

Evaluation of the Reproductive Toxicity of Dietary Fumonisin B₁ in Rats

Francis A. Gbore^{1,*}, Tolu J. Owolawi¹, Magdalene Erhunwunsee¹, Olatunbosun Akele² and Ruth A. O. Gabriel-Ajobiewe²

¹Department of Environmental Biology and Fisheries, ²Department of Microbiology, Adekunle Ajasin University, Akungba-Akoko, Nigeria

Received 21st February 2012; accepted 20th April 2012

Abstract

The toxicity of dietary fumonisin B₁ (FB₁), a mycotoxin from the common maize fungus *Fusarium verticillioides*, on serum gonadotropins, ovarian histopathology, and fertility were examined in female Wistar rats. Thirty-nine female rats were randomly assigned to two test diets containing 10.0 and 20.0 mg FB₁/kg and a control diet. After 14 days of feeding, blood samples were obtained by intracardiac puncture from 4 rats in each treatment for gonadotropins evaluation, and were then killed by cervical dislocation to collect samples of ovaries for histopathology. Also, each of the remaining nine females in each treatment was mated to one healthy adult male rat. The serum LH concentrations of rats fed diets containing 10 and 20 mg FB₁/kg were significantly lower ($P < 0.05$) than those fed the control diet, while the serum FSH level of rats fed diet containing 20 mg FB₁/kg only was significantly lower ($P < 0.05$) compared with the controls. Dietary FB₁ however failed to induce histopathological changes in the ovaries of the rats. Fertility, gestation lengths and foetal weights of the rats decreased significantly ($P < 0.05$) with increased dietary FB₁. The concentration of daily dietary FB₁/kgBW of 1.74 in this study is less than five times the estimated probable daily fumonisin intake of 355 µg/kg BW for person eating 'mouldy' maize in the high oesophageal cancer area of the Transkei region, South Africa. The apparent significant "safety factor" of about five times relative to human FB₁ exposure may be a cause for concern in areas where maize is a dietary staple.

Keywords: Fertility, Fumonisin B₁, Gonadotropins, Mycotoxin, Rats, Toxicity.

1. Introduction

Mycotoxins are natural contaminants of cereals and other food commodities throughout the world and they significantly impact human and animal health. Animals, as well as humans, are exposed to mycotoxins through consumption of contaminated diets, which can be considered the gateway to cases of natural intoxication by these fungal secondary metabolites (Gutema, 2000; Hennigen, 2000).

The economic consequences of mycotoxin contamination are profound, and exposure of people and livestock to mycotoxin-contaminated foods is particularly a serious problem in the tropics (Reddy and Raghavender, 2008). According to Lawlor and Lynch (2001), 25% of the global crop is contaminated with mycotoxins. Crops with large amounts of mycotoxins often have to be destroyed. Alternatively, contaminated crops are sometimes diverted into animal feeds. Giving contaminated feeds to susceptible animals poses a serious threat to the health and productivity of the animals and cause great economic losses (Griessler and Encarnaç o, 2009), by acting directly

or indirectly on fertility. Reproductive inefficiency is recognized as the most costly limiting constraint to efficient animal production as reproduction is the bedrock of animal production (Gbore, 2009; Ewuola and Egbunike, 2010). This makes the assessment of the effects of mycotoxins on livestock reproduction a unique challenge in an effort to improve livestock production.

Fusarium verticillioides (Sacc) Nirenberg (= *F. moniliforme* Sheld.), one of the most prevalent mycotoxigenic fungi reported to be associated with dietary staples such as maize intended for human and animal consumption throughout the world (Nelson *et al.*, 1991; Kedera *et al.*, 1992), produces the mycotoxin, fumonisin. *F. verticillioides* is present in virtually all maize samples (Marasas *et al.*, 2001). Maize is the major cereal utilized in the formulation of livestock feeds in several parts of the world, hence, the potential for fumonisins to be found in feeds and feedstuffs is high. Several naturally occurring fumonisins are known; FB₁ has been reported to be the most abundant and most toxic which represents approximately 70% of the total concentration in naturally contaminated foods and feeds, followed by fumonisins B₂ (FB₂) and B₃ (Murphy *et al.*,

* Corresponding author. e-mail: fgbore@yahoo.com.

1993; Norred, 1993). Consequently, toxicological studies on the fumonisins have been concentrated on FB₁.

The carcinogenicity, hepatotoxicity and mutagenicity as well as the effects on feed intake, live weight gain and blood abnormalities of fumonisin in animals have been well documented (Marasas *et al.*, 1988; Harrison *et al.*, 1990; Kellerman *et al.*, 1990; Gelderblom *et al.*, 1991, 1994; Colvin and Harrison, 1992; Voss *et al.*, 1998; Ewuola and Egbunike, 2008; Ewuola *et al.*, 2008; Gbore and Egbunike, 2008, 2009; Gbore, 2009). However, studies on depression of fertility and reproductive processes in rats by the toxin are rare.

