Changing Epidemiology of Classical and Emerging Human Fungal Infections: A Review

Khaled H. Abu-Elteen^{1,*} and Mawieh A. Hamad²

¹Department of Biology and Biotechnology, The Hashemite University, Zarqa, Jordan and ²Department of Medical Technology, Sharjah University, UAE

Received 17th August, 2012; accepted 8th October, 2012

Abstract

In recent decades, many fungal species have emerged as major causes of human disease. While invasive candidiasis, aspergillosis, and cryptococcosis remain very common, rates of infection by other opportunistic fungal pathogens such as *Histoplasma capsulatum*, *Coccidioides immitis, Fusarium* spp. and other yeasts and molds are on the rise. Adding to this bleak picture is the fact that treatment and control measures currently available to us are hardly able to keep up with current trends of morbidity and mortality that associate with common and emerging fungal infections. It is interesting to note the likelihood of emerging fungal pathogens exhibiting significant resistance to standard antifungal therapy is real. Hence, invasive infections due to previously rare fungi such as *Acremonium, Scedosporium, Paecilomyces*, and *Trichoderma* species are proving difficult to treat. The ever increasing number of hosts with compromised immunity, increased volume of surgeries and invasive medical procedures, limited repertoire of and increased resistance to available antifungals, and better diagnosis and pathogen identification procedures are largely to blame for these alarming trends. Improvements in managing patients with cancer, AIDS, diabetes, and transplantation that are significantly improving patient survival rates are also generously contributing to the pool of patients with compromised immunity. In this article, we review the changing spectrum of invasive mycosis, risk-factors for infections and susceptibility to available antifungals.

Keywords: Aspergillus, Antifungal Drugs, Candida, Compromised Immunity, Cryptococcus, Histoplasma, Invasive Mycosis.

1. Introduction

The incidence of invasive fungal infections (IFIs) and rates of morbidity and mortality due to such infections have all been on the rise over the last three decades (Binder et al., 2011; Low et al., 2011). Although Candida albicans, Aspergillus fumigatus and Cryptococcus neoformans are the most common causes of IFIs (Pfaller and Diekema, 2004a), the incidence of infection by Candida spp. other than C. albicans, Aspergillus spp. other than A. fumigatus, opportunistic yeast-like fungi (Trichosporon spp., Rhodotorula spp. and Geotrichum capitatum [Blastoschizomyces capitatus]), zygomycetes; hyaline molds (Fusarium, Acremonium, Scedosporium, Paecilomyces, and Trichoderma spp), and a wide variety of dematiaceous fungi are all on the rise (table 1). The emergence of organisms such as Fusarium spp., Histoplasma capsulatum, Coccidioides immitis as significant pathogens has important implications for diagnosis and management. On the one hand, the clinical presentation of infections caused by such pathogens can mimic more common diseases (e.g. aspergillosis). On the other hand, these emerging pathogens are resistant to conventional antifungals making infection management difficult to say the least (Fleming et al., 2002; Miceli et al., 2011).

1	Table	1.	Spectrum	of	opportur	nistic	human	fungal	pathogens	
	0					~				

Genus /	Species /	Genus /	Species /
Group	Members	Group	Members
Candida spp.	C. albicans	Other yeasts	Cryptococcus
	C. glabrata		neoformans
	C. gulliermondii		Trichosporon spp
	C. kefyr		Rhodotorula spp.
	C. krusei		Geotrichum
	C. lusitaniae		capitatum
	C. rugosa		Blastoschizomyces spp.
	C. parapsilosis		Malaseezia spp.
	C. tropicalis		Saccharomyces spp.
		Zygomycetes	Absidia spp.
			Cunninghamella spp.
			Mucor spp.
Aspergillus spp.	A. fumigtus		Rhizopus spp.
	A. niger	Dematiaceous	Rhizomucor spp.
	A. flavus	molds	Alternaria spp.
	A. terreus		Bipolaris spp.
Other hyaline	Scedosporium spp.		Curvularia spp.
molds	Fusarium spp		Cladophialophora spp.
	Acremonium spp.		Exophiala spp.
	Paecilomyces spp		Phialophora spp.
	. Trichoderm spp.		
	Scopulariopsis spp.		

^{*} Corresponding author e-mail: salma@hu.edu.jo

Continuous rise in the number of hosts with compromised immunity (cancer patients, transplant recipients on immunosuppressive therapy, AIDS patients, and diabetics among others) as a result of improved patients management procedures and drugs is partly to blame for this worrying trend (Richardson and Lass-Florl, 2008).

Increased use of antibiotics and immunosuppressive drugs (cyclosporine A, tacrolimus, etc.), hyperalimentation fluids, polyethylene catheters, pressure monitoring devices, heroin abuse, organ transplantation, abdominal surgeries, and prosthetic cardiac valves all disturb host immunity and predispose to opportunistic fungal infections. Lack of preventative measures (vaccines) against all human fungal infections and the limited efficacy of and increased resistance to the few (<15) available antifungal drugs further complicate the issue. Unfortunately, as risk-factors for IFIs continue to increase in type, frequency, and severity, it is likely that the rate of IFI will continue on its upward trend.

Here, we review recent epidemiologic trends of two major groups of human fungal infections. Namely, those caused by yeasts (Candida and Cryptococcus) and yeastlike fungi (Trichosporon, Rhodotorula, and Geotrichum) and those caused by filamentous molds (Aspergillus, Scedosporium, Fusarium, Acremonium, Paecilomyces, Trichoderma, Zygomycetes Mucormycetes, or Dematiaceous molds or Phaeohyphomycetes, and Histoplasma). Published data permitting, the focus in discussing each group will be on the incidence of infection by the pathogen(s), predisposing factors to infection, rates of morbidity and mortality that associate with it, and clinical manifestations.

2. Pathogenic Yeasts and Yeast-Like Fungi

2.1. Candida Species

Candida albicans and other *Candida* spp. are commonly found in the gastrointestinal tract, oral cavity, and genital areas as harmless commensals. Based on recent studies in healthy individuals, asymptomatic oral carriage of *Candida* spp. occurs in about 24-70% of children and adults with a reduced frequency in babies less than 1 year of age. *C. albicans* represents the majority (38-76%) of isolates identified in both adults and children. The frequency of *C. albicans* varies across different age groups with far greater proportions of isolates identified as *C. albicans* occurring in young babies and the elderly. Higher oral carriage rates are found in HIV positive patients (Liu *et al.*, 2008) and diabetics (Abu-Elteen *et al.*, 2006).

Asymptomatic vaginal carriage of *Candida* spp. is estimated to occur in 21-32% of healthy women, with *C. albicans* representing 20-98% of identified isolates (Ferrer, 2000; Pfaller and Diekema, 2007; Enoch *et al.*, 2006; Pirotta and Garland, 2006). Beigi *et.al.*, (2004) found that within a group of women repeatedly screened over 12 months, 30% were never colonized, 70% were colonized on at least one occasion, and 4% were persistently colonized. Higher rates of vaginal carriage have been found in pregnant women, women colonized by *Lactobacillus* spp (Beigi *et al.*, 2004; Hamad *et al.*, 2006), type I diabetics (de Leon *et al.*, 2002), and post-antibiotic treatment (Pirotta and Garland, 2006). Table 2 gives a brief summary of risk factors for localized and systemic candidiasis in general. It is worth noting that as well as being harmless commensals, *C. albicans* and other *Candida* spp. are opportunistic pathogens capable of causing a wide range of superficial, localized, and/or systemic infections (Pfaller and Diekema, 2007; 2004a).

Table 2. General factors predisposing to Candida infections

Factors	Comments	Representative
		examples
Immunological factors	Presence of	A cute leukemia,
Neutrophil defect	abnormally small	chemotherapy,
	numbers of	irradiation
	neutrophils in the	
	circulating blood,	
	doficiency	
T- lymphocyte.	Certain diseases	Auto immune
mononuclear	could lead to a defect	deficiency
phagocyte	in	syndrome (AIDS),
	T – lymphocyte	Hodgkin's disease,
	mononuclear	chemotherapy
	phagocyte	
Reticuloendothelial	Defect of RES causes	Congenital
system (RES)	impairment in the	absence
	infectious particles	OF defect of the
	from the blood; this	spleen.
	is due to congenital	splenectomy
	or surgical causes	
Chemotherapy and	Treatment with drugs	Immunosuppressiv
Radiotherapy	and/or irradiation that	e agents,
	alters the	antineoplastic
	composition of the	agents, antibiotics,
	endogenous microbial flora or	corticosteroius
	suppresses host	
	defenses against	
	infection	
Interruptes integument	Membrane trauma,	Finger or bone
	burns local occlusion	marrow punctures,
	or maceration of	gastrointestinal
	tissues	(GI) ulcers,
		catneters. 1 v
		dentures
Surgical procedures	Introduction of	Heart valves
C .	mechanical devices	replacements,
	and prostheses into	tracheostomy
	vessels or tissues	respiratory
		assistance,
		endoscopies, renai
		operation GI
		or gynecological
		surgery, blood
		transfusion
Physiological factors	Infectious, idiopathic,	Microbial
	congenital,	infections,
	or other debilitating	endocrine
	diseases	dystunctions,
	Digressions from	mediated
	normal physiological	immunity
	status	Pregnancy,
		infancy
Nutritional factors	Excessof deficiency	Carbohydrate -
	of food stuff that	rich diets, vitamin
	crate an environment	deficiency
	conductive to the	
	development of	
	mucosai canuluosis	

2.1.1. . Superficial Mucosal Infections

Superficial mucosal lesions (thrush) occur in the oral and vaginal cavities of immunocompetent as well as immunocompromised hosts. Oral candidiasis is common in infants and the elderly and in cancer patients undergoing chemotherapy to treat hematological malignancies and those undergoing head/neck radiation. It is characterized by white growth on the mucous membranes of the oral cavity that is usually underlined by red areas when the yeast growth is scraped off (MacCallum, 2007).

The majority of isolates associated with oral candidiasis are *C. albicans* (63-84%) (Davies *et al.*, 2006). Risk factors associated with oral candidiasis include xerostomia (dry mouth) and denture wearing (Abu-Elteen and Abu-Elteen, 1998; Davies *et al.*, 2006), poorly controlled diabetes mellitus (Abu-Elteen *et al.*, 2006), and immunosuppression (table 3). The frequency of oral candidiasis, but not oral carriage of *Candida* spp. (Sanchez-Vargas *et al.*, 2005), is higher in HIV positive patients with decreased CD4⁺ T cell count (Liu *et al.*, 2006).

Table 3. Specific factors predisposing to neonatal, oral

and vaginal candidiasis.

