., 2011).

2.5. Clostridium Perfringens

C. perfringens is a Gram-positive rod, sporeforming, anaerobic bacterium, which is highly associated with a wide array of gastrointestinal (GI) and histotoxic diseases in both humans and animals (McClane, 2007; Amimoto et al., 2007; Keyburn et al., 2008). The most common cause of C. perfringensassociated food poisoning in humans is the consumption of C. perfringens vegetative cells followed by sporulation in the gut (Paredes-Sabja and Sarker, 2009). The early steps in the development of C. perfringensassociated gas gangrene and other non-food-borne GI illnesses, such as antibiotic-associated diarrhoea, occur when C. perfringens spores ubiquitously found in the environment come in contact with the host, where spores undergo germination followed by outgrowth, cell proliferation and toxin secretion (Paredes-Sabja and Sarker, 2012).

3. Non-Pathogenic Clostridia

The biofuel producing clostridia are very important as they are nonpathogenic and produce biogenic fuels for industrial use (Köpke *et al.*, 2011). For instance, 2,3-Butanediol (2,3BD) is a commodity chemical usually produced from oil. It can be used as a precursor in the manufacture of a range of chemical products, including the solvents methyl ethyl ketone (MEK), gammabutyrolactone (GBL), and 1,3-butadiene. Commercially, the key downstream products of 2,3BD have a potential global market of around 32 million tons per annum, valued at approximately \$43 billion in sales (Xiu and Zeng, 2008). Three (3) acetogenic members of the Clostridium genus (C. autoethanogenum, C. ljungdahlii, and C. ragsdalei) have been reported to produce 2,3 BD using gases (carbon monoxide [CO] or hydrogen [H₂] and CO) as the source of carbon and energy. 2,3butanediol (2,3BD) is a high-value chemical usually produced petrochemically but can also be synthesized by some bacteria. To date, the best microbial 2,3BD production rates have been observed using pathogenic bacteria in fermentation systems that depend on sugars as the carbon and energy sources for product synthesis. Homologues of the genes involved in the requisite pathway have been identified (Köpke et al., 2011). In the report, a gene expression study demonstrated a correlation between mRNA accumulation from 2,3BD biosynthetic genes and the onset of 2,3BD production, while a broader expression study of Wood-Ljungdahl pathway genes has been shown to provide a transcription-level view of one of the oldest existing biochemical pathways. Genetic engineering is in its top form to develop new stains of clostridial organisms with non-pathogenic factors and great industrial uses (Zhang et al., 2012).

Acetone-butanone-ethanol (ABE) fermentation involves the production of these via fermentation. This is re-invigorated by the high and ever rising cost of crude and the desire for alternative energy sources to cut cost. As a result of increasing oil prices and extensive oil consumption accompanied by the potentially serious environmental impacts of oil extraction, the use of renewable biofuels as a partial replacement for traditional fossil fuels has gained recent worldwide attention. Among all of the alternatives, butanol obtained from the acetone-butanol- ethanol (ABE) process through a biological approach has become a potential replacement fuel (Yen et al., 2011; Bankar et al., 2011; Nakayama et al., 2011; Survase et al., 2011; Jones et al., 2011; Yu et al., 2011). The shortage of gasoline, even in some oil producing countries, has stimulated governments of countries like China to reawaken their biofuel programme (Dong et al., 2011). Clostridial organisms such as C. acetobutylicum, C. beijerinckii, С. saccharobutylicum, and С. saccharoperbutylacetonicum are known for producing biofuels (Jones and Kreis, 1995). Genetic manipulation of these bacteria through gene knockout/ down systems have been developed through homologues recombination methods based on replicable/ non replicable vectors (Harris et al., 2002; Tomas et al., 2004; Heap et al., 2007; Papoutsakis, 2008; Heap and Minton, 2009; Kuehne et al., 2011; Lütke-Eversloh and Bahl, 2011). Antisense RNA strategies have also been employed to manipulate the genetic code of clostridia for biofuel production (Desai and Papoutsakis, 1999; Tummala et al., 2003). C. acetobutylicum entire genome has been sequenced in an attempt to identify candidate genes for the purpose of genetic manipulation for biofuel production and other purposes (Bao et al., 2011). Biofuel tolerant mutants have been developed to enhance biofuel production by some non-pathogenic clostridia (Mao *et al.*, 2011). Apart from the usefulness of non-pathogenic clostridia in biofuel production, it is believed that their spores could serve as vehicles for systematic colonization of tumor cells in cancer therapy (Fox *et al.*, 2000; Barbe' *et al.*, 2006).

4. Clostridial Organisms as Agents of Bioterrorism

Clostridial organisms are targets for bioterrorism in our changing world where microorganisms have been used as agents of biological attack. Botulinum toxin in bioterrorism has been reviewed (Bossi et al., 2006). Aerosols of botulinum toxin could be used as a biological weapon (Franz et al., 1997; Lecour et al., 1998; Arnon et al., 2001). Deliberate release may also involve contamination of food or water supplies with toxin or C. botulinum bacteria. Botulinum toxin is extremely lethal and easy to produce. The Aum Shinrikyo cult in Japan attempted unsuccessfully to release an airborne form of botulinum toxin in Tokyo on three separate occasions in the early 1990s (Arnon et al., 2001). It is likely that several countries have developed and stockpiled botulinum toxin weapons (Arnon et al., 2001). It has been estimated that a point-source aerosol release of botulinum toxin could incapacitate or kill 10% of the population. C. botulinum is a large, Grampositive, strictly anaerobic bacillus that forms a subterminal spore. These spores can be found in soil samples and marine sediments throughout the world. Four groups of C. botulinum are described. Group I organisms are proteolytic in culture and produce toxin types A, B or F; group II organisms are nonproteolytic and produce toxin types B, E or F; group III produces toxin types C or D, and group IV toxins type G. These toxins are proteins of approximately 150 kD molecular weight and induce similar effects whether inhaled or ingested. When ingested, toxins are absorbed in the duodenum and jejunum, and enter into the bloodstream, and eventually reach peripheral cholinergic synapses. Botulinum toxin does not penetrate intact skin (Arnon et al., 2001). Toxins act by binding to the presynaptic nerve terminal at the neuromuscular junction and at cholinergic autonomic sites. This binding prevents release of acetylcholine and interrupts neurotransmission (Humeau et al., 2000). Human botulism is almost always caused by toxin types A, B, E and, in rare cases, F. Types C and D are associated with disease in birds and mammals. Type G is not associated with any disease in humans or animals. It has been estimated that, weight for weight, these toxins are the most toxic compounds known, with an estimated toxic dose, for toxin type A, of only 0.001 μ g/kg of body weight when administered intravenously, subcutaneously or intraperitoneally (Middlebrook and Franz, 1997). By inhalation, the dose that would kill 50% of exposed persons (LD50) is 0.003 μ g/kg of body weight. This toxin is 100,000 times more toxic than sarin gas (Fanz et al., 1997).

C. botulinum has been classified as a category A biological warfare agent (Johnson and Bradshaw, 2001). There are seven serologically distinct serotypes of the neurotoxin secreted by this bacterium (A–G), with types

A, B, E, and rarely F responsible for human intoxication. Botulinum neurotoxin (BoNT) A is the deadliest of the seven toxins with potency approximately one million times greater than cobra toxin, and one hundred billion times greater than cyanide. The LD50 for humans is approximately 1 ng/kg of body weight (Schantz and Johnson, 1992). Eubanks *et al.* (2007) outlined the detailed technological advancement for the detection and protection against botulism as a biological warfare agent.

C. perfringens epsilon toxin, has been classified as category B agents (http://www.niaid. nih.gov/ biodefense/bandc_priority.htmj for classification of biothreat agents) (Marks, 2004). Both C. perfringens spores and toxins have reportedly been considered as biological warfare agents (Mangold and Goldberg, 1999; Butler, 1999). Like B. anthracis spores, the spores of C. perfringens are robust and heat stable, but unlike B. anthracis spores there is no evidence that the spores can be delivered by the airborne route to cause disease. Rather, the spores have been considered as infective agents in weapons which cause traumatic injury. Flechette (shrapnel) weapons developed by Japan during World War II resulted in the delivery of spores into wounds and the subsequent establishment of gas gangrene (Mangold and Goldberg, 1999). The treatment of gas gangrene is notoriously difficult, because the infection becomes established in tissues that are deprived of blood. Evidence that the toxins produced by C. perfringens are potential biological warfare agents is still a course of dispute. However, it is believed that Iraq had a strong interest in C. perfringens toxins as biological weapons (Butler, 1999) and C. perfringens toxin is now placed on the list of CDC select agents as a toxin of particular concern. This toxin is active in a wide range of animal species and plays a central role in enterotoxaemia of sheep and lambs (Titball, 2009).