Flynn *et al.* (1996) reported growth inhibition of rat embryos exposed *in vitro* to FB₁ on gestation day (GD) 9.5. Although useful as screens, *in vitro* methods allow direct foetal exposure and avoid maternal gastrointestinal absorption, pharmacokinetics, placental transfer, and other potential barriers of *in utero* exposure. *In vivo* assessment of the effects of fumonisin is therefore necessary.

Consumption of lesser amounts of fungal toxins at levels below those that cause overt toxicity may result in impaired fertility and decreased reproduction in animals. In a preliminary study, Gbore and Olorunfemi (2009) reported significant concentration-dependent decline conception rates in rabbits exposed to dietary fumonisin prior to mating. Fumonisin inhibit sphingolipids metabolism, and a variety of biological activities for sphingolipids have been reported (Wang *et al.*, 1992). It was therefore hypothesized that fumonisin could alter the release of gonadotropins from the pituitary because the brain contains high levels of sphingolipids. Studies on the alterations of gonadotropins and subsequent fertility in rats exposed to dietary FB₁, to our understanding, are scarce. It is essential, therefore, to assess the gonadotrophic and ovarian histopathological effects of *F. verticillioides*-contaminated maize-based diets and subsequent fertility and reproductive processes in rats.

2. Materials and Methods

2.1. Experimental Site and Animals

Eight weeks old male and female Wistar rats (*Rattus norvegicus*) obtained from a commercial breeder of Wistar rats in Benin City, Nigeria were used. Females weighed 168.92 ± 1.41 g. Male rats (202 ± 2.21 g) were used as sires only and were not exposed to dietary fumonisin. Thirty-nine mature females were individually housed under standard housing conditions (22°C, light/dark cycle 12/12 hours) in wire mesh rat cages at the Animal House of the Department of Biochemistry, Adekunle Ajasin University, Akungba Akoko, Nigeria. Further laboratory analyses were carried out at the Department of Chemical Pathology of the University College Hospital, and Department of Veterinary Pathology, all of the University of Ibadan, Ibadan, Nigeria. This study was approved by the local Institutional Animal Ethics Committee and was performed in accordance with "Guide for the care and use of Laboratory Animals" (National Research Council, 1996).

2.2. Fumonisin B₁ Production and Experimental Diets

Maize grits in 500 g quantities were placed into autoclavable polypropylene bags and soaked with 200 ml of distilled water for 2 h, then autoclaved for 1 h at 121°C and 120 kPa. The autoclaved maize grits were then cultured with a toxigenic strain of *F. verticillioides* (MRC 286) obtained from the Plant Pathology Laboratory of the International Institute of Tropical Agriculture (IITA), Ibadan, Nigeria to produce FB₁ as described previously (Nelson *et al.*, 1994). Uncultured maize grits and the cultured maize grits were used to formulate three diets. Samples of homogeneously mixed diets were quantified in replicates for FB₁ and other common *Fusarium* mycotoxins including deoxynivalenol (DON, vomitoxin), T-2 toxin, and zearalenone using mycotoxin quantitative CD-ELISA test kits (Neogen, Lansing, MI, USA) and reconfirmed by using HPLC analyses as described by Shephard *et al.* (1990). The concentrations of FB₁ in the diets were adjusted to 0.2, 10.0 and 20.0 mg/kg constituting diet 1 (= control diet, which had no *Fusarium*-contaminated maize grits), diets 2 (medium FB₁-contaminated diet) and 3 (high FB₁-contaminated diet), respectively. The concentrations of all other common *Fusarium* mycotoxins screened were below the detection limit of 0.2mg/kg for the toxins. The dietary FB₁ doses in this study were based on a preliminary dose-range-finding study by Collins *et al.* (1998a). Lower doses were selected for this study, the highest dose being 20 mg/kg. The pelleted diets provided ~20% crude protein, 5% crude fibre and 2.9 kcal of digestible energy/g.

2.3. Experimental Model

After 3 weeks of physiological adjustment period, the rats were randomly allocated to each of the three diets (n = 13 rats per treatment). The rats were provided with fresh clean water and appropriately weighed feed daily, and the weights of feed portions given and left uneaten after 24 h were determined. The body weight was determined weekly on a weighing scale (Ohaus Corp., Pine Brook, NJ, USA) with a precision of 0.05 g. The body weight gain of each rat was determined weekly as the weight difference in comparison to the weight in the previous week.

After 14 days of feeding, blood samples were obtained by intracardiac puncture from 4 rats from each treatment and were killed by cervical dislocation to collect samples of ovaries. Blood samples were collected into vacutainer tubes, covered and centrifuged at 4000 rpm for 10 minutes. The separated sera were decanted and deep-frozen for serum gonadotropins analyses at the Chemical Pathology Unit of the University College Hospital, Ibadan, Nigeria.