Predisposing	Type of candidiasis				
factor(s)					
	Neonatal	Oral	Vaginal		
Interrupted	Birth trauma,	Burns, oral trauma,			
Nutritional factors	Malnutrition, breast and bottle feeding	Malabsorption	Malnutrition		
Chemotherapy and radiotherapy	Antibiotic and steroid therapy (taken by mother of infant after birth) effect (psychophamaceuticals), radiation therapy	Broad spectrum antibiotics, corticosteroids, irradiation, or drugs with xerostomic side	Antibiotic therapy, oral contraceptives		
Surgical procedures	Resuscitative procedure intubations (especially transplantation and heat surgery), tooth extraction	Parenteral nutrition, convalescence after operations			
Other factors	Unsterile delivery, male sex, unhygienic environment, low birth weight, thumb and dummy sucking,	Poor oral hygiene, oral leukoplakia, immunological factors	Environmental factors (humidity and warmth, wearing tight-		

Vulvovaginal candidiasis (VC) represents a real health problem to women of childbearing age worldwide. The majority of cases (>80%) of VC involve colonization by the genitourinary tract commensal *C. albicans* (Pfaller and Diekema, 2007; Enoch *et al.*, 2006). Occurrence of VC and recurrent VC has been attributed to compromised immunity and increased levels of estrogen in the reproductive tract milieu (Hamad *et al.*, 2004). Symptoms of VC include itching, burning, soreness, and abnormal vaginal discharge. *C. glabrata* is emerging as an important and potentially resistant opportunistic fungal pathogen in VC (Pfaller and Diekema, 2004a). Abu-Elteen (2001) has demonstrated that among the *Candida* spp. *C. glabrata* alone has increased in incidence as a cause of VC in Jordan since 1994. In many geographic regions, *C. glabrata* is becoming a common cause of VC and is gradually showing increased resistant to fluconazole (Richardson and Lass-Florl, 2008; Pfaller and Diekema, 2004a; Ray *et al.*, 2007).

2.1.2. Bloodstream Infections or Candidemia

Bloodstream infections by *Candida* spp. (candidemia) is the fourth leading cause of nosocomial bloodstream infections (BSI) in the United States (Pfaller and Diekema, 2004a; 2007, Wisplinghoff et al., 2004). The annual incidence of Candida associated BSIs have been reported at 6-23 cases/100,000 Americans (Clark and Hajjeh, 2004; Pfaller et al., 2006), and 2.5-11 cases/100 000 Europeans (Tortorano et al., 2006). In recent years, it has been noted that a gradual increase in the incidence of BSIs is taking place (Bougnuox et al., 2008; Vardakaz et al., 2009; Chow et al., 2008). More than 17 different Candida spp. have been identified as etiologic agents of BSIs. However, 95% or so of all Candida BSIs are caused by four Candida spp.; namely, C. albicans, C. glabrata, C. parapsilosis, and C. tropicalis (Hajjeh et al., 2004; Pfaller and Diekema, 2004b). The remaining 5% of Candida BSIs are caused by C. krusei, C. lusitaniae, C. guilliermondii, C. dubliniensis, and C. rugosa among others (Bassetti et al., 2009; Fridkin et al., 2006; Peman et al., 2008). C. albicans is the lead cause of BSIs as it accounts for 42-100% of cases depending on the patient group. For example, frequency ratio of C. albicans to non-albicans Candida in patients with hematological malignancies was about 1/2 that in patients with solid tumors (Tortorano et al., 2006; Pasqualotto et al., 2006). Although epidemiologic data reflects significant differences between countries with regard to Candida spp. distribution in general, C. albicans continues to be the most commonly encountered Candida spp. in Europe and the United States. The second most commonly encountered Candida spp. in Southern Europe and in France, Germany, and the UK are C. glabrata and C. parapsilosis respectively (Hajjeh et al., 2004; Almirante et al., 2005). C. parapsilosis occurs with high frequency in premature neonates and in patients with vascular catheters (Almirante et al., 2005; Arendrup et al., 2005). C. glabrata infections are rare in infants and children but occur with high frequency in the elderly (Pfaller et al., 2006; Hajjeh et al., 2004; Malani et al., 2005). C. tropicalis is an important cause of invasive diseases in patients with hematological malignancy (Pfaller and Diekema, 2004 a). With the increasing use of fluconazole as an anti-candidiasis agent in the USA, the emergence of C. glabrata and C. krusei has been reported (Pfaller and Diekema, 2004 a, Malani et al., 2005; Shao et al., 2007). In contrast, the frequency of C. glabrata as a cause of BSIs has decreased in Europe from 12.3 % to 8.8% and in Latin America from 10.2% to 4.7% (Hope et al., 2002). But overall, the frequency of non-albicans Candida spp. as causative agents of BSIs continues to rise (Pfaller et al., 2006; Clark and Hajjeh, 2004).

The majority of patients who develop candidemia are intensive care unit (ICU) patients and those undergoing abdominal surgery. Other patient groups at risk of candidemia are cancer patients (solid tumor or hematological malignancies), premature babies (<1 kg birth weight), patients on steroids therapy (MacCallum, 2007), and patients with catheters and other invasive medical devices. According to one study, 14% of screened central venous catheter (CVC) tips were positive mainly for coagulase-negative *Staphylococcus* spp. followed by *C. albicans* (Hammarskjold *et al.*, 2006). It should be noted that for critically-ill patients, several risk factors may be present in, or apply to, the same patient.

2.2. Cryptococcus Species

Cryptococcal infections occur with a near worldwide distribution in immunosuppressed hosts. They are the second most common cause of opportunistic fungal infections in AIDS patients. The incidence of infections caused by the encapsulated yeast Cryptococcus neoformans (C. neoformans) has risen markedly over the last 20 years (Bicanic and Harrison, 2004). Infections occur through inhalation of small diameter (<10 mm) yeast like organisms which enter small respiratory passages and become mostly dormant for a time (Bicanic and Harrison, 2004; Subramanian and Mathai, 2005) before reactivating in the lungs and/or lymph nodes. Clinical manifestations of infection can range from asymptomatic colonization of the respiratory tract to a widespread dissemination depending on host immune factors, inoculum, and degree of virulence (Mitchell and Perfect, 1995). As dissemination occurs, the central nervous system (CNS) is commonly involved. The basal meninges of the brain are preferentially affected causing thickening with subsequent invasion of the deeper brain tissues. In the meninges, the organism appears to be suspended in a mucoid-like material that is derived from the capsule (Mitchell and Perfect, 1995).

Cryptococcus neoformans is a basidomycete that normally grows as saprophytic haploid-budding yeast. Opposite mating types of C. neoformans do exist and the pathogen can undergo sexual reproduction and meiosis to produce spore. The yeast is spherical-oval in shape and is 5-10 µm in diameter. C. neoformans strains manifest antigenic differences that allow them to be grouped into five different serotypes (A, B, C, D, and an AD hybrid) as well as different varieties. C. neoformans var. neoformans includes serotypes D and AD while var. grubii includes serotype A and var. gattii includes serotypes B and C. C. neoformans var neoformans and var. grubii are responsible for majority of clinical the infections in immunocompromised host while var. gattii causes disease primarily in immunocompetent hosts (Fraser et al., 2005; Morrow and Fraser, 2009).

Cryptococcus neoformans has a number of virulence factors that enable it to survive and replicate in humans (Casadevall *et al.*, 2003), especially in cases where T-cell immunity is compromised. Fungal capsule , which is antiphagocytic and down-regulates cellular and humoral immune responses when shed into host tissues, and laccase and melanin, which interfere with oxidative killing by phagocytes, are among the prevalent virulence factors (Mitchell and Perfect, 1995). Production of melanin from 1-dopa by the enzyme laccase may account for the predilection of the organism for CNS. *C. neoformans* is both an intracellular and an extracellular pathogen; it can survive and replicate within acidic macrophage phagolysosomes (Levitz *et al.*, 1999). A host site with abundant carbon dioxide concentration favors capsule

bioformation (Subramanian and Mathai, 2005). While *C. neoformans* lives in soil and organic matter containing pigeon and bird excreta, *C. gattii* is found primarily in tropical and subtropical regions and has been associated with several spp. of eucalyptus trees an causes infection in immunocompetent hosts.

C. neoformans is neurotropic and most patients with cryptococcal meningitis suffer from defective cellular immunity. The infection is seen most frequently in association with lymphomas, AIDS, transplant recipients, and patients on corticosteroid therapy (Khawcharoenporn et al., 2007 a). In the 1980s (the age of AIDS), cryptococcosis emerged as an important opportunistic infection occurring in 5-10% of AIDS patients in the US, Europe, and Australia (Bicanic and Harrison, 2004). With increased use of fluconazole for oral candidiasis and the advent of highly active antiretroviral therapy (HAART) in the mid-1990s, the annual incidence of cryptococcosis has markedly decreased in the developed countries. For example, in Atlanta/Georgia in the US, the incidence of cryptococcosis dropped from 66 cases/1000 AIDS patients in 1993 to only 7 cases/1000 in 2000 (Mirzak et al., 2003). In a recent review of cryptococcal infections in HIVnegative patients, splenectomy was reported to be a risk factor in 3% of cases (Qazzafi et al., 2007). Other groups at risk of cryptococcosis are organ transplant recipients on immunosuppressive therapy and patients with sarcoidosis or lymphoproliferative disorders. In a cohort of 306 HIVnegative patients with cryptococcosis, the predisposing conditions were steroids (28%), organ transplantation (18%), chronic organ failure (Liver, kidney, lung)(18%), malignancy (18%) and rheumatological diseases (13%) (Pappas, et al., 2001); in 22% of patients in the study group no predisposing factor was identified.

Traditionally, non-neoformans cryptococci have been regarded as saprophytes and rarely reported as human pathogens (Khawcharoenporn et al., 2007b). However, the incidence of infection due to such organisms has increased over the last few decades with Cryptococcus albidus being responsible for 80% of reported cases. Impaired cell-mediated immunity is an important risk factor for non-neoformans cryptococcal infections and prior azole prophylaxis accounts for increased incidence of resistance being noted. Cryptococcus gattii causes disease in immunocompetent hosts in a geographically restricted area in Australia (Richardson and Lass-Florl, 2008; Bicanic and Harrison, 2004). Recently, invasive C. gattii infections in immunocompetent hosts have been reported in Western Canada, mainly Vancouver Island (Lindberg et al., 2007) and the North West region of the USA. The organism, which is thought to thrive only in tropical regions, has been recovered in some temperate climate zone countries.

2.3. Pathogenic Yeast-Like fungi

The frequency of invasive mycoses due to rare and emerging opportunistic yeast-like fungi has increased significantly over the last two decades (Richardson and Lass-Florl, 2008; Pfaller and Diekema, 2004a; Walsh *et al.*, 2004). Such organisms may occupy environmental niches, found in food and water, or exist as normal human microflora. The list of opportunistic yeast-like fungi is long; hence, this discussion will be limited to three genera that pose particular medical problems. Namely, *Trichosporon, Rhodotorula*, and *Geotrichum capitatum* (*G. capitatum* or *Blastoschizomyces capitatum* as it is commonly known) (Richardson and Lass-Florl, 2008; Pfaller and Diekema, 2004a; Konotoyiannis *et al.*, 2004; Girmenia *et al.*, 2005; Makela *et al.*, 2003).