5. Epidemiology and Economic Importance of Clostridial Infections

Clostridium, a eubacteria, consists of diverse species whose economic importance includes the ability to cause disease (pathogenic strains), industrial uses and therapeutic potentials. The pathogenic strains which cause very lethal infections include *C. difficile, botulinum, perfringens, septicum, chauvoei*, etc. Clostridial infections cut across animal and human populations, with zoonotic species/ strains posing a great threat to community health in endemic areas. *C. difficile* infection is an increasing burden to the health care system, totaling more than \$1 billion/ year in the United States of America alone (Leffler and Lamont, 2009).

Life-threatening soft tissue infections caused by *Clostridium* species have been described in the medical literature for hundreds of years largely because of their fulminant nature, distinctive clinical presentations and complex management issues. *C. perfringens, C. septicum* and *C. histolyticum* are the principal causes of trauma-associated gas gangrene and their incidence increases dramatically in times of war, hurricanes,

earthquakes and other mass casualty conditions (Stevens *et al.*, 2011). Currently, tetanus occurs in about 1,000,000 people annually worldwide, and most patients are newly born in developing countries located in tropical areas. The World Health Organization (WHO) estimated that there were 715,000 deaths from tetanus among newborns in 1990. Neonatal tetanus is considered endemic in 90 developing countries and resulted in 248,000 deaths in 1997 (World Health Organization; <u>http://www.who.int/vaccine-diseases/</u>NeonatallTetanus.shtml). Tetanus continues to cause ~250,000 deaths worldwide each year, predominantly in low- and middle income countries (Alam *et al.*, 2008).

The life threatening consequences of clostridial organisms have been reviewed (Stevens et al., 2011). During the civil war, 50% of soldiers who were wounded ultimately died. If the soldier was not killed outright, the extensive destruction of tissue inflicted by the 45 caliber lead bullets used at the time predisposed him to the development of gas gangrene, largely due to the resultant vascular damage. Civil war surgeons had few resources such as anaesthesia, intravenous fluids and antibiotics and thus "prophylactic amputation" became common place. During World War I, medical treatments had improved. However, weaponry also became more sophisticated. In this period, the incidence of gas gangrene was as high as 10% of wounded soldiers (MacLennan, 1962). Gas gangrene also claimed an estimated 100,000 German soldiers during this period.

In World War II, medical evacuation, improved surgical debridement techniques in field hospitals, gas gangrene anti-toxin and antibiotics were available and reduced the overall incidence of gas gangrene. For example, death from gas gangrene in US casualties in the western front of Europe was 8 cases/1000 wounded, for the Free French Forces it was 12.3/1000 and among prisoners of war, 51.9/1000 wounded. The much higher rate among prisoners was attributed to a delay in definitive surgical treatment which was 1 day for US casualties and 3.5 days for prisoners (MacLennan, 1962). In addition, the incidence of gas gangrene was higher in European theaters than in desert operations, likely because the fertile valleys of Europe were heavily contaminated with C. perfringens (50,000 per gram of soil), whereas they were rarely recovered in the Sahara Desert. Uniforms too were found to be contaminated with enteric clostridia and C. perfringens was isolated from 24% of shirts and 44% of trousers (McLennan, 1943a,b,c).

In the Korean, Vietnam, Gulf and Afghanistan wars, rapid evacuation, thorough (but not radical) surgical debridement, vascular reconstruction, improved supportive measures and greater antibiotic efficacy and availability greatly reduced the incidence of *C. perfringens* gas gangrene. Gas gangrene continues to be problematic in both military theaters and the civilian sectors, always associated with traumatic injuries. For instance, among nearly 2800 patients hospitalized following the 2008 Wenchuan earthquake in China, over 2.4% developed gas gangrene (Chen *et al.*, 2011). Interestingly, *C. septicum* as a cause of non-traumatic gas gangrene has increased in civilian populations in recent years and necrotizing infections associated with skin-popping black tar heroin have been attributed to *C. perfringens, C. novyi* and *C. sordellii. C. sordellii* has been reported not only in trauma cases but also associated with pregnant females undergoing normal vaginal delivery, caesarian sections or medical abortions. *C. difficile*, the causative agent of antimicrobial- associated diarrhoea and pseudomembranous colitis, is currently one of the most important nosocomial pathogens because of its zoonotic nature (Rupnik et al., 2009). Nosocomial infections with the Gram-positive pathogen *C. difficile* pose a major risk for hospitalized patients and result in significant costs to health care systems.

Blackleg is known to be an endemic disease in both developed and developing countries of the world and is a well-known cause of financial loss to cattle raisers in many parts of the world (Adams, 1998). The economic losses of ruminants to the disease have yet to be quantified in most countries, but in Nigeria, losses of Zebu cattle to the disease have been estimated at US\$4.3 million annually (Useh *et al.*, 2006b). In United States of America, Latin America, India and other parts of Asia and Europe, the economic losses of ruminants to blackleg have yet to be estimated, but it has been reported that the disease causes major economic losses in cattle and minor losses in sheep (Cottral, 1978; Ramarao and Rao, 1990; Troxel *et al.*, 1997; Flyod, 1994; Adams, 1998).

6. Management, Prevention and Control of Infections by Pathogenic Clostridia

Resistance of pathogenic clostridial organisms to therapeutic antimicrobials has been reported and is increasingly becoming an issue of concern to public health (Gamboa-Coronado et al., 2011). The resistance means that alternative therapeutic protocols will have to be developed in the future to treat resistant strains. Management of C. difficile infection in human beings has been reviewed (Leffler and Lamont, 2009). Drugs like metronidazole, (oral administration), vancomycin and several stand-alone therapies or as adjunct agents have been used. Alternative treatments include the use of rifaximin (Johnson et al., 2007; Garey et al., 2008). Immunotherapy (Kyne et al., 2001), microbiologic therapy (Seal et al., 1987) and probiotic therapy (McFarland, 2006) have also been carried out. Prevention and control is better achieved by appropriate use of antibiotics, especially broad spectrum agents and those shown to have a particular predisposition for C. difficile infection. Treatment of clostridial infections caused by the highly toxigenic strains involves neutralization of the toxins.

The drug of choice for treatment of blackleg is procaine penicillin (10 000 IU/kg) over a 3–5 day therapy period. It is generally advocated that the administration of crystalline penicillins intravenously should precede the long-acting preparations that should be administered directly in the affected tissues (Radostits *et al.*, 2000). The best preventive and control strategy is vaccination.

7. Natural Products as Potential Therapeutic Agents Against Clostridial Infections

Globally, millions of people in the developing world rely on medicinal plants for primary health care, income generation and livelihood improvement (WHO, 2002). The active substances present in many medicinal plants could be used as therapeutic alternatives against clostridial infections (Woodford and Livermore, 2009). Although the mechanisms of pharmacological actions of most medicinal plants with potentials to ameliorate clostridial infections have yet to be defined, there are strong indications that the results of most *in vitro* studies that seem to point to some beneficial actions of these plants is worth further investigations after all. Table 1 shows a list of medicinal plants from different parts of the world with potential therapeutic properties against clostridial infections.

C. perfringens is classified into five serotypes (A, B, C, D and E) based on the type of enterotoxins it produces. Delta-toxin is one of the five haemolysins released by Clostridium perfringens which play a most important role in gas gangrene and gastroenteritis. The native structure of the delta-toxin is not yet reported in the structural databases (Skariyachan et al., 2011). C. perfringens has been shown to develop multiple drug resistance, indicating that the treatment for this bacterium is quite challenging. There is need, therefore, to employ the use of alternative therapeutic agents. The rational design of improved therapeutics requires the crystal structure for the toxin. However, the structure for the toxin is not yet available in its native form. In a recent study, some authors modeled the toxin structure using a- haemolysin of Staphylococcus aureus (PDB: 3M4D chain A) as template. The docking of the toxin with the herbal extract curcumin (1,7-bis (4-hydroxy-3methoxyphenyl) hepta-1,6-diene-3,5-dione) showed a binding energy of -8.6 Kcal/mol, in comparison to the known antibiotic linezolid with binding energy of -6.1 Kcal/mol. This data is believed to be relevant in the future design and development of novel compounds against the delta-toxin produced by C. perfringens.