2.4. Evaluation of Reproductive Performance

To evaluate reproductive performance, after 2 weeks of feeding the respective experimental diets, each of the remaining nine females in each treatment was placed in a cage with one healthy adult male. The animals were kept together overnight, and then separated the following morning. Immediately after each separation, a vaginal smear examination was carried out to determine if sexual intercourse had occurred. When intercourse was positive (presence of spermatozoa in the vaginal smear), the night/day routine was discontinued and the female was housed individually during the estimated period of

gestation. At parturition, percentage of fertility with respect to the number of positive smears, gestation length, number of litters, mean litter size, number of live foetus per litter, and foetal weights were determined.

2.5. Examination of Ovaries

For examination by light microscopy, ovary samples collected were fixed in 10% neutral buffered formalin (pH 7.2) before dehydration in ten changes in ethanol of different concentrations ranging from 70 to 100% at 1-hr interval. After dehydration, the tissues were cleared in two changes of chloroform before infiltration and embedding in molten wax (60°C) for 12 h. Thereafter, the tissues were blocked in paraffin wax and later sectioned using a microtome. Paraffin sections (4µm) of the ovary samples were stained with haematoxylin and eosin. Photomicrographs were taken with a Zeiss Axiophot instrument using Kodak Plus-X pan (PX 135 to 24) film.

2.6. Determination of Serum Gonadotropins

Serum levels of gonadotropins; follicle-stimulating hormone (FSH) and luteinizing hormone (LH), were measured using a commercial ELISA kit (Habersham, Buckinghamshire, UK). All samples were run in duplicate in a single assay.

2.7. Statistical Evaluation

Data from this study were analyzed by one-way analysis of variance procedure of SAS (2001). The treatment means were compared using the Duncan procedure of the same software and results giving *P* values of <0.05 were considered significantly different.

3. Results

3.1. Feed Consumption and Body Weight Gain

Daily observation through the 35 days did not indicate detectable alterations in the general state of any of the animals. The average food consumptions were 17.64, 16.03, and 18.77 g for rats on diets 1, 2, and 3 respectively. Significant (*P*<0.05) differences in feed consumption and final weights were not observed (Figures 1 and 2), although the mean weights of the dams given 10 and 20 mg FB₁/kg (216 ± 17.6 to 218 ± 15.0 g) were 2.5 - 3.4% lower than that of the controls (223.56 ± 11.0 g) on day 35. Based upon feed consumption and body weight data, the diets provided 0.08, 0.74 and 1.74 mg FB₁/kg BW per day to the rats on diets 1, 2, and 3, respectively.

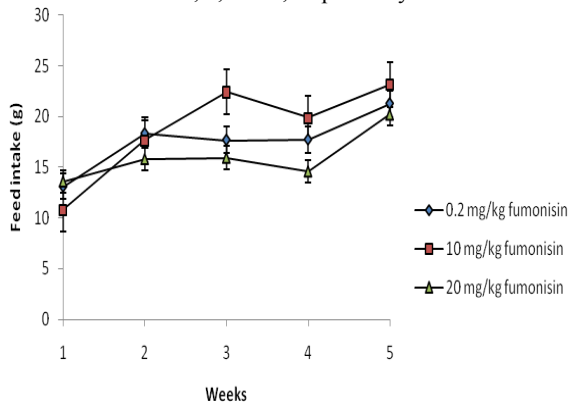


Figure 1. fed intake of rats fed different concentrations of dietary fumonisin B₁.

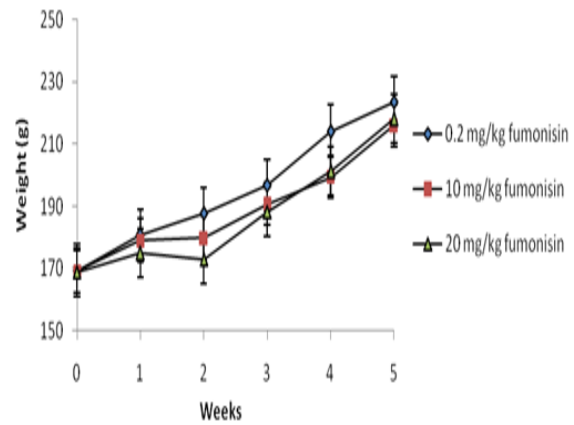


Figure 2. weights of rats fed different concentrations of dietary fumonisin B₁.

3.2. Serum Gonadotropins Levels

Dietary FB₁ significantly depressed serum gonadotropins in rats (Table 1). The serum LH concentrations of rats fed diets containing 10 and 20 mg FB₁/kg were significantly lower (*P*<0.05) than those fed the control diet. Similarly, the serum FSH levels of rats fed diet containing 20 mg FB₁/kg were significantly lower (*P*<0.05) than those fed the control diet and diet containing 10 mg FB₁/kg.