2.3.1. Trichosporon Species

Trichosporon is a genus of basidiomycetous yeasts that inhabits the soil and colonizes human skin and GI tract (Li et al., 2005). Previously, all pathogenic members of the genus Trichosporon were regarded as members of a single species (Trichosporon beigelii). More recently however, with biochemical and morphologic differences within the genus being increasingly appreciated, T. beigelii has been regrouped into several distinct species. Greater than eight of which have the potential to cause human disease; namely, T. asahii, T. inkin, T. asteroids, T. cutaneum, T. mucoides, T. ovoides, T. pullulans, and more recently T. loubieri (Li et al., 2005). While T. asahii and T. mucoides cause deep invasive and disseminated infections, T. asteroids and T. cutaneum cause superficial skin infections, T. ovoides causes white piedra of the scalp, and T. inkin causes white piedra of the pubic hair (Walsh et al., 2004; Flemming et al., 2002; Middelhoven, 2003; Marty et al., 2003). T. faecale was isolated from the skin of a patient with tinea pedis in Germany (Hahner et al., 2008) and T.loubieri has been associated with mycosis in patients with adult polycystic kidney disease (Padhye et al., 2003). T. mycotoxinivorans is a newly recognized human respiratory pathogen with high predilection for patients with cystic fibrosis (Hicket et al., 2009). Additionally, T. montevideense and T. domesticum have also been implicated in summer-type hypersensitivity pneumonitis (Nishiura et al., 1997; Sugita et al., 1998).

Risk factors for infection include immunosuppression, disruption of mucosal integrity, and CVCs. Neutropenic cancer patients on cytotoxic therapy are among the high risk groups of developing trichosporonosis. Furthermore, it has been reported that 63% of 287 *Trichosporon* cases had an underlying hematological malignancy (Girmenia *et al.*, 2005) suggesting that hematological malignancies represent a major risk factor for trichosporonosis. In contrast, disseminated trichosporonosis is less common in patients with solid-organ transplants (SOT), AIDS, or burns, and in premature babies.

Overall rates of mortality due to infections by *Trichosporon* spp are high; they range between 60-80% (Walsh *et al.*, 2004; Flemming *et al.*, 2002). However, recent improvement in diagnosis, treatment, and prevention measures are bringing these rates down.

2.3.2. Rhodotorula Species

Rhodotorula spp. are yeast-like fungi that belong to the family *Cryptococcaceae*, sub-family *Rhodotorulodea*. These encapsulated basidiomycetes are being increasingly recognized as important human pathogens (Lo Re, *el al*, 2003; Thakur *et al.*, 2007; De Almeida *et al.*, 2008; Baradkar and Kumar, 2008; Shinde *et al.*, 2008; Hsueh *et al.*, 2003; Fung *et al.*, 2008; Riedel *et al.*, 2007; Zaas *et al.*, 2003). Many species of the genus *Rhodotorula* have been described. *R. rubra, R. glutinis, R. mucilaginosa*, and *R. minuta* have been implicated as causes of meningitis,

endocarditis, ventriculitis, peritonitis, fungemia, CVCrelated infections, and keratitis (De Almeida et al., 2008; Baradkar and Kumar, 2008; Shinde et al., 2008; Hsueh et al., 2003; Fung et al., 2008; Riedel et al., 2007; Zaas et al., 2003). They exist as commensals on the skin, nails, and mucous membranes (Pfaller and Diekema, 2004a; Richardson and Lass-Florl, 2008). While Rhodotorula strains appear to be less virulent than the more common yeast pathogens (Candida and Cryptococcus neoformans), Rhodotorula infections have been associated with a crude mortality rate of up to 15% (De Almeida et al., 2008). They can also cause sepsis and other life-threatening complications (Pfaller and Diekema, 2004a; Richardson and Lass-Florl, 2008). Risk factors include CVC and malignancies (Pfaller and Diekema, 2004a). De Almeida et al. (2008) and Tuon et al. (2007) reported that in 88 cases of CVC-related fungemia due to Rhodotorula spp, all but one patient had an underlying disease state; most commonly cancer (78.4%). R. mucilaginosa was the species most frequently recovered (75%) followed by R. glutinis (6%). Rhodotorula BSIs can be successfully managed with line removal, antifungal therapy, or combinations of both.

2.3.3. Geotrichum capitatum

Geotrichum capitatum (formerly known as Trichosporon capitatum or Blastoschizomyces capitatus) is an uncommon, but frequently fatal, cause of IFIs in immunocompromised patients, particularly those with hematological malignancies (Pfaller and Diekema, 2004a; Richardson and Lass-Florl, 2008; Girmenia et al., 2005; Bouza and Munoz, 2004; Martino et al., 2004). It is widely distributed in nature and may be found as part of the normal skin flora. In a retrospective multicentre study from Italy, the incidence of G. capitatum infections among patients with acute leukemia was reported at 0.5% with a 55.7% crude mortality rate (Girmenia et al., 2005). Infection of neutropenic patients with G. capitatum presents in a manner similar to that of Trichosporon infections; i.e., frequent breakthrough infection (36% of episodes), frequent fungemia with multi-organ (including brain) dissemination, and a mortality rate of 60-80% (Martino et al., 2004); blood cultures are usually positive. As with Trichosporon infections, chronic disseminated G. capitatum infections may be seen upon resolution of neutropenia.

3. Pathogenic Filamentous Fungi

3.1. Aspergillus Species

Although the genus *Aspergillus* contains approximately 175 species, only *A. fumigatus, A. flavus, A. terreus, A. niger* and *A. nidulans* are associated with human disease (Pfaller and Diekema, 2004 a; Richardson and Lass-Florl, 2008; Maschmeyer *et al.*, 2007). The conidia or spores are easily released into the atmosphere to reach the lung alveoli (Richardson and Lass-Florl, 2008); breathed air is a major route of transmission. Invasive aspergillosis (IA) occurs almost exclusively in immunocompromised individuals. Infections have frequently been described in patients with hematological malignancies, SOT recipients, and patients undergoing chronic intermittent hemodialysis

(IA in the later case has been linked mainly to *Aspergillus* spp-contaminated hospital ventilation systems). Over the last 10 years, *A. fumigatus* has become the most prevalent airborne fungal pathogen accounting for >90% of human fungal infections (Maschmeyer *et al.*, 2007; Singh and Paterson, 2005; Denning, 1998; Rosenhagen *et al.*, 2009). IA is currently responsible for approximately 30% of fungal infections in patients dying of cancer; it also occurs in 10-25% of all leukemia patients where post-treatment mortality rate is 80-90% (Rosenhagen *et al.*, 2009; Denning, 1995; Verweij and Denning, 1997).

Risk factors for IA include prolonged and profound neutropenia, high-grade graft- versus-host diseases (GVHD), use of corticosteroids, age >40 years, and receipt of stem cells from HLA-mismatched donors (Shao et al., 2007; Nivoix et al., 2008; Marr et al., 2002) (table 4). particularly Aspergillosis remains common in hematopoietic stem cell transplant (HSCT) recipients and patients with advanced AIDS. It is also emerging as a serious outcome of immunosuppression, especially that rendered by new and more effective generations of immunosuppressives like infliximab (Pfaller et al., 2006; Patterson, 2005). Rates of Aspergillus infection in HSCT recipients and in SOT recipients have been reported at 2-26% and 1-15% respectively. Rates of mortality in transplant recipients due to IA range between 74-92%. Some have suggested that about 9-17% of deaths that occur in transplant recipients during the first year can be attributed to IA (Singh and Paterson, 2005). Pagano et al. (2007) have reported that IA rate of infection in 3000 transplant recipients was 2.8% (91 cases) and that the mortality rate within this subgroup was about 72%. Other studies have reported higher incidence; a Spanish study (Martino, et al., 2002) reported a rate of infection of 8.1%. Recent US series (Upton et al., 2007) have put IA-related mortality rates at 2.5%.

Globally speaking, the incidence of IA is about 1.4%; it is somewhat higher in lung transplant recipients (3%), heart transplant recipients (2.4%) (Gavalda et al., 2005), and liver transplant recipients (1.5-10%) (Rosenhagen et al., 2009). In general terms, IA is associated with high rates of mortality, which exceeds 50% according to many reports (Pfaller et al., 2006; Shao et al., 2002; Nivoix et al., 2008). Higher mortality rates were noted in HSCT recipients compared with SOT recipients (68% vs 41%) and in neutropenic patients compared with nonneutropenic counterparts (89% vs. 60%) (Cornillet et al., 2006). Aspergillosis is also emerging as a serious form of mycosis in the ICU; it has been reported that the incidence of aspergillosis in the ICU is 2.7-58 cases/1000 admissions with a mortality rate of 75-95% (Meersseman et al., 2007). Most infected patients present chronic obstructive pulmonary disease and receive high-dose corticosteroids (Meersseman et al., 2007).

Infections by non-funigatus Aspergillus spp. are becoming increasingly common as well (Shao et al., 2002; Singh and Paterson, 2005). This is especially true with regard to infections caused by *A. terreus* (Meersseman et al., 2007; Howard et al., 2009; Steinbach et al., 2004; Lass- Florl et al., 2005), which has been recently recognized as a cause of frequently lethal infections and which tends to be resistant to amphotericin B (Richardson

and Lass-Florl, 2008; Pfaller and Diekema, 2004a). In some cases, hospital-born A. flavus infections are becoming more common than those caused by A. fumigatus; the reason(s) for this trend are not readily apparent (Hedayati et al., 2007). Common clinical syndromes that associate with A. flavus infections include chronic granulomatous sinusitis, keratitis, cutaneous aspergillosis, wound infections, and osteomyelitis (Shao et al., 2002; Richardson and Lass-Florl, 2008; Pfaller and Additionally, A. flavus produces Diekema, 2004a). aflatoxin, which is an extremely toxic and potent hepatocarcinogen. Infections caused by a recently recognized spp. of Aspergillus (A. lentulus) have been reported (Balajee et al., 2006; Ando et al., 2008). Recent studies have shown that new triazoles like voriconazole and posaconazole are more effective than fluconazole in preventing and treating IA cases in HSCT recipients or those with GVHD (Shao et al., 2002; Richardson and Lass-Florl, 2008; Abu-Elteen and Hamad, 2007).