Natural product	possible clostridial target	Reference
Combretum fragrans	C. chauvoei	Useh et al. (2004)
Aristolochia paucinervis	C. perfringens/ difficile	Gadhi et al. (1999)
Melia toosendan	C. botulinum	Shi & Li (2007);Nakai et al. (2009)
Propolis (bee glue)	Clostridium perfringens	Boyanova et al. (2006)
Clostridium difficile		"
Clostridium tertium		"
Other Clostridium species		"
Phyllanthus muellerianus	C. sporogenes	Brusotti et al. (2011)
Tamarindus indicus	C. chauvoei	Useh et al. (2004)
Galla Rhois	Clostridium perfringens, C. paraputrif	<i>icum</i> Ahn <i>et al.</i> (1998)
Glycyrrhiza glabra linn	Clostridium spp	Saxena (2005)
Ghee & cocoanut	C. tetanii	Hlady et al. (1992; Bennett et al. (1997)
Teas & tea flavonoids	C. perfringens	Friedman, 2007
Korean dung beetle	C. difficile	Kang et al. (2011)
Clematis hirsute	C. chauvoei	Yineger et al. (2007)
Rumex nepalensis	C. chauvoei	"
Leonotis ocymifolia	Anthrax bacillus	"
Cucumis ficifolius	C. chauvoei	"
Nigella sativa L.	C.chauvoei	"
Discopodium		
eremanthum Chiov.	C. chauvoei	"
Sonchus bipontini	C. chauvoei	"
Crepis rueppellii	C. chauvoei	"
Nicotiana tabaccum L.	C. chauvoei	"
Ruta chalepensis L	"	"
Sida schimperiana.	"	"
Sonchus bipontini	"	"
Cymbopogon citrates	"	"
Alchemilla abyssinica	"	"
Salvia nilotica	"	"
Vernonia myrantha	"	"
Senecio fresenii	"	"
Curcumin (from Curcuma	C. perfringens δ- toxin	Skariyachan et al. (2011)
<i>longa</i>) docked with α -toxin		
of S. aureus		
Tea phenolics	C. difficile	Lee et al. (2006)
Thearubigin	C. botulinum	Satoh <i>et al.</i> (2001)

Table 1. List of some natural products with ethnopharmacological potentials against clostridial infections

It has been reported that tea catechins strongly inhibit the growth of *C. perfringens in vitro* (Hara *et al.*, 1989; Ahn *et al.*, 1991; Hara *et al.*, 1995). Green tea catechins also reduced the heat-resistance characteristics of *C. hermoaceticum*, which proliferate in vending machines, causing sour spoilage in milk and other drinks (Sakanaka *et al.*, 2000). Some other unpublished data (cited by Friedman, 2007) also suggest that concentrated green tea extracts inhibit sporulation and growth of *C. perfringens* in ground meat and turkey products during chilling. These observations suggest that the use of polyphenols or other natural antimicrobials may result in reduction of temperatures used in thermal processing of foods.

Coprisin, an antibacterial peptide has been isolated from *Copris tripartitus*, a Korean dung beetle, and a nineamino-acid peptide identified in its α -helical region (Hwang *et al.*, 2009). Kang *et al.* (2011) investigated the clinical significance of treatment with a coprisin analogue (a disulfide dimer of the nine peptides) on inflammation and mucosal damage in a mouse model of acute gut inflammation, established by administration of antibiotics followed by *C. difficile* infection. Coprisin treatment significantly ameliorated decreases in body weight, improved survival rate, decreased mucosal damage and pro-inflammatory cytokine production. In contrast, the coprisin analogue had no apparent antibiotic activity against commensal bacteria, including Lactobacillus and Bifidobacterium spp, which are known to inhibit the colonization of the gut by C. difficile. The exposure of C. difficile to the coprisin analogue caused a marked increase in nuclear propidium iodide (PI) staining, indicating membrane damage. The staining levels were similar to those seen with bacteria treated with a positive control for membrane disruption (EDTA). In contrast, coprisin analogue treatment did not trigger increases in the nuclear PI staining of Bifidobacterium thermophilum. This observation suggests that the antibiotic activity of the coprisin analogue may occur through specific membrane disruption of C. difficile. Thus, these results indicate that the coprisin analogue may prove useful as a therapeutic agent for C. difficile -associated inflammatory diarrhoea and pseudo-membranous colitis.

Chinese traditional medicine has a long history which dates back thousands of years ago. To date, Chinese people still believe in traditional medicine and herbal medications are very popular in Chinese societies around the world. The latest encyclopedia of Chinese traditional medicine lists over 5500 natural sources (82.8% of which are plants) that form the basis for the 100,000-500,000 prescriptions of Chinese traditional medicine (Tang and Eisenbrand, 1992; Zhu and Woerdenbag, 1995). However, Chinese traditional medicine is unacceptable to Western society partly because the bases of Chinese traditional medicine like the yin-yang theory and five element theory (Cheng, 2000) are difficult to understand. Chinese herbal medicine has shown that the fruit and bark of plants belonging to the family Melia could be used as digestive tract-parasiticide and agricultural insecticide. Toosendanin (TSN, C30H38O11, FW=574), triterpenoid derivative, was extracted from the bark of Melia toosendan Sieb. et Zucc. by Chinese scientists in 1950's and used as an ascarifuge in China instead of imported sendanin. Studies have demonstrated that TSN possesses special biological actions and perhaps considerable pharmacological values in scientific research, clinical medicine and agriculture. The first is that by interfering with neurotransmitter release by causing an initial facilitation, TSN eventually blocks synaptic transmission at both the neuromuscular junction and central synapses. The action might result from TSNinduced Ca²⁺-sensitivity change and final elimination of transmitter release machinery. The second is that despite sharing many similar actions with botulinum neurotoxin (BoNT) on blocking neuromuscular transmission, TSN has a markedly anti-botulismic action in vivo and in vitro: TSN-treatment is reported to prevent death in mice and monkeys with botulism. TSN-incubation in vitro or TSNinjection in vivo endows neuromuscular junction with a high tolerance to BoNT. Studies suggest that the antibotulismic action is achieved by preventing BoNT from approaching its enzymatic substrate, SNARE protein (Shi and Li, 2007).

Other studies on the role of TSN as anti-botulism natural product have been reported (Nakai et al., 2009).

Experiments carried out on phrenic nerve-diaphragm preparations of rats showed that the neuromuscular blocking action of TSN was similar to that of botulinum neurotoxin (BoNT) in many respects (Shi et al., 1980, 1981a,b, 1982). For example, they were both concentration- and temperature dependent; and the temperature coefficients were both high. Also, their dosage-response relationships were similar and the blocking effects were both not only irreversible but also related to the nerve activity and concentration of Ca²⁺ There was always a delay between drug application and transmission block. Even if the drugs were washed out during the latent period, the blockade still occurred, indicating that both TSN and BoNT bound to their binding sites very fast. Similar to BoNT, TSN eventually blocked neurotransmitter release by abolishing Ca²⁺sensitivity of the transmitter release machinery. Moreover, some drugs, like guanidine and 4aminopyridine, which facilitates neurotransmitter release and antagonize botulism, also have anti-TSN effect (Shi et al., 1981a). Except for sharing these similarities, the action of TSN is different from that of BoNT in some respects. For example, TSN changed excitability and fine structure of nerve endings and affected neurotransmitter release at central synapses as it did at the neuromuscular junction (Huang et al., 1996; Shi and Chen, 1999; Chen et al., 1999; Xu et al., 2004). The most prominent difference was that TSN always facilitated neurotransmitter release before blocking it. During the facilitatory phase, both the transmitter release (either spontaneous or evoked), and the Ca²⁺- sensitivity of transmitter release machinery were enhanced (Shi et al., 1981a, 1982; Xu et al., 2004). The similarities between the actions of TSN and BoNT suggest that TSN and BoNT may act at an adjacent site by a similar approach. However, analysis of their differences may give us a clue with which to reveal the antibotulismic mechanism of TSN.