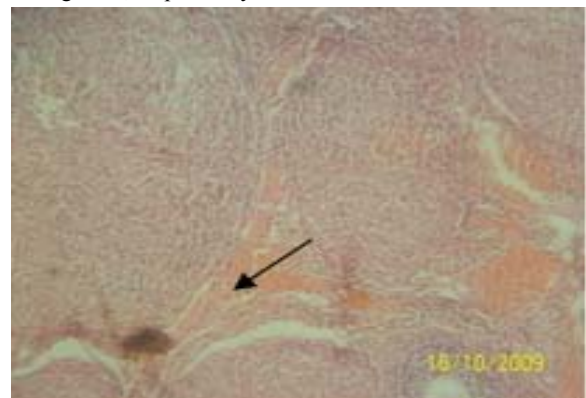
Table 1. Serum Gonadotropins (ng/ml) of Rats Exposed to Different Concentrations of Dietary Fumonisin B₁ (Mean ± SEM)

Parameters	Dietary fumonisin B ₁ concentrations (mg/kg)		
	0.2 (Control Diet)	10.0 (Diet 1)	20.0 (Diet 2)
Luteinizing Hormone	23.00 ± 0.01 ^a	10.90 ± 2.50 ^b	10.03 ± 2.80 ^b
Follicle Stimulating Hormone	215.00 ± 21.01 ^a	214.90 ± 20.07 ^a	199.80 ± 22.16 ^b

^{ab}: Means on same row with different superscripts differ significantly (*P* < 0.05)

3.3. Histopathology of the Ovaries

The histopathological examination of the ovaries showed no modification by the dietary FB₁ concentrations. No visible lesion was observed in the ovaries (Figures 3a, b and c) for the control rats, those fed 10 and 20 mg FB₁/kg diets, respectively.



a

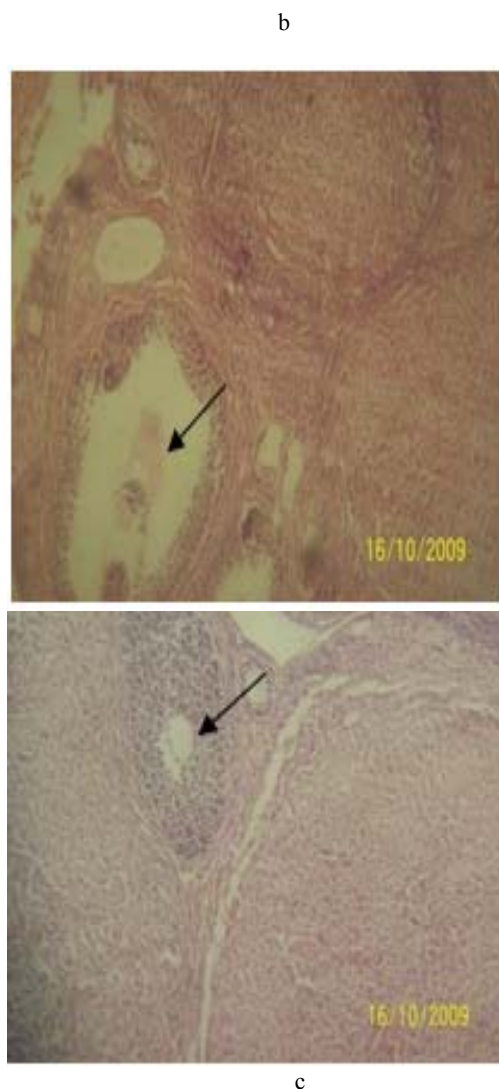


Figure 3. Haematoxylin-and-eosine stained sections of rat ovaries exposed to diets containing different concentrations of dietary FB_1 . No visible lesion was observed in the ovaries of rats fed diets containing different concentrations of FB_1 as shown in Figures 3a, b and c for the control animals, those fed 10 and 20 mg/kg dietary FB_1 respectively. Note the oocytes (arrowed). Mag. X 200.

3.4. Fertility Assessment

The summary of the reproductive indices of female rats exposed to different concentrations of dietary FB_1 are shown in Table 2. The results revealed that fertility, gestation length and foetal weight/litter were dose-dependent. These parameters decreased with increased dietary FB_1 , which were significant ($P < 0.05$) at both feed contamination levels. Out of the 9 rats mated in each group (as determined using the vaginal smears), only 5 had pups in the group fed diet containing 20 mg FB_1 /kg as against 8 and 7 for rats fed the control diet and diet containing 10 mg FB_1 /kg, respectively. No statistically significant difference was found across the treatment groups for number of live foetus/litter. The total number of pups obtained from the dams resulted in relative number of pups/dam of 6.78 for those fed the control diet as against 6.11 and 3.78 for those fed 10 and 20 mg FB_1 /kg diet, respectively.