 Table 4. Co-morbid conditions for aspergillosis and mold infections in high-risk patients

Hematological	Organ transplant patients		
malignancies			
Leukaemia	Lung, Liver, heart, renal		
Myelodysplastic	Acute and chronic		
syndrome	rejection		
Stem –	Steroids		
cell transplant	Haemodialysis		
GvHD ^a (acute	Tacrolimus		
and chronic)	Renal failure		
Prolonged	Cytomegalo virus		
neutropenia	(CMV)		
Induction	Re – transplantation		
chemotherapy	Splenectomy		
Fungal	Alemutuzumab		
colonization	Local epidemiology		
Local	Diabetic ketoacidosis ^b		
epidemiology	Iron overload ^b		
Steroid	Diabetes mellitus ^b		
prophylaxis	Deferoxamine therapy ^b		
Neutrophil	skin breakdown ^b		
dysfunction			
Cytotoxic drugs			
Infliximab			
Alemtuzumab			
T- cell depletes			
stem – cell			
products			
CD34-selected			
stem cell products			
Diabetic			
ketoacidosis ^b			
Iron overload ^b			
Diabetes mellitus ^b			
Deferoxamine			
therapy ^b			
Skin breakdown ^b			

^aGvHD, Graft vs. host disease. ^bRelates to mucormycosis

3.2. Filamentous Fungi-Beyond Aspergillus

Other genera of filamentous fungi such as *Scedosporium* spp., *Fusarium* spp., *Paecilomyces* spp., dematiaceous fungi (e.g. *Alternaria* spp.), and the mucorales group (*Mucor* spp., *Rhizopus* spp., *Rhizomucor* spp., *Absidia* spp., and *Cunninghamella* spp.) are currently recognized as emerging opportunistic human pathogens (Shao *et al.*, 2002; Richardson and Lass-Florl, 2008; Cortez *et al.*, 2008; Nucci and Anaissie, 2007). Infections due to these opportunistic molds are usually marked by poor responses to antifungal therapy, *in vitro* resistance to most available antifungals, and an overall poor outcome with excessive mortality.

3.2.1. Scedosporium Species

Within the genus Scedosporium, Scedosporium apiospermum (teleomorph, Pseudallescheria boydii) and S. prolificans are ubiquitous filamentous fungi that live in soil, sewage, and polluted waters. Scedosporiosis represents a broad spectrum of clinical diseases caused by agents of the genus Scedosporium. Infections caused by these organisms can be localized, extend to the surrounding tissues, or disseminate to distant organs. The range of diseases caused by these fungi is broad, ranging from transient colonization of the respiratory tract to saprophytic involvement of abnormal airways, allergic bronchopulmonary reaction, and invasive localized disease. These infections occur mainly in the skin and soft tissues but could extend to tendons, ligaments, and bone (mycetoma). Septic arthritis. osteomyelitis, lymphocutaneous syndrome, pneumonia, endocarditis, peritonitis, chorioretinitis, and endophthalmitis are possible outcomes. In individuals who experience neardrowning accidents, P. boydii and S. apiospermum should always be considered in the differential diagnosis of any post-accident infections, especially if pneumonia or brain abscess ensues. Scedosporium apiospermum and S. prolificans, represent two medically-important antifungalresistant opportunistic pathogens. S. apiospermum causes mycetoma and deep-seated infections (e.g. CNS abscesses) and could disseminate in neutropenic bone marrow transplant (BMT) recipients and immunosuppressed individuals; crude mortality rate is about 55% (Cortez et al., 2008, Mellinghoff et al., 2002; Nesky et al., 2000; Perlroth et al., 2007). S. prolificans causes bone and soft tissue infections in immunocompetent individuals and deeply invasive and disseminated infections in immunocompromised patients with a crude mortality rate of 90% (Perlroth et al., 2007). Surgical resection remains the only definitive therapy for S. prolificans infections (Walsh et al., 2004; Cortez et al., 2008).

3.2.2. Fusarium Species

Like Aspergillus, Fusarium spp. are fungi with hyalinebranched septated hyphae (Richardson and Lass-Florl, 2008; Pfaller and Diekema, 2004 a; Cuenca-Estrella *et al.*, 2008; Caston-Osorio *et al.*, 2008; Nucci and Anaissie, 2007). Of all filamentous fungi, *Fusarium* spp. remain the second most common cause of invasive disease in immunosuppressed patients (Nucci, and Anaissie, 2007). Besides classical risk factors (neutropenia, GVHD, and immunosuppression), recent findings suggest that hospital water systems may play a significant role in the transmission of these pathogens (Nucci and Anaissie, 2007; Dignani and Anaissie, 2004; Anaissie et al., 2001). Furthermore, fusariosis frequency is higher in patients with hematological malignancies and in HSCT recipients (Nucci and Anaissie, 2007; Dignani and Anaissie, 2004). Clinical manifestations of fusariosis are more often characterized by cutaneous involvement and fungemia than those of Aspergillus spp. However, fusariosis cannot be always distinguished from IA with high enough confidence on the basis of clinical manifestations alone (Nucci and Anaissie, 2002). Typical presentation of disseminated fusariosis includes positive blood culture (up to 75%) and the appearance of multiple purpuric cutaneous nodules with central necrosis (Nucci and Anaissie, 2007; Nucci et al., 2004; Dignani and Anaissie, 2004; Walsh et al., 2004). Infections by Fusarium spp. usually associate with high mortality that is due in part to high rates of resistance to available antifungals.

3.2.3. Acremonium

Acremonium spp. are becoming increasingly recognized as opportunistic fungal pathogens. Maior predisposing factors for infection include prolonged corticosteroid therapy, splenectomy, and bone marrow transplantation with subsequent tacrolimus-dependent immunosuppression (Richardson and Lass-Florl, 2008; Pfaller and Diekema, 2004a). Following entry through penetrating injuries, they can cause foot mycetomas and corneal infections even in immunocompetent hosts. Greater than 35 cases of Acremonium-related infections have been described in adults (Schinabeck and Ghannoum, 2003) and >15 cases (excluding mycetoma and keratitis) have been documented in children (Miyakis et al., 2006). Among Acremonium spp., A. strictum is the most commonly identified species in children and adults. The presence of adventitious forms of A. strictum provides a mechanism for hematological spread and dissemination. Fungemias caused by A. strictum has been reported mainly in neutropenic patients (Schinabeck and Ghannoum, 2003).

3.2.4. Paecilomyces

Paecilomyces are cosmopolitan filamentous fungi that inhabit the soil, decaying plants, and food products. Member species are usually considered as contaminants; however, some can cause infection in humans and animals. The genus *Paecilomyces* contains several species including the emerging pathogens P. lilacinus and P. variotii (Richardson and Lass-Florl, 2008; Pfaller and Diekema, 2004a; Pastor and Guarro, 2006). P. lilacinus tends to cause devastating oculomycosis and other severe human infections (Pastor and Guarro, 2006). It usually shows low susceptibility to conventional antifungals and variable susceptibility to novel triazoles in vitro. Around 120 cases of human P. lilacinus infections have been reported between 1964 and 2004. Most of which were oculomycosis (51.3%) and cutaneous and subcutaneous infections (35.3%); the rest (13.4%) were miscellaneous infections. Direct cutaneous inoculation can lead to infections that involve a variety of human organ systems. Pulmonary and cutaneous infections, cellulitis, onychomycosis, otitis media, endocarditis, osteomyelitis,

and catheter-related fungemia have all been reported (Pastor and Guarro, 2006). Peritonitis and sinusitis are the most common infections caused by *P. variotii*.

Different infections are usually associated with varying sets of predisposing factors. In that, while oculomycosis associates with lens implantation, cutaneous and subcutaneous infections occur in SOT and BMT recipients, neutropenic and immunodeficient hosts, and patients Infections in apparently undergoing surgery. immunocompetent hosts have also been reported. The following reported case is cited to serve as an illustration of the predisposition to, infection by, and manifestation and diagnosis of P. lilacinus infections. A male of 56 years of age presented with a 2-month history of painful erythematous nodules over the right knee 12 months after receiving a liver transplant. Several biopsies yielded a mold that was initially (phenotypically) identified as a Penicillium, subsequent molecular sequence analysis however determined the etiologic agents to be P. lilacinus. Skin and soft tissue infections were the most common presentation (Pastor and Guarro, 2006; van Schooneveld et al., 2008; Pfaller and Diekema, 2004a). Surgical debridement combined with drug therapy or correction of the predisposing factor(s) is usually required for measurable improvement.

3.2.5. Trichoderma

Trichoderma spp. have traditionally been employed in the biotechnology industry as sources of enzymes and antibiotics. They have also been used in agriculture as plant growth promoters and biofungicides. However, mounting epidemiological data suggests that these previously nonpathogenic spp., are emerging as important opportunistic pathogens in immunocompromised patients and in patients undergoing peritoneal dialysis (Pfaller and Diekema, 2004a; Richardson and Lass-Florl, 2008). It is now recognized that fatal disseminated disease due to *Trichoderma longibrachiatum* occurs in patients with hematologic malignancies and in BMT or SOT transplant recipients (Chouaki *et al.*, 2002).

3.3. Mucormycosis

Mucormycosis (formerly zygomycosis) is the term used to describe a group of frequently lethal mold infections that have a predilection for diabetic patients, patients steroid therapy, and on severely immunocompromised hosts (such as HSCT recipients) (Bitar et al., 2009; Spellberg et al., 2005; Chayakulkeeree et al., 2006). The majority of human infections are due to fungi that mostly belong to the genera (or principal species) Rhizopus (R. arrhizus), Mucor (M. circinelloides), Rhizomucor (R. pusillus), Cunninghamella (C. bertholletiae), and Absidia (A. corymbifera). Despite the emergence of mucormycosis as a significant cause of mycosis, it remains much less frequent than other (more common) forms like invasive aspergillosis. The incidence figures are difficult to collect as few national studies have been undertaken; however, the annual incidence rate of 1.7 cases/million is the estimated figure in the US (Pfaller and Diekema, 2004a; Richardson and Lass-Florl, 2008; Bitar et al., 2009, Spellberg et al., 2005; Chayakulkeeree et al., 2006). Mucormycosis are generally acute and rapidly progressive with mortality rates of 70-100% (Gonzalez et *al.*, 2002). The molds that cause entomophthoramycosis (*Conidiobolus* spp. and *Basidiobolus* spp) also belong to the class Zygomycetes. They principally cause nasal, facial, and other subcutaneous infections, which may become persistent but rarely disseminate. Such infections are rarely encountered outside of West Africa, India, and Central and South America (Pfaller and Diekema, 2004a; Richardson and Lass-Florl, 2008; Spellberg *et al.*, 2005; Chayakulkeeree *et al.*, 2006).