C. chauvoei, which is a very important veterinary pathogen is known to produce sialidase (neuraminidase), an enzyme reported to facilitate its spread in host tissues (Useh et al., 2004a; Useh et al., 2006c). Many herbal remedies have been reported to ameliorate blackleg, caused by C. chauvoei, but the rational basis for this has yet to be demonstrated in the literature (Yineger et al., 2007). The Fulani pastoralists of rural Nigeria, who own livestock resources in the country, prefer the use of herbal remedies to treat animal diseases, including blackleg, because they are more natural, cheaper and safer (Jagun et al., 1996; Gammaniel, 2000). Tamarindus indicus and Combretum fragrans are two common herbal remedies used recurrently by the nomads of rural Nigeria to treat blackleg (Abdu et al., 2000). A study by some Nigerian authors to investigate the possible role of the herbs on blackleg showed that methanolic extracts of the stem barks of these plants inhibited sialidase (neuraminidase) activity in vitro in a dose-dependent manner. The authors therefore suspected that neuraminidase inhibition, leading to inability of the bacteria to spread in tissues, could be a major mechanism by which the herbs ameliorated the disease. The results revealed a target of action by the medicinal plants, thus corroborating their use in traditional medicine practices in Nigeria (Useh et al.,

2004b). Sialidases (neuraminidases) have been implicated in the pathologies of several diseases, including cell invasion in Chaga's disease, anaemia in trypanosomosis and viral invasion in Newcastle disease. The foregoing, according to the authors, makes the sourcing of neuraminidase inhibitors mandatory for treatment and amelioration of clinical symptoms related to the physiological activity of the enzyme. In most cases, the inhibitors are synthetic and indeed costly, e.g. Zanamivir used in the treatment of influenza virus infection. Moreover, some of the synthetic inhibitors become less effective on account of mutations, which are rampant in sialidases. This makes recourse to plants as source of neuraminidase inhibitors an appealing alternative, because their active compounds are synthesized in direct response to bacterial and viral invasion. In the said study, the two medicinal plants were tested, because of their reported role in ameliorating blackleg in traditional veterinary practice. These authors concluded that the active principles of these medicinal plants should be further characterized, to exploit the findings in drug development.

8. Conclusion

The future use of natural products in the clinical management of clostridial infections, although laced with a lot of potentials, is a puzzle that research should be intensified to resolve, considering the economic importance, with regards to the pathogenic, industrial and potential therapeutic uses of members of this genus. The emergence of drug resistant strains, with enhanced pathogenicity is deadly and requires concerted efforts to pull through this quagmire. Medical and veterinary research should be re-jigged to evolve active principles that are not so easily rendered inconsequential by the surge in drug resistant strains. Future research should focus on the possibility of targeting the critical virulence factors of clostridial agents using active principles from herbs, which have proved to be useful and potential therapeutic agents over time.

Acknowledgements

A United States Senior Fulbright Research Award to NM Useh at the Department of Population Medicine and Diagnostic Sciences, College of Veterinary Medicine, Cornell University, Ithaca, New York, where the manuscript was prepared is gratefully acknowledged. NM Useh also expresses appreciation to German Research Foundation (DFG), Bonn, Germany, The Academy of Sciences for the developing world (TWAS), Trieste, Italy and Deutscher Akademischer Austausch Dienst (DAAD), Bonn, Germany for generous financial support via clostridial research grants at various times.

References

Abdu PA, Jagun AG, Gefu JO, Mohammed AK, Alawa CBI and Omokanye AT. 2000. Ethnoveterinary practices, research and development. In: Proceedings of National Animal Production Research Institute (NAPRI), Ahmadu Bello University Zaria, Nigeria, 163. Adams LG. 1998. Animal health issues in South Texas cattle. Workshop on beef cattle production systems and natural resources conservation in semi-arid lands of South Texas and Northern Mexico held at Universidad Auto'noma dr Tamaulipas, Cd. Victoria, Tamaulipas, Mexico, 26–27. February 1998.

Ahn Y- J, Kawamura T, Kim M, Yamamoto T and Mitsuoka T. 1991. Tea polyphenols: selective growth inhibitors of *Clostridium* spp. *Agric Biol Chem.*, **55**: 1425-1426.

Ahn Y-J, Lee C -O, Kweon J-H, Ahn V and Park J-H. 1998. Growth-inhibitory effects of Galla Rhois-derived tannins on intestinal bacteria. *J Appl Microbiol.*, **84**: 439-443.

Alam SI, Bansod S and Singh L. 2008. Immunization against *Clostridium perfringens* cells elicits protection against *Clostridium tetani* in mouse model: identification of cross-reactive proteins using proteomic methodologies. *BMC Microbiol.*, **8**: 194.

Al-Barrak A, Embil J, Dyck B, Olekson K, Nicoll D, Alfa M and Kabani A. 1999. An outbreak of toxin A negative, toxin B positive *Clostridium difficile*-associated diarrhoea in a Canadian tertiary-care hospital. *Canadian Comm Dis Rep.*, **25**: 65-69.

Aldape M J, Bryant A E and Stevens D L. 2006. *Clostridium sordellii* infection: epidemiology, clinical findings, and current perspectives on diagnosis and treatment. *Clin Infec Dis.*, **43**: 1436-1446.

Al- Saif N and Brazier JS. 1996. The distribution of *Clostridium difficile* in the environment of South Wales. *J Med Microbiol.*, **45**: 133-137.

Amimoto K, Noro T, Oishi E and Shimizu M. 2007. A novel toxin homologous to large clostridial cytotoxins found in culture supernatant of *Clostridium perfringens* type C. *Microbiol.*, **153**: 1198-1206.

Armstrong H L and MacNamee J K. 1950. Blackleg in deer. J Amer Vet Med Assoc., 117: 212-214.

Arnon, S S, Schechter R, Inglesby T V, Henderson D A, Bartlett J G, Ascher M S, Eitzen E, Fine A D, Hauer J, Layton M, Lillibridge S, Osterholm M T, O'Toole T, Parker G, Perl T M, Russell P K, DSwerdlow D L, Tonat K and Working Group on Civilian Biodefense. 2001. Botulinum toxin as a biological weapon. *J Amer Med Assoc.*, **285**: 1059-1070.

Arroyo L G, Kruth S A, Willey B M, Staempfli H R, Low D E and Weese J S. 2005. PCR ribotyping of *Clostridium difficile* isolates originating from human and animal sources. *J Med Microbiol.*, **54**: 163-166.

Bagadi H O. 1977. The production and counting of spores of *Clostridium chauvoei*. Appl Environ Microbiol., **33**:1287-1288.

Bakri M. 2009. *Clostridium difficile* in ready to eat salads, Scotland. *Emerg Infect Dis.*, **15**: 817-818.

Bandelj P, Trilar T, Racnik J, Zadravec M, Pirs T, Avbersek J, Micunovic J, Ocepek M and Vengust M. 2011. Zero prevalence of *Clostridium difficile* in wild passerine birds in Europe. *FEMS Microbiol Lett.*, **321**, 183-185.

Bankar S B, Survase S A, Singha R S and Granström T. 2011. Continuous two stage acetone-butanol ethanol fermentation with integrated solvent removal using *Clostridium acetobutylicum* B 5313. *Biores Technol*. doi:10.1016/j.biortech.2011.12.

Bao G, Wang R, Zhu Y, Dong H, Mao S, Zhang Y, Chen Z, Li Y and Ma Y. 2011. Complete genome sequence of *Clostridium acetobutylicum* DSM 1731, a solvent-producing strain with multireplicon genome architecture. *J Bacteriol.*, **193**: 5007-5008.

Barbe[´] S S, van Mellaert L and Anne J. 2006. The use of clostridial spores for cancer treatment. *J Appl Microbiol.*, **101**: 571-578.

Bartlett J G. 2007. Changing trends in bacterial infections: *Staphylococcus aureus*, bacterial pneumonia, *Clostridium difficile*. *Top HIV Med.*, **15**: 94-98.

Bennett J, Macia J, Traverso H, Banoagha S, Malooly C and Boring J. 1997. Protective effects of topical antimicrobials against neonatal tetanus. *Int J Epidemiol.*, **26**: 897-903.

Bossi P, Garin D, Guihot A, Gay F, Crance J-M, Debord T, Autran B and Bricaire F. 2006. Bioterrorism: management of major biological agents. *Cell Mol Life Sci.*, **63**: 2196-2212.

Boyanovaa L, Kolarovb R, Gergovaa G and Mitov I. 2006. *In vitro* activity of Bulgarian propolis against 94 clinical isolates of anaerobic bacteria. *Anaerobe*, **12**: 173177.

Brusottia G, Cesaria I, Frassàa G, Grisoli P, Dacarroa C and Caccialanza G. 2011. Antimicrobial properties of stem bark extracts from *Phyllanthus muellerianus* (Kuntze) Excell. *J Ethnopharmacol.*, **135**: 797-800.