Table 2. Reproductive Performance of Rats Exposed to Different Concentrations of Dietary Fumonisin B_1 (Mean \pm SEM)

Parameters	Dietary fumonisin B_1 concentrations (mg/kg)		
	0.2 (Control Diet)	10.0 (Diet 1)	20.0 (Diet 2)
Fertility (%)	88.89 \pm 33.33 ^a	77.78 \pm 44.10 ^b	55.56 \pm 52.70 ^c
Gestation length (days)	21.60 \pm 0.53 ^a	23.33 \pm 0.51 ^b	23.00 \pm 0.74 ^b
No of live foetus/litter	7.67 \pm 0.33	8.00 \pm 0.33	7.25 \pm 0.38
Foetal weight/litter (g)	5.33 \pm 0.01 ^a	5.23 \pm 0.04 ^b	5.22 \pm 0.04 ^b
No of litters	8	7	5
Total no of live foetus	61	55	34
No of dead foetus	0	1	2
Rel.* no of live foetus/dam	6.78	6.11	3.78

*Relative to total no of rats with positive scrapes in each treatment.

^{abc}: Means on same row with different superscripts differ significantly ($P < 0.05$).

4. Discussion

Although, initial studies using laboratory animal (Voss *et al.*, 1996; LaBorde *et al.*, 1997; Collins *et al.*, 1998a, b) provided no evidence that FB_1 is teratogenic. However, more recent observations and experimental findings have again drawn attention to FB_1 as a possible risk factor for birth defects (Merrill *et al.*, 2001, Marasas *et al.*, 2004, Voss *et al.*, 2006). In this present study, the decrease in fertility of 77.78 and 55.56% for the rats fed diets 2 and 3 respectively compared with 88.89% for the control rats revealed adverse effect of the dietary fumonisin on fertility processes in the rats. The total number of pups obtained from the 9 mated female rats in each treatment showed that the 34 pups obtained from rats fed 20 mg/kg dietary FB_1 was only 55.74% of the pups obtained from the controls. This is quite significant in breeding programmes. Effects of fumonisin on developing foetuses were expressed most often by foetal death and resorption (Floss *et al.*, 1994). Reduced weight gain is often the earliest indicator of maternal illness in developmental toxicity studies, and would be expected as a sign of maternal effect. While the cause of foetal deaths in rats exposed to diets containing 10 and 20 mg FB_1 /kg was not evident in this study, it was clear that the dose-dependent significant decline in foetal weight observed in this study was as a result of developmental toxicity produced by FB_1 and not secondary to maternal toxicity. In a study (LaBorde *et al.*, 1997), the observed reduced foetal weight from rabbits gavaged daily on GD 3 – 19 with purified FB_1 at 0.5 – 1 mg/kg/day was ascribed to maternal toxicity, rather than any developmental toxicity produced by FB_1 . Dose-responsive significant decrease in foetal weight observed in this study correlates with reports of other studies. Pregnant rats dosed by gavage on GD 8 – 12 with a

semipurified extract of culture material containing FB₁ with a purity of 80% resulted in lower foetal weight at dose of 60 mg/kg (Lebepe-Mazur *et al.*, 1995). Similarly, decreased body weight of live foetuses obtained from pregnant Syrian hamsters gavaged with FB₁ in a dose-dependent manner was reported in a study (Penner *et al.*, 1998). Voss *et al.* (1996) reported lower litter weights from rats fed *F. moniliforme* culture material providing 1 – 55 ppm FB₁ from two weeks before mating compared to the control group.

The dose-dependent significant decrease in fertility observed in this study may be due to an increase in gonadal steroid inhibition or suppression of the hypothalamus and/or pituitary gland resulting in a decline in serum FSH and LH levels. These two gonadotropins are the most important regulatory hormones of ovarian and uterine function (Everett, 2006). Suppression in secretion of hypothalamic gonadotropin releasing hormone (GnRH) causes reduced secretion of LH and FSH from pituitary (Rai *et al.*, 2004). Fumonisin is structurally similar to sphinganine and sphingosine and inhibit sphingosine metabolism in tissues, leading to an accumulation of sphingoid bases, which are intermediates in sphingolipid biosynthesis (Wang *et al.*, 1991). Because the brain contains high levels of sphingolipids, the disruption of sphingosine metabolism was speculated to be the mechanism behind the degeneration of neuronal cells seen in equine leukoencephalomalacia (Wang *et al.*, 1991). A variety of biological activities for sphingolipids have been reported (Wang *et al.*, 1992). Alterations in the amounts of any of these by fumonisins could potentially result in a variety of biological and pathological effects (Penner *et al.*, 1998). These may be responsible for the significant decline in serum gonadotropins resulting in reduced fertility with increased dietary FB₁ observed in this study. The mechanisms involve is not clear further investigations are therefore necessary to elucidate the mechanism.