Inhaled infectious spores may establish an infection in the sinuses; less common routes of acquisition include the intestinal tract (by ingestion) or the skin (through breaches). The fact that mucormycosis is less common than IA suggests that these pathogens possess fewer (and/or milder) virulence factors (Spellberg et al., 2005; Chayakulkeeree et al., 2006). A review of 929 cases of mucormycosis reported in the literature up to 2004 has indicated that a majority of the cases associate with type 2 diabetes mellitus, a sizable portion associate with unknown risk factors, and a minority of cases associate with malignancy (Roden et al. (2005). That said, the number of cases that associate with malignancy, HSCT, and intravenous drug abuse has been on the rise for over three Rhinocerebral mucormycosis was more decades. commonly associated with diabetes, whereas pulmonary infection occurred more often in those with malignancy. Multivariate analysis revealed that independent risk factors for increased mortality include disseminated infection with Cunninghamella spp. as the causative agent and renal failure Antifungal therapy and surgery were independently associated with decreased mortality risks (Rogers, 2008).

In a retrospective review of 15 patients with mucormycosis diagnosed at a non-oncology tertiary referral centre between 1999 and 2004 (Sims and Ostrosky-Zeichner, 2007), it was found that 9/16 episodes were associated with diabetes mellitus, whereas trauma, vascular disease, steroid therapy, and neutropenia constituted the rest of contributory conditions. Ten episodes were due to Rhizopus spp. and six were due to Mucor spp. Common sites of infection include wounds, rhinocerebrum, and pulmonary and peritoneal areas. The noticeably significant increase in the incidence of mucormycosis between 2000 and 2003 coincided with increased use of voriconazole; hence the possible link between drug overuse and predisposition to mucormycosis (Trifilio et al., 2007; Almyroudis et al., 2007).

3.4. Dematiaceous molds (Phaeohyphomycosis)

The long and taxonomically-diverse list of infections caused by dematiaceous (pigmented thick-walled) fungi are grouped under phaeohyphomycosis. Dematiaceous molds are characterized by the presence of a pale brown-dark melanin-like pigment in the cell wall. They may cause a variety of cutaneous and subcutaneous infections in immunocompetent hosts and invasive or disseminated infections in immunocompetent and immunocompromised hosts (Pfaller and Diekema, 2004a; Richardson and Lass-Florl, 2008; Caston-Osorio *et al.*, 2008; Negroni *et al.*, 2004; Revankar *et al.*, 2004). The number of dematiaceous molds being reported as etiologic agents of phaeohyphomycosis is growing; several of which target the nervous system (Caston-Osorio *et al.*, 2008;

Walsh et al., 2004; Revankar et al., 2004); hence the descriptive name "neurotropic fungi". Common neurotropic fungi include Cladophialophora bantiana, Bipolaris spicifera, Exophiala spp., Wangiella dermatitidis, Ramichloridium obovoideum, and Chaetomium atrobrunneum (Revankar et al., 2004). Brain abscess is the most common CNS presentation. However, Bipolaris spp. and Exerohilum rostratum infections may initially present as sinusitis to then extend into the CNS (Revankar et al., 2004; Yehia et al., 2004).

3.5. Histoplasmosis

Histoplasmosis or Darling's disease is pulmonary mycosis caused by the soil-inhabiting dimorphic fungus Histoplasma capsulatum. There are two varieties of H. capsulatum that are pathogenic to humans; H. capsulatum var. capsulatum and H. capsulatum var. duboisii. H. capsulatum var. farciminosum represents a third variety that is recognized as an equine pathogen (Kauffman, 2007; 2009). Although H. capsulatum var. capsulatum occurs in many different parts of the world, it is most commonly encountered in North and Central America and in Europe (Kauffman, 2007; 2009). H. capsulatum var. duboisii occurs in Africa; cases that have been reported in Europe were related to Africans visiting Europe for treatment purposes. In the United States, H. capsulatum is endemic in the Mississippi and the Ohio River valleys. It also exists in localized foci in many Middle Eastern countries. Soil containing large amounts of bird or bat guano supports the growth of these molds (Kauffman, 2009).

Humans acquire H. capsulatum infections during occupational or recreational activities in areas where the pathogen is highly endemic (disrupted soil, accumulated dirt and guano in old buildings and bridges, or in caves where bats roost) (Kauffman, 2007; 2009). Most individuals with histoplasmosis are asymptomatic; symptomatic episodes manifest within 3-17 days after exposure. Most affected individuals have clinically silent manifestations and show no apparent ill effects (Kauffman, 2007; 2009). The acute phase is characterized by nonspecific respiratory (cough or flu-like) symptoms. Chest X-ray findings are unremarkable in 40-70% of cases. In some cases, chronic histoplasmosis may resemble tuberculosis; disseminated histoplasmosis affects multiple organ systems and is often fatal unless treated. Severe infections can cause hepatosplenomegaly, lymphadenopathy, and adrenal enlargement. Leakage from scar tissues left on the retina following ocular histoplasmosis damages the retina and could result in loss of vision. Immunosuppressed patients and those unable to develop effective cell-mediated immunity against the organism are likely to manifest symptomatic disease during acute/disseminated episodes (table 5) (Kauffman, 2007; 2009).

Table 5. A tentative tally of the common risk factors for disseminated histoplasmosis

Risk factor
Age (infants)
AIDS
Hematologic malignancies
Solid organ transplant
Hematopoietic stem cell transplant
Immunosuppressive agents
Corticosteroids
Tumor necrosis factor antagonists
Congenital T-cell deficiencies
Gamma interferon receptor deficiency
Hyperimmunoglobulin M syndrome

4. Recent Advances in Fungal Species Identification

Correct and timely identification of fungal clinical isolates is an essential component in the management of patients with invasive fungal infections: this is particularly true for the immunocompromised and the critically-ill. Recent advances in fungal genomics is helping in this regard as PCR-based identification of clinical isolates is proving to be far superior as compared to conventional biochemical identification panels (e.g. API-20C-AUX, VITEK ID-YST, and so on). Pyrosequencing is а relatively inexpensive, extremely rapid DNA sequencing method that uses novel chemistry to sequence short (>70bp) fragments within pre-selected regions of the genome in question (Borman et al., 2010; Montero et al., 2009). Several studies suggest that pyrosequencing could be a very productive approach for the identification of medically important yeasts (Boyanton et al., 2008; Gharizadeh et al., 2004; Montero et al., 2008). Pyrosequencing of a short segment within the internal transcribed spacer 2 region (ITS2) was shown capable of accurately distinguishing C. glabrata from its close genetic relative C. nivariensis (Borman et al., 2008b). ITS2 pyrosequencing has also been reported capable of discriminating between C. parapsilosis, C. orthopsilosis, and C. metapsilosis (Borman et al., 2009). Another promising target region for comparative pyrosequencing of various Candida species is the D1-D2 segment of the nuclear 28S large rRNA gene (Andrew et al., 2008b; Borman et al., 2010). The utility of pyrosequencing in large-scale comparative studies aiming at distinguishing closely-related and disparate pathogenic fungi and at identifying rare yeast species has been demonstrated (Ghannoum et al., 2010; Borman et al., 2010; Montero et al., 2009). Employing a pyrosequencing approach, Ghannoum and co-workers (2010) were able to characterize the profile of the oral microbiome (mycobiome) in healthy subjects. The mycobiome characterized in their study consisted of 85 different fungal genera and 101 different fungal species.

5. Conclusion

Common human fungal infections are on the rise and fungal species that have been classically labeled as mildlypathogenic or nonpathogenic are emerging as serious pathogens. Coccidioidomycosis (Cox and Magee, 2004), paracoccidioidomycosis (Travassos et al., 2007), blastomycosis, and unusual fungal and pseudofungal infections (Pfaller and Diekema, 2005) are cases in point. This trend is strongly associated with improvements in disease management, improvements in the diagnosis of infections and infectious diseases, overuse/misuse of antifungals and antibiotics, and resistance to existing antifungal drugs. It is also aided by increased resistance to and limited efficacy of existing antifungal drugs. Slow progress in developing more effective and safer antifungals and the striking lack of fungal vaccines are not helping either. Therefore, use of definitive diagnostic procedures, rational application of available antifungals, and prudent management of patients at risk are the more imperative.

Acknowledgments

This work was supported –in part- by research grant (UOS-MH-120515) from the deanship of graduate studies and research, University of Sharjah, UAE. Khaled H. Abu-Elteen wishes to thank the Hashemite University, Jordan for help and support throughout this project.

References

Abu-Elteen KH. 2001. Increased incidence of vulvovaginal candidiasis caused by *Candida glabrata* in Jordan. *Jpn J Infect Dis.*, **54**: 103-107.

Abu-Elteen KH and Abu-Elteen RM. 1998. The prevalence of *Candida albicans* populations in the mouths of complete denture wearers. *New Microbiol.*, **21**:41-48.

Abu-Elteen KH and Hamad M. 2007. Novel antifungal therapies. In: Kevin Kavanagh, (Eds.), **New Insights in Medical Mycology**, Springer, The Netherlands, pp. 69-98.

Abu-Elteen KH, Hamad MA and Salah SA. 2006. Prevalence of oral *Candida* infections in diabetic patients. *Bahrain Med Bull.*, **28**: 12-17.

Almirante B, RodríguezD, Park BJ, Cuenca-Estrella M, Planes AM, Almela M, Mensa J, Sanchez F, Ayats J, Gimenez M, Saballs P, Fridkin SK, Morgan J, Rodriguez-Tudela JL, Warnock DW, Pahissa A, the Barcelona Candidemia Project Study Group . 2005. Epidemiology and predictors of mortality in cases of *Candida* bloodstream infection: results from population-based surveillance, Barcelona, Spain, from 2002 to 2003. *JClin Microbiol.*, **43**: 1829-1835.

Almyroudis NG, Sutton DA, Fothergill AW, Rinaldi MG and Kusne S. 2007. *In vitro* susceptibilities of 217 clinical isolates of zygomycetes to conventional and new antifungal agents. *Antimicrob Agents Chemother.*, **51**: 2587–2590. Anaissie EJ, Kuchar RT, Rex JH, Francesconi A, Kasai M, Müller F-M C, Lozano-Chiu M, Summerbell RC, Dignani MC, Chanock ST and Walsh TJ. 2001. Fusariosis associated with pathogenic *Fusarium* species colonization of a hospital water system: A new paradigm for the epidemiology of opportunistic mold infections. *Clin Infect Dis.*, **33**: 1871-1878.

Ando T, Moriya A and Shibuya K. 2008. Aspergillosis. *Nippon Rinsho*, **66**: 2345-2349.

Arendrup MC, Fuursted K, Gahrn-Hansen B, Jensen IM, Knudsen JD, Lundgren B, Schønheyder HC and Tvede M. 2005. Seminational surveillance of fungemia in Denmark: notably high rates of fungemia and numbers of isolates with reduced azole susceptibility. *J Clin Microbiol.*, **43**: 4434-4440.

Balajee SA, Gribskov JL, Hanley E, Nickle D and Marr KA. 2006. *Aspergillus lentulus* sp. nov., a new sibling species of *A. fumigatus*. *Eukaryot Cell*, **4**: 625-632.

