# Glutathione as Potential Target for Cancer Therapy; More or Less is Good? (Mini-Review)

Maher Y. Abdalla

Department of Biological Sciences and Biotechnology, The Hashemite University, P.O. Box 150459, Zarqa 13115, Jordan Received 3 April 2011; received in revised form 11 May 2011; accepted 13 May 2011

#### Abstract

Glutathione (GSH) plays important roles in antioxidant defense, nutrient metabolism, and regulation of cellular processes, including cell differentiation, proliferation and apoptosis. Glutathione deficiency contributes to oxidative stress, which plays a key role in aging and pathogenesis of many diseases, one of which is cancer. The GSH content of cancer cells is associated with multidrug and radiation resistance. Just as low intracellular GSH levels decrease cellular antioxidant capacity, elevated GSH levels generally increase antioxidant capacity and resistance to oxidative stress, and this phenomenon is observed in may cancer cells as compared to normal ones. The present review will address the following questions: what is cancer-glutathione relation? Can glutathione play a role in treating or preventing cancer?

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Keywords: Glutathione, antioxidant, cancer, prevention.

## 1. Introduction

It is amazing how a tripeptide composed of cysteine, glutamic acid and glycine can be of this importance for cellular function. Glutathione (L-g-glutamyl-L-cysteinylglycine) is the principal tripeptide thiol involved in the antioxidant cellular defense (Clark et al., 1984; Vojislav et al., 2001; Ganesaratnam et al., 2004). The most two important structural features of GSH are:  $\gamma$ -glutamyl linkage and sulphydryl group (–SH). It is the thiol of cysteine residue that composes the active group (Figure 1) (Kaplowitz et al., 1985).

GSH is a tripeptide produced by the liver and is able to detoxify the lungs, RBCs, liver and the intestinal tract; it also removes a wide range of toxins, including those produced by heavy metals, cigarette smoke, alcohol, radiation and cancer chemotherapy. Glutathione neutralizes oxygen molecules before they cause damage to cells. It is found in two forms: free or bound to proteins. Free form is present mainly in its reduced form (GSH), which can be converted to the oxidized form (GSSG) during oxidative stress, and can be reverted to the reduced form by the action of the enzyme glutathione reductase (Ames 1989; Ames et al., 1993) (Figure 2).

In normal conditions, the GSH concentrations in mammalian cells can range between 1 and 10 mM, with the reduced GSH predominating over the oxidized form (Hassan and Fridovich, 1980). Maintaining optimal GSH:GSSG ratios in the cell are critical for survival, and a deficiency of GSH can result in oxidative damage. This ratio exceeds 100 in a normal resting cell, whereas in various models of oxidative stress, this ratio was reported to decrease to values between 10 and 1 (Hassan and Fridovich, 1980).

Glutathione is synthesized in the cell by the sequential actions of  $\gamma$ -glutamylcysteine synthetase (GCS) and glutathione synthetase (GS) in a series of six-enzyme-catalysed reactions (Meister and Anderson, 1983). This review will highlight the importance of GSH homeostasis in cancer therapy.

#### 2. Reactive Oxygen Species and Human Diseases

Due to different roles of reactive oxygen species (ROS) in cell signaling and many human pathological processes, imbalance of GSH is observed in a wide range of pathologies, including cancer, neurodegenerative disorders, cystic fibrosis (CF), HIV, and aging (Townsend and Tew, 2003; Ganesaratnam et al., 2004; Ken et al., 2004; Hayes et al., 2005). Maintaining proper GSH levels and oxidation state are important for cell function and their disruptions are observed in many human diseases. GSH deficiency leads to an increased susceptibility to oxidative stress and, thus, progression of many disease states (Townsend and Tew, 2003; Ganesaratnam et al., 2004; Ken et al., 2004; Hayes et al., 2005). On the other hand, elevated GSH levels increase antioxidant capacity and resistance to oxidative stress and this is observed in many types of cancer (Townsend and Tew, 2003; Ganesaratnam et al., 2004; Ken et al., 2004; Hayes et al., 2005).