The elongated gestation lengths observed in this study further leads credibility to the fact that FB₁ potentially affect reproductive development. As in this study, seemingly longer gestation length from 21.9 ± 0.35 days was observed in rats fed control diet to 22.7 ± 1.00 days in rats fed *F. moniliforme* culture material providing 10 ppm FB₁ from two weeks before mating (Voss *et al.*, 1996). Adverse effects of mycotoxins on sexual and reproductive developments have been reported. Green *et al.* (1990) and Rainey *et al.* (1990) observed that 1.5 mg zearalenone/kg diet disturbed the hypothalamo-hypophyseal function of prepubertal gilts, but after withdrawal of the contaminated diets, the animals attained puberty without delay, and their fertility was unimpaired. Also, dietary zearalenone levels as low as 0.05–0.06 mg/kg DM have been shown to increase the number of ovarian follicles and to decrease the serum concentration of the gonadotropic hormone FSH in female piglets (Döll *et al.*, 2003), thus potentially affecting their sexual development. Recently, dietary FB₁ have been reported to delay attainment of sexual maturity in growing pigs (Gbore, 2009) and rabbits (Ewuola and Egbunike, 2010).

Impaired action of FSH and LH on the ovary has as a primary consequence, a concomitant alteration in the capacity of this organ to synthesize ovarian reproductive hormones; mainly estrogens from follicular cells and

progesterone from luteal cells (Everett, 2006). Since the histopathological examination of the ovary did not reveal any lesion, it is suggested that the decline in fertility rates could not have resulted from impaired ovaries but impaired action of the gonadotropins on the organs.

The data from this study suggest that dietary FB₁ of ≥10 mg /kg significantly reduced serum gonadotropin levels and lowered fertility without adverse effect on the ovarian histology of rats. The concentration of daily dietary FB₁/kgBW (1.74) which resulted in significant decline in fertility of female rats in this study is less than five times the estimated probable daily fumonisin intake of 355 µg/kg BW (Gelderblom *et al.*, 1996) for person eating ‘mouldy’ maize in the high oesophageal cancer area of the Transkei region, South Africa. The apparent significant “safety factor” of about five times relative to human FB₁ exposure may be a cause for concern in areas where maize is a dietary staple. However, further studies on whether the toxin disrupts the hypothalamic production or release of GnRH or the absorbed FB₁ acts directly on the pituitary, at doses that are not maternally toxic, to lower serum gonadotropins are warranted.

Acknowledgements

The authors wish to thank Dr. R. Bandyopadhyay (Plant Pathologist) of the International Institute for Tropical Agriculture (IITA), Ibadan, Nigeria, and Mr. E. A. Agunloye of the University Health Services, Adekunle Ajasin University, Akungba-Akoko, Nigeria for their technical assistance in the study.

References

- Collins TF, Shackelford ME, Sprando RL, Black TN, LaBorde JB, Hansen DK, Eppley RM, Trucksess MW, Howard PC, Bryant MA, Ruggles DI, Olejnik N and Rorie JI. 1998a. Effects of fumonisin B₁ in pregnant rats. *Food Chem Toxicol.*, **36**: 397-408.
- Collins TF, Sprando RL, Black TN, Shackelford ME, LaBorde JB, Hansen DK, Eppley RM, Trucksess MW, Howard PC, Bryant MA, Ruggles DI, Olejnik N and Rorie JI. 1998b. Effects of fumonisin B₁ in pregnant rats. Part 2. *Food Chem Toxicol.*, **36**: 673-685.
- Colvin BM and Harrison LR. 1992. Fumonisin-induced pulmonary edema and hydrothorax in swine. *Mycopathologia*, **117**: 79-82.
- Döll S, Dänicke S, Ueberschaer KH, Valenta H, Schnurrbusch U, Ganter M, Klobasa F and Flachowsky G. 2003. Effects of graded levels of Fusarium toxin contaminated maize in diets for female weaned piglets. *Arch Anim Nutr.*, **57**: 311-334.
- Everett JW. 2006. Pituitary and hypothalamus: perspectives and overview. In: Knobil E and Neill JD, editors. **Knobil and Neill's Physiology of Reproduction**. 3rd Edition. London: Academic Press Elsevier Inc, pp. 1289-307.
- Ewuola EO and Egbunike GN. 2008. Haematological and serum biochemical response of growing rabbit bucks fed dietary fumonisin B₁. *Afr J Biotechnol.*, **7**: 4304-4309.
- Ewuola EO and Egbunike GN. 2010. Effects of dietary fumonisin B₁ on the onset of puberty, semen quality, fertility rates and testicular morphology in male rabbits. *Reproduction*, **139**: 439-445.
- Ewuola EO, Gbore FA, Ogunlade JT, Bandyopadhyay R, Niezen J and Egbunike GN. 2008. Physiological response of rabbit bucks to