Baradkar VP and Kumar S. 2008. Meningitis caused by *Rhodotorula mucilaginosa* in human immunodeficiency virus seropositive patient. *Ann Indian Acad Neurol.*, **11**: 245-247.

Bassetti M, Filippo A, Laura N, Emanuele M, Maria Pia M, Michele M, Barbara R, Franco BP, GiancarloIand Claudio V. 2009. Incidence of candidaemia and relationship with fluconazole use in an intensive care unit. *J Antimicrob Chemother.*, **64(3)**:625-629.

Beigi RH, Meyn LA, Moore DM, Krohn MA and Hillier SL. 2004. Vaginal yeast colonization in nonpregnant women: A Longitudinal Study. *Obstet Gynecol.*, **104**:926-930.

Bicanic T and Harrison TS. 2004. Cryptococcal meningitis. *British Med Bull.*, **72**: 99-118.

Bitar D, Van Cauteren D, Lanternier F, Dannaoui E, Che D, Dromer F, Desenclose GC and LortholaryO. 2009. Increasing incidence of zygomycosis (mucormycosis), France, 1997-2006. *Emerg Infect Dis.*, **15**: 1395-1401.

Binder U and Lass-Flörl C. 2011. Epidemiology of Invasive Fungal Infections in the Mediterranean Area. *Medit J Hemat Infect Dis.*, 3: e20110016, DOI 10.4084/MJHID.2011.016.

Borman A M, Petch R, Linton C J, Palmer M D, Bridge P D and Johnson E M. 2008. *Candida nivariensis*, an emerging pathogenic fungus with multidrug resistance to antifungal agents. *J Clin Microbiol.*, **46**:933–938.

BormanA M, Linton C J, Oliver D, Palmer M D, Szekely A, Odds F C and Johnson E M. 2009. Pyrosequencing analysis of 20 nucleotides of the internal transcribed spacer 2 discriminates *Candida parapsilosis, Candida metapsilosis, and Candida orthopsilosis. J Clin Microbiol.*, **47**:2307–2310.

BormanAM, Linton CJ, Oliver O, Palmer MD, Szekely A and Johnson EM. 2010. Rapid Molecular Identification of Pathogenic Yeasts by Pyrosequencing Analysis of 35 Nucleotides of Internal Transcribed Spacer 2. *J Clin Microbiol.*, **48(10)**: 3648–3653.

Bougnuox ME, Kac G, Aegerter Pand Fagan JY.2008. Candidaemia and candiduria in criticially ill patients admitted to intensive care units in France: incidence, modecular diversity, management and outcome. *Intensive Care Med*., **34**: 292-299. Bouza E and MunozP. 2004. Invasive infection caused by *Blastoschizomyces capitatus* and *Scedosporium* spp. *Clin Microbiol Infect.*, **10**(Suppl. 1):76-85.

Boyanton B L, Luna R A, Fasciano L R, Menne K Gand Versalovic J. 2008. DNA pyrosequencing-based identification of pathogenic *Candida* species by using the internal transcribed spacer 2 region. *Arch Pathol Lab Med.*, **132**:667–674

Casadevall A, Steenbergen JN and Nosanchuk JD. 2003. 'Ready made' virulence and ' dual use' virulence factors in pathogenic environmental fungi- the *Cryptococcus neoformans* paradigm. *Curr Opin Microbiol.*, **6**: 332-337.

Caston-Osorio JJ, Rivero A and Torre-CisnerosJ.2008. Epidemiology of invasive fungal infection. *Inter J Antimicrobial Agents*, **32** (Suppl. 2): 103-109.

Chayakulkeeree M, Ghannoum MA and Perfect JR.2006. Zygomycosis: the re-emerging fungal infection. *Eur J Microbiol Infect Dis*., **25**:215–229.

Chouaki T, Lavarde V, Lachaud L, RaccurtCP and Hennequin C. 2002. Invasive infections due to *Trichoderma* species: report of 2 cases, findings of in vitro susceptibility testing, and review of the literature. *Clin Infect Dis.*, **35**:1360-1367.

ChowJK, Golan Y, Ruthazerm R, Karchmer A, GarmeliY, Lichtenberg D, Chawla V, Young JA and Hadley S. 2008. Risk factors for albicans and non albicans candidaemia in the intensive care unit. *Crit Care Med.*, **36**, 1993-1998.

ClarkTA and Hajjeh RA. 2004. Recent trends in the epidemiology of invasive mycoses. *Curr Opin Infect Dis.*, **17**: 511-515.

Cornillet A,Camus C, Nimubona S, Gandemer V, Tattevin P, Belleguic C, Chevrier S, Meunier C, Lebert C, Aupée M, Caulet- Maugendre S, Faucheux M, Lelong B, Leray E, Guiguen C and Gangneux JP. 2006. Comparison of epidemiological, clinical, and biological features of invasive Aspergillosis in neutropenic and nonneutropenic patients: A 6Year Survey. *Clin Infect Dis.*, **43**: 577-584.

Cortez KJ, Roilides E, Quiroz-Telles F, Meletiadis J, Antachopoulos C, Knudsen T, Buchanan W, Milanovich J, Sutton DA, Fothergill A, RinaldiMG, Shea YR, Zaoutis T, Kottilil S and Walsh TJ. 2008. Infections caused by *Scedosporium* spp. *Clin Microbiol Rev.*, **21**: 157-197.

Cox RA and MageeDM. 2004. Coccidioidomycosis : Host response and vaccine development. *Clin Microbiol Rev.*, **17**: 804-839.

Cuenca-Estrella M, Bernal-Martinez L, Buitrago MJ, Castelli MV, Gomez-Lopez A, ZaragozaO and Rodriguez-Tudela JL.2008.Update on the epidemiology and diagnosis of invasive fungal infection. *Inter J Antimicrobial Agents*, **32** (Suppl.2): 143-147.

Davies AN, Brailsford SR and BeightonD. 2006. Oral candidiasis in patients with advanced cancer. *Oral Oncol.*, **42**:698-702.

De Almeida GMD, Costa SF, Melhem M, MottaAL, Szeszs MW, MiyashitaF, Pierrotti LC, Rossi F and Burattini MN. 2008. *Rhodotorula* spp. isolated from blood cultures: clinical and microbiological aspects. *Medical Mycology*, **46**:547-556. deLeon EM, Jacober SJ, Sobel JD and Foxman B. 2002. Prevalence and risk factors for vaginal Candida colonization in women with type 1 and type 2 diabetes. *BMC Infect Dis.*, **2**:1.

Denning D W. 1998. Invasive aspergillosis. *Clin Infect Dis.*, **26**:781-805.

Denning D W. 1995. Issues in the management of invasive aspergillosis. *Ann Med Interne.*, **146**:106-110.

Dignani MC and Anaissie E. 2004. Human fusariosis. *Clin Microbiol Infect.*, **10** (Suppl 1): 67-75.

Enoch DA, Ludlam HA, and Brown NM. 2006. Invasive fungal infections: a review of epidemiology and management options. *J Med Microbiol.*, **55**: 809-818.

Ferrer J. 2000. Vaginal candidiasis: epidemiology and etiological factors. *Intl J Gynecol Obstet.*, **71**: 521-527.

Flemming R V, Walsh TJ and Anaissie EJ. 2002. Emerging and less common fungal pathogens. *Infect Dis Clin N Am.*, **16**:915-933.

Fraser JA, Giles SS, Wenink EC, Geunes-Boyer SG, Wright JR, Diezmann S, Allen A, Stajichm JE, Dietrich FS, Perfect JR and Heitman J. 2005. Same-sex mating and the origin of the Vancouver Island *Cryptococcus gattii* outbreak. *Nature*, **437**: 1360-1364.

Fridkin SK, Kaufman D, Edwards JR, Shetty S and Horan T. 2006. Changing incidence of *Candida* bloodstream infections among NICU patients in the United States: 1995–2004. *Pediatrics*, **117**: 1680-1687.

Fung HB, Martyn CA, Shahidi A and Brown ST. 2008. *Rhodotorula mucilaginosa* lymphadenitis in an HIV- infected patient. *Inter J Infect Dis.*, **13**: e27-e29.

Gavalda J, Len O, San Juan R, AguadoJM, Fortun J, Lumbreras C, MorenoA, Munoz P, Blanes M, Ramos A, Rufi G, Gurgui M, Torre- CisnerosT, Montejo M, Cuenca- Estrella M, Rodriguez- Tudela JL and Pahissa A for RESITRA (Spanish Network for Research on Infection in Transplantation).2005. Risk factors for invasive Aspergillosis in solid organ transplant recipients: A case control study. *Clin Infect Dis.*, **41**: 52-59.

Gharizadeh B, Norberg E, Loffler J, Jalal S, Tollemar J, EinseleH, Klingspor L and Nyren P. 2004. Identification of medically important fungi by the Pyrosequencing technology. *Mycoses*, **47**:29–33.

Ghannoum MA, Jurevic RJ, Mukherjee PK, Cui F, Sikaroodi M, Naqvi M and Gillevet PM. 2010. Characterization of the oral fungal microbiome (Mycobiome) in healthy individuals. *PLoS Pathog*, **6(1):** 1000713. doi:10.1371.

Girmenia C, Pagano L, Martino B, D'Antonio D, Fanci R, Specchia G, Melillo L, Buelli M, Pizzarelli G, Venditti M, Martino P and the GIMEMA Infection Program. 2005. Invasive infections caused by *Trichosporon* species and *Geotrichum capitatum* in patients with hematological malignancies: a retrospective multicenter study from Italy and review of literature. *J Clin Microbiol.*, **43**: 1818-1828. Gonzalez C E, Rinaldi MG and Sugar AM. 2002. Zygomycosis. *Infect Dis Clin N. Am.*, **16**:895-914.

Hahner D, Kirschner R, Piepenbring M and Schofer H. 2008. First isolation of the anamorphic basidiomycetous *yeast Trichosporon faecale* in Germany, from the skin of a patient with tinea pedis. *Mycopathologia*, **165**: 149-153.

Hajjeh RA, Sofair AN, Harrison LH, Lyon GM, Arthington-Skaggs BA, Mirza SA, Phelan M, Morgan J, Lee-Yang W, Ciblak MA, Benjamin LE, Thompson Sanza L, Huie S, Yeo SF, Brandt ME and Warnock DW. 2004. Incidence of bloodstream infections due to *Candida* species and *in vitro* susceptibilities of isolates collected from 1998 to 2000 in a population-based active surveillance program. *JClin Microbiol.*, **42**: 1519-1527.

HamadM, Muta'ebE, Abu-Shaqra Q, Fraij A, Abu-Elteen K and Yasin SR. 2006. Utility of the oestrogen-dependent vaginal candidiasis murine model in evaluating the efficacy of various therapies against vaginal *Candida albicans* infection. *Mycoses*, **49**: 104-108.