Free radicals produced by normal cellular metabolism can lead to extensive damage to DNA, protein, and lipid (Olinski *et al.*, 1992; Okamoto *et al.*, 1994; Devi *et al.*, 2000; Wu *et al.*, 2004).

<sup>\*</sup> Corresponding author: maher\_abdalla@yahoo.com

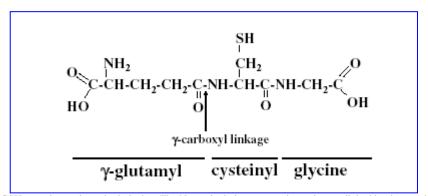


Figure 1. Structure of GSH or γ-glutamylcysteinyl glycine. The N-terminal glutamate and cysteine are linked by the γ-carboxyl group of glutamate (Kaplowitz, *et al.*, 1985).

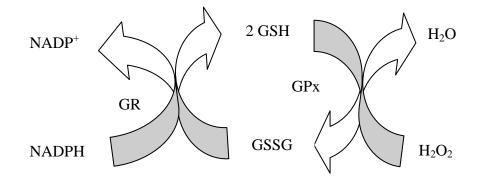


Figure 2. Pathway of ROS clearance (Adopted from Droge, 2002). Oxidants such as H<sub>2</sub>O<sub>2</sub> is converted to H<sub>2</sub>O by the action of GPX (or Catalase) using GSH. Regeneration of GSH requires NADPH and GR enzyme.

GR=glutathione reductase

GPx=glutathione peroxidase

DNA accumulates oxidative damage induced by ROS generated by endogenous and exogenous sources. This damage is a major contributor to diseases such as cancer, heart disease, cataracts, brain dysfunction, and aging (Ames, 1989; Ames, Shigenaga et al., 1993). It is estimated that the number of oxidative hits to DNA per cell per day is around 100,000 in the rat and 10,000 in the human (Ames, Shigenaga et al., 1993). It is possible that oxidative lesions in endogenous mammalian DNA exceeds 100 different types, of which 8-hydroxyguanine (8-oxoG) is one of the most abundant (Ames, 1989; Ames, Shigenaga et al., 1993). In normal functional cells, DNA repair enzymes efficiently remove most of the lesions formed by ROS. Several different methods are used to remove any mutation or mismatch (Demple and Harrison, 1994). In 2008, Petta et al., have shown the role of human DNA polymerase iota in protecting cells against oxidative stress (Petta et al., 2008). However, increased ROS generation in cancer cells leads to the accumulation of oxidative products of DNA, proteins, and lipids in tissues, and their release into the blood and urine. DNA oxidative products (8-oxoG), and lipid peroxidation have been detected in many cancer tissues, such as colorectal adenocarcinomas, mammary ductal carcinomas, renal cell carcinoma, and blood samples from leukemia patients (Olinski et al., 1992; Okamoto et al., 1994; Devi et al., 2000; Wu et al., 2004).

#### 3. GSH and Cancer

Many types of cancer cell have increased levels of free radicals and ROS compared with their normal counterparts (Toyokuni, Okamoto *et al.*, 1995; Kawanishi, Hiraku *et al.*, 2006). However, several studies using primary cancer tissues have revealed increased levels of ROS-scavenging enzymes and antioxidant compounds (Goodwin and Baylin, 1982; Oltra *et al.*, 2001). This increase could be a result of an adaptive response to intrinsic ROS stress.

While GSH is important in the detoxification of carcinogens, its elevated state in many types of tumors may also increase resistance or alters the cytotoxicity of many chemotherapy drugs or radiation (Clark *et al.*, 1984; Vojislav *et al.*, 2001; Ganesaratnam *et al.*, 2004). One example is human fibroblast tumor cell lines which has higher levels of cellular GSH than did normal human fibroblasts (Goodwin and Baylin, 1982; Carney *et al.*, 1983). This increased GSH may be an important factor in chemo- or radiotherapy resistance seen in these cells (Yu and Brown, 1984; Guichard *et al.*, 1986).