Butler R. 1999. UNSCOM's Comprehensive Review. Available at: http://cns.miis.edu/iraq/ ucreport/.

Cato E P, George W L and Finegold S M. 1986. Genus *Clostridium*. In: Sneath PHA (Ed),. **Bergy's Manual of Systematic Bacteriology**. Volume 2. William and Wilkins, Baltimore, MD; pp. 1141-1182.

Centers for Disease Control and Prevention (CDC): Botulism in the United States, 1899-1996, handbook for epidemiologists, clinicians, and laboratory workers Atlanta, GA: CDC; 1998.

Chen E, Deng L, Liu Z, Zhu X, Chen X and Tang H. 2011. Management of gas gangrene in Wenchuan earthquake victims. J Huazhong Uni Sci Technol Med Sci., **31**: 83e7.

Chen W Y, Yin P B, Ye W L and Shi Y L. 1999. Toosendanininduced change of dopamine level detected by microdialysis *in vivo* at rat striatum. *Chinese Sci Bull.*, **44**: 1106-1110.

Cheng J T. 2000. Review: drug therapy in Chinese traditional medicine. *J Clin Pharmacol.*, 40: 445-450.

Cottral G E. 1978. Clostridia. Manual of Standard Methods for Veterinary Microbiology. 1st ed. Cornell University Press, Ithaca, NY; p. 510–25.

Date K, Fagan R, Crossland S, Maceachern D, Pyper B, Bokanyi R, Houze Y, Andress E and Tauxe R. 2011. Three outbreaks of foodborne botulism caused by unsafe home canning of vegetables-Ohio and Washington, 2008 and 2009. *J Food Prot.*, **74(12)**: 2090-2096.

Desai R and Papoutsakis E. 1999. Antisense RNA strategies for metabolic engineering of *Clostridium acetobutylicum*. *Appl Environ Microbiol.*, **65**: 936-945.

Dong H, Tao W, Dai Z, Yang L, Gong F, Zhang Y and Li Y. 2011. **Biobutanol. Advances in Biochemical Engineering/Biotechnology** DOI: 10.1007/10_2011_128. Springer-Verlag Berlin Heidelberg 2011.

Dong H, Zhang Y, Dai Z and Li Y. 2010. Engineering *Clostridium* strain to accept unmethylated *DNA*. *PLoS One*, **5**: e9038.

Eubanks L M, Dickerson T J and Janda K D. 2007. Technological advancements for the detection of and protection against biological and chemical warfare agents. *Chem Soc Rev.*, **36**: 458-470.

Ezeji T, Milne C, Price N D and Blaschek H P. 2010. Achievements and perspectives to overcome the poor solvent resistance in acetone and butanol-producing microorganisms. *Appl Microbiol Biotechnol.*, **85**: 1697-1712. Ezeji T C, Qureshi N and Blaschek H P. 2007. Bioproduction of butanol from biomass: from genes to bioreactors. *Curr Opin Biotechnol.*, **18**: 220-227.

Fasanella A, Galante D, Garofolo G and Jones M H. 2010. Anthrax undervalued zoonosis. *Vet Microbiol.*, **140**: 318-331.

Floyd J G. 1994. Blackleg and other clostridial diseases in cattle. Extension Veterinarian, ANR-888, A Publication of the Alabama Cooperative Extension System; 3 pp.

Forgetta V, Oughton MT, Marquisi P, Brukner I, Blanchette R, Haub K, Magrini V, Mardis ER, Gerding DN, Loo VG, Miller MA, Mulvey MR, Rupnik M, Dascal A and Dewar K. 2011. Fourteen-genome comparison identifies DNA markers for severe-disease-associated strains of *Clostridium difficile*. J Clin Microbiol., **49**: 2230-2238.

Fox M E, Lemmon M J, Giaccia A J, Minton N P and Brown J M. 2000. Genetically modified *Clostridium* for gene therapy of tumors. *Meth Mol Med.*, **35**: 413-418.

Franz D R, Jahrling P B, Friedlander A M, McClain D J, Hoover D L, Bryne W R, Pavlin J A, Christopher G W and Eitzen E M Jr. 1997. Clinical recognition and management of patients exposed to biological warfare agents. *J Amer Med Assoc.*, **278**: 399-411.

Friedman M. 2007. Overview of antibacterial, antitoxin, antiviral, and antifungal activities of tea flavonoids and teas. *Mol Nutr Food Res.*, **51**: 116-134.

Gadhi C A, Weber M, Mory F, Benharref A, Lion V, Jana V and Lozniewski A. 1999. Antibacterial activity of *Aristolochia paucineris* Pomel. *J Ethnopharmacol.*, **67**: 87-92.

Gamboa-Coronado M M, Mau-Inchaustegui S and Rodríguez-Cavallini E. 2011. Molecular characterization and antimicrobial resistance of *Clostridium perfringens* isolates of different origins from Costa Rica. *Rev Biologie Tropica*, **59**:1479-85.

Gammaniel K S. 2000. Toxicity from medicinal plants and their products. *Nig J Prod Med.*, **4**: 4-8.

Garey K W, Salazar M, Shah D, Rodrigue R and DuPont H L. 2008. Rifamycin antibiotics for treatment of *Clostridium difficile*-associated diarrhoea. *Ann Pharmacother.*, **42**: 827-835.

Garofolo G, Galante D, Serrecchia L, Buonavoglia D and Fasanella A. 2011. Development of a real time PCR taqman assay based on the TPI gene for simultaneous identification of *Clostridium chauvoei* and *Clostridium septicum*. J Microbiol Meth., **84**: 307-311.

Gilca R, Fortin E, Frenette C, Longtin Y and Gourdeau M. 2012. Seasonal variations in *Clostridium difficile* infections are associated with influenza and respiratory syncytial virus activity independently of antibiotic prescriptions: a time-series analysis in Quebec, Canada. *Antimicrobial Agents Chemother.*,**56(2):** 639-646

Gnerlich J L, Ritter J H, Kirby J P and Mazuski J E. 2011. Simultaneous necrotizing soft tissue infection and colonic necrosis caused by *Clostridium septicum*. *Surg Infect.*, **12**: 501-506.

Goohuis A, Bakker D, Corver J and Debast S B. 2008. Emergence of *Clostridium difficile* infection due to a new hypervirulent strain, polymerase chain reaction Ribotype 078. *Clin Infect Dis.*, **47**: 1162-1170.

Halm A, Wagner M, Köfer J and Hein I. 2010. Novel real-time PCR assay for simultaneous detection and differentiation of *Clostridium chauvoei* and *Clostridium septicum* in clostridial myonecrosis. *J Clin Microbiol.*, **48**: 1093-1098.

Hara Y, Luo S J, Wickremasinghe R L and Yamanishi T. 1995. Use and benefits of tea. *Food Rev Int.*, **11**: 527-542.

Hara Y, Watanabe M and Sakaguchi G. 1989. The fate of *Clostridium botulinum* spores inoculated into tea drinks. *Shokuhin Kogyo Gakkaishi*, **36**: 375-379.

Harris, L M, Welker N E and Papoutsakis E T. 2002. Northern, morphological, and fermentation analysis of *spo0A* inactivation and overexpression in *Clostridium acetobutylicum* ATCC 824. *J Bacteriol.*, **184**: 3586-3597.

Harvey R B, Norman K N, Andrews K, Hume M E, Scanlan C M, Callaway T R, Anderson R C, and Nisbet D J. 2011a. *Clostridium difficile* in poultry and poultry meat. *Roger Food borne Pathogens and Dis.*, **8**: 1321-1323.

Harvey R B, Norman K N, Andrews K, Norby B, Hume M E, Scanlan C M, Hardin M D and Scott H M. 2011b. *Clostridium difficile* in retail meat and processing plants in Texas. *J Vet Diag Invest.*, **23**: 807-811.

Hatheway C L. 1990. Toxigenic clostridia. *Clin Microbiol Rev.*, **3**: 66-98.

Heap J T, Pennington O J, Cartman S T, Carter G P and Minton N P. 2007. The ClosTron: a universal gene knock-out system for the genus *Clostridium. J Microbiol Meth.*, **70**: 452-464.

Heap J T and Minton N P: Methods, 2009. International patent WO 2009/ 101400.

Heuermann D, Roggentin P, Kleinneidam R G and Schauer R. 1991. Purification and characterization of a sialidase from *Clostridium chauvoei* NC08596. *Glycoconj J.*, **8**: 95-101.