- dietary fumonisin: Performance, haematology and serum biochemistry. *Mycopathologia*, **165**: 99-104.
- Floss JL, Casteel SW, Johnson GC, Rottinghaus GE and Krause GF. 1994. Developmental toxicity of fumonisin in Syrian hamsters. *Mycopathologia*, **128**: 33-38.
- Flynn TJ, Pritchard D, Bradlaw JA, Eppley R and Page S. 1996. *In vitro* embryotoxicity of fumonisin B₁ evaluated with cultured postimplantation staged rat embryos. *In vitro Toxicol*, **9**: 271-279.
- Gbore FA. 2009. Growth performance and puberty attainment in growing pigs fed dietary fumonisin B₁. *J Anim Physiol Anim Nutr.*, **93**: 761-767.
- Gbore FA and Egbunike GN. 2008. Testicular and epididymal sperm reserves and sperm production of pubertal boars fed dietary fumonisin B₁. *Anim Reprod Sci*, **105**: 392-397.
- Gbore FA and Egbunike GN. 2009. Toxicological evaluation of dietary fumonisin B₁ on serum biochemistry of growing pigs. *J Cent Eur Agric*, **10**: 255-262.
- Gbore FA and Olorunfemi OS. 2009. Reproductive performance of rabbit does fed dietary fumonisin. Proceedings of 14th Annual Conference of Animal Science Association of Nigeria (ASAN). LAUTECH Ogbomoso, Nigeria, pp. 234-236.
- Gelderblom WCA, Kriek NPJ, Marasas WFO and Thiel PG. 1991. Toxicity and carcinogenicity of the *Fusarium moniliforme* metabolite, fumonisin B₁, in rats. *Carcinogenesis*, **12**: 1247-1251.
- Gelderblom WCA, Cawood ME, Snyman SD and Marasas WFO. 1994. Fumonisin B₁ dosimetry in relation to cancer initiation in rat liver. *Carcinogenesis*, **15**: 209-214.
- Gelderblom WCA, Snyman SD, Abel S, Lebepe-Mazur S, Smuts CM, Van der Westhuizen L, Marasas WFO, Victor TC, Knasmuller S and Huber W. 1996. Hepatotoxicity and carcinogenicity of the fumonisins in rats: a review regarding mechanistic implications for establishing risk in humans. *Advan Experiment Med Biol*, **392**: 279-296.
- Green ML, Diekman MA, Malayer JR, Scheidt AB and Long GG. 1990. Effect of prepubertal consumption of zearalenone on puberty and subsequent reproduction of gilts. *J Anim Sci*, **68**: 171-178.
- Griessler K and Encarnaçao P. 2009. Fumonisin - mycotoxins of increasing importance in fish. *Aquacult Asia Magazine*, XIV (2): 24-26.
- Gutema T, Munimbazi C and Bullerman LB. 2000. Occurrence of fumonisins and moniliformin in corn and corn-based food products of U.S. origin. *J Food Protect*, **63**: 1732-1737.
- Harrison LR, Colvin BM, Greene JT, Newman LE and Cole JR, Jr. 1990. Pulmonary edema and hydrothorax in swine produced by fumonisin B₁, a toxic metabolite of *Fusarium moniliforme*. *J Vet Diagn Invest*, **2**: 217-221.
- Hennigen MR, Sanchez S, Di Benedetto NM, Longhi A, Torroba JE and Valente Soares LM. 2000. Fumonisin levels in commercial corn products in Buenos Aires, Argentina. *Food Addit Contam*, **17**: 55-58.
- Kedera CJ, Leslie JF and Claflin LE. 1992. Systemic infection of corn by *Fusarium moniliforme*. (Abstr.) *Phytopathology*, **82**: 1138.
- Kellerman TS, Marasas WFO, Thiel PG, Gelderblom WCA, Cawood M and Coetzer JAW. 1990. Leukoencephalomalacia in two horses induced by oral dosing of fumonisin B₁. *Onderstepoort J Vet Res*, **57**: 269-275.
- LaBorde JB, Terry KK, Howard PC, Chen JJ, Collins FX, Shackelford ME and Hansen DK. 1997. Lack of embryotoxicity of fumonisin B₁ in New Zealand white rabbits. *Fundam Appl Toxicol*, **40**: 120-128.
- Lawlor PG and Lynch PB. 2001. Mycotoxins in pig feeds – 1: Source of toxins, prevention and management of mycotoxicosis. *Irish Vet J*, **54**: 117-120.
- Lebepe-Mazur S, Bal H, Hopmans E, Murphy P and Hendrich S. 1995. Fumonisin B₁ is fetotoxic in rats. *Vet Human Toxicol*, **37**: 126-130.
- Marasas WFO, Jaskiewicz K, Venter FS and VanSchalkwyk DJ. 1988. *Fusarium moniliforme* contamination of maize in oesophageal cancer areas in Transkei. *South Afr Med J*, **74**: 110-114.