Manipulation of intracellular GSH using drugs such as 2-oxothiazolidine-4-carboxylate (OTZ) (a compound that stimulates GSH synthesis (Williamson *et al.*, 1982) or L-buthionine-(S,R)-sulfoximine (BSO) (a compound that inhibits GSH synthesis (Griffith *et al.*, 1979)) has been used to increase the sensitivity of different tumor cell lines to therapy and showed that selective differential

chemotherapy responses of normal versus tumor cells is possible (Griffith *et al.*, 1979; Williamson *et al.*, 1982).

It has been shown that manipulating intracellular oxidant status of tumor cells can be of clinical value. Increasing ROS or decreasing free radical scavengers such as GSH was shown to be toxic to tumor cells. Weydert *et. al.*(2008) have shown that combining GSH depletion using 1,3-Bis(2-chloroethyl)-1-nitrosourea (BCNU) chemotherapy with superoxide dismutase (SOD) gene therapy could be extremely successful in the treatment of breast cancer. Simon *et al.* (2007) have also shown that BSO sensitizes cancer cells to chemotherapy agents. Combining agents that induce mitochondrial dysfunction, such as AZT, and GSH depletion with BSO causes significant toxicity in head and neck cancer. However, it's important to note that different cells respond differently to oxidative stress inducing therapies (Mattson *et al.*, 2009).

Other factors might play an important role in the GSH therapy mechanism and should be considered when using GSH manipulation drugs. One important factor is a group of transferases enzymes called glutathione-S-transferases (GSTs). Elevated levels of GST in many tumor cell types have been demonstrated to limit the effectiveness of chemotherapy (Blair et al., 1997; Cullen et al., 2003). Moreover, GSTs have been associated with multidrug resistance of tumor cells, and over expression of GSTs can increase susceptibility to carcinogenesis and inflammatory disease (Townsend and Tew, 2003; Ganesaratnam et al., 2004; Ken et al., 2004; Hayes et al., 2005). One mechanism by which chemotherapy resistance may occur is by gene amplification of GST(s). It has been shown that over expression of the gene products of GST- $\pi$ , can provide a tumor cells with survival advantage relative to normal cells. High GST- $\pi$ , expression was associated with poor overall survival and may be associated with a more aggressive phenotype in head and neck squamous cell carcinoma (Shiga et al., 1999; Ulrike et al., 2002; Cullen et al., 2003).

### 4. GSH level, more or less is better?

GSH is involved in a variety of cellular functions such as DNA repair, cell cycle, regulation of cell signaling and transcription factors (Arrigo, 1999). It has been reported that GSH can modulate the activity of multiple stress genes which act to regulate the genes of cell proliferation, differentiation and apoptosis (Wiseman and Halliwell, 1996). The fact that the changes in the intracellular GSH/GSSG ratio are critical for activation of cell proliferation and cell death makes it a very important to consider when using any treatment that has an effect on intracellular GSH levels. As shown in Figure 3, a higher level of GSH (left side of Figure 3) is important for normal cellular functions, signal transduction and protection against certain carcinogens. However, this high level (whether induced by certain drugs or as normal response to stimulants) can slow down any effective cancer treatment that works by increasing intracellular ROS (Figure 3).

On the other hand, when intracellular GSH levels are low (using certain drugs such as BSO), the cells are more vulnerable to ROS attacks. Increased ROS might activate different intracellular oncogenic pathways or mutate a tumor suppressor gene pathway, which will activate a tumorigenesis process (Irani et al., 1997; Komatsu et al., 2008). Because the increase of ROS in cancer cells maybe part of the initiation and progression of cancer, such intrinsic oxidative stress is often viewed as an adverse event. However, as excessive levels of ROS stress can also be toxic to the cancer cells and cells are likely to be more vulnerable to damage by further ROS induced by exogenous drugs and make them more responsive to ROS producing cancer treatments (Figure 3). Therefore, changing ROS levels by GSH modulation is a way to selectively kill cancer cells without causing significant toxicity to normal cells (Trachootham et al., 2009). It is important to take into consideration that under increased levels of ROS, certain cancer cells may acquire some proliferation, cancerous measures such as: immortalization, and metastasis (Behrend et al., 2003; Hu et al., 2005; Makiya 2008).