Hlady W G, Bennette J V, Samadi A R, Begum J, Hafez A, Tarafdar AI and Boring J R. 1992. Neonatal tetanus in rural Bangladesh: risk factors and toxoid efficacy. *Amer J Public Hlth.*, **82**: 1365-1369.

Holderman L V, Cato E P and More W E C. 1977. **Anaerobe Laboratory Manual**. 4th ed. Anaerobe Laboratory Virginia Polytechnic Institute and State University, Blacksburg, VA; pp. 1-156.

Hopman N E, Keessen E C, Harmanus C, Sanders I M, vanLeengoed L A, Kuijper E J and Lipman L J. 2011. Acquisition of *Clostridium difficile* by piglets. *Vet Microbiol.*, **149(1-2):** 186-192.

Huang H Y, Zhou C W and Shi Y L. 1996. Toosendanin facilitates [3H] noradrenaline release from rat hippocampal slices. *Nat Toxins*, **4**: 92-95.

Humeau Y, Doussau F, Grant N and Poulain B. 2000. How botulinum and tetanus neurotoxins block neurotransmitter release. *Biochimie.*, **82**: 427-446.

Hwang J-S, Lee J, Kim Y J, Bang H S, Yun E Y, Kim S R, Suh H J, Kang B R, Nam S H, Jeon J P, Kim I and Lee D G. 2009. Isolation and characterization of a defensin-like peptide (corprisin) from the dung beetle *Corpris tripartitus*. *Int J Peptides* doi:10.1155/2009/136284.

Jagun A G, Abdu P A, Mohammed A K, Alawa C B I, Omokanye A K, 1996. Survey of ethnoveterinary practices in Nigeria. Technical progress report on the survey conducted on ethnoveterinary practices in Nigeria, sponsored by International Development Research Centre (IDRC), Canada. pp 77-79.

Jhung M A, Thompson A D, Killgore G E, Zukowski W E, Songer G, Warny M, Johnson S, Gerding D N, McDonald L C and Limbago B M. 2008. Toxinotype V *Clostridium difficile* in humans and food animals. *Emerg Infect Dis.*, **14**: 1039-1045.

Jia K, Zhu Y, Zhang Y and Li Y. 2011. Group II intron-anchored gene deletion in *Clostridium*. *Plos One*, **6**: e16693.

Johnson E A and Bradshaw M. 2001. *Clostridium botulinum* and its neurotoxins: a metabolic and cellular perspective. *Toxicon*, **39**: 1703-1722.

Johnson S and Gerding D N. 1998. *Clostridium difficile*associated diarrhoea. *Clin Infect Dis.*, **26**: 1027-1034.

Johnson S, Schriever C, Galang M, Kelly C P and Gerding D N. 2007. Interruption of recurrent *Clostridium difficile*-associated diarrhoea episodes by serial therapy with vancomycin and rifaximin. *Clin Infect Dis.*, **44**: 846-848.

Jones D T and Kreis S. 1995. Origins and relationships of industrial solvent-producing clostridial strains. *FEMS Microbiol Rev.*, **17**: 223-232.

Jones S W, Tracy B P, Gaida S M and Papoutsakis E T. 2011. Inactivation of δ^{G} F in *Clostridium acetobutylicum* ATCC 824 blocks sporulation prior to asymmetric division and abolishes δ^{E} and δ^{G} protein expression but does not block solvent formation. *J Bacteriol.*, **193**: 2429-2440.

Kang J K, Hwang J S, Nam H J, Ahn K J, Seok H, Kim S-K, Yun E Y, Pothoulakis C, Lamont J T and Kim V. 2011. Mechanisms of action: physiological effects of the insect peptide coprisin prevents *Clostridium difficile*-mediated acute inflammation and mucosal damage through selective antimicrobial activity. *Antimicrobial Agents Chemother.*, **55**: 4850-4857.

Keel K, Brazier J, Post K, Weese S and Songer, J. 2007. Prevalence of PCR ribotypes among *Clostridium difficile* isolates from pigs, calves, and other species. *J Clin Microbiol.*, **45**: 1963-1964.

Keessen E C, Donswijk C J, Hol S P, Hermanus C, Kuijper E J and Lipman L J A. 2011. Aerial dissemination of *Clostridium difficile* on a pig farm and its environment. *Environ Res.*, **111**: 1027-1032.

Keessen E C, Leengoed L A, Bakker D, vandenBrink K M, Kuijper E J and Lipman L J. 2010. Prevalence of *Clostridium difficile* in swine thought to have *Clostridium difficile* infections (CDI) in eleven swine operations in the Netherlands. *Tijdschr Diergeneeskd* **135**:134–137.

Keyburn AL, Boyce JD, Vaz P, Bannam TL, Ford ME, Parker D, Di Rubbo A, Rood J I and Moore R J. 2008. NetB, a new toxin that is associated with avian necrotic enteritis caused by *Clostridium perfringens*. *PLoS Pathog.*, **4**: 26.

Kiel N, Ho V and Natalie A P. 2011. A case of gas gangrene in an immunosuppressed Crohn's patient. *World Gastroenterol.*, **17**: 3856-3858.

Köpke M, Mihalcea C, Liew F, Tizard JH, Ali M S, Conolly J J, Al-Sinawi B and Simpson S D. 2011. 2,3-Butanediol Production by Acetogenic bacteria, an alternative route to chemical synthesis, using Industrial waste gas. *Appl Environ Microbiol.*, **77**: 5467-5475.

Koene M G J, Mevius D J, Wagenaar A, Harmanus C, Hensgens M P M, Meetsma A M, Putirulan F F, van Bergen M A P and Kuijper E J. 2012. *Clostridium difficile* in Dutch animals: their presence, characteristics and similarities with human isolates. *Clin Microbiol Infect.*,**18**: 778-784.

Kuehne S A, Heap J T, Cooksley C M, Cartman S T and Minton N P. 2011. *Strain Engineering: Methods and Protocols*, James A. Williams (Ed), **Methods in Molecular Biology**, vol. 765, DOI 10.1007/978-1-61779-197-0-23, Springer Science and Business Media,

Kuijper E J, Coignard B and Tull P. 2006. Emergence of *Clostridium difficile*-associated disease in North America and Europe. *Clin Microbiol Infect.*, **12**: 2-18.

Kuske C R. 2006. Current and emerging technologies for the study of bacteria in the outdoor air. *Curr Opin Biotechnol.*, **17**: 291-296.

Kyne L, Hamel M B, Polavaram R and Kelly C P. 2002. Health care costs and mortality associated with nosocomial diarrhoea due to *Clostridium difficile*. *Clin Infect Dis.*, **34**: 346-353.

Kyne L, Warny M, Qamar A and Kelly C P. 2001. Association between antibody response to toxin A and protection against recurrent *Clostridium difficile* diarrhoea. *Lancet*, **357**: 189-193.

Lamanna C. 1959. The most poisonous poison. Science, 130: 763-772.

Lange M, Neubauer H and Seyboldt C. 2010. Development and validation of a multiplex real-time PCR for detection of *Clostridium chauvoei* and *Clostridium septicum*. *Mol Cell Probes.*, **24**: 204-210.

Lecour H, Ramos H, Almeida B and Barbosa R. 1998. Food borne botulism. A review of 13 outbreaks. *Arch Int Med.*, **148**: 578-580.

Lee H C, Jemmer A M, Low C S and Lee Y K. 2006. Effects of tea phenolics and their aromatic faecal bacterial metabolites on intestinal microbiota. *Res Microbiol.*, **157**: 876-884.

Leffler D A and Lamont J T. 2009. Treatment of *Clostridium difficile*-associated disease. *Gastroenterol.*, **136**: 1899-1912.

Legaria M C, Lumelsky G and Rosetti S, 2003. *Clostridium difficile*-associated diarrhoea from a general hospital in Argentina. *Anaerobe*, **9**: 113-116.

Lütke-Eversloh T and Bahl H. 2011. Metabolic engineering of *Clostridium acetobutylicum*: recent advances to improve butanol production. *Curr Opin Biotechnol.*, **22**: 634-647.

MacLennan J D. 1943a. Anaerobic infections of war wounds in the Middle East. *Lancet*, **242**: 94e9.

MacLennan J D. 1943b. Anaerobic infections of war wounds in the Middle East. *Lancet*, **242**: 123e6.

MacLennan J D. 1943c. Anaerobic infections of war wounds in the Middle East. *Lancet*, **242**: 63e6.

MacLennan J D. 1962. The histotoxic clostridial infections of man. *Bacteriol Rev.*, **26**: 177e276.