Hamad M, Abu-Elteen KH and Ghaleb M. 2004. Estrogendependent induction of vaginal candidiasis in naïve mice. *Mycoses*, **47**: 304-309.

Hammarskjold F, Wallen G and Malmvall BE. 2006. Central venous catheter infections at a county hospital in Sweden: a prospective analysis of colonization, incidence of infection and risk factors. *Acta Anaesthesiol Scand.*, **50**: 451-460.

Hedayati MT, Pasqualotto AC, Warn PA, Bowyer P and Denning DW. 2007. *Aspergillus flavus*: human pathogen, allergen and mycotoxin producer. *Microbiology*, **153**: 1677-1692.

Hickey PW, Sutton DA, Fothergill AW, Rinaldi MG, Wickes BL, SchmidtHJ and Walsh TJ.2009. *Trichosporon mycotoxinivorans*: Anovel respiratory pathogen in patients with cystic fibrosis. *J Clin Microbiol.*, **47**: 3091-3097.

Hope W, Morton A and Eisen DP. 2002. Increase in prevalence of nosocomial non-*Candida albicans* candidaemia and the association of *Candida krusei* with fluconazole use. *J Hosp Infect.*, **50**: 56-65.

HowardSJ, Cerar D, Anderson MJ, Albarrag A, Fisher MC, Pasqualotto AC, Laverdiere M, Arendrup MC, Perlin DS and Denning DW. 2009. Frequency and evolution of azole resistance in *Aspergillus fumigatus* associated with treatment failure. *Emerg Infect Dis.*, **15**: 1068-1076.

Hsueh P-R, Teng L-J, Ho S-W and Luh K-T. 2003. Catheterrelated sepsis due *to Rhodotorula glutinis*. *J Clin Microbiol.*, **41**: 857-859.

Kauffman CA. 2007. Histoplasmosis: a clinical and laboratory update. *Clin Microbiol Rev.*, **20**: 115-132.

Kauffman CA. 2009. Histoplasmosis. *Clin Chest Med.*, **30**: 217-225.

Khawcharoenporn T, Apisarnthanarak A and MundyLM. 2007a. Treatment of cryptococcosis in the setting of HIV coinfection. *Expert Rev Anti Infect Ther.*, **5**: 1019-1030.

Khawcharoenporn T, Apisarnthanarak A and MundyLM. 2007b. Non-neoformans cryptococcal infections : a systematic review. *Infection*, **35**: 51-58. Kojic EM and Darouiche RO. 2004. *Candida* infections of medical devices. *Clin Microbiol Rev.*, **17**: 255-267.

Kontoyiannis DP, Torres HA and ChaguaM. 2004. Trichosporonosis in a tertiary care cancer center: risk factors, changing spectrum and determinants of outcome. *Scand J Infect Dis.*, **36**: 564-569.

Lass-Florl C, Griff K, Mayr A, Petzer A, Gastl G, Bonatti H, Freund M, Kropshofer G, Dierich M and Nachbaur D. 2005. Epidemiology and outcome of infections due to *Aspergillus terreus*: 10-year single centre experience. *Br J Haematol.*, **131**: 201-207.

Levitz SM, Nong S, Seetoo K, Harrison TS, Speizer R and Simons E. 1999. *Cryptococcus neoformans* resides in an acidic phagolysosome of human macrophages. *Infect Immun.*, **67**: 885-890.

Li H-M, Du H-T, Liu W, Wan Z and Li R-Y. 2005. Microbiological characteristics of medically important *Trichosporon* species. *Mycopathologia*, **160**: 217-225.

LindbergJ, Hagen F, Laurson A, Stenderup J and Boekhout T. 2007. *Cryptococcus gattii* risk for tourists visiting Vancouver Island, Canada. *Emerg Infect Dis.*, **13**: 178-179.

Liu X, Liu H, Guo Z and Luan W. 2006. Association of asymptomatic oral candidal carriage, oral candidiasis and CD4+ lymphocyte count in HIV-positive patients in China. *Oral Dis.*, **12**: 41-44.

Lo Re V,III, Fishman NO and Nachamkin I. 2003. Recurrent catheter- related *Rhodotorula rubra* infection. *Clin Microbiol Infect.*, **9**: 897-900.

Low CY and Rotstein C. 2011. Emerging fungal infections in immunocompromised patients. *F1000 Medicine Reports*, **3**:14, doi:10.3410/M3-14.

MacCallum D. (007. *Candida albicans*: New insights in infection, disease, and treatment. In: Kevin Kavanagh (Eds.). **New Insights in Medical Mycology.** Springer, The Netherlands, pp.99-129.

Makela P, Leaman D and Sobel JD. 2003. Vulvovaginal trichosporonosis. *Infect Dis Obst Gynecol.*,**11**: 131-133.

Malani A, HmoudJ, ChiuL, Carver PL, Bielaczyc A and Kauffman CA. 2005. *Candida glabrata* fungemia: experience in a tertiary care center. *Clin Infect Dis.*, **41**: 975-981.

Marr KA, Carter RA, Crippa F, Wald A and Corey L. 2002. Epidemiology and outcome of mold infections in hematopoietic stem cell transplant recipients. *Clin Infect Dis.*, **34**: 909-917.

Martino R, Salavert M, Parody R, Tomas JF, de la Camara R, VazquezL, Jarque I, Prieto E, Sastre JL, Gadea I, Peman J and Sierra J. 2004. *Blastoschizomyces capitatus* infection in patients with leukemia: report of 26 cases. *Clin Infect Dis.*, **38**:335-341.

Martino R, Subirá M, Rovira M, Solano C, Vázquez L, Sanz GF, Urbano-Ispizua A, Brunet S and De la Cámara R. 2002. Invasive fungal infections after allogeneic peripheral blood stem cell transplantation: incidence and risk factors in 395 patients. *Br J Haematol.*, **116**: 475-482.

Marty FM, Barouch DH, Coakley EP and Baden LR. 2003. Disseminated trichosporonosis caused by *Trichosporon loubieri*. J *Clin Microbiol.*, **41**: 5317-5320. Maschmeyer G, Haas A and Cornely OA. 2007. Invasive aspergillosis: epidemiology, diagnosis and management in immunocompromised patients. *Drugs*, **67**: 1567-1601.

Meersseman W, LagrouK, Maertens J and Van Wijngaerden E. 2007. Invasive Aspergillosis in the Intensive Care Unit. *Clin Infect Dis.*, **45**: 205-216.

Mellinghoff I K, Winston DJ, Mukwaya G and Schiller GJ. 2002. Treatment of *Scedosporium apiospermum* brain abscesses with posaconazole. *Clin Infect Dis.*, **34**:1648-1650.

Miceli MH, Díaz JA and Lee SA. 2011. Emerging opportunistic yeast infections. *Lancet Infect Dis.*, **11(2)**:142-51.

Middelhoven WG. 2003. Identification of clinically relevant *Trichosporon* species. *Mycoses*, **46**: 7-11.

Mirza SA, Phelan M and Rimland D. 2003. The changing epidemiology of cryptococcosis : an update from populationbased active surveillance in 2 large metropolitan areas,1992-2000. *Clin Infect Dis.*, **36**: 789-794.

Mitchell TG and Perfect JR. 1995. Cryptococcosis in the era of AIDS-100 years after the discovery of *Cryptococcus neoformans*. *Clin Microbiol Rev.*, **8**: 515-548.

Miyakis S, Velegraki A, Delikou S, Parcharidou A, Papadakis V, Kitra V, Papadatos I and Polychronopoulou S.2006. Invasive *Acremonium strictum* infection in a bone marrow transplant recipient. *Pediatr Infect Dis.*, **25**: 273–275.

Montero C I, Shea Y R, Jones P A, Harrington S M, Tooke N E, Witebsky F G and Murray P R. 2008. Evaluation of Pyrosequencing® technology for the identification of clinically relevant non-dematiaceous yeasts and related species. *Eur J Clin. Microbiol Infect Dis.*, **27**:821–830.

Morrow CA and Fraser JA. 2009. Sexual reproduction and dimorphism in the pathogenic basidiomycetes, *FEMS Yeast Res.*, **9(2):1**61-77.

Negroni R, Helou SH, Petri N, Robles AM, Arechavala A and Bianchi MH. '2004. Case study: posaconazole treatment of disseminated phaeohyphomycosis due to *Exophiala spinifera*. *Clin Infect Dis.*, **38**:e15-e20.

Nesky M A, McDougal C and Peacock JE Jr. 2000. *Pseudallescheria boydii* brain abscess successfully treated with voriconazole and surgical drainage: case report and literature review of central nervous system pseudallescheriosis. *Clin Infect Dis.*, **31**:673-677.

Nishiura Y, Nakagawa-Yoshida K, Suga M, ShinodaT, Gueho E and Ando M. 1997. Assignment and serotyping of *Trichosporon* species: the causative agents of summer-type hypersensitivity pneumonitis. *J Med Vet Mycol.*, **35**: 45-52.

Nivoix Y, Velten M, Letscher-Bru V, Moghaddam A, Natarajan-Ame S, Fohrer C, Lioure B, Bilger K, Lutun P, Marcellin L, Launoy A, Freys G, Bergerat J-P and Herbrecht R. 2008. Factors associated with overall and attributable mortality in invasive aspergillosis. *Clin Infect Dis.*, **47**: 1176-1184.

Nucci M and Anaissie E. 2002. Cutaneous Infection by *Fusarium* species in healthy and immunocompromised hosts: Implications for diagnosis and management. *Infect Dis.*, **35**: 909-920.

Nucci M and Anaissie E. 2007. *Fusarium* infections in immunocompromised patients. *Clin Microbiol Rev.*, **20**:695-704.

Nucci M, Marr KA, Queiroz- Telles F, Martins CA, Trabasso P, Costa S, Voltarelli JC, Colombo AL, Imhof A, Pasquini R, Maiolino A, Souza CA and Anaissie E. 2004. *Fusarium* infection in hematopoietic stem cell transplant recipients. *Clin Infect Dis.*, **38**: 1237-1242.

Padhye A, Verghese S, Ravichandran P, Balamurugan G, Hall L, Padmaja P and Fernandez MC. 2003. *Trichosporon loubieri* infection in a patient with adult polycystic kidney disease. *J Clin Microbiol.*, **41**: 479-482.

Pagano L, Caira M, Nosari A, Van Lint MT, Candoni A, Offidani M, Aloisi T, Irrera G, Bonini A, Picardi M, Caramatti C, Invernizzi R, Mattei D, Melillo L, de Waure C, Reddiconto G, Fianchi L, ValentiniCG, Girmenia C, Leone G and Aversa F. 2007. Fungal infections in recipients of hematopoietic stem cell transplants: results of the SEIFEM B 2004 Study—Sorveglianza Epidemiologica Infezioni Fungine Nelle Emopatie Maligne. *Clin Infect Dis.*, **45**: 1161-1170.