## 5. Recommendations

It is clear that different cancer cells respond differently to certain cancer therapies. This difference could be due to inherent features of these cells or could be due to the nature of the action of drugs. In general, future drugs should be able to increase ROS production and used in combination with other drugs that interfere with ROS scavenging at the same time. It is important to find out whether these cells have a drug resisting mechanisms. These mechanisms can reverse the drug effect and implicate the need of higher doses.

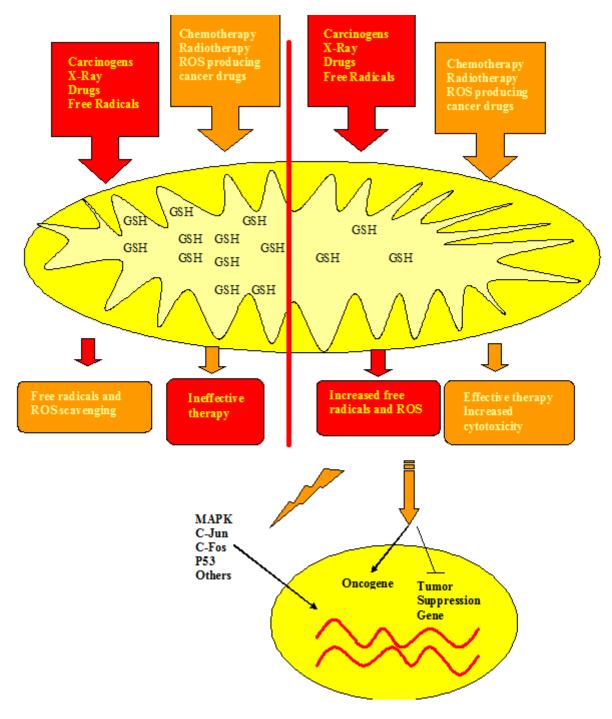


Figure 3. Glutathione level can affect expected outcome. High level of GSH (left side) is needed for cellular functions, signal transduction and protection against certain carcinogens. However, it can slow down any effective cancer treatment that works by increasing intracellular ROS. Low GSH levels (right side) renders cells more vulnerable to ROS attacks. Increased ROS activates intracellular oncogenic pathways or mutate a tumor suppressor gene pathway, which will activate a tumorigenesis process.

#### References

Ames BN. 1989. Endogenous oxidative DNA damage, aging, and cancer. Free Radical Res., **7(3)**: 121-128.

Ames BN, Shigenaga MK and Hagen TM. 1993. Oxidants, antioxidants, and the degenerative diseases of aging. Proc. National Academy of Sci. of the United States of Amer., **90(17)**: 7915-7922.

Arrigo A.-P. 1999. Gene expression and the thiol redox state. Free Radical Biol. Med., **27(9-10)**: 936-944.

Behrend L, Henderson G and Zwacka RM. 2003. Reactive oxygen species in oncogenic transformation. Biochem. Soc. Trans., **31(6)**: 1441-1444.

Blair S L, Heerdt P, Sachar S, Abolhoda A, Hochwald S, Cheng H and Burt M. 1997. Glutathione metabolism in patients with non-small cell lung cancers. Cancer Res., **57**(1): 152-155.

Carney D N, Mitchell J B and Kinsella TJ. 1983. *In Vitro* radiation and chemotherapy sensitivity of established cell lines of human small cell lung cancer and its large cell morphological variants. Cancer Res., **43(6)**: 2806-2811.

Clark EP, Epp ER, Biaglow JE, Morse-Gaudio M. and Zachgo E. 1984. Glutathione depletion, radiosensitization, and misonidazole potentiation in hypoxic chinese hamster ovary cells by buthionine sulfoximine. Radiation Res., **98(2)**: 370-380.

Cullen K J, Newkirk K A, Biaglow JE, Morse-Gaudio M and Zachgo E. 2003. Glutathione S-transferase {pi} amplification is associated with cisplatin resistance in head and neck squamous cell carcinoma cell lines and primary tumors. Cancer Res., **63(23)**: 8097-8102.