MacLennan J D and MacFarlane R G. 1945. Toxin and antitoxin studies of gas gangrene in man. *Lancet*, **246**: 301e5.

Mangold T and Goldberg J. 1999. Plague Wars: A True Story of Biological Warfare. London: Macmillan.

Mao S, Luo Y, Bao G, Zhang Y, Li Y and Ma Y. 2011. Comparative analysis on the membrane proteome of *Clostridium acetobutylicum* wild type strain and its butanol-tolerant mutant. *Mol BioSystems*, **7**: 1660-1677.

Marks J D. 2004. Medical aspects of biologic toxins. *Anesthesiol Clin North Amer.*, **22**: 509- 532.

Masuyer G, Stancombe P, Chaddock J A and Acharya K R. 2011. Structures of engineered *Clostridium botulinum* neurotoxin derivatives. *Acta Crystallographica*, **F67**, 1466-1472.

McClane BA. 2007. *Clostridium perfringens*. In: Doyle MP, Beuchat LR, (Eds),**Food Microbiology: Fundamentals and Frontiers**, 3rd ed. Washington, D.C.: ASM Press; 2007. pp. 423-444.

McDonald LC. 2005. *Clostridium difficile*: responding to a new threat from an old enemy. *Infect Control Hosp Epidemiol.*, **26**: 672-675.

McFarland LV. 2006. Meta-analysis of probiotics for the prevention of antibiotic associated diarrhoea and the treatment of *Clostridium difficile* disease. *Amer J Gastroenterol.*, **101**: 812-822.

Merchant I A and Barner R D. 1964. An Outline of the Infectious Diseases of Domestic Animals. Iowa State University Press, Ames, IA, pp. 22-26.

Metcalf D S, Costa M C, Dew W M V and Weese J S. 2010. *Clostridium difficile* in vegetables, Canada. *Lett Appl Microbiol.*, **51**: 600-602.

Mhoma J R L, Rwehabula P, Kundy D J and Msangi N F. 1995. Assessment of blackquarter vaccine (AWASA strain) by mass vaccination of cattle and sheep in Tanzania. *Bull Anim Hlth Prod Afr.*, **43**: 227-30.

Middlebrook J and Franz D. 1997. Botulinum toxins. In: Zajtchuk R. (ed.) ,**Textbook of Military Medicine: Medical Aspects of Chemical and Biological Warfare**, US Department of the Army, Surgeon General, and the Borden Institute, Washington, DC. pp. 643–654

Minton N P. 2003. Clostridia in cancer therapy. *Nature Rev Microbiol.*, **1**: 237-242.

Mulvey M R, Boyd D A, Gravel D, Hutchinson J, Kelly S, McGeer A, Moore D, Simor A, Suh K N, Taylor G, Weese J S and Miller M. 2010. Hypervirulent *Clostridium difficile* strains in hospitalized patients, Canada. *Emerg Infect Dis.*, **16**: 678-681.

Nagano N, Isomine S, Kato H, Sasaki Y, Takahashi M, Sakaida K, Nagano Y and Arakawa Y. 2008. Human fulminant gas gangrene caused by *Clostridium chauvoei*. *J Clin Microbiol.*, **46**: 1545-1547.

Nakai Y, Tepp W H, Dickerson T J, Johnson E A and Janda K D. 2009. Function-oriented synthesis applied to the antibotulinum natural product toosendanin. *Bioorg Med Chem.*, **17**: 1152-1157.

Nakayama S, Kiyoshi K, Kadokura T and Nakazato A. 2011. Butanol Production from crystalline cellulose by cocultured *Clostridium thermocellum* and *Clostridium saccharoperbutylacetonicum* N1-4. *Appl Environ Microbiol.*, **77**: 6470-6475.

Nwokediuko S, Bababode B, Chinedu OU and Ozoh G. 2002. Peculiarities of chronic diarrhoea in Enugu, Southeastern Nigeria. *J Hlth Sci.*, **48**: 435-440.

Otte B, Grunwaldt E, Mahmoud O and Jennewein S. 2009. Genome shuffling in *Clostridium diolis* DSM 15410 for improved 1,3-propanediol production. *Appl Environ Microbiol.*, **75**: 7610-7616.

Papoutsakis E T. 2008. Engineering solventogenic clostridia. *Curr Opin Biotechol.*, **19**: 420-429.

Paredes-Sabja D and Sarker M R. 2012. Interactions between *Clostridium perfringens* spores and Raw 264.7 macrophages interactions between *Clostridium perfringens* spores and Raw 264.7 macrophages. *Anaerobe*, **18**(1): 148-156.

Paredes-Sabja D and Sarker M R. 2009. *Clostridium perfringens* sporulation and its relevance to pathogenesis. *Future Microbiol.*, **4**: 519-525.

Radostits O M, Gay C C, Blood D C and Hinchcliff KW. 2000. Veterinary Medicine. A Textbook of the Diseases of Cattle, Sheep, Pigs, Goats and Horses. 9th ed. W.B Saunders Company Ltd, London, pp. 760-763.

Ramarao D and Rao BU. 1990. Studies on the incidence of blackquarter in Karnataka during 1979-1985. *Indian Vet J.*, **67**:795-801.

Rodloff A C and Krüger M. 2012. Chronic *Clostridium* botulinum infections in farmers. *Anaerobe*, **18**(2): 226-228.

Rotz L D, Khan A S, Lillibridge S R, Ostroff S M and Hughes J M. 2002. Public health assessment of potential biological terrorism agents. *J Amer Med Assoc.*, **8**: 225-230.

Rupnik M, Wilcox M H and Gerding D N. 2009. *Clostridium difficile* infection: new developments in epidemiology and pathogenesis. *Natures Rev Microbiol.*, **7**: 526-36.

Rupnik M. 2007. Is *Clostridium difficile*-associated infection a potentially zoonotic and food borne disease ? *Clin Microbiol Infect.*, **13**: 457-459.

Sakanaka S, Juneja L R and Taniguchi M. 2000. Antimicrobial effects of green tea polyphenols on thermophilic spore-forming bacteria. *J Biosci Bioeng.*, **90**: 81-85.

Satoh E, Ishii T, Shimizu Y, Sawamura S and Nishimura M. 2001. Black tea extract, thearubigin fraction, counteracts the effects of botulinum neurotoxins in mice. *Exper Biol Med.*, **226**: 577-580.

Saxena S. 2005. *Glycyrrhiza glabra* medicine over the millennium. *Nat Prod Rad.*, **4**: 358-367.

Schantz E J and Johnson E A. 1992. Properties and use of botulinum toxin and other microbial neurotoxins in medicine. *Microbiol Rev.*, **56**: 80-99.

Seal D, Borriello SP, Barclay F, Welch A, Piper M and Bonnycastle M. 1987. Treatment of relapsing *Clostridium difficile* diarrhoea by administration of a non-toxigenic strain. *Eur J Clin Microbiol.*, **6**: 51-53.

Shapiro R L, Hatheway C and Swerdlow D L. 1998. Botulism in the United States: a clinical and epidemiologic review. *Ann Int Med.*, **129**: 221-228.

Shi Y L and Chen W Y. 1999. Effect of toosendanin on acetylcholine release from rat brain, a microdialysis study. *Brain Res.*, **850**: 173-178.

Shi Y-L and Li M-F. 2007. Biological effects of toosendanin, a triterpenoid extracted from Chinese traditional medicine. *Prog Neurobiol.*, **82**: 1-10.

Shi Y L, Wang W P and Xu K. 1981a. Electrophysiological analysis on the presynaptic blocking effects of toosendanin on neuromuscular transmission. *Acta Physiol Sin.*, **33**: 259-265.

Shi Y L, Wang W P, Yan S Q and Xu K. 1982. Effects of calcium ions and nerve impulses on changes in miniature endplate potential frequency produced by toosendanin. *Acta Physiol Sin.*, **34**: 304-309.

Shi Y L, Wei N S, Yang Y Q and Wang Z X. 1980. Toosendanin-a presynaptic neuromuscular blocking agent. *Acta Physiol Sin.*, **32**: 293-297.

Shi Y L, Yang Y Q, Wang W P and Xu K. 1981b. Influence of stimulus frequency, temperature and Ca²⁺ on onset of toosendanin paralysis of isolated neuromuscular preparations of rats. *Acta Physiol Sin.*, **33**: 141-146.

Simango C and Mwakuruddwa S. 2008. *Clostridium difficile* in broiler chickens sold at market places in Zimbabwe and their antimicrobial susceptibility. *Int J Food Microbiol.*, **10**: 268-270.