- Marasas WFO, Miller JD, Riley RT and Visconti A. 2001. Fumonisin—occurrence, toxicology, metabolism and risk assessment. In: Summerell BA, Leslie JF, Backhouse D, Bryden WL and Burgess LW, editors. Paul E. Nelson Memorial Symposium. St. Paul, Minn: APS Press. pp. 332-359.
- Marasas WF, Riley RT, Hendricks KA, Stevens VL, Sadler TW, Gelineau-van Waes J, Missmer SA, Cabrera J, Torres O, Gelderblom WC, Allegood J, Martinez C, Maddox J, Miller JD, Starr L, Sullards MC, Roman AV, Voss KA, Wang E and Merrill AH, Jr. 2004. Fumonisin disrupt sphingolipid metabolism, folate transport, and neural tube development in embryo culture and in vivo: a potential risk factor for human neural tube defects among populations consuming fumonisin-contaminated maize. *J Nutr*, **134**: 711-716.
- Merrill AH Jr, Sullards MC, Wang E, Voss KA and Riley RT. 2001. Sphingolipid metabolism: roles in signal transduction and disruption by fumonisins. *Environ Health Perspect*, **109** (Suppl 2): 283-289.
- Murphy PA, Rice LG and Ross PF. 1993. Fumonisin B₁, B₂ and B₃ content of Iowa, Wisconsin, and Illinois corn and corn screenings. *J Agric Food Chem*, **41**: 263-266.
- National Research Council Institute of Laboratory Animal Resources, 1996. **Guide for the Care and Use of Laboratory Animals**. Washington, DC: National Academy Press.
- Nelson PE, Plattner RD, Shackelford DD and Desjardins AE. 1991. Production of fumonisins by *Fusarium moniliforme* strains from various substrates and geographic areas. *Appl Environ Microbiol*, **57**: 2410-2412.
- Nelson PE, Juba JH, Ross PF and Rice LG. 1994. Fumonisin production by *Fusarium* species on solid substrates. *J Assoc Of Anal Chem Int* **77**: 522-524.
- Norred WP. 1993. Fumonisin: mycotoxins produced by *Fusarium moniliforme*. *J Toxicol Environ Health*, **38**: 309-328.
- Penner JD, Casteel SW, Pittman L, Jr., Rottinghaus GE and Wyatt RD. 1998. Developmental toxicity of purified fumonisin B₁ in pregnant Syrian hamsters. *J Appl Toxicol*, **18**: 197-203.
- Rai J, Pandey SN and Srivastava RK. 2004. Testosterone hormone level in albino rats following restraint stress of long duration. *J Anat Soc India*, **53**: 17-19.
- Rainey MR, Tubbs RC, Bennett LW and Cox NM. 1990. Prepubertal exposure to dietary zearalenone alters hypothalamo-hypophysial function but does not impair postpubertal reproductive function of gilts. *J Anim Sci*, **68**: 2015-2022.
- Reddy BN and Raghavender CR. 2008. Outbreaks of fusarial-toxicoses in India. *Cereal Res Commun*, **36** (Suppl. B): 321-325.
- SAS Institute Inc. 2001. SAS/STAT User's Guide. Version 8 for Windows. SAS Institute Inc., SAS Campus Drive Cary, NC, USA.
- Shephard GS, Sydenham EW, Thiel PG and Gelderblom WCA. 1990. Quantitative detection of FB₁ and B₂ by HPLC with fluorescence detection. *J Liq Chromatogr*, **12**: 2077-2078.
- Voss KA, Bacon CW, Herbert RA, Chapin RE, Chamberlain WJ, Plattner RD and Merdith FI. 1996. Studies on the reproductive

effects of *Fusarium moniliforme* culture material in rats and the biodistribution of [¹⁴C] fumonisin B₁ in pregnant rats. *Nat Toxins*, **4**: 24-33.

Voss KA, Riley RT, Bacon CW, Meredith FI and Norred WP. 1998. Toxicity and sphinganine levels are correlated in rats fed fumonisin B₁ (FB₁) or hydrolysed FB₁. *Environ Toxicol Pharmacol*, **5**: 101-104.

Voss KA, Gelineau-van Waes JB and Riley RT. 2006. Fumonisin: current research trends in developmental toxicology. *Mycotox Res*, **22**, 61-68.

Wang E, Norred WP, Bacon CW, Riley RT and Merrill AH, Jr. 1991. Inhibition of sphingolipid biosynthesis by fumonisins. Implications for diseases associated with *Fusarium moniliforme*. *J Biol Chem*, **266**: 14486-14490.

Wang E, Ross PF, Wilson TM, Riley RT and Merrill AH, Jr. 1992. Increases in serum sphingosine and sphinganine and decreases in complex sphingolipids in ponies given feed containing fumonisins, mycotoxins produced by *Fusarium moniliforme*. *J Nutr*, **122**: 1706-1716.