Demple B and Harrison L. 1994. Repair of oxidative damage to dna: enzymology and biology. Annual Review of Biochem., **63(1)**: 915.

Devi GS, Prasad MH, Saraswathi I, Raghu D, Rao DN and Reddy PP. 2000. Free radicals antioxidant enzymes and lipid peroxidation in different types of leukemias. Clin. Chim. Acta **293**: 53–62.

Droge W. 2002. Free radicals in the physiological control of cell function. Physiological Review **82**:47-95.

Ganesaratnam K, Balendiran, Dabur R., Fraser D. 2004. The role of glutathione in cancer. Cell Biochem. Function **22(6)**: 343-352.

Goodwin G and Baylin SB. 1982. Relationships between Neuroendocrine Differentiation and Sensitivity to {gamma}radiation in culture line oh-1 of human small cell lung carcinoma. Cancer Res., **42(4)**: 1361-1367.

Griffith O W, Anderson ME and Meister A. 1979. Inhibition of glutathione biosynthesis by prothionine sulfoximine (S-n-propyl homocysteine sulfoximine), a selective inhibitor of gamma-glutamylcysteine synthetase. J. Biol. Chem., **254(4)**: 1205-1210.

Guichard M, Lespinasse F and Malaise EP. 1986. Influence of buthionine sulfoximine and misonidazole on glutathione level and radiosensitivity of human tumor xenografts. Radiation Res., **105(1)**: 115-125.

Hassan HM and Fridovich I. 1980. Mechanism of the antibiotic action pyocyanine. J. Bacteriol., **141(1)**: 156-163.

Hayes JD., Flanagan JU and Jowsey IR. 2005. Glutathione transferases. Annual Review of Pharmacol Toxicol., **45(1)**: 51.

Hu Y, Rosen DG, Zhou Y, Feng L, Yang G, Liu J and Huang P. 2005. Mitochondrial manganese-superoxide dismutase expression in ovarian cancer: role in cell proliferation and response to oxidative stress. J. Biol. Chem. **280(47)**: 39485-39492.

Irani K, Xia Y, Zweier JL, Sollott SJ, Der CJ, Fearon ER, Sundaresan M, Finkel T and Goldschmidt-Clermont PJ. 1997. Mitogenic signaling mediated by oxidants in ras-transformed fibroblasts. Science **275(5306)**: 1649-1652.

Kaplowitz N, Aw TY and Ookhtens M. 1985. The regulation of hepatic glutathione. Annual Review of Pharmacol. Toxicol., **25(1)**: 715-744.

Kawanishi S, Hiraku Y, Pinlaor S and Ma N. 2006. Oxidative and nitrative DNA damage in animals and patients with inflammatory diseases in relation to inflammation-related carcinogenesis. Biological Chem., **387(4)**: 365-372.

Higashi K, Hiai H, Higashi T and Muramatsu M.2004. Regulatory mechanism of glutathione S-transferase P-form during chemical hepatocarcinogenesis: old wine in a new bottle. Cancer letters **209(2)**: 155-163.

Komatsu D, Kato M, Nakayama J, Miyagawa S and Kamata T. 2008. NADPH oxidase 1 plays a critical mediating role in oncogenic Ras-induced vascular endothelial growth factor expression. Oncogene **27(34)**: 4724-4732.

Makiya N. 2008. Reactive oxygen species in tumor metastasis. Cancer letters **266(1)**: 53-59.

Mattson D M, Ahmad IM, Dayal D, Parsons AD, Aykin-Burns N, Li L, Orcutt KP, Spitz DR, Dornfeld KJ and Simons AL. 2009. Cisplatin combined with zidovudine enhances cytotoxicity and oxidative stress in human head and neck cancer cells via a thiol-dependent mechanism. Free Radical Biol. and Med., **46(2)**: 232-237.

Meister A and Anderson ME. 1983. Glutathione. Annual Review of Biochem., **52(1)**: 711.