Pappas PG, Perfect JR, Cloud GA, Larsen RA, Pankey GA, Lancaster DJ, HendersonH, Kauffman CA, Haas DW, Saccente M, Hamill RJ, Holloway MS, WarrenR M and Dismukes WE. 2001. Cryptococcosis in human immunodeficiency virus- negative patients in the era of effective azole therapy. *Clin Infect Dis.*, **33**: 690-699.

Pasqualotto AC, Rosa DD, Medeiros LR and Severo LC. 2006. Candidaemia and cancer: patients are not all the same. *BMC Infect Dis.*, **6**: 50-56.

Pastor FJ and Guarro J. 2006. Clinical manifestations, treatment and outcome of *Paecilomyces lilacinus* infections. *Clin Microbiol Infect.*,**12**: 948–960.

Patterson TF. 2005. Advances and challenges in management of invasive mycoses. *Lancet*, **366**: 1013-1025.

Peman J, Bosch M, Canton E, Viudes A, Jarque I, Gomez-Garcia M, Garcia-MartinezJM and Gobernado M. 2008. Fungemia due to *Candida guilliermondii* in a pediatric and adult population during a 12-year period. *Diag Microbiol Infect Dis.*, **60**: 109-112.

Perlroth J, Choi B and Spellberg B. 2007. Nosocomial fungal infections: epidemiology, diagnosis, and treatment. *Med Mycol.*, **45**: 321-346.

Pfaller MA and Diekema DJ. 2004 a. Rare and emerging opportunistic fungal pathogens: concern for resistance beyond *Candida albicans* and *Aspergillus funigatus*. J Clin Microbiol., **42**: 4419-4431.

Pfaller MA and Diekema DJ. 2004 b. Twelve years of fluconazole in clinical practice: global trends in species distribution and fluconazole susceptibility of bloodstream isolates of *Candida*. *Clin Microbiol Infect.*, **10** (Suppl.1):11-23.

Pfaller MA and Diekema DJ. 2005. Unusual fungal and pseudofungal infections of humans. *J Clin Microbiol.*, **43**: 1495-1504.

Pfaller MA and Diekema DJ. 2007. Epidemiology of invasive candidiasis: a persistent public health problem. *Clin Microbiol Rev.*, **20**: 133-163.

Pfaller MA, Pappas PG and Wingard JR. 2006. Invasive fungal pathogens: current epidemiological trends. *Clin Infect Dis.*, **43** (Suppl.1) S3-14.

Pirotta MV and Garland SM. 2006. Genital *Candida* species detected in samples from women in Melbourne, Australia, before and after treatment with antibiotics. *J Clin Microbiol.*, **44**:3213-4217.

Qazzafi Z, Thiruchunapalli D, Birkenhead D, Bell D and Sandoe JA. 2007. Invasive *Cryptococcus neoformans* infection in an asplenic patient. *J Infect.*, **55**:566-568.

Ray D, Goswami R, Banerjee U, Dadhwal V, Goswami D, Mandal P, Sreenivas V and Kochupillai N. 2007. Prevalence of *Candida glabrata* and its response to boric acid vaginal suppositories in comparison with oral fluconazole in patients with diabetes and vulvovaginal candidiasis. *Diabetes Care*, **30**: 312-317.

Revankar S G, Sutton DA and Rinaldi MG. 2004. Primary central nervous system phaeohyphomycosis: a review of 101 cases. *Clin Infect Dis.*, **38**:206-216.

Richardson, M and Lass-Florl C. 2008. Changing epidemiology of systemic fungal infections. *Clin Microbiol Infect.*, **14** (Suppl. 4): 5-24.

Riedel DJ, Johnson JK and Forrest GN. 2007. *Rhodotorula glutinis* fungemia in a liver-kidney transplant patient. *Transplant Infect Dis.*, **10**: 197-200.

Roden MM, Zaoutis TE, Buchanan WL, Knudsen TA, Sarkisova TA, Schaufele RL, Sein M, Sein T, Chiou CC, Chu JH, Kontoyiannis DP and WalshTJ. 2005. Epidemiology and outcome of zygomycosis: a review of 929 reported cases. *Clin Infect Dis.*, **41**:634–653.

Rogers T. 2008. Treatment of zygomycosis: current and new options. *J Antimicrob Chemother.*, **61**(Suppl 1):35-39.

Rosenhagen M, FeldhuesR, Schmidt J, Hoppe-Tichy T and Geiss HK. 2009. A risk profile for invasive aspergillosis in liver transplant recipients. *Infection*, **37**: 313-319.

Sánchez-VargasLO, Ortiz-LópezNG, Villar M, Moragues MD, Aguirre JM, Cashat-Cruz M, Lopez-Ribot JL, Gaitán-Cepeda LA and Quindós G. 2005. Oral *Candida* isolates colonizing or infecting human immunodeficiency virus-infected and healthy persons in Mexico. *J Clin Microbiol.*, **43**: 4159-4162.

Schinabeck MK and Ghannoum MA. 2003. Human hyalohyphomycoses: a review of human infections due to *Acremonium* spp., *Paecilomyces* spp., *Penicillium* spp., and *Scopulariopsis* spp. *J Chemother.*, **15** (suppl 2): 5–15.

ShaoP-L, Huang L-M and HsuehP-R. 2007. Recent advances and challenges in the treatment of invasive fungal infections. *Inter Antimicrob Agents*, **30:** 487-495.

Shinde RS, Mantur BG, Patil G, Parande MV and Parande AM. 2008. Meningitis due to *Rhodotorula glutinis* in an HIV infected patients. *Indian J Med Microbiol.*, **26**: 375-377.

Sims CR and Ostrosky-Zeichner L. 2007. Contemporary treatment and outcomes of zygomycosis in a non-oncologic tertiary care center. *Arch Med Res.*, **38**:90–93.

Singh Nand PatersonDL. 2005. *Aspergillus* infections in transplant recipients. *Clin Microbiol Rev.*, **18**: 44-69.

Soni Nand Wagstaff A. 2005. Fungal infection. *Curr Anaesthesia and Crit Care*, **16**: 231-241.

Spellberg B, Edwards J Jr and Ibrahim A. 2005. Novel perspectives on mucormycosis: pathophysiology, presentation, and management. *Clin Microbiol Rev.*, **18**:556–569.

Steinbach WJ, Benjamin DK Jr, Kontoyiannis DP, Perfect JR, Lutsar I, Marr KA, Lionakis MS, Torres HA, Jafri H and Walsh TJ. 2004. Infections due to *Aspergillus terreus*: A multicenter retrospective analysis of 83 cases. *Clin Infect Dis.*, **39**: 192-198.

Subramanian S and Mathai D. 2005. Clinical manifestations and management of cryptococcal infection. *J Postgrad Med.*, **51**(Suppl 1): 21-26.

Sugita T, Nishikawa A, Ikeda R, Sakashita H, Sakai Y and Yoshizawa Y. 1998. First report of *Trichosporon ovoides* isolated from the home summer-type hypersensitivity pneumonitis patient. *Microbiol Immunol.*, **42**: 475-478.

Thakur K, SinghG, Agarwal S and Rani L. 2007. Meningitis caused by *Rhodotorula rubra* in an human immunodeficiency virus infected patient. *Indian J Med Microbiol.*, **25**: 166-168.

Tortorano AM, Kibbler C, Peman J, Bernhardt H, Klingspor L and Grillot R. 2006. Candidaemia in Europe: epidemiology and resistance. *Int J Antimicrob Agents*, **27**: 359-366.

Travassos LR, Goldman G, TabordaC and Puccia R. 2007. Insights in *Paracoccidioides barasiliensis* pathogenicity. In: Kevin Kavanagh, (Eds). **New Insights in Medical Mycology**, Springer, The Netherlands, pp.241-265.

Trifilio SM, Bennett CL, Yarnold PR, McKoy JM, Parada J, Mehta J, Chamilos G, PalellaF, Kennedy L, Mullane K, Tallman MS, Evens A, Scheetz M H, Blum W and Kontoyiannis DP. 2007. Breakthrough zygomycosis after voriconazole administration among patients with hematologic malignancies who receive hematopoietic stem-cell transplants or intensive chemotherapy. *Bone Marrow Transplant*, **39**: 425–429.

Tuon FF, de Almeida GM and Costa SF. 2007. Central venous catheter-associated fungemia due to *Rhodotorula* spp. a systematic review. *Med Mycol*., **45**: 441–447.

UptonA and MarrAK. 2006. Emergence of opportunistic mold infections in the hematopoietic stem cell transplant patient. *Curr Infect Dis Rep.*, **8**: 434-441.

Upton A, Kirby KA, Carpenter P, Boeckh M and Marr KA. 2007. Invasive aspergillosis following hematopoietic cell transplantation: Outcomes and prognostic factors associated with mortality. *Clin Infect Dis.*, **44**: 531-540.

van SchooneveldT, Freifeld A, Lesiak B, Kalil A, Sutton AD and IwenPC. 2008 *Paecilomyces lilacinus* infection in a liver transplant patient: case report and review of the literature. *Transpl Infect Dis.*, **10**:117-122.

Vardakaz KZ, Michalopoulos A, Kiriakidou KG, Siampl EP, Samonis G and Falagas M E. 2009. Candidaemia: incidence, risk factors, characteristics and outcomes in immunocompetent critically ill patients. *Clin Microbiol Infect.*, **15**: 289-292. Verweij P E and Denning DW. 1997. Diagnostic and therapeutic Yehia

strategies for invasive aspergillosis. *Respir Crit Care Med.*, **18**:203-215.

Walsh T J, Groll A, Hiemenz J, Flemming R, Roilides E and Anaissie E. 2004. Infections due to emerging and uncommon medically important fungal pathogens. *Clin Microbiol Infect.*, **10**(Suppl. 1):48-66.

Wisplinghoff H, Bischoff T, Tallent SM, Seifert H, Wenzel RP and Edmond MB. 2004. Nosocomial bloodstream infections in US hospitals: Analysis of 24,179 cases from a prospective nationwide surveillance study. *Clin Infect Dis.*, **39**: 309-317.

Yehia M, Thomas M, Dilmore H, Van der Merwe W and Dittner I. 2004. Subcutaneous black fungus (phaeohyphomycosis) infection in renal transplant recipients: three cases. *Transplantation*, **77**: 140-142.

Zaas AK, Boyce M, Schell W, Alexander lodge B, Miller JL and Perfect JR. 2003. Risk of fungemia due to *Rhodotorula* and antifungal susceptibility testing of *Rhodotorula* isolates. *J Clin Microbiol.*, **41**: 5233-5235.