Skariyachan S, Mahajanakatti A B, Sharma N and Sevanan M. 2011. Selection of herbal therapeutics against deltatoxin mediated clostridial infections. *Bioinfo.*, **6**: 375-379.

Songer J G and Uzal F A. 2005. Clostridial enteric infections in pigs. J Vet Diag Investig., **17**: 528-536.

Songer J G, Trinh H T, Killgore G E, Thompson A D, McDonald L C and Limbago B M. 2009. *Clostridium difficile* in retail meat products, USA, 2007. *Emerg Infect Dis.*, **15**:819-821.

Sosa A. 2005. Antibiotic policies in developing countries. In: Gould IM, van der Meer J W M (Eds), Antibiotic Policies: Theory and Practice. New York: Kluwer Academic/Plenum, pp. 591-616.

Stabler R A, Gerding D N, Songer J G, Drudy D, Brazier J S, Trinh H T, Witney A A, Hinds J and Wren B W. 2006. Comparative phylogenomics of *Clostridium difficile* reveals clade specificity and microevolution of hypervirulent strains. *J Bacteriol.*, **188**:7297-7305.

Stevens D L, Aldape M J and Bryant A E. 2012. Life-threatening clostridial infections. *Anaerobe*, **18(2)**: 254-259.

Survase S A, Sklavounos E, Jurgens G, van Heiningen A and Granström T. 2011. Continuous acetone-butanol-ethanol fermentation using SO_2 -ethanol-water spent liquor from spruce. *Biores Technol.*, **102**: 10996-11002.

Tang W and Eisenbrand G. 1992. Chinese Drugs of Plant Origin: Chemistry, Pharmacology, and Use in Traditional and Modern Medicine. Springer, Heidelberg.

Titball R W. 2009. *Clostridium perfringens* vaccines. *Vaccine*, **27**: D44-D47.

Tomas C A, Beamish J and Papoutsakis E T. 2004. Transcriptional analysis of butanol stress and tolerance in *Clostridium acetobutylicum. J Bacteriol.*, **186**: 2006-2018.

Troxel T R, Burke G L, Wallace W T, Keaton L W, McPeake S R, Smith D and Nicholson I. 1997. Clostridial vaccination efficacy on stimulating and maintaining an immune response in beef cows and calves. *J Anim Sci.*, **75**:19-25.

Tummala S B, Welker N E and Papoutsakis E T. 2003. Design of antisense RNA constructs for downregulation of the acetone formation pathway of *Clostridium acetobutylicum*. *J Bacteriol.*, **185**: 1923-1934.

Twine S M, Reid C W, Aubry A, McMullin D R, Fulton K M, Austin J and Logan S M. 2009. Motility and flagellar glycosylation in *Clostridium difficile*. *J Bacteriol.*, **191**: 7050-7062.

Useh N M, Ajanusi J O, Esievo K A N and Nok A J. 2006c. Characterization of a sialidase (neuraminidase) from *Clostridium chauvoei* (Jakari strain). *Cell Biochem Funct.*, **24**: 347-352.

Useh N M, Ibrahim N D G, Nok A J and Esievo K A N. 2006b. Relationship between outbreaks of blackleg of cattle and annual rainfall in Zaria, Nigeria. *Vet Rec.*, **158**:100-101.

Useh N M, Nok A J, Ajanusi O J, Balogun E O, Oladele S B and Esievo K A N. 2004a. *In vitro* production of neuraminidase by *Clostridium chauvoei* (Jakari strain). *Vet Arh.*, **74**: 289-298.

Useh N M, Nok A J, Ambali S F and Esievo K A N. 2004b. Inhibition of *Clostridium chauvoei* (Jakari strain) neuraminidase activity by methanolic extracts of the stem barks of *Tamarindus indicus* and *Combretum fragrans*. *J Enzyme Inhib Med Chem.*, **19**: 339-42.

Useh N M, Nok A J and Esievo K A N. 2006a. Blackleg in ruminants. *Persp Agric Vet Sci Nutr Nat Res.*, 1: 040.

Uzal F A, Paramidani M, Assis R, Morris W and Miyakawa M F. 2003. Outbreak of clostridial myositis in calves. *Vet Rec.*, **152**: 134-136.

von Abercron S M M, Karlsson F, Wigh G T, Wierup M and Krovacek K. 2009. Low Occurrence of *Clostridium difficile* in retail ground meat in Sweden. *J Food Prot.*, **72**: 1732-1734.

Warny M, Pepin J, Fang A, Killgore G, Thompson A, Brazier, Frost E and McDonald L C J. 2005. Toxin production by an emerging strain of *Clostridium difficile* associated with outbreaks of severe disease in North America and Europe. *Lancet*, **366**: 1079-84. Weatherhead J E and Tweardy D J. 2012. Lethal human neutropenic entercolitis caused by *Clostridium chauvoei* in the United States: Tip of the iceberg? *J Infect.*, **64**: 225-227. doi:10.1016/j.jinf.2011.09.004.

Weese J S, Finley R, Reid-Smith R R, Janecko N and Rousseau J. 2010. Evaluation of *Clostridium difficile* in dogs and the household environment. *Epidemiol Infect.*, **138**: 1100-1104.

Weese J S. 2010. *Clostridium difficile* in food-innocent by stander or serious threat? *Clin Microbiol Infect.*, **16**:3-10.

Weese J S, Staempfli H R, Prescott J F, Kruth S A, Greenwood S J and Weese H E. 2001a. The roles of *Clostridium difficile* and enterotoxigenic *Clostridium perfringens* in diarrhoea in dogs. *J Vet Int Med.*, **15**: 374-378.

Weese J S, Weese H E, Bourdeau T L and Staempfli H R. 2001b. Suspected *Clostridium difficile* associated diarrhoea in two cats. *J Amer Vet Med Assoc.*, **218**: 1436-1439.

Weingart O G, Schreiber T, Mascher C, Pauly D, Dorner M B, Berger T F H, Egger C, Gessler F, Loessner M J, Avondet M-A and Dorner B G. 2010. The case of botulinum toxin in milk: experimental data. *Appl Environ Microbiol.*, **76**: 3293-3300.

Williams B M. 1977. Clostridal myositis in cattle: bacteriology and gross pathology. *Vet Rec.*, **100**: 90-91.

Williams B M, Andrews A H. Bacterial conditions. 1992. In: Andrews AH, Lowrey RW, Boyd H, Eddy RG, editors. **Bovine Medicine, Diseases and Husbandry of Cattle**. Blackwell Scientific Publications, Oxford, pp. 551-557.

Woodford N and Livermore D M. 2009. Infections caused by Gram-positive bacteria: a review of the global challenge. *J Infect.*, **59** Suppl 1: S4.

World Health Organization (WHO), 2002. WHO Traditional Medicine Strategy 2002–2005. World Health Organization, Geneva.

Xiu Z L and Zeng A P. 2008. Present state and perspectives of downstream processing of biologically produced 1,3-propanediol and 2,3-butanediol. *Appl Microbiol Biotechnol.*, **78**: 917-926.

Xu T H, Shi Y L, Ren X M and Zhou J Y. 2004. Inhibition of glutamate release from rat cerebral synaptosomes by a prejunctional blocker toosendanin. *Chinese J Pharmacol Toxicol.*, **18**: 321-326.

Yen H-W, Lin S-F and Yang I-K. 2012. Use of poly (etherblock-amide) in pervaporation coupling with a fermentor to enhance butanol production in the cultivation of *Clostridium acetobutylicum. J Biosci Bioeng.*, **113(3):** 372-377.

Yineger H, Kelbessa E, Bekele T and Lulekal E. 2007. Ethnoveterinary medicinal plants at Bale Mountains National Park, Ethiopia. *J Ethnopharmacol.*, **112**: 55-70.

Yu M, Zhang Y, Tang I-C and Yang S-T. 2011. Metabolic engineering of *Clostridium tyrobutyricum* for n-butanol production. *Metabolic Eng.*, **13**: 373-382.

Zhang L, Leyn S A, Gu Y, Jiang W, Dmitry A and Yang R C. 2012. Ribulokinase and transcriptional regulation of arabinose metabolism in *Clostridium acetobutylicum*. *J Bacteriol.*, **194(5)**: 1055-1064.

Zhu Y P and Woerdenbag, H J. 1995. Traditional Chinese herbal medicine. Pharmacy World. *Science*, 17: 103-112.