Okamoto K, Toyokuni S, Uchida K, Ogawa O, Takenewa J, Kakehi Y, Kinoshita H, Hattori-Nakakuki, Y, Hiai H and Yoshida O. 1994. Formation of 8-hydroxy-2-deoxyguanosine and 4-hydroxy-2-nonenalmodified proteins in human renal-cell carcinoma. Inter. J. Cancer **58**: 825–829.

Olinski R, Zastawny T, Budzbon J, Skokowski J, Zegarski W and Dizaroglu M. 1992. DNA base modifications in chromatin of human cancerous tissues. FEBS Lett., **309:** 193–198.

Oltra AM, Carbonell F, Tormos C, Iradi A and Sáez G. 2001. Antioxidant enzyme activities and the production of MDA and 8oxo-dG in chronic lymphocytic lerkenia. Free Radical Biol. Med., **30(11)**: 1286-1292.

Petta TB, Nakajima S, Zlatanou A, Despras E, Couve-Privat S, Ishchenko A, Sarasin A, Yasui A and Kannouche P. 2008. Human DNA polymerase iota protects cells against oxidative stress. EMBO J., **27(21)**: 2883-2895.

Shiga H, Heath EI, Rasmussen AA, Trock B, Johnston PG, Forastiere AA, Langmacher M, Baylor A, Lee M and Cullen KJ. 1999. Prognostic Value of p53, Glutathione S-transferase {{pi}}, and thymidylate synthase for neoadjuvant cisplatin-based chemotherapy in head and neck cancer. Clin Cancer Res., **5(12)**: 4097-4104.

Simons AL, Ahmad IM, Mattson DM, Dornfeld KJ and Spitz DR. 2007. 2-deoxy-D-glucose combined with cisplatin enhances cytotoxicity via metabolic oxidative stress in human head and neck cancer cells. Cancer Res., **67**(7): 3364-3370.

Townsend DM and Tew KD. 2003. The role of glutathione-Stransferase in anti-cancer drug resistance. Oncogene **22(47)**: 7369-7375.

Toyokuni S, Okamoto K, Yodoi J and Hiai H. 1995. Persistent oxidative stress in cancer. FEBS Letters **358(1)**: 1-3.

Trachootham D, Alexandre J and Huang P. 2009. Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach? Nat Rev Drug Discov., **8**(7): 579-591.

Bockmühl U, Schlüns K, Schmidt S, Matthias S and Petersen I. 2002. Chromosomal alterations during metastasis formation of head and neck squamous cell carcinoma. Genes, Chromosomes and Cancer **33(1)**: 29-35.

Vojislav V, Trudey N and Hedley DW. 2001. Differential effects of buthionine sulphoximine in hypoxic and non-hypoxic regions of human cervical carcinoma xenografts. Radiotherapy and oncology . J. of the European Soc. Therapeutic Radiology and Oncology **60(1)**: 69-73.

Weydert CJ, Zhang Y, Sun W, Waugh TA, Teoh ML, Andringa KK, Aykin-Burns N, Spitz DR, Smith BJ and Oberley LW. 2008. Increased oxidative stress created by adenoviral MnSOD or CuZnSOD plus BCNU (1,3-bis(2-chloroethyl)-1-nitrosourea) inhibits breast cancer cell growth. Free Radical Biol. Med., **44**(5): 856-867. Williamson JM, Boettcher B and Meister A. 1982. Intracellular cysteine delivery system that protects against toxicity by promoting glutathione synthesis. Proceedings of the National Academy of Sciences of the United States of America. **79(20)**: 6246-6249.

Wiseman H and Halliwell B. 1996. Damage to DNA by reactive oxygen and nitrogen species: role in inflammatory disease and progression to cancer. Biochem. J., **313(1)**: 17-29.

Wu LL, Chiou CC, Chang PY and Wu JT. 2004. Urinary 8-OHdG: a marker of oxidative stress to DNA and a risk factor for cancer, atherosclerosis and diabetics. Clin. Chim. Acta **339**: 1–9.

Yu NY and Brown JM. 1984. Depletion of glutathione *in vivo* as a method of improving the therapeutic ratio of misonidazole and SR 2508. Inter. J. Radiat. Oncol. Biol. Phys., **10**: 1265-1269.